

Supporting information

Heterogeneous Synergistic Catalysis for Promoting Aza-Michael-Henry Tandem Reaction for the Synthesis of Chiral 3-Nitro-1,2-Dihydroquinoline

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Figures and Descriptions:

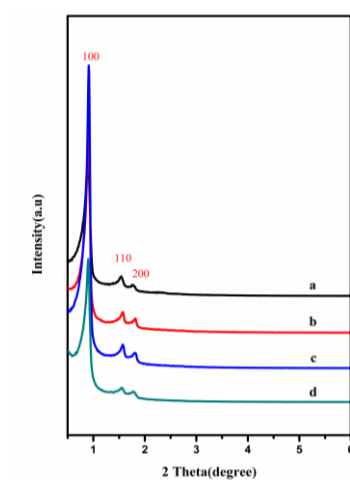


Fig. S1. XRD patterns of (a) SBA-15, (b) SBA-15-SH, (c) SBA-15-PY, and (d) SBA-15-Q.

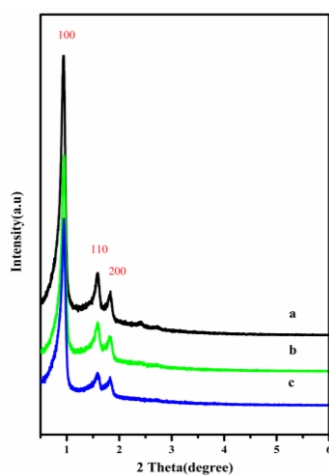


Fig. S2. XRD patterns of (a) SBA-15, (b) SBA-15-Br, and (c) SBA-15-AEP.

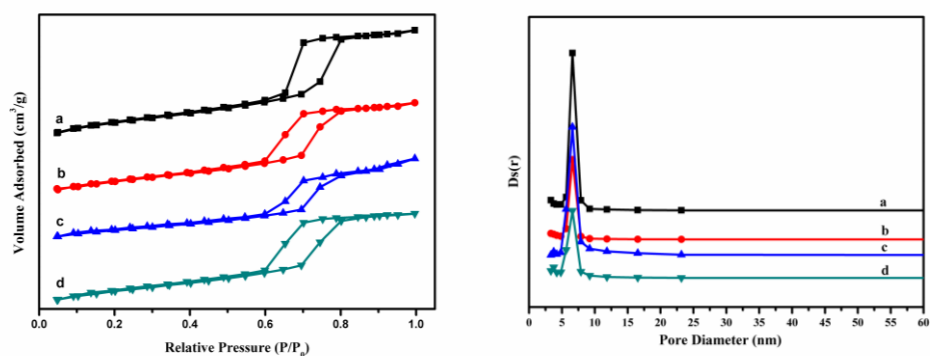


Fig. S3. Nitrogen adsorption-desorption isotherms and the pore size distribution of (a) SBA-15, (b) SBA-15-SH, (c) SBA-15-PY, and (d) SBA-15-Q.

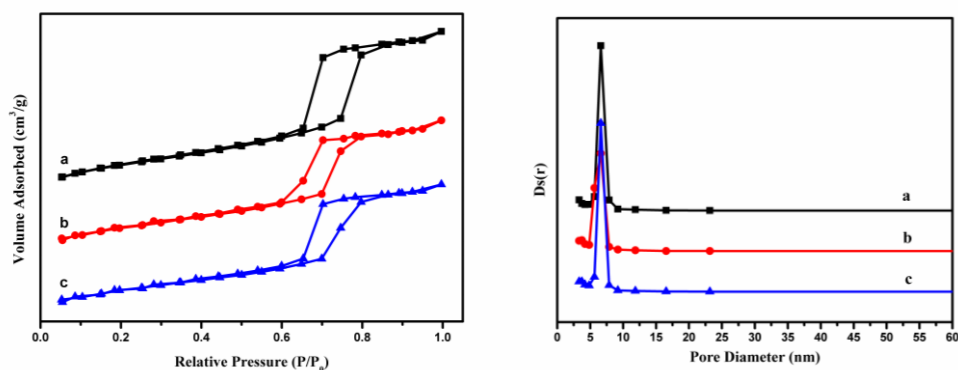


Fig. S4. Nitrogen adsorption-desorption isotherms and the pore size distribution of (a) SBA-15, (b) SBA-15-Br, and (c) SBA-15-AEP.

