



Article Drug Repurposing of the Antiviral Drug Acyclovir: New Pharmaceutical Salts

Paulo Nunes, Pedro Henrique de Oliveira Santiago, Cecilia Carolina Pinheiro da Silva and Javier Ellena *🗅

Instituto de Física de São Carlos, Universidade de São Paulo, CP 369, São Carlos 13560-970, SP, Brazil; paulo.ns@usp.br (P.N.); pedrophs.santiago@gmail.com (P.H.d.O.S.) * Correspondence: javiere@ifsc.usp.br; Tel.: +55-1633738096

Abstract: Drug repurposing is becoming interesting in terms of offering advantages over the traditional drug development, once drug discovery is a costly, time-consuming, and highly risky process. In particular, with the coronavirus disease (COVID-19) declared by World Health Organization as a global pandemic, there has emerged a considerable need to develop therapeutic agents capable of preventing viral outbreaks. Concomitantly, well-known and long-used drugs such as acyclovir (Acv) have been tested against COVID-19. Acv is a guanosine analogue that acts as an antiviral drug, commonly used to treat herpes simplex virus (HSV), genital herpes, and varicella zoster virus (VZV). Acv showed to inhibit viral proteases, multiple viral genes expression, and RNA-Dependent RNA Polymerase, helping to recover COVID-19 patients. However, ACV is a BCS class III/IV drug, with low permeability and/or slight water solubility (concentration-dependent). Given the repurposing eligibility of Acv, in this work, two new salts of this drug are presented (nitrate and sulfate), with the aim of improving its pharmacokinetic properties. The new salts were evaluated by X-ray diffraction, and thermal and spectroscopic analyses. A third salt, a chloride one, was also characterized and used for comparison.

Keywords: pharmaceutical salts; COVID-19; drug repurposing



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1. Introduction

Active Pharmaceutical Ingredients (APIs) are usually available in a crystalline or amorphous state, spread through several dosage forms such as capsules, powders, suspensions, or gels, and are largely orally administered as pills or tablets, which represent the most convenient, compact, and economical way to deliver them to the population [1,2]. In particular, crystalline APIs are more attractive than non-crystalline APIs (amorphous), due to their thermodynamic stability, purity, and manufacturability. However, in the crystalline state, APIs may exist in several different solid forms, such as salts, co-crystals, hydrates, and solvates. Beyond exhibiting polymorphism, each of them presenting specific properties (melting point, solubility, hygroscopicity, etc.) that can affect the quality and safety of a medicine [3].

More than 40% of commercially available APIs exhibit bioavailability issues. The development of new crystalline solid forms of APIs could be an effective strategy to improve their pharmacokinetic and pharmacodynamic properties [4,5]. Scientists worldwide have noticed that modifications in the properties of APIs promote, in some cases, secondary health effects, thus opening doors to a new area inside the pharmacy field: Drug Repurposing (DR). In this sense, DR has become a good strategy in terms of offering advantages over traditional drug development, as it is a costly, time-consuming, and risky process [6]. Recently, given the pandemic caused by the new coronavirus, SARS-CoV-2, termed by the World Health Organization as coronavirus disease 2019 (COVID-19), a huge need for the development of therapeutic agents capable of preventing virus outbreak emerged. In this scenario, DR gained force and several APIs, especially antivirals, were investigated in an attempt to be used in the treatment of COVID-19 [6,7].

For example, remdesivir, initially designed for treating Hepatitis C and Ebola virus, has shown activity against SARS-CoV-2 and was approved for use in the COVID-19 treatment. Research on COVID-19 has shown that SARS-CoV-2 replication involves protein RNA-dependent RNA polymerase (RdRp) [8]. In this way, APIs that act by interfering with viral replication were selected for testing against COVID-19 with a greater therapeutic benefit for mild and usual patients, as well as in severe patients [9,10]. Deserving notice among them is the antiviral drug acyclovir (Acv), a cheap, common and easily accessible antiviral used worldwide. Acv, (2-amino-1,9-dihydro-9-[(2-hydroxyethoxy)methyl]-6H-purin-6-one, or 9-[(2-hydroxyethoxy)methyl]-guanine) (Figure 1), is a well-known antiviral drug, guanosine analogue, discovered in mid for the treatment of herpes simplex virus (HSV), genital herpes, and varicella-zoster virus (VZV).



Figure 1. Molecular structure of acyclovir (Acv) showing the torsion angles δ , χ and τ .

