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Supporting Information

A Feasible Way to Produce Carbon Nanofiber by Electrospinning from Sugarcane Bagasse

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Figure S1. FT-IR spectra of SCB and different SCB esters.

32 In this study, the effect of chemical modification on the structure of SCB was demonstrated by 33 FT-IR in the region of 4000–400 cm⁻¹. As shown in Figure S1, the intensity in broad region peak for 34 -OH stretching vibrations at 3450 cm⁻¹ of SCB-A showed an obvious decrease comparing to that of 35 unmodified SCB. It indicated that the hydroxyl groups were involved in the homogeneous 36 esterification [1]. In addition, the intensities of C-H bending vibrations (such as 1460, 1421 and 1377 37 cm⁻¹) were also enhanced after the introduction of aliphatic acyl groups. Meantime, a significant 38 increase of peak intensity in the broad region at 1748 cm⁻¹ for C=O stretching vibrations was observed. 39 In general, these results indicated that the esterification of SCB with three acid anhydrides occurred 40 successfully in the homogeneous system, which was consistent with the previous literatures [2-4].





Figure S2. (a) ¹H NMR and (b) ¹³C NMR spectra of different SCB esters.

43 To further confirm the esterification of SCB with acid anhydrides, SCB and SCB-A were 44 characterized with ¹H NMR and ¹³C NMR spectroscopy, and the spectra are shown in Figure S2. In the 45 ¹H NMR spectra (Figure S2a), the signals at 2.5 and 3.3 ppm are assigned to DMSO-d₆ and water, respectively, and the signals in the range of 0.5 to 2.4 ppm are attributed to the acyl moieties [2,4]. In 46 47 the ¹H NMR spectrum of SCB-AA, there was a wide signal between 1.6 and 2.2 ppm that was assigned 48 to the hydrogen proton in methyl group (α -CH₃) from the attached acetyl group. The proton signals at 49 2.0-2.4 ppm and 0.5-1.2 ppm attributed to β -CH₂ and α -CH₃ of SCB-PA. In the ¹H NMR spectrum of 50 SCB-BA, three distinct signals emerging in the chemical shift between 0.5 and 2.4 ppm corresponds to 51 γ -CH₂ (δ H=2.0-2.4 ppm), β -CH₂ (δ H=1.2-1.7 ppm), and α -CH₃ (δ H=0.5-1.0 ppm) groups [4]. The signals 52 range from 173.0 to 169.0 ppm corresponds to the carbonyl of C=O were emerged in ¹³C NMR 53 spectrum of SCB-A (Figure S2b), which provided the direct evidence of the successful attachment of 54 the corresponding aliphatic side chains onto SCB-A. The signal at 21.1 ppm is assigned to α -C of acetyl 55 group in SCB-AA, the signals at 27.2 and 9.5 ppm are assigned to β -C and α -C of propionyl group in 56 SCB-PA and the signals at 35.6, 18.2 and 13.9 ppm are assigned to the carbons of γ -C, β -C and α -C of 57 butyryl group in SCB-BA, respectively. Those results confirmed the successful esterification of SCB, 58 which was consistent with the results of FT-IR analysis above and other similar literatures [1,2,5]. 59



- - Figure S3. SEM images of electrospun PAN nanofiber mats in different stages (scale bar=1 μ m).

Sample	T_{in} (°C) ¹	T ₅₀ (°C) ²	T _{max} (°C) ³	Residue (%) ⁴
SCB	275.8±1.7	332.2±3.1	345.5±6.7	16.1±7.0
SCB-AA	272.2±6.2	345.1±5.5	337.8±1.5	24.8±2.1
SCB-PA	281.9±2.6	355.5±2.1	356.1±2.9	17.6±4.8
SCB-BA	292.2±2.5	359.3±2.7	360.4±1.1	17.1±1.2
PAN-SNF	301.3±4.9	411.0±6.4	311.3±3.5	36.1±1.5
AA-SNF-50%	303.3±1.3	363.0±9.9	328.1±4.0	36.1±1.6
PA-SNF-50%	303.4±5.4	366.9±6.4	323.9±4.8	32.6±1.2
BA-SNF-50%	308.8±0.4	372.3±4.3	331.3±2.8	30.8±1.2

65 Table S1. The TGA/DTG data and residue content of different samples

⁶⁶ ¹ T_{in}: Initial decomposition temperature; ²T₅₀: 50% weight loss temperature; ³T_{max}: temperature of the

67 DTG peak; ⁴Residue: the residue at 700 °C.

Sample	<i>d</i> (002) (Å)	Crystallite size, L_c (nm)	Lateral size, <i>L</i> ^a (nm)
PAN-CNF	3.64±0.02	1.02±0.07	2.98±0.21
AA-CNF-50%	3.64±0.04	0.94±0.08	2.65±0.01
PA-CNF-50%	3.63±0.08	0.96±0.02	2.78±0.67
BA-CNF-50%	3.61±0.05	1.02±0.04	3.51±0.28

Table S2. The characterization of graphite structure derived from XRD of different CNFs

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