N₂ adsorption-desorption isotherms for all samples are typical of type IV, and H1-type hysteresis loop with delayed capillary evaporation located at a P/P_0 of about 0.7 is observed, revealing the predominance of well-defined mesopores with uniform size. The pore size shows a narrow distribution with the maximum at 6.5-6.5 nm. Table S1 summarizes the pore sizes along with the BET surface areas and pore volumes. The decreases in surface area and pore volume clearly indicate that the grafted (3-mercaptopropyl)trimethoxysilane/3-bromopropyltrichlorosilane groups and chiral amines are located inside the mesopore channels.

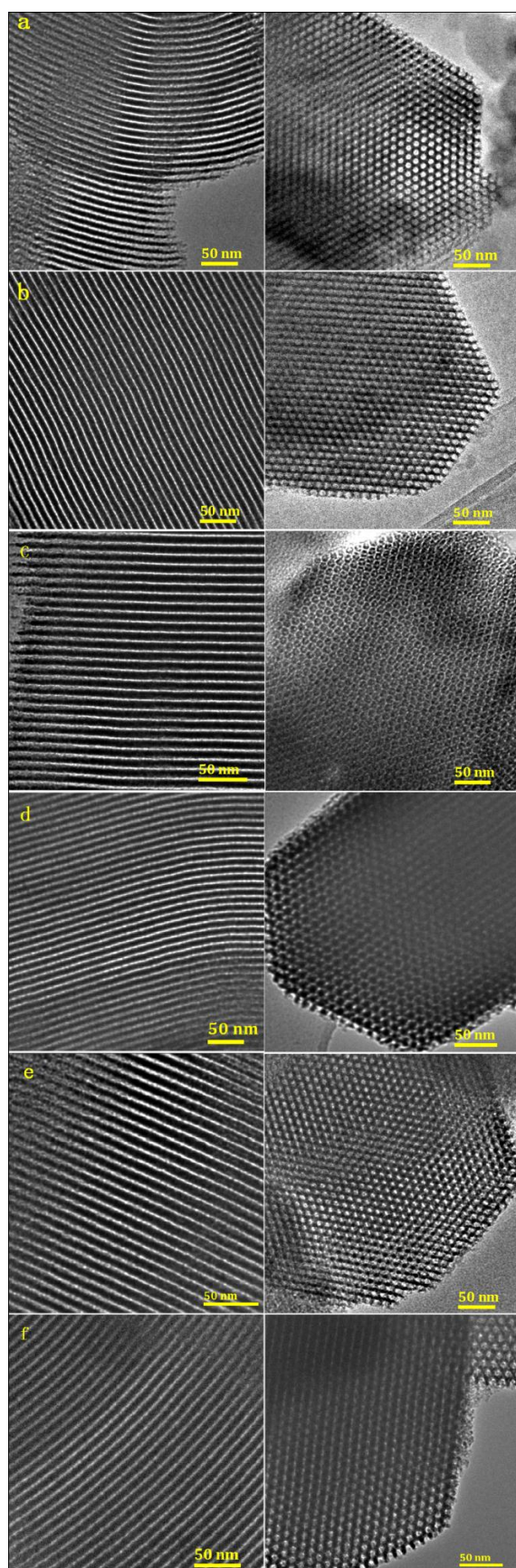


Fig. S5. TEM images of (a) SBA-15, (b) SBA-15-SH, (c) SBA-15-Br, (d) SBA-15-PY, (e) SBA-15-AEP, and (f) SBA-15-Q, perpendicular (left) and parallel (right) to the channels.

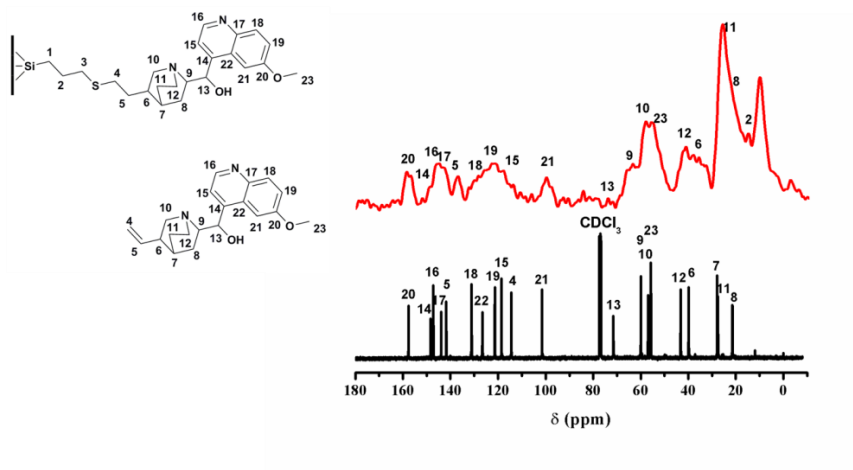


Fig. S6. ^{13}C CP/MAS NMR spectrum of SBA-15-Q and ^{13}C NMR spectrum of quinine.