Acv has as the major mechanism of action and its conversion to acyclovir monophosphate by virally encoded thymidine kinase and its subsequent conversion to acyclovir triphosphate by cellular enzymes, thus inhibiting viral DNA polymerase by acting as an analogue to deoxyguanosine triphosphate (dGTP) [11]. The tests involving Acv showed evidence for promoting some effect against COVID-19 infection [12–15]. However, despite being used for many years, this drug has few crystalline solid forms reported [15–18]. Acv is commercialized as acyclovir sesquihydrate, a solid form known since 1980s that shows to be more stable than the others polymorphs, as well as the anhydrous forms [19], which exhibit solubility/permeability issues [20]. These facts place this drug as a BCS class III or IV API, depending on the mg/tablet (drug concentration) [21,22].

Since pharmaceutical cocrystals [21] and salts [22,23] are interesting choices to improve the physico-chemical properties, and the last ones can be synthetized by acid-base reactions [24–26], in this work we report two new Acv salts (nitrate and sulfate), aiming to improve its pharmacokinetics properties. These new salts were evaluated by single crystal X-ray diffraction (SCXRD), differential scanning calorimetry (DSC), Fourier transform infrared (FTIR), Raman spectroscopy, Thermogravimetric analysis (TGA) and Hirshfeld surface analysis (HS) and Full Interaction Maps (FIM). In addition, acyclovir hydrochloride [22], whose X-ray powder diffraction study has already been reported in the literature, was studied by SCXRD for the first time and included for comparison reasons [4,24,25].

2. Materials and Methods

2.1. Supramolecular Synthesis

Acv salts were obtained by suspending 22.5 mg (0.1 mmol) of the API in 1 mL of alcohol for 5 min, after that, 0.2 mL of a 1.0 M solution of the respective inorganic acid (1:1 molar ratio) was added to the suspension, and the reaction was stirred for another 15 min. The solutions were cooled to room temperature and left to slowly evaporate. The yield of the salts synthesis is about 78%, 82% and 74% for the HAcv·HSO₄, HAcv·Cl and HAcv·NO₃, respectively.

2.2. Fourier Transform Infrared

FT–IR spectra were collected using a Bomem Michelson FT MB-102 spectrometer in the 4000-400 cm⁻¹ range. To record the IR spectra, 1 mg of each compound in 100 mg of KBr was used to prepare the pellets.

2.3. Raman Spectroscopy

Raman spectra were collected using a Horiba LabRAM HR Evolution UV-VIS-NIR spectrometer (Palaiseau, Saclay Campus, France) and measured in the 200–4000 cm⁻¹ range with a 532 nm diode laser and a Horiba Synphony CCD detector (Palaiseau, Saclay Campus, France).

2.4. Single Crystal X-ray Diffraction (SCXRD)

SCXRD measurements were carried out on a Rigaku XtaLAB Synergy-S Dualflex diffractometer equipped with a HyPix-6000HE hybrid photon-counting detector using Cu-K α radiation (λ = 1.54184 Å). The crystals were kept at 100 K during data collection. Cell refinement, data collection, data reduction, and absorption correction were performed using CrysAlisPro [27]. he intrinsic phasing method was employed to solve the structures using the SHELXT-2018/2 solution program [28], while the refinement was performed using least-squares methods in the SHELXL-2019/2 program [29] both within Olex2 software [28]; non-hydrogen atoms were located in subsequent Fourier-difference map analyses and refined anisotropically. The hydrogen atoms positions were located in idealized positions and refined with isotropic displacement parameters using the riding model [Uiso(h) = 1.2 Ueq(N, C) or 1.5 Ueq(O)] [29]. The Olex2 software [30] was also used to generate graphical illustrations. The data collection and refinement parameters of the salts are listed in Table 1.

Table 1. Crystallographic parameters of Acv Salts.

	HAcv·HSO ₄	HAcv·NO ₃	HAcv·Cl
Chemical formula	C ₈ H ₁₃ N ₅ O ₇ S	$C_8H_{12}N_6O_6$	C ₈ H ₁₂ N ₅ O ₃ Cl
Molar mass (g mol ^{-1})	323.29	288.24	261.68
Crystal system	Triclinic	Monoclinic	Triclinic
Space group	P-1	$P2_1/c$	P-1
a (Å)	7.7475 (1)	11.9342 (10)	6.6360 (5)
b (Å)	8.6389 (1)	14.8161 (10)	8.6147 (6)
c (Å)	10.4881 (1)	6.7449 (8)	10.2881 (8)
α (ο)	74.2290 (10)	90	83.137 (6)
β (0)	85.7930 (10)	90.4810 (90)	71.696 (7)
γ (o)	69.1930 (10)	90	80.607 (6)
V (Å3)	631.28 (1)	1192.58 (19)	549.43 (7)
Z	2	4	2
Density/g cm ^{-3}	1.701	1.605	1.582
θmin, θmax/°	8.764, 140.084	9.516, 148.988	9.078, 140.024
Index ranges	$-9 \le h \le 9$	$-14 \leq h \leq 14$	$-8 \le h \le 8$
-	$-10 \leq k \leq 10$	$-18 \leq k \leq 18$	$-9 \le k \le 10$