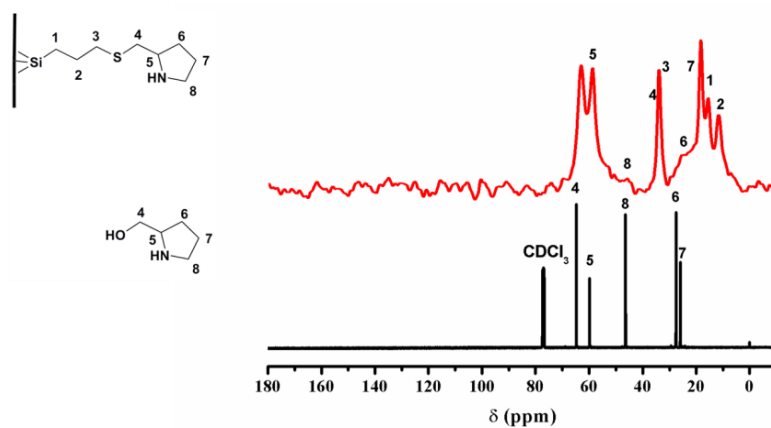


Fig. S7. ^{13}C CP/MAS NMR spectrum of SBA-15-PY and ^{13}C NMR spectrum of (S)-(+)-prolinol.

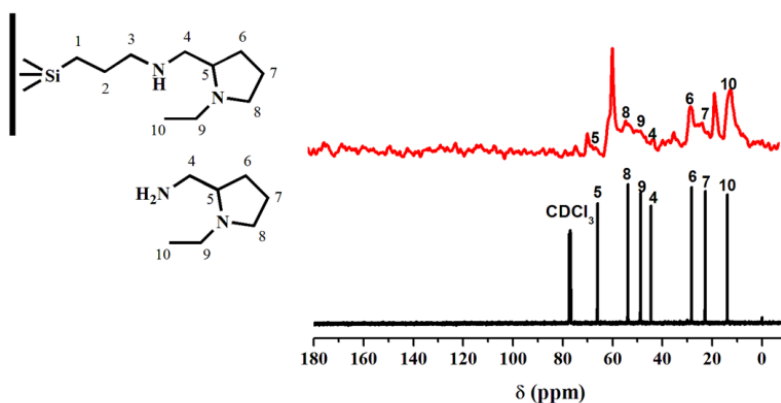


Fig. S8. ^{13}C CP/MAS NMR spectrum of SBA-15-AEP and ^{13}C NMR spectrum of (S)-(-)-2-aminomethyl-1-ethylpyrrolidine.

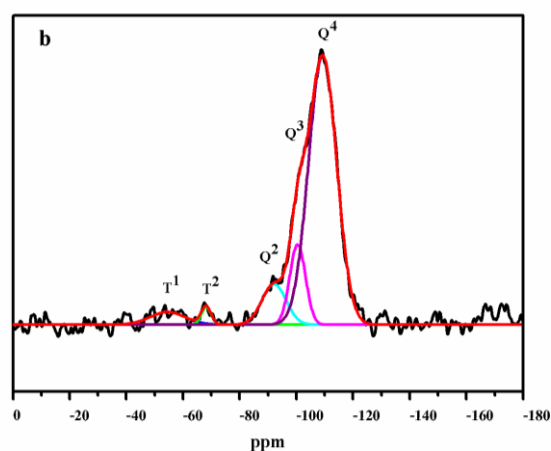
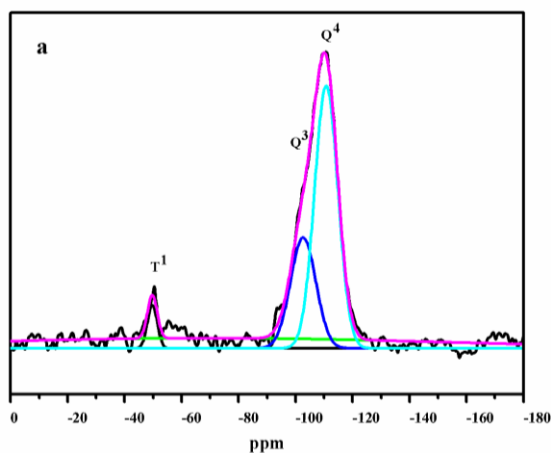


Fig. S9. ^{29}Si MAS NMR spectra of (a) SBA-15-SH and (b) SBA-15-Br.

^{29}Si BD/MAS NMR spectra were recorded for SBA-15-SH/SBA-15-Br. The resonances attributed to the silicon in Q^4 [$\text{Si}(\text{SiO})_4$], Q^3 [$\text{Si}(\text{SiO})_3\text{OH}$], and Q^2 [$\text{Si}(\text{SiO})_3(\text{OH})_2$] linkages are observed at -110, -100 and -92 ppm. The silanol density was quantified by curve fitting and deconvolution of ^{29}Si MAS NMR signals according to the reference method [Palkovits, R.; Yang, C. M.; Olejnik, S.; Schüth, F. Active sites on SBA-15 in the Beckmann rearrangement of cyclohexanone oxime to ϵ -caprolactam, *J. Catal.* 2006, 243, 93-98.] and the calculated formula is illustrated as follows:

$$\text{Silanol density (}\mu\text{mol g}^{-1}\text{)} = \sum W_{Qn} \cdot M_{Qn}$$

Wherein W and M respectively represent the peak area percentage and the molar mass of Q^i ($i = 2, 3, 4$). The results are shown in Table S3. The final silanol density based on the specific surface area of SBA-15-SH/SBA-15-Br is calculated as 4.46 and $8.87\mu\text{mol/m}^2$.

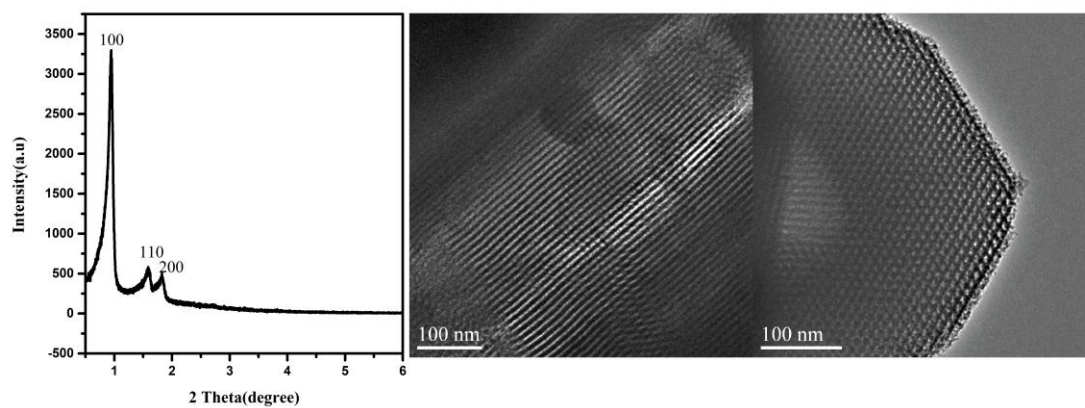


Fig. S10. XRD pattern and TEM images of reused SBA-15-AEP in five runs.

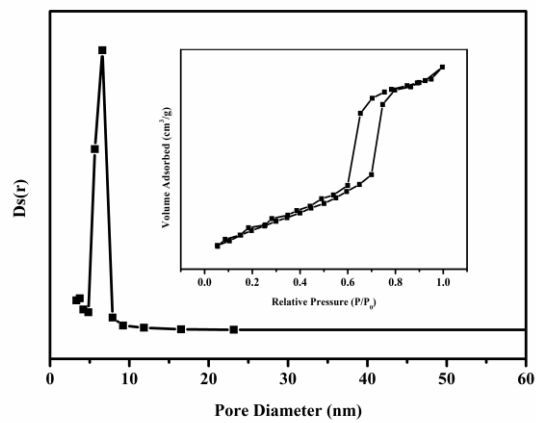


Fig. S11. Nitrogen adsorption-desorption isotherms and the pore size distribution of reused SBA-15-AEP in five runs.

Tables:

Table S1. The specific surface area, pore volume, and pore diameter of mesoporous silica.