	HAcv·HSO ₄	HAcv·NO ₃	HAcv·Cl	
	$-12 \le l \le 12$	$-8 \le l \le 8$	$-12 \le l \le 12$	
F (000)	336.0	600.0	272.0	
μ/mm^{-1}	2.753	1.204	3.183	
Absorption correction	Gaussian	Multi-scan	Gaussian	
Max./min. transmission	1.000/0.433	1.000/0.474	1.000/0.370	
Measured reflections	26,012	3585	5287	
Independent reflections/R _{int}	2400/0.029	3585/	2022/0.039	
Refined parameters	192	183	155	
Final R indexes $[I \ge 2\sigma(I)]$	$R_1 = 0.0258 / wR_2 = 0.0672$	$R_1 = 0.0622 / wR_2 = 0.1606$	$R_1 = 0.0334 / wR_2 = 0.0846$	
Final R indexes [all data]	$R_1 = 0.0260 / wR_2 = 0.0674$	$R_1 = 0.0901 / wR_2 = 0.1811$	$R_1 = 0.0361 / wR_2 = 0.0865$	
GooF	1.071	1.050	1.062	
Largest diff. peak and hole (eÅ ⁻³)	0.20/-0.44	0.31/-0.33	0.35/-0.29	
CCDC number	2247004	2247005	2247006	

Table 1. Cont.

The CIF file of structures of HAcv·HSO₄, HAcv·NO₃ and HAcv·Cl were deposited in the Cambridge Structural Data Base with CCDC number 2247004. 2247005 and 2247006, respectively. Copies of the data can be obtained, free of charge, via www.ccdc.cam.ac.uk (accessed on 4 April 2023).

2.5. Thermal Analysis

Thermogravimetric analysis was performed using a Shimadzu TGA-50 thermobalance instrument. Approximately 4.0 mg of each sample was used for the measurements in an alumina crucible and heated at 10 °C min⁻¹ under N2 atmosphere (50 mL min⁻¹). DSC data were acquired according to previous TGA data, that is, up to the degradation temperature of each compound. These experiments were performed using a Shimadzu DSC-60 calorimeter. The samples were heated at a rate of 10 °C min⁻¹ with a crimped sealed aluminum pan using nitrogen as the purge gas (50 mL min⁻¹). The data were processed using Shimadzu TA-60 thermal data analysis software.

2.6. Hirshfeld Surface

The Hirshfeld surfaces and two-dimensional fingerprint plots for the salts were obtained from the crystallographic information files (CIFs) generated by SCXRD analyses using the CrystalExplorer 17.5 program package [31]. The d_{norm} surfaces were mapped over the color scale from -0.7 (red) to 1.3 (blue), and the shape index surfaces were obtained in the range of -1.0 (red) to 1.0 (blue). The bidimensional fingerprint plots were generated with the combination of the d_i and d_e distances, in the scale of 0.4 to 3.0 Å [31,32].

2.7. Full Interaction Maps

This analysis is useful to explore the intermolecular interactions in solid state and were calculated using the Mercury program [33] based on the molecular interactions library in the Cambridge Structural Database (CSD).

2.8. H Nuclear Magnetic Resonance

The ¹H Nuclear Magnetic Resonance (NMR) spectra of the studied compounds were collected on a 9.4 T Bruker Avance III (400 MHz) spectrometer using DMSO-d₆ as solvent.

2.9. Equilbirum Solubility Studies

The aqueous solubility of the commercial Acv and its salts was investigated at room temperature using a UV-Vis 1800 Shimadzu spectrophotometer. Saturated solutions of the compounds were prepared, suspending each compound in deionized water (Milli-Q water) and left to stir for 24 h. After 24 h, the solutions were filtered using a 0.45 μ m filter,

diluted, and measured in the 200–600 nm range. The concentrations of the compounds were determined in comparison with calibration curves constructed using the absorbance values at $\lambda = 251$ nm for the standard solution prepared with concentrations ranging from 0.025 to 0.2 mg mL⁻¹.

3. Results and Discussion

3.1. Single Crystal X-ray Diffraction (SCXRD)

Acyclovir is a guanine derivative and possess amphoteric characteristic, acting as weak acid (pk_a between 2.16 and 2.27) or base (pK_a between 9.04 and 9.25), depending on the system which is present. Three different salts were synthesized, considering the basicity of the purine group, using the following inorganic acids: hydrochloric, nitric, and sulfuric. The asymmetric unit of the salts and the geometric parameters of the hydrogen bonds are shown in Figure 2 and Table 1, respectively, while the hydrogen bonds and bond lengths of Acv and its salts are available in Tables S1 and S2.