Sample	Surface area/ m ² g ⁻¹	Pore volume/cm ³ g ⁻¹	Pore diameter/nm
SBA-15	618	1.043	6.6
SBA-15-SH	549	0.995	6.6
SBA-15-Q	386	0.608	6.5
SBA-15-PY	452	0.727	6.5
SBA-15-Br	539	0.798	6.6
SBA-15-AEP	454	0.783	6.6

Table S2. The density of chiral basic sites and the ratio of silanol to basic sites.

Catalyst	SBA-15-Q	SBA-15-PY	SBA-15-AEP
The content of C, unit mass (mmol/g)	11.69	7.49	6.03
The content of H, unit mass (mmol/g)	20.50	17.70	16.90
The content of S, unit mass (mmol/g)	0.800	0.938	--
The content of N, unit mass (mmol/g)	0.829	0.639	0.629
The content of N, unit area (μmol/m ²)	2.148	1.414	1.385
The density of basic sites (μmol/m ²) ^a	1.074	0.707	0.693
The density of silanol (μmol/m ²) ^b	4.46	4.46	8.87
The molar ratio of silanol to basic sites ^c	4:1	6:1	13:1

^a The density of basic sites = The content of N (unit area, μmol/m²)/N atom number in the basic site (2 for SBA-15-Q and SBA-15-AEP; 1 for SBA-15-PY); ^b determined from the ²⁹Si BD-MAS NMR spectra; ^c The molar ratio of silanol to basic sites = The density of silanol (μmol/m²)/The density of basic sites (μmol/m²)

Table S3. The molecular weight, molar fractions of Qⁱ sites and the density of silanols for SBA-15-SH and SBA-15-Br.

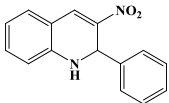
Sample	Q ⁴ (%)	Q ³ (%)	Q ² (%)	MW ^a (g/mol)	Content of OH (mmol/g)	Density of OH (μmol/m ²)
SBA-15-SH	69.35	30.65	-	62.75	2.45	4.46
SBA-15-Br	77.17	13.60	9.24	62.89	4.78	8.87

^a determined as sum of molar weights and fractions of Q⁴, Q³, and Q².

Product analysis:

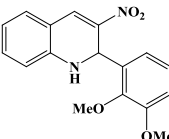
3-nitro-2-phenyl-1,2-dihydroquinoline: ^1H NMR (400 MHz, CDCl_3): δ =7.98 (s, 1H), 7.40–7.35 (m, 2H), 7.32–7.28 (m, 3H), 7.20–7.16 (m, 2H), 6.71 (t, J = 7.5 Hz, 1H), 6.46 (d, J = 8.1 Hz, 1H), 5.99 (s, 1H), 4.70 (s, 1H); ^{13}C NMR (101 MHz, CDCl_3): δ = 144.38, 142.16, 134.13, 131.29, 131.26, 130.92, 129.01, 128.85, 128.77, 126.28, 118.65, 114.93, 113.41, 55.51. HRMS (ESI): calcd for $\text{C}_{15}\text{H}_{12}\text{N}_2\text{NaO}_2$ $[\text{M}]^+$ m/z 275.26; found 275.05.

HPLC analysis of 3-nitro-2-phenyl-1,2-dihydroquinoline

Structure	Column	Eluent	Flow rate	Retention time
	CHIRALPAK OD-H	Hexane: 2-PrOH = 85: 15	1.0 mL/min	13.36 min
				16.03 min

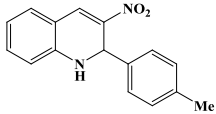
2-(2,3-dimethoxyphenyl)-3-nitro-1,2-dihydroquinoline: ^1H NMR (400 MHz, CDCl_3): δ =8.13 (s, 1H), 7.26–7.17 (m, 2H), 7.13–7.06 (m, 1H), 6.94 (m, 1H), 6.74 (dd, J = 7.7, 1.3 Hz, 1H), 6.65 (t, J = 7.4 Hz, 1H), 6.37 (d, J = 8.2 Hz, 1H), 6.34 (s, 1H), 4.99 (s, 1H), 4.00 (s, 3H), 3.87 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3): δ =152.89, 144.96, 144.64, 139.36, 134.33, 133.98, 132.96, 131.17, 124.48, 118.59, 118.44, 115.20, 113.85, 112.48, 60.99, 55.79, 49.36. HRMS (ESI): calcd for $\text{C}_{17}\text{H}_{16}\text{N}_2\text{NaO}_4$ $[\text{M}]^+$ m/z 335.31; found 335.19.