Figure 2. ORTEP type illustration of the asymmetric unit of (**a**) Acv hydrogen sulfate, (**b**) Acv nitrate and (**c**) Acv hydrochloride with crystallographic labelling. Thermal ellipsoids represented at 50% probability level.

The formation of the Acv cation occurs through protonation of the N5 atom of the imidazole ring due of the significant differences in the pKa values between Acv and the inorganic acids ($\Delta pK_a > 3$) [34,35]. Fourier map analyses and vibrational spectroscopy studies confirmed the presence of hydrogen in N5. The three salts are composed of one protonated ACV molecule and one counterion: hydrogen sulfate, nitrate, or chloride for HAcv·HSO₄, HAcv·NO₃ and HAcv·Cl, respectively.

The Acv molecule presents conformational flexibility around the (2-hydroxyethoxy)methyl group attached to N4, where the orientation of this group influences the organization of the Acv molecules on the crystal lattice. Figure 3 shows the overlay of the Acv molecular conformations observed in the structure of the salts, as well as the free Acv (CCDC code MECWIC) [36]. It can be noticed that the Acv molecule in HAcv·NO₃ has the most different conformation for the (2-hydroxyethoxy)methyl group, which is positioned in the opposite direction when compared to Acv and the other salts. This conformational

difference involves the δ angle and is a consequence of the difference in the intermolecular interactions between Acv and the anions. The conformational analyses the structures shows that the N5-C5 bond length is stable along the three salts, meanwhile the N5-C3 bond length shows different values: 1.391 (3) Å in Acv and 1.383 (2) Å, 1.385 (2) Å and 1.386 (5) Å in HAcv·HSO₄, HAcv·Cl and Hacv·NO₃, respectively. In contrast, the C3–N5–C5 P5 bond angles show small differences: 107.970° (11), 108.221° (14) and 108.669° (3) for HAcv·HSO₄, HAcv·Cl and HAcv·NO₃, respectively. However, no other substantial changes in the imidazole ring were verified due to the protonation.



Figure 3. Overlay of the Acv molecule with the HAcv⁺ cations obtained by superposing of pyrimidine rings. Hydrogen atoms were omitted for clarity.

The HAcv·HSO₄ salt crystallizes in the triclinic P-1 space group, with one HAcv⁺ cation and one hydrogen sulfate anion per asymmetric unit. The presence of only one O–H group in the HSO₄⁻ counterion is evidenced by the S1–O5 bond length, which has a single bond character and is at least 0.11 Å longer than the others S–O bonds [37]. Figure 4a,b show the crystal packing of HAcv·HSO₄ with the representation of the N–H···O, N–H···N and O–H···O intermolecular interactions.



Figure 4. Crystal packing of HAcv·HSO₄ (**a**) through the ac plane (**b**) viewed through the bc plane (**c**) Graphic set of the three motifs $R_2^2(8)$, involving the interaction of Acv⁺ molecules mediated by HSO₄.

The crystal packing of HAcv·HSO₄ is composed by HAcv⁺ and HSO₄⁻ intercalated layers along the [001] direction (Figure 4a), connected through N–H···O and O–H···O hydrogen bonds. The N–H···N and O–H···O intermolecular interactions between two HAcv⁺ cations and two hydrogen sulfate anions, respectively, are responsible for the formation of homosynthons, while the N–H···O interactions between the 2-aminopyrimidine groups and the hydrogen sulfate anions form heterosynthons that form ribbons with $R_2^2(8)$ motifs (Figure 4c). These ribbons are linked by N5–H5···O3 interactions. In this structure the hydrogen sulfate anions participate in HSO₄⁻··· π interactions with the acyclovir rings with distance 2.969 (12) Å and 3.083 (12) Å as reported previously [38].

The asymmetric unit of the HAcv·NO₃ salt is composed of one HAcv⁺ cation and one nitrate anion and crystallizes in the monoclinic P2₁/c space group with a twinned structure along the [001] direction. The crystal structure of HAcv·NO₃ is stabilized by classical O–H···O and N–H···O hydrogen bonds (Figure 5) that form a bi-dimensional supramolecular network along the [100] plane. Dimeric arrangements are formed with the association of two HAcv⁺ cations through the N1–H1···O3' hydrogen bond as well as $\pi \cdot \cdot \cdot \pi$ stacking interaction (3.624 Å) involving the pyrimidine rings (Figure 5b). The dimers are connected through the N2–H2a···O1'' as well as the bifurcated N2–H2b···O1'···H5''–N5'' interactions (C¹₂(9) graph set). In addition, the interactions between the HAcv⁺ and NO₃⁻ ions form synthons with $R^3_3(10)$ and $R^2_3(9)$ graph sets (Figure 5c).