HPLC analysis of 2-(2,3-dimethoxyphenyl)-3-nitro-1,2-dihydroquinoline

Structure	Column	Eluent	Flow rate	Retention time
	CHIRALPAK OD-H	Hexane: 2-PrOH = 85: 15	1.0 mL/min	12.35min
				12.81min

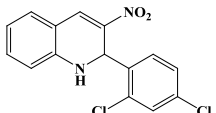
3-nitro-2-p-tolyl-1,2-dihydroquinoline: ^1H NMR (400 MHz, CDCl_3): δ = 7.96 (s, 1H), 7.31–7.05 (m, 6H), 6.69 (t, 1H), 6.44 (d, J = 8.0 Hz, 1H), 5.94 (s, 1H), 4.68 (s, 1H), 2.30 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3): δ =152.89, 144.96, 144.64, 139.36, 134.33, 133.98, 132.96, 131.17, 124.48, 118.59, 118.44, 115.20, 113.85, 112.48, 60.99, 55.79. HRMS (ESI): calcd for $\text{C}_{16}\text{H}_{14}\text{N}_2\text{NaO}_2$ $[\text{M}]^+$ m/z 289.28; found 289.20.

HPLC analysis of 3-nitro-2-p-tolyl-1,2-dihydroquinoline

Structure	Column	Eluent	Flow rate	Retention time
	CHIRALPAK OD-H	Hexane: 2-PrOH = 85: 15	1.0 mL/min	9.25min
				12.97min

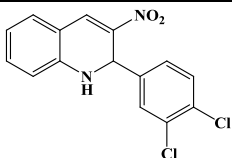
2-(2,4-chlorophenyl)-3-nitro-1,2-dihydroquinoline: ^1H NMR (400 MHz, CDCl_3): δ =8.20 (s, 1H), 7.57–7.16 (m, 5H), 6.76 (t, 1H), 6.47 (d, J = 8.0 Hz, 1H), 6.46(s, 1H), 4.98 (s, 1H); ^{13}C NMR (101 MHz, CDCl_3): δ =143.67, 138.63, 134.98, 134.38, 133.15, 131.96, 131.37, 130.10, 128.69, 127.97, 119.15, 115.06, 114.03, 51.05. HRMS (ESI): calcd for $\text{C}_{15}\text{H}_{10}\text{Cl}_2\text{N}_2\text{O}_2$ $[\text{M}]^+$ m/z 321.02; found 321.23.

HPLC analysis of 2-(2,4-chlorophenyl)-3-nitro-1,2-dihydroquinoline

Structure	Column	Eluent	Flow rate	Retention time
	CHIRALPAK OD-H	Hexane: 2-PrOH = 85: 15	1.0 mL/min	9.67min
				19.18min

2-(3,4-chlorophenyl)-3-nitro-1,2-dihydroquinoline: ^1H NMR (400 MHz, CDCl_3): δ =8.00 (s, 1H), 7.46 (dd, J = 7.2, 2.2 Hz, 1H), 7.38 (dd, J = 13.1, 7.1 Hz, 1H), 7.19–7.21 (m, 3H), 6.81–6.70 (m, 1H), 6.55–6.42 (m, 1H), 5.98 (s, 1H), 4.71 (s, 1H). ^{13}C NMR (101 MHz, CDCl_3): δ =143.81, 142.05, 140.02, 134.87, 135.73, 133.01, 132.19, 130.38, 128.35, 125.66, 119.15, 114.64, 113.51, 54.57. $\text{C}_{15}\text{H}_{10}\text{Cl}_2\text{N}_2\text{O}_2$ $[\text{M}]^+$ m/z 321.02; found 321.26

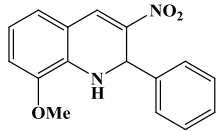
HPLC analysis of 2-(3,4-chlorophenyl)-3-nitro-1,2-dihydroquinoline

Structure	Column	Eluent	Flow rate	Retention time
	CHIRALPAK OD-H	Hexane: 2-PrOH = 85: 15	1.0 mL/min	9.79min
				17.25min

8-methoxy-3-nitro-2-phenyl-1,2-dihydroquinoline: ^1H NMR (400 MHz, CDCl_3): δ =7.99 (s, 1H), 7.44 – 7.26 (m, 5H), 6.83 (dd, J = 7.8, 1.0 Hz, 1H), 6.76 (dd, J = 7.9, 0.8 Hz, 1H), 6.63 (t, J = 7.9 Hz, 1H), 6.08 (s, 1H), 5.29 (s, 1H), 3.79 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3): δ =145.45, 142.52, 140.91, 135.00, 131.39, 128.94, 128.62, 126.27, 122.77, 117.39, 114.37, 113.63, 55.59, 55.08.