Figure 5. Crystal packing of HAcv·NO₃. (a) 2D supramolecular arrangement viewed through the bc plane (b) $\pi \cdots \pi$ interactions forming a dimeric arrangement (c) Graphic set motifs $R_3^3(10)$, R_3^2 (9) and C_2^1 (9) involving the interaction of Acv⁺ molecules mediated by NO₃⁻.

The crystal structure of HAcv·Cl belongs to the triclinic P-1 space group with the asymmetric unit formed by one HAcv⁺ cation and one chloride ion. The HAcv⁺ ··· HAcv⁺ and HAcv⁺ ··· Cl⁻ interactions stabilize the bidimensional network along the [100] plane, as showed in Figure 6a. The formation of Cl⁻ and HAcv⁺ chain-like arrangements, which were intercalated along the [001] direction, was observed. HAcv⁺ homosynthon, with $R_2^2(16)$ motif, are obtained through the N2–H2a···O1^{''} interactions, while the N1–H1···Cl1^{'''} and N2–H2b···Cl1^{'''} interactions form a heterosynthon with $R_2^1(6)$ motif (Figure 6c). A $\pi \cdot \cdot \cdot \pi$

stacking interaction with distance 3.417 (2) Å was also observed for HAcv·Cl (Figure 6c), which connects two HAcv⁺ cations through purine groups with the formation of a pseudo-dimeric structure.



Figure 6. Crystal packing of HAcv·Cl. (a) 2D supramolecular arrangement viewed through the bc plane (b) Graphic set motif $R_2^2(16)$ and R_2^1 (6) involving the interaction of Acv⁺ molecules and mediated by Cl⁻ (c) $\pi \cdots \pi$ interactions forming a dimeric arrangement.

3.2. Hirshfeld Surfaces Analysis

Hirshfeld surfaces (HS) analysis and the 2D-fingerprint plots (FP) allow the investigation of intermolecular interactions in crystal structures. Normalizing the distances from the HS to the nearest atom inside (d_i) and outside (d_e) the surface by the van der Waals radii of the atoms gives the origin to the d_{norm} HS, which shows the contacts in the crystal structure of a compound. The d_{norm} surface shows regions in red, white, and blue to indicate the contacts with distances shorter and longer than the sum of the vdW radii of the involved atoms, respectively. The shape index (S) is another HS property that helps evaluate the $\pi \cdots \pi$ stacking interactions, which are indicated by pairs of blue and red triangles. The HS mapped on d_{norm} property are represented in Figure 7, while the FP and shape index surfaces are shown in Figures S1–S4.

The red spots observed in the d_{norm} HS of the salts indicate that $N-H\cdots O$ hydrogen bonds stabilize the three structures. The surfaces also show the presence of $O-H\cdots N$ hydrogen bonds in the HAcv·HSO₄ and HAcv·NO₃ structures, due to the presence of oxygen atoms in the counterions as well as $N-H\cdots N$ interaction in HAcv·HSO₄ and $N-H\cdots Cl$ and $O-H\cdots Cl$ interactions in HAcv·Cl. Furthermore, it was possible also to observe the presence of non-classical $C-H\cdots O$ interactions in the structure of the salts. Pairs of red and blue triangles are present on the shape index surface (Figure S4) of HAcv·NO₃ and HAcv·Cl, indicating the presence of $\pi \cdots \pi$ interactions in the crystal structure of both salts.

The 2D-fingerprint plots of the salts (Figures S1–S3) clearly show the intermolecular contacts present in the structures. The decomposition of these fingerprints allows the quantification of the contribution of each contact to the crystal packing. The FPs of the synthesized compounds, presented in Figure S7, show that the $H \cdots H$ and $O \cdots H$ contacts were the major contributors to the crystal packing of these salts. $H \cdots H$ comprise 28.9%,

27.0% and 36.7% for HAcv·HSO₄, HAcv·NO₃; and HAcv·Cl, respectively. Meanwhile, the O···H contacts represented 44.8% (HAcv·HSO₄), 49.6% (HAcv·NO₃), and 27.2% (HAcv·Cl). The H···H contribution of HAcv·Cl was higher than the O···H one due to the absence of oxygen atoms in its anion (Cl⁻). An opposite situation was observed for the other two salts. The HAcv·NO₃ and HAcv·Cl have more C···C and C···N contacts contribution than the one observed for HAcv·HSO₄. This is due to the presence of the π ··· π stacking interactions in the HAcv·NO₃ and HAcv·Cl structures, as verified in the shape index surfaces. The O···H contacts are observed in regions of smaller d_i and d_e values in the FPs for the three compounds, an indicative that these interactions are shorter and stronger than the other ones.