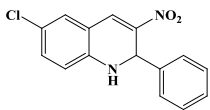
HRMS (ESI): calcd for $C_{16}H_{14}N_2NaO_3[M]^+$ m/z 305.28; found 305.27.

HPLC analysis of 8-methoxy-3-nitro-2-phenyl-1,2-dihydroquinoline

Structure	Column	Eluent	Flow rate	Retention time
	CHIRALPAK OD-H	Hexane: 2-PrOH = 85: 15	1.0 mL/min	13.41min
				16.07min

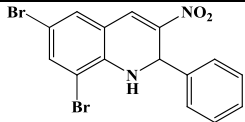
6-chloro-3-nitro-2-phenyl-1,2-dihydroquinoline: 1H NMR (400 MHz, $CDCl_3$): δ 7.89 (s, 1H), 7.41–7.28 (m, 6H), 7.12 (dd, J = 8.6, 2.4 Hz, 1H), 6.42 (d, J = 8.6 Hz, 1H), 5.98 (s, 1H), 4.72 (s, 1H); NMR (101 MHz, $CDCl_3$): δ 142.74, 142.08, 141.71, 133.65, 130.05, 129.76, 129.11, 128.99, 126.25, 116.05, 114.66, 55.48. HRMS (ESI): calcd for $C_{15}H_{11}ClN_2NaO_2 [M]^+$ m/z 309.70; found 309.50.

HPLC analysis of 6-bromo-3-nitro-2-phenyl-1,2-dihydroquinoline

Structure	Column	Eluent	Flow rate	Retention time
	CHIRALPAK OD-H	Hexane: 2-PrOH = 85: 15	1.0 mL/min	10.83min
				17.53min