Figure 7. HS of (a) Acv hydrogen sulfate, (b) Acv nitrate and (c) Acv hydrochloride mapped in d_{norm}.

3.3. Full Interaction Maps Analysis

A search in the CSD database only show a few reported Acv solid form crystal structures, and none of them show a further evaluation of the interactions of the functional groups. The display of the Full Interaction Maps (FIMs) created using carbonyl oxygen and alcohol oxygen probes for the Acv, and with an additional charged (NH) nitrogen probe for the salts; Providing an overview of the preferred directionality and the strength of the potential interactions that the functional groups in the molecule can form.

The FIM of the anhydrous Acv form can be observed in Figure S5a. The blue regions (related to the donor probe) indicate a higher propensity of the O1, O2, and N5 atoms (donor region) to donate electron density to the O3, N2, and N1 atoms (acceptor region). These regions are connected by $O1 \cdots H5-O3$, $O1 \cdots H2B-N2$, $N5 \cdots H1-N1$ and $O2 \cdots H2A-N2$ interactions. Meanwhile, the red regions (related to the acceptor probe maps) involve the N3, N2, N1, O3, and N5 atoms and show a significant propensity to receive electrons, which stabilize the crystal structure through the functional groups involving the O1, O2, N3, and N1 atoms, as discussed previously. Figures S5b and S6 also show similar donor and acceptor groups for the Acv salts HAcv·HSO₄, HAcv·NO₃ and HAcv·Cl. Table 2 summarizes the hydrogen bonds in the studied compounds.

	Blue		Red		
Acv	01···H5-O3 01···H2B-N5 N5···H1-N1	O2 ···· H2A -N5	N2-H2A · · · O2 N1-H1 · · · N5 O3-H5 · · · O1		
HAcv·HSO ₄	O3-H5 · · · · N5		N2-H2A · · · N3 N5-H2B · · · O5 N1-H1 · · · O4	N5-H5 ····O3 O3-H3 ····O6	
HAcv·NO ₃	O1-H2A · · · N2 O3-H1 · · · N2		N2-H2A ····O1 N5-H2B ····O5 N1-H1 ····O3	N5-H5 · · · O5 O3-H3 · · · O4 O3-H3 · · · O5	
HAcv·Cl	O3-H5 · · · N5		N2-H2A · · · O2 N5-H2B · · · Cl	N5-H5 · · · O5 N1-H1 · · · Cl	

Table 2. Hydrogen bonds by region (Donor and Acceptors).

Atoms in bold are related to the region detached.

3.4. Thermal Analysis

The thermal behaviour of the Acv salts was studied using a combination of DSC/TGA techniques. The curves are presented in Figure 8. The phase purity of the salts was evaluated by observing the presence of a single endothermic peak associated with the melting point of the compounds, shown in the corresponding DSC curves at 134 °C, 149 °C, and 163 °C for the HAcv·HSO₄, HAcv·Cl and HAcv·NO₃, respectively. It is possible to identify one more endothermic event in HAcv·HSO₄ and HAcv· at 49 °C and 47 °C, which could be associated to the loss of a small amount of water that could be present in the samples due to the hydroscopic tendency showed by these salts. To the enthalpy evaluated in the salts the values are presented in order: HAcv·Cl (192.2 J g⁻¹) > HAcv·HSO₄ (111.4 J g⁻¹) > HAcv·NO₃ (110.8 J g⁻¹).



Figure 8. DSC/TGA curves of (a) Acv hydrogen sulfate, (b) Acv hydrochloride and (c) Acv nitrate.

According to the TGA data, the thermal degradation of $HAcv \cdot HSO_4$, $HAcv \cdot Cl$ and $HAcv \cdot NO_3$ began at 165 °C, 187 °C and 180 °C, respectively. Thermal stability can be associated with the rupture of the crystal structure upon exposure to the heating process in an

inert environment as an experimental condition in the DSC/TGA experiments. The thermal behaviour observed with TGA for Acv salts allows to establish a thermal stability based in the degradation of the compounds in order: $HAcv \cdot Cl$, $HAcv \cdot NO_3$ and $HAcv \cdot HSO_4$.

3.5. Spectroscopic Analysis

The FT-IR and Raman studies of Acv and its salts were performed to confirm the formation of salts (Figures 9 and 10). FT-IR and Raman spectra of the Acv sample confirmed that the API used in the supramolecular synthesis corresponded to Acv sesquihydrate [19]. The salt formation was identified by evaluating the changes in the frequency of some vibrational modes related to the promoted N5 atom from the imidazolium group, as well as the formation of new hydrogen bonds between the API and anions. The band assignments are listed in Table 3.



Figure 9. FT-IR spectra of Acv (green) and its salts, HAcv·HSO₄ (black), HAcv·NO₃ (blue), and HAcv·Cl (red).

The FT-IR spectra show that ν (N-H) stretching frequencies associated with the imidazolium group are present in HAcv·HSO₄ (3417 cm⁻¹), HAcv·Cl (3523 cm⁻¹) and HAcv·NO₃ (3425 cm⁻¹), but not in Acv (Figure 9). The inorganic anions used as coformers show different FT-IR characteristics, starting with the HSO₄⁻ and NO₃⁻ anions, which show ν (SO) and ν (NO₃⁻) bands at 1054 cm⁻¹ and 1384 cm⁻¹, respectively.

The Raman spectra of Acv and its salts present clear differences that can be attributed to the protonation of the imidazolium group and the vibrational modes arising from the inorganic anions (see Figure 10). The HSO₄⁻ and NO₃⁻ anions show v(SO) and v(NO₃) stretching bands at 1020 cm⁻¹ and 1282 cm⁻¹, respectively. As expected, no bands corresponding to the Cl⁻ anion were observed in the Raman spectrum of HAcv·Cl. A new δ (N–H) band was present in the Raman spectra of the salts HAcv·HSO₄ (3152 cm⁻¹), HAcv·Cl (3312 cm⁻¹) and HAcv·NO₃ (3321 cm⁻¹), which confirms the protonation of the imidazolium ring from Acv.



Figure 10. Raman spectra of Acv (green) and its salts $Hacv \cdot HSO_4$ (blue), $Hacv \cdot NO_3$ (red), and $Hacv \cdot Cl$ (black).

	Acv		HAcv·HSO ₄		HAcv·Cl		HAcv·NO ₃	
	IR	Raman	IR	Raman	IR	Raman	IR	Raman
ν(C–N)	1185	1355	1164	1341	1182	1338	1151	1336
ν(C=N)	1485	1611	1487	1610	1483	1604	1485	1600
ν(C=O)	1693	1630	1714	1703	1693	1703	1708	1717
ν(N–H)	—	3475	3417	3152	3523	3312	3425	3321
v(NO ₃)	—	_	—	_	_	—	1384	1282
v(SO)	—	_	1054	1020	_	—	_	_

Table 3. Main bands (cm^{-1}) for IR and Raman spectra of Acv and its salts.

3.6. H NMR Spectroscopy

The ¹H NMR spectra for Acv and its salts were obtained using DMSO-d₆ as solvent and reference. The ¹H spectra for each compound and the chemical shifts (δ) are presented in the Supplementary material (Figures S8–S10 and Table S2).

The spectrum of the free Acv shows the signals of the guanine group in the range of 10.70 to 6.50 ppm, appearing as singlets with the integration of 1, 1, and 2 for the - NH-C(O)- (10.64 ppm), N=CH-N (7.82 ppm) and -NH₂ (6.51 ppm) groups, respectively. The hydrogens atoms from the methylene carbons of the (2-hydroxyethoxy)methyl group appear as signals in 5.35 and 3.47 ppm, with the first one as a singlet (integration of 2) correspondent to the -NCH₂O- group, while the last one as a multiplet integrating for 4, being a superposing of the signals correspondent to the hydrogens from the ethyl group. These results are comparable to some data reported previously for Acyclovir [39].

The spectra of the salts also showed the signals corresponding to the hydrogens from the N=CH-N, -NH₂ and -NCH₂O- groups, but in regions with higher chemical shifts, which occurred due to the protonation of the Acv molecule. The signal related to the ethyl group in the free Acv spectrum is seen as two multiplets in the salts spectra, with the hydrogens from the carbon bonded to the hydroxyl being a little more deshielded than the other carbon. The protonation of the imidazolium ring is verified with the appearance of a new singlet (integration of 1) in the 11.07 to 11.29 ppm in the spectra of the salts. Unfortunately, the signal corresponding to the amide was not observed in the spectra of the salts, probably due to the possible exchange of the proton between the Acv cation and the solvent.

3.7. Equilibrium Solubility

Solubility is an important property of an API because of your strong relation with its oral absorption and pharmaceutical work. The Acv (Commercial form) and his anhydrous form exhibit solubility/permeability issues [20] that place this drug as a BCS class III or IV API, depending on the mg/tablet (drug concentration) [21,22]. In attempt to evaluate the aqueous solubility of the acyclovir salts, the synthesized compounds were measured in an aqueous solution (the concentration of acyclovir compounds in a saturated aqueous solution in the presence of undissolved solid) of the HAcv·HSO₄, HAcv·NO₃ salts. According to the Figure S12, the HAcv·HSO₄ and HAcv·NO₃ shows to be 2.5 and 1.8 times more soluble when compared to the commercial form of Acv (5.9 mM of solubility), presenting an aqueous solubility of 7.7 mM and 15.7 mM, respectively. Meanwhile the HAcv·HCl present a solubility of 241 mM [22].

4. Conclusions

Within our effort to develop new solid forms with improved pharmacokinetic properties of repurposed drugs as a subsidiary to the combat Covid-19 pandemic, two new salts of Acv were studied: HAcv·HSO₄ and HAcv·NO₃. These new solid forms and the HAcv·Cl salt were further characterized by Single-crystal X-ray diffraction, Hirshfield surface (HS) studies, and thermal and spectroscopic analyses. The evaluation of the crystal structures revealed that the three salts showed protonation of the imidazolium nitrogen N5. The DSC/TGA results obtained for the Acv salts allowed the establishment of the following thermal stability order: HAcv·Cl > HAcv·NO₃ > HAcv·HSO₄. The evaluation of aqueous solubility allows to determine that the new salts are more soluble that the commercial form of the Acv showing solubility of 7.7 mM (HAcv·HSO₄) and 15.7 mM (HAcv·NO₃). Thus, based on the supramolecular features/interactions presented in this work, a better comprehension of the pharmacologic properties of the salts is needed as a research proposal for future studies.

Supplementary Materials: The following supporting information can be downloaded at: https:// www.mdpi.com/article/10.3390/cryst13050782/s1, Figure S1: Fingerprint images (a) C-C (b) C-N (c) H-H (d) N-H (e) O-H and (f) Total of Acv hydrogen-sulfate salt; Table S1: Hydrogen Bonds for the Acv salts; Figure S2: Fingerprint images (a) C-C (b) C-N (c) H-H (d) N-H (e) O-H and (f) Total of Acv nitrate salt; Table S2: Bond Lengths for the Acv salts; Figure S3: Fingerprint images (a) C-C (b) C-N (c) H-H (d) N-H (e) O-H (f) H-Cl and (g) Total of Acv hydrochloride salt; Table S3: ¹H NMR data for commercial acyclovir and its salts; Figure S4: Detail Hirshfeld surfaces mapped in shape index (a) Acv nitrate and (b) Acv hydrochloride salt; Figure S5: Full interaction Maps of (a) Acyclovir free solvent and (b) Acv hydrogen-sulfate salt; Figure S6: Full interaction Maps of (a) Acv hydrochloride (b) Acv nitrate salts; Figure S7: % of contacts contribution in Acv salts; Figure S8: ¹H NMR (400 MHz) spectrum of commercial HAcv·NO₃ in DMSO-d₆; Figure S10: ¹H NMR (400 MHz) spectrum of commercial HAcv·NO₃ in DMSO-d₆; Figure S11: ¹H NMR (400 MHz) spectrum of commercial HAcv·NO₃ in DMSO-d₆; Figure S11: ¹H NMR (400 MHz) spectrum of commercial HAcv·NO₃ in DMSO-d₆; Figure S11: ¹H NMR (400 MHz) spectrum of commercial HAcv·NO₃ in DMSO-d₆; Figure S11: ¹H NMR (400 MHz) spectrum of commercial HAcv·NO₃ in DMSO-d₆; Figure S11: ¹H NMR (400 MHz) spectrum of commercial HAcv·NO₃ in DMSO-d₆; Figure S11: ¹H NMR (400 MHz) spectrum of commercial HAcv·NO₃ in DMSO-d₆; Figure S11: ¹H NMR (400 MHz) spectrum of commercial HAcv·NO₃ in DMSO-d₆; Figure S11: ¹H NMR (400 MHz) spectrum of commercial HAcv·O Acv salts. Author Contributions: Conceptualization, P.N. and P.H.d.O.S.; methodology, P.H.d.O.S. and J.E.; validation, P.N., P.H.d.O.S., C.C.P.d.S. and J.E.; formal analysis, P.N., P.H.d.O.S. and J.E.; investigation, P.N.; resources, J.E.; data curation, P.H.d.O.S. and J.E.; writing—original draft preparation, P.N., P.H.d.O.S. and C.C.P.d.S.; writing—review and editing, P.N., P.H.d.O.S., C.C.P.d.S. and J.E.; supervision, J.E.; project administration, J.E.; funding acquisition, J.E. All authors have read and agreed to the published version of the manuscript.

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