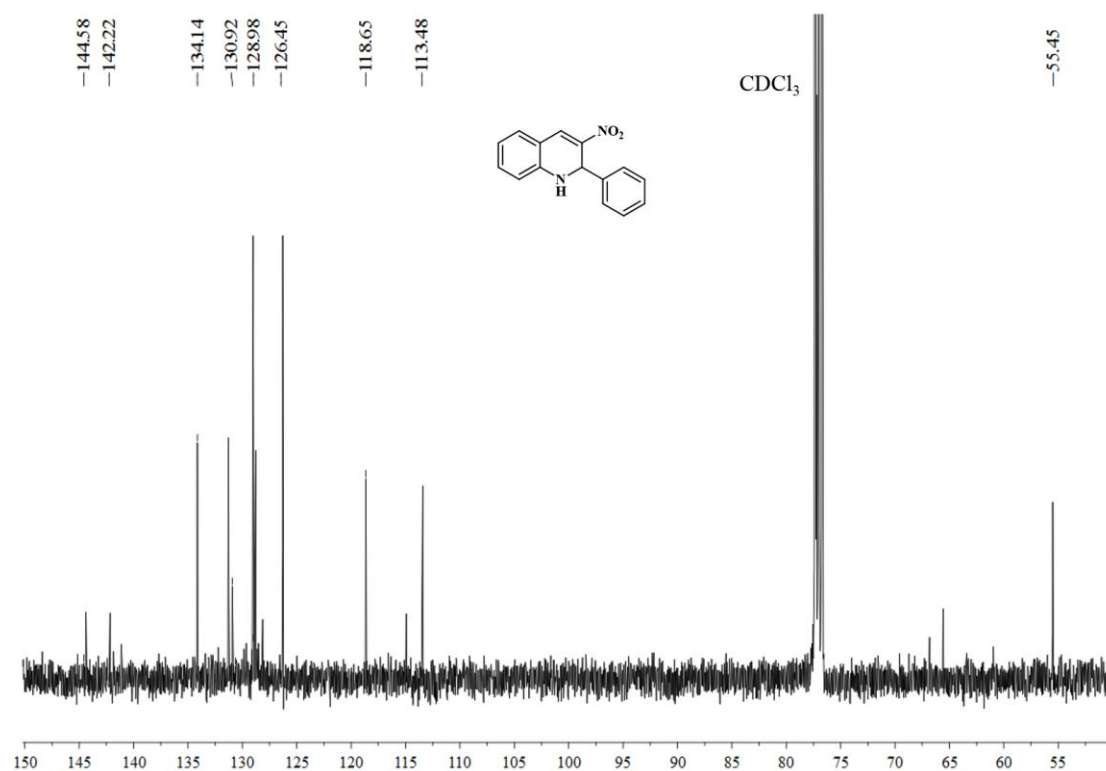
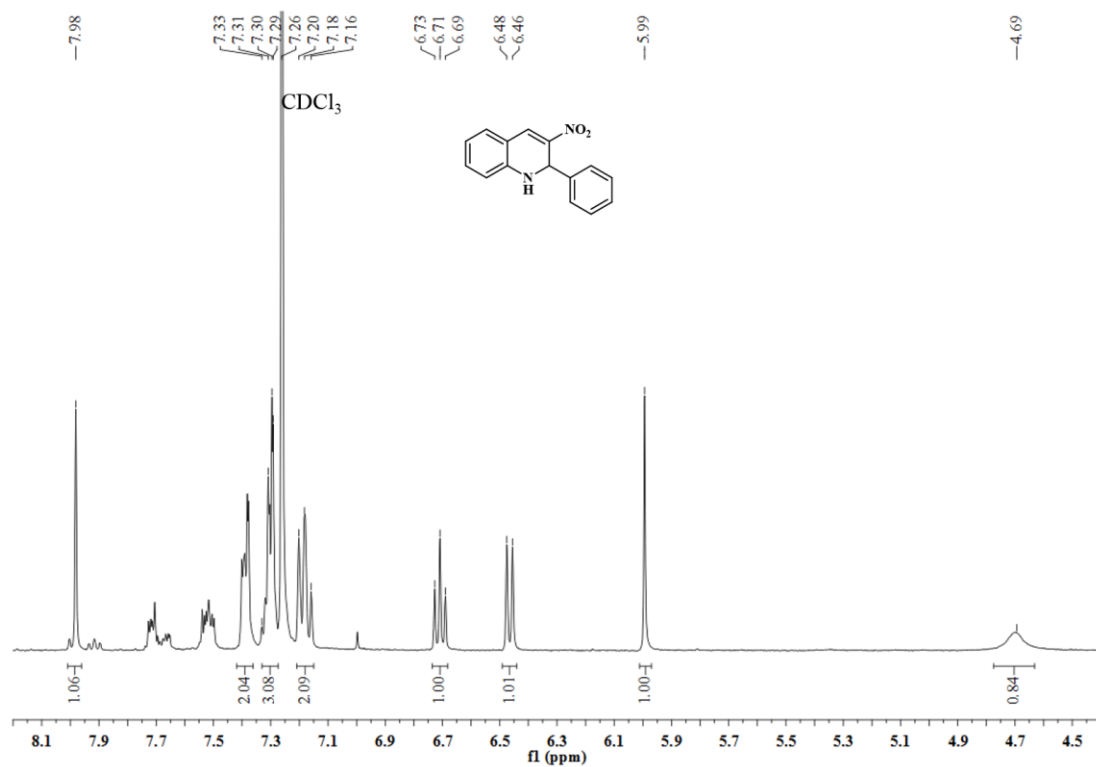
6,8-dibromo-3-nitro-2-phenyl-1,2-dihydroquinoline: 1H NMR (400 MHz, $CDCl_3$): δ 7.75 (s, 1H), 7.62 (s, 1H), 7.56 (s, 1H), 7.45–7.31 (m, 6H), 5.63 (s, 1H); ^{13}C NMR (101 MHz, $CDCl_3$): δ 191.98, 139.86, 137.10, 135.10, 134.64, 134.51, 134.16, 131.33, 131.15, 129.96, 128.75, 127.67, 127.02, 59.31, 49.35, 23.44. HRMS (ESI): calcd for $C_{15}H_{11}Br_2N_2NaO_2 [M]^+$ m/z 433.05; found 433.27.

HPLC analysis of 6-bromo-3-nitro-2-phenyl-1,2-dihydroquinoline

Structure	Column	Eluent	Flow rate	Retention time
	CHIRALPAK OD-H	Hexane: 2-PrOH = 85: 15	1.0 mL/min	8.97min
				10.87min

Appendix

$^1\text{H}/^{13}\text{C}$ NMR spectra



For yield calculation:

