

**Table S1.** List of sources and biochemical characteristics of purified microbial type I pullulanases.

Source	TE	MM (kDa)	T <sub>opt</sub> (°C)	pH <sub>opt</sub>	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	β-limit dextrin	Amylose	
<b>Type I pullulanase</b>											
<i>Anaerobranca gottschalkii</i> DSM 13577 <sup>t</sup>	R	96	70	8.0	G3	ND	G1–G <sub>o</sub>	ND	G5–G <sub>o</sub>	-	[1]
<i>Anoxybacillus</i> sp. AR-29	R	85	55	6.0	RD	ND	ND	ND	ND	ND	[2]
<i>Anoxybacillus</i> sp. LM14-2 <sup>b</sup>	R	82	70	6.0	RD	RD	RD	RD	ND	-	[3]
<i>Anoxybacillus</i> sp. LM18-11 <sup>c</sup>	R	82	60	6.0	G3	ND	ND	ND	ND	-	[4]
<i>Anoxybacillus</i> sp. SK3-4	R	80	60	6.0	G3	-	-	-	G3, G4, G5	-	[5]
<i>Aureobasidium pullulans</i>	N	73	ND	ND	G3	G2, G3	ND	G2, G3	ND	-	[6]
<i>Bacillus acidopullulyticus</i> <sup>c</sup>	R	101	60	5.0	G3, G4	RD	RD	RD	ND	-	[7,8]
<i>Bacillus acidopullulyticus</i>	N	115	60–65	5.0	G3, G4	RD	RD	RD	RD	-	[9]
<i>Bacillus acidopullulyticus</i>	N	116	60–65	5.0	G3, G4	RD	RD	RD	RD	-	[9]
<i>Bacillus amyloliquefaciens</i> HxP-21	N	51	55	4.5	RD	RD	RD	RD	ND	-	[10]
<i>Bacillus cereus</i> Nws-bc5	R	80	40	6.0	G3	G2, G3	RD	ND	ND	-	[11]
<i>Bacillus cereus</i>	N	110	50	6.0–6.5	G3	ND	RD	ND	G2, G3, G4	ND	[12]
<i>Bacillus deramificans</i>	R	101	55	4.5	RD	ND	ND	ND	ND	ND	[13]
<i>Bacillus flavocaldarius</i> KP 1228	R	55	83–85	ND	RD	RD	RD	RD	ND	-	[14]
<i>Bacillus flavocaldarius</i> KP 1228	N	55	75–80	6.3	G3	RD	RD	ND	RD	-	[15]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltopentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>o</sub>: oligosaccharides ( $\geq$  G8); TE: type of enzyme; MM: molecular mass; T<sub>opt</sub>: optimum temperature; pH<sub>opt</sub>: optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

**Table S1.** *Continued.*

Source	TE	MM (kDa)	T <sub>opt</sub> (°C)	pH <sub>opt</sub>	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	β-limit dextrin	Amylose	
<b>Type I pullulanase</b>											
<i>Bacillus megaterium</i> WW1210	R	112	55	6.5	G3	ND	G3–G7	ND	ND	ND	[16]
<i>Bacillus methanolicus</i> PB1	R	66	50	5.5	G3	RD	RD	ND	ND	-	[17]
<i>Bacillus naganoensis</i> ATCC 53909 <sup>T</sup>	R	101	60	4.5	RD	ND	ND	ND	ND	ND	[18]
<i>Bacillus subtilis</i> 168 <sup>c</sup>	R	81	40	6.0	RD	ND	ND	ND	ND	ND	[19]
<i>Bacillus subtilis</i> BK07	R	90	40	8.0	RD	ND	ND	ND	ND	ND	[20]
<i>Bacillus subtilis</i> PY22	R	90	40	6.0	RD	ND	ND	ND	ND	ND	[20]
<i>Bacillus subtilis</i> WY-34	R	76.2	40	6.0	RD	RD	ND	ND	ND	-	[21]
<i>Bacillus pseudofirmus</i> 703	R	87	45	7.0–8.0	G3	RD	RD	ND	ND	ND	[22]
<i>Bacillus</i> sp. AN-7	N	106	90	6.0	G2, G3	G2	G2	ND	ND	-	[23]
<i>Bacillus</i> sp. CICIM 263	R	101	70	6.5	G3	G2, G3, G4	G2, G3	G2, G3	ND	-	[24]
<i>Bacillus</i> sp. KSM-1876	N	120	50	10.0–10.5	G3	RD	RD	RD	ND	-	[25]
<i>Bacillus</i> sp. S-1	N	180	60	9.0	G3	RD	RD	RD	RD	-	[26]
<i>Bacillus</i> sp. S-1	N	140	60	9.0	G3	RD	RD	RD	RD	-	[26]
<i>Clostridium thermosulfurogenes</i> SV2	N	80	75	6.0	G3	ND	ND	ND	ND	ND	[27]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltopentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>o</sub>: oligosaccharides ( $\geq$  G8); TE: type of enzyme; MM: molecular mass; T<sub>opt</sub>: optimum temperature; pH<sub>opt</sub>: optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

**Table S1.** *Continued.*

Source	TE	MM (kDa)	T <sub>opt</sub> (°C)	pH <sub>opt</sub>	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	β-limit dextrin	Amylose	
<b>Type I pullulanase</b>											
<i>Exiguobacterium acetyllicum</i> YH5	R	100	50	6.0	G3	G2, G3, G <sub>o</sub>	-	[28]			
<i>Exiguobacterium</i> sp. SH3	R	108	45	8.5	G3	RD	RD	ND	ND	-	[29]
<i>Fervidobacterium nodosum</i> Rt17-B1	R	95	80	5.0	G3	G2–G6	G2–G6	ND	ND	ND	[30]
<i>Fervidobacterium pennavorans</i> Ven5	R	93	80	6.0	G3	G2, G3	G2, G3	G2, G3	ND	-	[31]
<i>Fervidobacterium pennavorans</i> Ven5	N	240 <sup>d</sup>	85	6.0	G3, G <sub>o</sub>	ND	ND	ND	ND	-	[32]
<i>Geobacillus kaustophilus</i> DSM 7263 <sup>t</sup>	R	80	65	6.0	G3	RD	RD	RD	ND	-	[33]
<i>Geobacillus thermopakistaniensis</i>	R	80	ND	ND	ND	ND	ND	ND	ND	ND	[34]
<i>Geobacillus stearothermophilus</i> TRS128	N	83	65	6.0	G3	ND	ND	ND	ND	ND	[35]
<i>Geobacillus thermoleovorans</i> US105	R	160 <sup>d</sup>	70	6.0	G3	ND	ND	ND	ND	-	[36]
<i>Klebsiella pneumoniae</i> <sup>c</sup>	R	145	30–37	5.0	G3	RD	RD	RD	RD	-	[37– 39]
<i>Klebsiella variicola</i> SHN-1	R	118	55	5.0	G3	ND	ND	ND	ND	ND	[40]
<i>Laceyella sacchari</i> No. 15	N	79	70	7.0	G3	G2, G3	G2, G3	ND	G2, G3	ND	[41]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltpentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>o</sub>: oligosaccharides ( $\geq$  G8); TE: type of enzyme; MM: molecular mass; T<sub>opt</sub>: optimum temperature; pH<sub>opt</sub>: optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

**Table S1.** *Continued.*

Source	TE	MM (kDa)	$T_{opt}$ (°C)	$pH_{opt}$	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	$\beta$ -limit dextrin	Amylose	
<b>Type I pullulanase</b>											
<i>Lactococcus lactis</i> IBB 500	N	73	45	4.5	G2, G3	G2, G3	ND	ND	ND	-	[42]
<i>Lactobacillus acidophilus</i> NCFM	R	129	50	5.5	G3	G2, G3, G4	ND	ND	RD	ND	[43]
<i>Micrococcus</i> sp. 207	N	120	50	8.0	G3, G4	RD	RD	RD	ND	-	[44]
Metagenome sample (Pulm)	R	80	40	6.0–7.0	RD	RD	ND	ND	ND	ND	[45]
Metagenome sample (PersiPul1)	R	71	60	7.0	RD	RD	RD	ND	ND	-	[46]
Mixed culture bacterium (AmyA1)	R	81.7	60	6.0	G3	RD	ND	RD	ND	ND	[47]
<i>Paenibacillus barengoltzii</i> CAU904	R	75	50	5.5	G3	RD	RD	ND	ND	-	[48]
<i>Paenibacillus lautus</i> DSM 3035 <sup>T</sup>	R	87.9	40	7.0	G3	RD	RD	RD	ND	-	[49]
<i>Paenibacillus polymyxa</i> Nws-pp2	R	99	35	6.0	G3	G2, G3	G2, G3	G2, G3	G2, G3	ND	[50]
<i>Thermotoga maritima</i> MSB8	N	89	90	5.9	G3	ND	ND	ND	ND	ND	[51]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltopentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>n</sub>: oligosaccharides ( $\geq G8$ ); TE: type of enzyme; MM: molecular mass;  $T_{opt}$ : optimum temperature;  $pH_{opt}$ : optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

**Table S1.** *Continued.*

Source	TE	MM (kDa)	T <sub>opt</sub> (°C)	pH <sub>opt</sub>	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	β-limit dextrin	Amylose	
<b>Type I pullulanase</b>											
<i>Thermotoga neapolitana</i> KCCM 41025	R	93	80–85	6.5	G3	G2, G3, G <sub>o</sub>	G2, G3, G <sub>o</sub>	G2, G3, G <sub>o</sub>	ND	-	[52]
<i>Thermus aquaticus</i> YT-1	N	83	85	6.5	G3	G1, G2, G3	G1, G2, G3	ND	ND	ND	[53]
<i>Thermus caldophilus</i> GK-24	N	65	75	5.5	G3	G1, G2, G3	G1, G2, G3	RD	RD	ND	[54]
<i>Thermus thermophilus</i> HB8	R	80	70	5.5–6.5	G3	ND	ND	ND	ND	ND	[55]
<i>Shewanella arctica</i> 40-3	R	155	35	6.0–7.0	G3	ND	ND	ND	ND	ND	[56]
<i>Shewanella arctica</i> 40-3	N	150	45	7.0	G3	RD	RD	ND	ND	-	[57]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltpentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>o</sub>: oligosaccharides ( $\geq$  G8); TE: type of enzyme; MM: molecular mass; T<sub>opt</sub>: optimum temperature; pH<sub>opt</sub>: optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits

**Table S2:** List of sources and biochemical characteristics of purified microbial type II pullulanases.

Source	TE	MM (kDa)	$T_{opt}$ (°C)	$pH_{opt}$	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	$\beta$ -limit dextrin	Amylose	
<b>Type II pullulanase (amylopullulanase)</b>											
<i>Alkalilimnicola</i> sp. NM-DCM-1	R	80	55	9.5	G3	G2, G3, G4	ND	ND	ND	G4, G5	[58]
<i>Anoxybacillus</i> sp. SK3-4	N	225	60	7.5	G1, G2, G3	G1, G2, G3	G1, G2, G3	G1, G2, G3	ND	G2, G3	[59]
<i>Anoxybacillus</i> sp. WB42	R	83	55–65	5.8	G3	G2–G5	G2–G5	ND	ND	G2–G5	[60]
<i>Bacillus cereus</i> H1.5	N	93	55	6.0	G3	RD	RD	ND	ND	RD	[61]
<i>Bacillus circulans</i> F-2	N	220	50	7.0	G3	G3–G <sub>o</sub>	ND	ND	ND	ND	[62]
<i>Bacillus megaterium</i> Y103	R	110	45	6.5	G3	G1–G4	G1–G4	-	-	G1–G4	[63]
<i>Bacillus</i> sp. DSM 405	N	126	70	6.0	G3	G2, G3, G4	G2, G3, G4	G2, G3, G4	ND	G2, G3, G4	[64]
<i>Bacillus</i> sp. XAL601	R	224	70	9.0	G3	G2, G3, G4	ND	ND	ND	G2, G3, G4	[65]
<i>Caldisericum exile</i>	R	139	75	5.5, 8.5	G3	G1–G6	G1–G6	ND	ND	G1–G6	[66]
<i>Cohnella</i> sp. A01	R	127	70	8.0	G3	G2, G3, G4	ND	ND	ND	ND	[67]
<i>Cohnella</i> sp. A01	R	70	60	6.0	G3	G2, G3, G4	ND	ND	ND	ND	[67]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltopentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>o</sub>: oligosaccharides ( $\geq$  G8); TE: type of enzyme; MM: molecular mass;  $T_{opt}$ : optimum temperature;  $pH_{opt}$ : optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

**Table S2. Continued.**

Source	TE	MM (kDa)	T <sub>opt</sub> (°C)	pH <sub>opt</sub>	Reaction product <sup>a</sup>					Ref.
					Pullulan	Starch	Amylopectin	Glycogen	β-limit dextrin	
<b>Type II pullulanase (amylopullulanase)</b>										
<i>Caldivirga maquilingensis</i> IC-167 <sup>T</sup>	R	62.7	100	5.0	RD	G6-G <sub>o</sub>	G6-G <sub>o</sub>	ND	ND	G6-G <sub>o</sub> [68]
<i>Clostridium thermosulfurigenes</i> EM1	N	100	60–65	5.5–6.0	G3	G2, G3	ND	G2, G3	ND	G2, G3, G4 [69]
<i>Clostridium thermosulfurigenes</i> SVM17	N	97	70	5.5–6.0	G3	G1, G2	ND	ND	ND	ND [70]
<i>Desulfurococcus mucosus</i> DSM 2162 <sup>T</sup>	R	110 <sup>d</sup>	85	5.0	G3	G1, G2, G3	RD	-	ND	G1, G2, G3 [71]
<i>Geobacillus stearothermophilus</i> ATCC 12980 <sup>T</sup>	R	184	ND	ND	ND	ND	ND	ND	ND	ND [72]
<i>Geobacillus stearothermophilus</i> G-82	N	56	60	7.0	G1, G2, G3	ND	G1, G2, G3	G1, G2, G3	ND	G1, G2, G3 [73]
<i>Geobacillus stearothermophilus</i> TS-23	R	220	ND	ND	ND	ND	ND	ND	ND	ND [74]
<i>Geobacillus thermoleovorans</i> NP1	N	112	50	6.0	G1, G2, G3, G <sub>o</sub>	G2, G3, G <sub>o</sub>	ND	ND	ND	ND [75]
<i>Geobacillus thermoleovorans</i> NP33	R	182	60	7.0	G3	G2, G3, G4	G2, G3, G4	G2, G3, G4	ND	G2, G3, G4 [76]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltopentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>o</sub>: oligosaccharides ( $\geq$  G8); TE: type of enzyme; MM: molecular mass; T<sub>opt</sub>: optimum temperature; pH<sub>opt</sub>: optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

**Table S2. Continued.**

Source	TE	MM (kDa)	T <sub>opt</sub> (°C)	pH <sub>opt</sub>	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	β-limit dextrin	Amylose	
<b>Type II pullulanase (amylopullulanase)</b>											
<i>Geobacillus thermoleovorans</i> NP33	N	48	80	ND	RD	RD	ND	ND	ND	ND	[77]
<i>Geobacillus thermoleovorans</i> NP33	N	105	80	7.0	G1, G2, G3	G1, G2, G3	ND	ND	ND	ND	[78]
<i>Geobacillus</i> sp. L14	N	100	65	5.5	G1, G2, G3	G1, G2	ND	ND	ND	ND	[79]
<i>Halorubrum</i> sp. Ha25	N	140	50	7.5	G1	G1	ND	ND	ND	ND	[80]
<i>Lactobacillus amylophilus</i> GV6	N	90	37	6.5	G3	G1, G2, G3	ND	ND	ND	ND	[81]
Metagenome sample (PulSS4)	R	164	40	9.0	G3	G1–G4	G1–G4	G1–G4	ND	G1–G4	[82]
<i>Paenibacillus puldeungensis</i> LK18	R	76	45	6.0	RD	ND	ND	ND	ND	ND	[83]
<i>Pyrococcus furiosus</i> DSM 3638 <sup>T</sup>	R	90	105	6.0	G3, G4	G1–G6	RD	RD	ND	RD	[84]
<i>Pyrobaculum calidifontis</i> VA1	R	111	90	5.6	RD	ND	ND	ND	ND	ND	[85]
<i>Pyrococcus furiosus</i> DSM 3638 <sup>T</sup>	N	119	100	5.5	G3	G1–G6	ND	G1–G6	ND	G1–G6	[86]
<i>Pyrococcus woesei</i> DSM 3773 <sup>T</sup>	R	90	100	6.0	G3	G1–G6	ND	G1–G5	ND	G1–G4	[87]
<i>Pyrococcus yayanosii</i> CH1	R	125	95	6.6	G3	ND	ND	ND	ND	G5–G <sub>o</sub>	[88]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltopentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>o</sub>: oligosaccharides ( $\geq$  G8); TE: type of enzyme; MM: molecular mass; T<sub>opt</sub>: optimum temperature; pH<sub>opt</sub>: optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

Table S2. *Continued.*

Source	TE	MM (kDa)	$T_{opt}$ (°C)	$pH_{opt}$	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	$\beta$ -limit dextrin	Amylose	
<b>Type II pullulanase (amylopullulanase)</b>											
<i>Staphylothermus marinus</i> DSM 3639 <sup>T</sup>	R	75.3	105	5.0	G3	ND	ND	ND	ND	ND	[89]
<i>Streptomyces erumpens</i> MTCC 7317	N	45	ND	ND	RD	G1, G2	ND	ND	ND	ND	[90]
<i>Sulfolobus acidocaldarius</i> DSM 639 <sup>T</sup>	R	100 <sup>d</sup>	100	3.0	G1, G2, G3	G1, G2, G3	G1, G2, G3	G1, G2, G3	ND	G1, G2, G3	[91]
<i>Thermoanaerobacter pseudethanolicus</i> ATCC 33233 <sup>T</sup>	R	140	90	5.5	G3	RD	RD	RD	RD	ND	[92-94]
<i>Thermoanaerobacter thermohydrosulfuricus</i> E101	R	165	80	ND	G3	ND	ND	ND	ND	ND	[95]
<i>Thermoanaerobacter</i> sp. B6A	N	450 <sup>d</sup>	75	5.0	G3	RD	RD	RD	RD	G2, G3, G4	[96]
<i>Thermoanaerobium brockii</i>	R	105	70	5.0	G3	G3-G <sub>o</sub>	ND	ND	RD	ND	[97]
<i>Thermoanaerobacterium</i> <i>saccharolyticum</i> B6A-RIT <sup>T</sup>	R	140	70-75	5.5-6.0	G3	RD	RD	RD	ND	RD	[98]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltpentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>o</sub>: oligosaccharides ( $\geq$  G8); TE: type of enzyme; MM: molecular mass;  $T_{opt}$ : optimum temperature;  $pH_{opt}$ : optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

**Table S2. Continued.**

Source	TE	MM (kDa)	T <sub>opt</sub> (°C)	pH <sub>opt</sub>	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	β-limit dextrin	Amylose	
<b>Type II pullulanase (amylopullulanase)</b>											
<i>Thermoanaerobacterium saccharolyticum</i> NTOU1	R	100	70	5.0	G2, G3	G2, G3	G2, G3	G2, G3	ND	G2, G3	[99]
<i>Thermoanaerobacterium thermosaccharolyticum</i> DSM 571 <sup>T</sup>	N	150	65	5.0–5.5	G3, G <sub>o</sub>	RD	RD	RD	ND	RD	[100]
<i>Thermoanaerobacterium thermosulfurigenes</i> EM1	R	205	60	5.5	G3	G2, G3	ND	G2, G3	ND	G2, G3	[101–103]
<i>Thermoanaerobium</i> sp. Tok6-B1	N	120	80	5.5	G3	G2, G3, G4	G2, G3	ND	ND	G2, G3, G4	[104]
<i>Thermococcus hydrothermalis</i>	R	105	105	5.75	G3	G1–G7	ND	ND	ND	G1–G7	[105]
<i>Thermococcus hydrothermalis</i> AL662	N	128	95	5.5	G3	G4–G7	ND	G4–G7	ND	ND	[106]
<i>Thermococcus kodakarensis</i> KOD1 <sup>T</sup>	R	86	100	5.5	G3	G2, G3	ND	ND	ND	ND	[107]
<i>Thermococcus kodakarensis</i> KOD1 <sup>T</sup>	R	120	105	5.5	RD	RD	ND	ND	ND	ND	[108]
<i>Thermococcus litoralis</i> DSM 5473 <sup>T</sup>	N	110	100	5.5	G3	G1–G6	ND	G1–G6	ND	G1–G6	[86]
<i>Thermococcus siculi</i> HJ21	R	148.6	95	6.0	RD	ND	ND	ND	ND	ND	[109]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltopentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>o</sub>: oligosaccharides ( $\geq$  G8); TE: type of enzyme; MM: molecular mass; T<sub>opt</sub>: optimum temperature; pH<sub>opt</sub>: optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

**Table S2. Continued.**

Source	TE	MM (kDa)	$T_{opt}$ (°C)	$pH_{opt}$	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	$\beta$ -limit dextrin	Amylose	
<b>Type II pullulanase (amylopullulanase)</b>											
<i>Thermofilum pendens</i> Hrk 5	R	65.5	95–100	3.5	RD	ND	G6–G <sub>o</sub>	ND	ND	ND	[110]
<i>Thermus thermophilus</i> HB27	R	52	70	6.5	G1, G2, G3	G1, G2, G3	G1–G4	ND	ND	G1, G2, G3	[111]
Uncultured bacterium (Env Npu193A)	R	66	75	7.0	G3	G2–G <sub>o</sub>	ND	ND	ND	ND	[112]
<b>Type II pullulanase (<math>\alpha</math>-amylase-pullulanase)</b>											
<i>Alkalibacterium</i> sp. SL3	R	226	50	9.0	G3	G1–G4	G1–G4	ND	ND	ND	[113]
<i>Bacillus</i> sp. KSM-1378	N	210	50	9.5	G3, G <sub>o</sub>	G1–G6	G1–G6	G1–G6	ND	G1–G6	[114]
<i>Bifidobacterium breve</i> UCC2003	R	182.3	ND	ND	G3	G2, G3	G2, G3	G2, G3	ND	ND	[115]
<i>Bifidobacterium adolescentis</i> P2P3	R	180	55	5.0	G3	G3	RD	RD	ND	RD	[116]
<i>Lactobacillus plantarum</i> L137	R	211	40	4.0	G3	G1–G4	ND	ND	ND	G4–G <sub>o</sub>	[117]
<i>Streptococcus suis</i> P1/7	R	230	ND	ND	ND	ND	ND	ND	ND	ND	[118]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltpentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>o</sub>: oligosaccharides ( $\geq$  G8); TE: type of enzyme; MM: molecular mass;  $T_{opt}$ : optimum temperature;  $pH_{opt}$ : optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

**Table S3.** List of sources and biochemical characteristics of purified microbial pullulan hydrolases.

Source	TE	MM (kDa)	$T_{opt}$ (°C)	$pH_{opt}$	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	$\beta$ -limit dextrin	Amylose	
<b>Pullulan hydrolase type I (neopullulanase)</b>											
<i>Anoxybacillus flavigilans</i>	R	72.8	60	5.5–6.5	P	G1, G2, G3	ND	ND	ND	ND	[119]
<i>Alicyclobacillus acidocaldarius</i> ATCC 27009 <sup>T</sup>	R	66	55	5.5	P	ND	ND	ND	ND	ND	[120]
<i>Bacillus polymyxa</i> CECT 155 <sup>b</sup>	R	58	50	6.0	P	RD	RD	ND	ND	RD	[121,122]
<i>Bacillus</i> sp. KSM-1876	R	68	40	7.5	P	ND	ND	ND	ND	ND	[123]
<i>Bacteroides thetaiotaomicron</i> 95-1	R	70	ND	ND	P	ND	ND	ND	ND	RD	[124]
<i>Geobacillus stearothermophilus</i> TRS40	R	62	60–65	6.0	G1, G2, P	G1, G2	G2	ND	ND	G1, G2	[125,126]
<i>Geobacillus stearothermophilus</i> IMA6503	R	140 <sup>d</sup>	55	6.0	P	G1, G2	ND	ND	ND	ND	[127]
<i>Laceyella sacchari</i>	R	69.1	ND	5.5–6.5	P	G1, G2, G3	ND	ND	ND	ND	[119]
<i>Lactobacillus mucosae</i> LM1	R	67	37	6.0	P	ND	ND	ND	ND	ND	[128]
<i>Micrococcus halobius</i> OR-1	N	53	50	8.0	P	G2, G3	G3, P	ND	ND	G2	[129]
<i>Paenibacillus</i> sp. KCTC 8848P	R	58	ND	ND	P	RD	ND	ND	ND	ND	[130]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltpentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>n</sub>: oligosaccharides ( $\geq$  G8); P: panose; I: isopanose; TE: type of enzyme; MM: molecular mass;  $T_{opt}$ : optimum temperature;  $pH_{opt}$ : optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

Table S3. *Continued.*

Source	TE	MM (kDa)	T <sub>opt</sub> (°C)	pH <sub>opt</sub>	Reaction product <sup>a</sup>						Ref.
					Pullulan	Starch	Amylopectin	Glycogen	β-limit dextrin	Amylose	
<b>Pullulan hydrolase type I (neopullulanase)</b>											
<i>Thermoactinomyces vulgaris</i> R-47 <sup>c</sup>	R	74	70	5.0	P	G1, G2	ND	ND	ND	RD	[131,132] ]
<i>Thermoactinomyces vulgaris</i> R-47 <sup>c</sup>	R	67	40	5.5	P	G1, G2	ND	ND	ND	ND	[133]
Uncultured bacterium (Amy 132)	R	69	55	5.5–6.5	P	G1, G2, G3	ND	ND	ND	ND	[119]
<b>Pullulan hydrolase type II (isopullulanase)</b>											
<i>Aspergillus brasiliensis</i> ATCC 9642 <sup>T</sup> <sup>c</sup>	R	90	40–45	3.5	I	ND	ND	ND	ND	ND	[134,135] ]
<i>Aspergillus brasiliensis</i> ATCC 9642 <sup>T</sup>	N	71	45	3.5	I	ND	ND	ND	ND	ND	[136]
<i>Aspergillus brasiliensis</i> ATCC 9642 <sup>T</sup>	N	69	40	3.5	I	ND	ND	ND	ND	ND	[136]
<i>Bacillus</i> sp. US149	N	200 <sup>d</sup>	60	5.0	I	-	-	-	ND	ND	[137]
<b>Pullulan hydrolase type III</b>											
<i>Thermococcus aggregans</i> DSM 10597	R	80	95	6.5	G1, G2, G3, P	G1, G2, G3	G1, G2, G3	ND	ND	G1, G2, G3	[138]
<i>Thermococcus kodakarensis</i> KOD1 <sup>T</sup>	R	80	95–100	3.5	G1, G2, G3, P	G1, G2, G3	G1, G2, G3	G1, G2, G3	ND	G2	[139]

G1: glucose; G2: maltose; G3: maltotriose; G4: maltotetraose; G5: maltpentaose; G6: maltohexaose; G7: maltoheptaose; G<sub>n</sub>: oligosaccharides ( $\geq$  G8); P: panose; I: isopanose; TE: type of enzyme; MM: molecular mass; T<sub>opt</sub>: optimum temperature; pH<sub>opt</sub>: optimum pH; R: recombinant; N: native; ND: not determined; RD: substrate was reduced as inferred from enzymatic assay; -: no product detected; T: type strain; a: reaction products were determined using high-performance liquid chromatography or thin-layer chromatography in the respective reference; b: the enzyme was characterized from crude; c: the crystal structures were solved and the data were available in RCSB Protein Data Bank (PDB) database; d: consists of several subunits.

## References

1. Bertoldo, C.; Armbrecht, M.; Becker, F.; Schäfer, T.; Antranikian, G.; Liebl, W. Cloning, sequencing, and characterization of a heat- and alkali-stable type I pullulanase from *Anaerobranca gottschalkii*. *Appl. Environ. Microbiol.* **2004**, *70*, 3407–3416.
2. Li, S.-F.; Xu, S.-Y.; Wang, Y.-J.; Zheng, Y.-G. Tailoring pullulanase PULAR from *Anoxybacillus* sp. AR-29 for enhanced catalytic performance by a structure-guided consensus approach. *Bioresour. Bioprocess.* **2022**, *In press*, doi: 10.21203/rs.21203.rs-1189094/v1189091.
3. Shaojing, S.; Fuping, L.; Nan, J.; Li, L.; Jianyong, X.; Muchen, C.; Hui, S. Study of a novel thermostable pullulanase producing strain *Anoxybacillus* sp. LM14-2. *Biotechnol. Bull.* **2011**, *9*, 136–141.
4. Xu, J.; Ren, F.; Huang, C.-H.; Zheng, Y.; Zhen, J.; Sun, H.; Ko, T.-P.; He, M.; Chen, C.-C.; Chan, H.-C., et al. Functional and structural studies of pullulanase from *Anoxybacillus* sp. LM18-11. *Proteins* **2014**, *82*, 1685–1693.
5. Kahar, U.M.; Ng, C.L.; Chan, K.-G.; Goh, K.M. Characterization of a type I pullulanase from *Anoxybacillus* sp. SK3-4 reveals an unusual substrate hydrolysis. *Appl. Microbiol. Biotechnol.* **2016**, *100*, 6291–6307.
6. Moubasher, H.; Wahsh, S.S.; El-Kassem, N.A. Purification of pullulanase from *Aureobasidium pullulans*. *Microbiology* **2010**, *79*, 759–766.
7. Svendsen, A.; Andersen, C.; Borchert, T. *International Publication No. WO/2001/051620*. Retrieved on October 11, 2017, from <http://www.freepatentsonline.com/>. **2001**.
8. Turkenburg, J.P.; Brzozowski, A.M.; Svendsen, A.; Borchert, T.V.; Davies, G.J.; Wilson, K.S. Structure of a pullulanase from *Bacillus acidopullulyticus*. *Proteins* **2009**, *76*, 516–519.
9. Kusano, S.; Nagahata, N.; Takahashi, S.-I.; Fujimoto, D.; Sakano, Y. Purification and properties of *Bacillus acidopullulyticus* pullulanase. *Agr. Biol. Chem.* **1988**, *52*, 2293–2298.
10. Gao, Z.; Hu, X.; Song, Y.; Ding, F.; Zhao, Y.; Chen, T. Purification and enzymatic characterization of thermolabile type I pullulanase from *Bacillus amyloliquefaciens*. *Sh. Kexue/Food Sci.* **2021**, *42*, 130–137.
11. Wei, W.; Ma, J.; Guo, S.; Wei, D.-Z. A type I pullulanase of *Bacillus cereus* Nws-bc5 screening from stinky tofu brine: functional expression in *Escherichia coli* and *Bacillus subtilis* and enzyme characterization. *Process Biochem.* **2014**, *49*, 1893–1902.
12. Takasaki, Y. Purifications and enzymatic properties of β-amylase and pullulanase from *Bacillus cereus* var. *mycoides*. *Agr. Biol. Chem.* **1976**, *40*, 1523–1530.
13. Duan, X.; Chen, J.; Wu, J. Improving the thermostability and catalytic efficiency of *Bacillus deramificans* pullulanase by site-directed mutagenesis. *Appl. Environ. Microbiol.* **2013**, *79*, 4072–4077.
14. Kashiwabara, S.; Ogawa, S.; Miyoshi, N.; Oda, M.; Suzuki, Y. Three domains comprised in thermostable molecular weight 54,000 pullulanase of type I from *Bacillus flavocaldarius* KP1228. *Biosci. Biotechnol. Biochem.* **1999**, *63*, 1736–1748.
15. Suzuki, Y.; Hatagaki, K.; Oda, H. A hyperthermostable pullulanase produced by an extreme thermophile, *Bacillus flavocaldarius* KP 1228, and evidence for the proline theory of increasing protein thermostability. *Appl. Microbiol. Biotechnol.* **1991**, *34*, 707–714.
16. Yang, S.; Yan, Q.; Bao, Q.; Liu, J.; Jiang, Z. Expression and biochemical characterization of a novel type I pullulanase from *Bacillus megaterium*. *Biotechnol. Lett.* **2017**, *39*, 397–405.
17. Zhang, S.-Y.; Guo, Z.-W.; Wu, X.-L.; Ou, X.-Y.; Zong, M.-H.; Lou, W.-Y. Recombinant expression and characterization of a novel cold-adapted type I pullulanase for efficient amylopectin hydrolysis. *J. Biotechnol.* **2020**, *313*, 39–47.
18. Wang, X.; Nie, Y.; Mu, X.; Xu, Y.; Xiao, R. Disorder prediction-based construct optimization improves activity and catalytic efficiency of *Bacillus naganoensis* pullulanase. *Sci. Rep.* **2016**, *6*, 24574.
19. Malle, D.; Itoh, T.; Hashimoto, W.; Murata, K.; Utsumi, S.; Mikami, B. Overexpression, purification and preliminary X-ray analysis of pullulanase from *Bacillus subtilis* strain 168. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.* **2006**, *62*, 381–384.
20. Erden-Karaoglu, F.; Karakaş-Budak, B.; Karaoglu, M.; Inan, M. Cloning and expression of pullulanase from *Bacillus subtilis* BK07 and PY22 in *Pichia pastoris*. *Protein Expr. Purif.* **2019**, *162*, 83–88.
21. Peng, H.; Peng, Z.; Qiao-Juan, Y.; Shao-Qing, Y.; Zheng-Qiang, J. Expression of a pullulanase gene from the thermophilic *Bacillus subtilis* and characterization of the recombinant enzyme. *Microbiology China* **2011**, *38*, 1755–1761.
22. Lu, Z.; Hu, X.; Shen, P.; Wang, Q.; Zhou, Y.; Zhang, G.; Ma, Y. A pH-stable, detergent and chelator resistant type I pullulanase from *Bacillus pseudofirmus* 703 with high catalytic efficiency. *Int. J. Biol. Macromol.* **2018**, *109*, 1302–1310.

23. Kunamneni, A.; Singh, S. Improved high thermal stability of pullulanase from a newly isolated thermophilic *Bacillus* sp. AN-7. *Enzyme Microb. Technol.* **2006**, *39*, 1399–1404.
24. Li, Y.; Zhang, L.; Niu, D.; Wang, Z.; Shi, G. Cloning, expression, characterization, and biocatalytic investigation of a novel bacilli thermostable type I pullulanase from *Bacillus* sp. CICIM 263. *J Agric. Food Chem.* **2012**, *60*, 11164–11172.
25. Ara, K.; Igarashi, K.; Saeki, K.; Kawai, S.; Ito, S. Purification and some properties of an alkaline pullulanase from alkalophilic *Bacillus* sp. KSM-1876. *Biosci. Biotechnol. Biochem.* **1992**, *56*, 62–65.
26. Lee, M.-J.; Lee, Y.-C.; Kim, C.-H. Intracellular and extracellular forms of alkaline pullulanase from an alkaliphilic *Bacillus* sp. S-1. *Arch. Biochem. Biophys.* **1997**, *337*, 308–316.
27. Reddy, P.R.M.; Swamy, M.V.; Seenayya, G. Purification and characterization of thermostable  $\beta$ -amylase and pullulanase from high-yielding *Clostridium thermosulfurogenes* SV2. *World J. Microbiol. Biotechnol.* **1998**, *14*, 89–94.
28. Qiao, Y.; Peng, Q.; Yan, J.; Wang, H.; Ding, H.; Shi, B. Gene cloning and enzymatic characterization of alkali-tolerant type I pullulanase from *Exiguobacterium acetylicum*. *Lett Appl Microbiol* **2015**, *60*, 52–59.
29. Rajaei, S.; Noghabi, K.A.; Sadeghizadeh, M.; Zahiri, H.S. Characterization of a pH and detergent-tolerant, cold-adapted type I pullulanase from *Exiguobacterium* sp. SH3. *Extremophiles* **2015**, *19*, 1145–1155.
30. Yang, Y.; Zhu, Y.; Obaroakpo, J.U.; Zhang, S.; Lu, J.; Yang, L.; Ni, D.; Pang, X.; Lv, J. Identification of a novel type I pullulanase from *Fervidobacterium nodosum* Rt17-B1, with high thermostability and suitable optimal pH. *Int. J. Biol. Macromol.* **2020**, *143*, 424–433.
31. Bertoldo, C.; Duffner, F.; Jorgensen, P.; Antranikian, G. Pullulanase type I from *Fervidobacterium pennavorans* Ven5: cloning, sequencing, and expression of the gene and biochemical characterization of the recombinant enzyme. *Appl. Environ. Microbiol.* **1999**, *65*, 2084–2091.
32. Koch, R.; Canganella, F.; Hippe, H.; Jahke, K.D.; Antranikian, G. Purification and properties of a thermostable pullulanase from a newly isolated thermophilic anaerobic bacterium, *Fervidobacterium pennavorans* Ven5. *Appl. Environ. Microbiol.* **1997**, *63*, 1088–1094.
33. Li, L.; Dong, F.; Lin, L.; He, D.; Chen, J.; Wei, W.; Wei, D. Biochemical characterization of a novel thermostable type I pullulanase produced recombinantly in *Bacillus subtilis*. *Starch/Stärke* **2018**, *70*, 1700179.
34. Iqrar, U.; Javaid, H.; Ashraf, N.; Ahmad, A.; Latief, N.; Shahid, A.A.; Ahmad, W.; Ijaz, B. Structural and functional analysis of pullulanase type 1 (PulA) from *Geobacillus thermopakistaniensis*. *Mol. Biotechnol.* **2020**, *62*, 370–379.
35. Kuriki, T.; Park, J.-H.; Okada, S.; Imanakai, T. Purification and characterization of thermostable pullulanase from *Bacillus stearothermophilus* and molecular cloning and expression of the gene in *Bacillus subtilis*. *Appl. Environ. Microbiol.* **1988**, *54*, 2881–2883.
36. Ayadi, D.Z.; Ali, M.B.; Jemli, S.; Mabrouk, S.B.; Mezghani, M.; Messaoud, E.B.; Bejar, S. Heterologous expression, secretion and characterization of the *Geobacillus thermoleovorans* US105 type I pullulanase. *Appl. Microbiol. Biotechnol.* **2008**, *78*, 473–481.
37. Abdullah, M.; Catley, B.J.; Lee, E.Y.C.; Robyt, J.; Wallenfels, K.; Whelan, W.J. The mechanism of carbohydrase action. 11. Pullulanase, an enzyme specific for the hydrolysis of alpha-1 $\rightarrow$ 6-bonds in amylose oligo- and polysaccharides. *Cereal Chem.* **1966**, *43*, 111–118.
38. Abdullah, M.; French, D. Substrate specificity of pullulanase. *Arch Biochem Biophys* **1970**, *137*, 483–493.
39. Mikami, B.; Iwamoto, H.; Malle, D.; Yoon, H.-J.; Demirkhan-Sarikaya, E.; Mezaki, Y.; Katsuya, Y. Crystal structure of pullulanase: evidence for parallel binding of oligosaccharides in the active site. *J. Mol. Biol.* **2006**, *359*, 690–707.
40. Chen, W.-B.; Nie, Y.; Xu, Y. Signal peptide-independent secretory expression and characterization of pullulanase from a newly isolated *Klebsiella variicola* SHN-1 in *Escherichia coli*. *Appl. Biochem. Biotechnol.* **2013**, *169*, 41–54.
41. Odibo, F.J.C.; Obi, S.K.C. Purification and characterization of a thermostable pullulanase from *Thermoactinomyces thalpophilus*. *J. Ind. Microbiol.* **1988**, *3*, 343–350.
42. Waśko, A.; Polak-Berecka, M.; Targoński, Z. Purification and characterization of pullulanase from *Lactococcus lactis*. *Prep. Biochem. Biotechnol.* **2011**, *41*, 252–261.
43. Møller, M.S.; Goh, Y.J.; Rasmussen, K.B.; Cypryk, W.; Celebioglu, H.U.; Klaenhammer, T.R.; Svensson, B.; Hachem, M.A. An extracellular cell-attached pullulanase confers branched  $\alpha$ -glucan utilization in human gut *Lactobacillus acidophilus*. *Appl. Environ. Microbiol.* **2017**, *83*, e00402-00417.
44. Kimura, T.; Horikoshi, K. Characterization of pullulan-hydrolysing enzyme from an alkalopsychrotrophic *Micrococcus* sp. *Appl. Microbiol. Biotechnol.* **1990**, *34*, 52–56.

45. Thakur, M.; Sharma, N.; Rai, A.K.; Singh, S.P. A novel cold-active type I pullulanase from a hot-spring metagenome for effective debranching and production of resistant starch. *Bioresour. Technol.* **2021**, *320*, 124288.
46. Motahar, S.F.S.; Salami, M.; Ariaeenejad, S.; Emam-Djomeh, Z.; Mamaghani, A.S.A.; Kavousi, K.; Moghadam, M.; Salekdeh, G.H. Synergistic effect of metagenome-derived starch-degrading enzymes on quality of functional bread with antioxidant activity. *Starch/Stärke* **2021**, *2100098*.
47. Jaslionis, A.; Petkauskaitė, R.; Kuisiene, N. A novel type I thermostable pullulanase isolated from a thermophilic starch enrichment culture. *Microbiology* **2014**, *83*, 227–234.
48. Liu, J.; Liu, Y.; Yan, F.; Jiang, Z.; Yang, S.; Yan, Q. Gene cloning, functional expression and characterisation of a novel type I pullulanase from *Paenibacillus barengoltzii* and its application in resistant starch production. *Protein Expr. Purif.* **2016**, *121*, 22–30.
49. Chen, S.-Q.; Cai, X.-H.; Xie, J.-L.; Wei, W.; Wei, D.-Z. Structural and biochemical properties of a novel pullulanase of *Paenibacillus lautus* DSM 3035. *Starch/Stärke* **2017**, *69*, 1500333.
50. Wei, W.; Ma, J.; Chen, S.-Q.; Cai, X.-H.; Wei, D.-Z. A novel cold-adapted type I pullulanase of *Paenibacillus polymyxa* Nws-pp2: In vivo functional expression and biochemical characterization of glucans hydrolyzates analysis. *BMC Biotechnol.* **2015**, *15*, 96.
51. Kriegshäuser, G.; Liebl, W. Pullulanase from the hyperthermophilic bacterium *Thermotoga maritima*: purification by  $\beta$ -cyclodextrin affinity chromatography. *J. Chromatogr. B Biomed. Appl.* **2000**, *737*, 245–251.
52. Kang, J.; Park, K.-M.; Choi, K.-H.; Park, C.-S.; Kim, G.-E.; Kim, D.; Cha, J. Molecular cloning and biochemical characterization of a heat-stable type I pullulanase from *Thermotoga neapolitana*. *Enzyme Microb. Technol.* **2011**, *48*, 260–266.
53. Plant, A.R.; Morgan, H.W.; Daniel, R.M. A highly stable pullulanase from *Thermus aquaticus* YT-1. *Enzyme Microb. Technol.* **1986**, *8*, 668–672.
54. Kim, C.-H.; Nashiru, O.; Ko, J.H. Purification and biochemical characterization of pullulanase type I from *Thermus caldophilus* GK-24. *FEMS Microbiol. Lett.* **1996**, *138*, 147–152.
55. Tomiyasu, K.; Yato, K.; Yasuda, M.; Tonozuka, T.; Ibuka, A.; Sakai, H. Cloning and nucleotide sequence of the pullulanase gene of *Thermus thermophilus* HB8 and production of the enzyme in *Escherichia coli*. *Biosci. Biotechnol. Biochem.* **2001**, *65*, 2090–2094.
56. Elleuche, S.; Krull, A.; Lorenz, U.; Antranikian, G. Parallel N- and C-terminal truncations facilitate purification and analysis of a 155-kDa cold-adapted type-I pullulanase. *Protein J.* **2017**, *36*, 56–63.
57. Qoura, F.; Elleuche, S.; Brueck, T.; Antranikian, G. Purification and characterization of a cold-adapted pullulanase from a psychrophilic bacterial isolate. *Extremophiles* **2014**, *18*, 1095–1102.
58. Mesbah, N.M.; Wiegel, J. Biochemical characterization of halophilic, alkalithermophilic amylopullulanase PulD7 and truncated amylopullulanases PulD7 $\Delta$ N and PulD7 $\Delta$ C. *Int. J. Biol. Macromol.* **2018**, *111*, 632–638.
59. Kahar, U.M.; Chan, K.-G.; Salleh, M.M.; Hii, S.M.; Goh, K.M. A high molecular-mass *Anoxybacillus* sp. SK3-4 amylopullulanase: characterization and its relationship in carbohydrate utilization. *Int. J. Mol. Sci.* **2013**, *14*, 11302–11318.
60. Wang, J.; Liu, Z.; Zhou, Z. Cloning and characterization of a novel thermophilic amylopullulanase with a type I pullulanase structure from *Anoxybacillus* sp. WB42. *Starch/Stärke* **2018**, *70*, 1700265.
61. Ling, H.S.; Ling, T.C.; Mohamad, R.; Ariff, A.B. Characterization of pullulanase type II from *Bacillus cereus* H1.5. *Am. J. Biochem. Biotechnol.* **2009**, *5*, 170–179.
62. Sata, H.; Umeda, M.; Kim, C.-H.; Taniguchi, H.; Maruyama, Y. Amylase-pullulanase enzyme produced by *B. circulans* F-2. *Biochim. Biophys. Acta* **1989**, *991*, 388–394.
63. Liu, X.; Chen, H.; Tao, H.-y.; Chen, Z.; Liang, X.-b.; Han, P.; Tao, J.-h. Cloning and characterization of a novel amylopullulanase from *Bacillus megaterium* Y103 with transglycosylation activity. *Biotechnol. Lett.* **2020**, *42*, 1719–1726.
64. Brunswick, J.M.; Kelly, C.T.; Fogarty, W.M. The amylopullulanase of *Bacillus* sp. DSM 405. *Appl. Microbiol. Biotechnol.* **1999**, *51*, 170–175.
65. Lee, S.-P.; Morikawa, M.; Takagi, M.; Imanaka, T. Cloning of the aapT gene and characterization of its product,  $\alpha$ -amylase-pullulanase (AapT), from thermophilic and alkaliphilic *Bacillus* sp. strain XAL601. *Appl. Environ. Microbiol.* **1994**, *60*, 3764–3773.
66. Li, X.; Ji, H.; Zhai, Y.; Bai, Y.; Jin, Z. Characterizing a thermostable amylopullulanase from *Caldisericum exile* with wide pH adaptation and broad substrate specificity. *Food Biosci.* **2021**, *41*, 100952.
67. Roodi, F.Z.; Aminzadeh, S.; Farrokhi, N.; Karkhane, A.; Haghbeen, K. *Cohnella* amylopullulanases: biochemical characterization of two recombinant thermophilic enzymes. *PLoS ONE* **2017**, *12*, e0175013.

68. Li, X.; Li, D. Preparation of linear maltodextrins using a hyperthermophilic amylopullulanase with cyclodextrin- and starch-hydrolysing activities. *Carbohydr. Polym.* **2015**, *119*, 134–141.
69. Spreinat, A.; Antranikian, G. Purification and properties of a thermostable pullulanase from *Clostridium thermosulfurogenes* EM1 which hydrolyses both  $\alpha$ -1,6 and  $\alpha$ -1,4-glycosidic linkages. *Appl. Microbiol. Biotechnol.* **1990**, *33*, 511–518.
70. Soma, M.; Reddy, G.; Gunda, S. Purification and characterization of highly thermostable amylopullulanase from a thermophilic, anaerobic bacterium *Clostridium thermosulfurogenes* SVM17. *Malays. J. Microbiol.* **2011**, *7*, 97–106.
71. Duffner, F.; Bertoldo, C.; Andersen, J.T.; Wagner, K.; Antranikian, G. A new thermoactive pullulanase from *Desulfurococcus mucosus*: cloning, sequencing, purification, and characterization of the recombinant enzyme after expression in *Bacillus subtilis*. *J. Bacteriol.* **2000**, *182*, 6331–6338.
72. Ferner-Ortner-Bleckmann, J.; Huber-Gries, C.; Pavkov, T.; Keller, W.; Mader, C.; Ilk, N.; Sleytr, U.B.; Egelseer, E.M. The high-molecular-mass amylase (HMMA) of *Geobacillus stearothermophilus* ATCC 12980 interacts with the cell wall components by virtue of three specific binding regions. *Mol. Microbiol.* **2009**, *1448*–1461.
73. Kambourova, M.S.; Emanuilova, E.I. Purification and general biochemical properties of thermostable pullulanase from *Bacillus stearothermophilus* G-82. *Appl. Biochem. Biotechnol.* **1992**, *33*, 193–203.
74. Chen, J.-T.; Chen, M.-C.; Chen, L.-L.; Chu, W.-S. Structure and expression of an amylopullulanase gene from *Bacillus stearothermophilus* TS-23. *Biotechnol. Appl. Biochem.* **2001**, *33*, 189–199.
75. Arabaci, N.; Arikan, B. An amylopullulanase (ApuNP1) from *Geobacillus thermoleovorans* NP1: Biochemical characterization and its potential industrial applications. *Prep. Biochem. Biotechnol.* **2019**, *49*, 127–135.
76. Nisha, M.; Satyanarayana, T. Characterization of recombinant amylopullulanase (gt-apu) and truncated amylopullulanase (gt-apuT) of the extreme thermophile *Geobacillus thermoleovorans* NP33 and their action in starch saccharification. *Appl. Microbiol. Biotechnol.* **2013**, *97*, 6279–6292.
77. Satyanarayana, T.; Noorwez, S.M.; Kumar, S.; Rao, J.L.U.M.; Ezhilvannan, M.; Kaur, P. Development of an ideal starch saccharification process using amylolytic enzymes from thermophiles. *Biochem. Soc. Trans.* **2004**, *32*, 276–278.
78. Nisha, M.; Satyanarayana, T. Characterization and multiple applications of a highly thermostable and  $\text{Ca}^{2+}$ -independent amylopullulanase of the extreme thermophile *Geobacillus thermoleovorans*. *Appl. Biochem. Biotechnol.* **2014**, *174*, 2594–2615.
79. Zareian, S.; Khajeh, K.; Ranjbar, B.; Dabirmanesh, B.; Ghollasi, M.; Mollania, N. Purification and characterization of a novel amylopullulanase that converts pullulan to glucose, maltose, and maltotriose and starch to glucose and maltose. *Enzyme Microb. Technol.* **2010**, *46*, 57–63.
80. Siroosi, M.; Amoozegar, M.A.; Khajeh, K.; Fazeli, M.; Rezaei, M.H. Purification and characterization of a novel extracellular halophilic and organic solvent-tolerant amylopullulanase from the haloarchaeon, *Halorubrum* sp. strain Ha25. *Extremophiles* **2014**, *18*, 25–33.
81. Vishnu, C.; Naveena, B.J.; Altaf, M.; Venkateshwar, M.; Reddy, G. Amylopullulanase - a novel enzyme of *L. amylophilus* GV6 indirect fermentation of starch to L(+) lactic acid. *Enzyme Microb. Technol.* **2006**, *38*, 545–550.
82. Lee, Y.-S.; Seo, S.-H.; Yoon, S.-H.; Kim, S.-Y.; Hahn, B.-S.; Sim, J.-S.; Koo, B.-S.; Lee, C.-M. Identification of a novel alkaline amylopullulanase from a gut metagenome of *Hermetia illucens*. *Int. J. Biol. Macromol.* **2016**, *82*, 514–521.
83. Su, H.-y.; Cui, T.-b. Gene cloning, expression and characterization of a pullulanase from *Paenibacillus puldeungensis* LK18 Strain. *Mod. Food Sci. Technol.* **2019**, *35*, 107–113.
84. Dong, G.; Viele, C.; Zeikus, J.G. Cloning, sequencing, and expression of the gene encoding amylopullulanase from *Pyrococcus furiosus* and biochemical characterization of the recombinant enzyme. *Appl. Environ. Microbiol.* **1997**, *63*, 3577–3584.
85. Habib-ur-Rehman; Siddiqui, M.A.; Qayyum, A.; Bano, A.; Rashid, a.N. Gene expression in *Escherichia coli* and purification of recombinant type II pullulanase from a hyperthermophilic archaeon, *Pyrobaculum calidifontis*. *Pakistan J. Zool.* **2018**, *50*, 1381–1386.
86. Brown, S.H.; Kelly, R.M. Characterization of amylolytic enzymes, having both  $\alpha$ -1,4 and  $\alpha$ -1,6 hydrolytic activity, from the thermophilic archaea *Pyrococcus furiosus* and *Thermococcus litoralis*. *Appl. Environ. Microbiol.* **1993**, *59*, 2614–2621.
87. Rüdiger, A.; Jorgensen, P.L.; Antranikian, G. Isolation and characterization of a heat-stable pullulanase from the hyperthermophilic archaeon *Pyrococcus woesei* after cloning and expression of its gene in *Escherichia coli*. *Appl. Environ. Microbiol.* **1995**, *61*, 567–575.

88. Pang, B.; Zhou, L.; Cui, W.; Liu, Z.; Zhou, S.; Xu, J.; Zhou, Z. A hyperthermostable type II pullulanase from a deep-sea microorganism *Pyrococcus yayanosii* CH1. *J. Agric. Food Chem.* **2019**, *34*, 9611–9617.
89. Li, X.; Li, D.; Park, K.-H. An extremely thermostable amylopullulanase from *Staphylothermus marinus* displays both pullulan- and cyclodextrin-degrading activities. *Appl. Microbiol. Biotechnol.* **2013**, *97*, 5359–5369.
90. Kar, S.; Ray, R.C.; Mohapatra, U.B. Purification, characterization and application of thermostable amylopullulanase from *Streptomyces erumpens* MTCC 7317 under submerged fermentation. *Ann. Microbiol.* **2012**, *62*, 931–937.
91. Choi, K.H.; Cha, J. Membrane-bound amylopullulanase is essential for starch metabolism of *Sulfolobus acidocaldarius* DSM639. *Extremophiles* **2015**, *19*, 909–920.
92. Saha, B.C.; Mathupala, S.P.; Zeikus, J.G. Purification and characterization of a highly thermostable novel pullulanase from *Clostridium thermohydrosulfuricum*. *Biochem. J.* **1988**, *252*, 343–348.
93. Mathupala, S.P.; Zeikus, J.G. Improved purification and biochemical characterization of extracellular amylopullulanase from *Thermoanaerobacter ethanolicus* 39E. *Appl. Microbiol. Biotechnol.* **1993**, *39*, 487–493.
94. Mathupala, S.P.; Lowe, S.E.; Podkowyrov, S.M.; Zeikus, J.G. Sequencing of the amylopullulanase (apu) gene of *Thermoanaerobacter ethanolicus* 39E, and identification of the active site by site-directed mutagenesis. *J. Biol. Chem.* **1993**, *268*, 16332–16344.
95. Melasniemi, H.; Paloheimo, M. Cloning and expression of the *Clostridium thermohydrosulfuricum*  $\alpha$ -amylase-pullulanase gene in *Escherichia coli*. *J. Gen. Microbiol.* **1989**, *135*, 1755–1762.
96. Saha, B.C.; Lamed, R.; Lee, C.-Y.; P.Mathupala, S.; Zeikus, J.G. Characterization of an endo-acting amylopullulanase from *Thermoanaerobacter* strain B6A. *Appl. Environ. Microbiol.* **1990**, *56*, 881–886.
97. Coleman, R.D.; Shio-Shong Yang; McAlister, M.P. Cloning of the debranching-enzyme gene from *Thermoanaerobium brockii* into *Escherichia coli* and *Bacillus subtilis*. *J. Bacteriol.* **1987**, *169*, 4302–4307.
98. Ramesh, M.V.; Podkowyrov, S.M.; Lowe, S.E.; Zeikus, J.G. Cloning and sequencing of the *Thermoanaerobacterium saccharolyticum* B6A-RI apu gene and purification and characterization of the amylopullulanase from *Escherichia coli*. *Appl. Environ. Microbiol.* **1994**, *60*, 94–101.
99. Lin, F.-P.; Ma, H.-Y.; Lin, H.-J.; Liu, S.-M.; Tzou, W.-S. Biochemical characterization of two truncated forms of amylopullulanase from *Thermoanaerobacterium saccharolyticum* NTOU1 to identify its enzymatically active region. *Appl. Biochem. Biotechnol.* **2011**, *165*, 1047–1056.
100. Ganghofner, D.; Kellermann, J.; Staudenbauer, W.L.; Bronnenmeier, K. Purification and properties of an amylopullulanase, a glucoamylase, and an  $\alpha$ -glucosidase in the amyloytic enzyme system of *Thermoanaerobacterium thermosaccharolyticum*. *Biosci. Biotechnol. Biochem.* **1998**, *62*, 302–308.
101. Spreinat, A.; Antranikian, G. Analysis of the amyloytic enzyme system of *Clostridium thermosulfurogenes* EM1: purification and synergistic action of pullulanases and maltohexaose forming  $\alpha$ -amylase. *Starch/Stärke* **1992**, *44*, 305–312.
102. Burchhardt, G.; Wienecke, A.; Bahl, H. Isolation of the pullulanase gene from *Clostridium thermosulfurogenes* (DSM 3896) and its expression in *Escherichia coli*. *Curr. Microbiol.* **1991**, *22*, 91–95.
103. Matuschek, M.; Burchhardt, G.; Sahm, K.; Bahl, H. Pullulanase of *Thermoanaerobacterium thermosulfurogenes* EM1 (*Clostridium thermosulfurogenes*): molecular analysis of the gene, composite structure of the enzyme, and a common model for its attachment to the cell surface. *J. Bacteriol.* **1994**, *176*, 3295–3302.
104. Plant, A.R.; Clemens, R.M.; Daniel, R.M.; Morgan, H.W. Purification and preliminary characterization of an extracellular pullulanase from *Thermoanaerobium Tok6-B1*. *Appl. Microbiol. Biotechnol.* **1987**, *26*, 427–433.
105. Erra-Pujada, M.; Chang-Pi-Hin, F.; Debeire, P.; Duchiron, F.; O'Donohue, M.J. Purification and properties of the catalytic domain of the thermostable pullulanase type II from *Thermococcus hydrothermalis*. *Biotechnol. Lett.* **2001**, *23*, 1273–1277.
106. Gantelet, H.; Duchiron, F. Purification and properties of a thermoactive and thermostable pullulanase from *Thermococcus hydrothermalis*, a hyperthermophilic archaeon isolated from a deep-sea hydrothermal vent. *Appl. Microbiol. Biotechnol.* **1998**, *49*, 770–777.
107. Han, T.; Zeng, F.; Li, Z.; Liu, L.; Wei, M.; Guan, Q.; Liang, X.; Peng, Z.; Liu, M.; Qin, J., et al. Biochemical characterization of a recombinant pullulanase from *Thermococcus kodakarensis* KOD1. *Lett. Appl. Microbiol.* **2013**, *57*, 336–343.
108. Guan, Q.; Guo, X.; Han, T.; Wei, M.; Jin, M.; Zeng, F.; Liu, L.; Li, Z.; Wang, Y.; Cheong, G.-W., et al. Cloning, purification and biochemical characterisation of an organic solvent-, detergent-, and thermostable amylopullulanase from *Thermococcus kodakarensis* KOD1. *Process Biochem.* **2013**, *48*, 878–884.

109. Jiao, Y.-L.; Wang, S.-J.; Lv, M.-S.; Xu, J.-L.; Fang, Y.-W.; Liu, S. A GH57 family amylopullulanase from deep-sea *Thermococcus siculi*: expression of the gene and characterization of the recombinant enzyme. *Curr. Microbiol.* **2011**, *62*, 222–228.
110. Li, X.; Zhao, J.; Fu, J.; Pan, Y.; Li, D. Sequence analysis and biochemical properties of an acidophilic and hyperthermophilic amylopullulanase from *Thermoflum pendens*. *Int. J. Biol. Macromol.* **2018**, *114*, 235–243.
111. Wu, H.; Yu, X.; Chen, L.; Wu, G. Cloning, overexpression and characterization of a thermostable pullulanase from *Thermus thermophilus* HB27. *Protein Expr. Purif.* **2014**, *95*, 22–27.
112. Tang, K.; Kobayashi, R.S.; Champreda, V.; Eurwilaichitr, L.; Tanapongpipat, S. Isolation and characterization of a novel thermostable neopullulanase-like enzyme from a hot spring in Thailand. *Biosci. Biotechnol. Biochem.* **2008**, *72*, 1448–1456.
113. Huang, H.; Lin, Y.; Wang, G.; Lin, J. Gene cloning, expression and biochemical characterization of a new multidomain, halotolerant and SDS-resistant alkaline pullulanase from *Alkalibacterium* sp. SL3. *Process Biochem.* **2020**, *96*, 1–10.
114. Ara, K.; Saeki, K.; Igarashi, K.; Takaiwa, M.; Uemura, T.; Hagihara, H.; Kawai, S.; Ito, S. Purification and characterization of an alkaline amylopullulanase with both  $\alpha$ -1,4 and  $\alpha$ -1,6 hydrolytic activity from alkalophilic *Bacillus* sp. KSM-1378. *Biochim. Biophys. Acta* **1995**, *1243*, 315–324.
115. Motherway, M.O.C.; Fitzgerald, G.F.; Neirynck, S.; Ryan, S.; Steidler, L.; Van Sinderen, D. Characterization of ApuB, an extracellular type II amylopullulanase from *Bifidobacterium breve* UCC2003. *Appl. Environ. Microbiol.* **2008**, *74*, 6271–6279.
116. Kim, S.-Y.; Kim, H.; Kim, Y.-J.; Jung, D.-H.; Seo, D.-H.; Jung, J.-H.; Park, C.-S. Enzymatic analysis of truncation mutants of a type II pullulanase from *Bifidobacterium adolescentis* P2P3, a resistant starch-degrading gut bacterium. *Int. J. Biol. Macromol.* **2021**, In-press, <https://doi.org/10.1016/j.ijbiomac.2021.1010.1193>.
117. Kim, J.-H.; Sunako, M.; Ono, H.; Murooka, Y.; Fukusaki, E.; Yamashita, M. Characterization of gene encoding amylopullulanase from plant-originated lactic acid bacterium, *Lactobacillus plantarum* L137. *J. Biosci. Bioeng.* **2008**, *106*, 449–459.
118. Ferrando, M.L.; Fuentes, S.; Greeff, A.d.; Smith, H.; Wells, J.M. ApuA, a multifunctional  $\alpha$ -glucan-degrading enzyme of *Streptococcus suis*, mediates adhesion to porcine epithelium and mucus. *Microbiology* **2010**, *156*, 2818–2828.
119. Nordberg Karlsson, E.; Labes, A.; Turner, P.; Fridjonsson, O.H.; Wennerberg, C.; Pozzo, T.; Hreggvidson, G.O.; Kristjansson, J.K.; Schönheit, P. Differences and similarities in enzymes from the neopullulanase subfamily isolated from thermophilic species. *Biologia* **2008**, *63*, 1006–1014.
120. Matzke, J.; Herrmann, A.; Schneider, E.; Bakker, E.P. Gene cloning, nucleotide sequence and biochemical properties of a cytoplasmic cyclomaltodextrinase (neopullulanase) from *Alicyclobacillus acidocaldarius*, reclassification of a group of enzymes. *FEMS Microbiol. Lett.* **2000**, *183*, 55–61.
121. Yebra, M.J.; Arroyo, J.; Sanz, P.; Prieto, J.A. Characterization of novel neopullulanase from *Bacillus polymyxa*. *Appl. Biochem. Biotechnol.* **1997**, *68*, 113–120.
122. Yebra, M.J.; Blasco, A.; Sanz, P. Expression and secretion of *Bacillus polymyxa* neopullulanase in *Saccharomyces cerevisiae*. *FEMS Microbiol. Lett.* **1999**, *170*, 41–49.
123. Igarashi, K.; Ara, K.; Saeki, K.; Ozaki, K.; Kawai, S.; Ito, S. Nucleotide sequence of the gene that encodes a neopullulanase from an alkalophilic *Bacillus*. *Biosci. Biotechnol. Biochem.* **1992**, *56*, 514–516.
124. Smith, K.A.; Salyers, A.A. Characterization of a neopullulanase and an  $\alpha$ -glucosidase from *Bacteroides thetaiotomicron* 95-1. *J. Bacteriol.* **1991**, *173*, 2962–2968.
125. Kuriki, T.; Okada, S.; Imanaka, T. New type of pullulanase from *Bacillus stearothermophilus* and molecular cloning and expression of the gene in *Bacillus subtilis*. *J. Bacteriol.* **1988**, *170*, 1554–1559.
126. Kamasaka, H.; Sugimoto, K.; Takata, H.; Nishimura, T.; Kuriki, T. *Bacillus stearothermophilus* neopullulanase selective hydrolysis of amylose to maltose in the presence of amylopectin. *Appl. Environ. Microbiol.* **2002**, *68*, 1658–1664.
127. Park, K.; Yoon, J.; Kim, T.; Cheong, K.A.; Park, C.; Lee, T.; Kim, Y. Catalytic activities of intracellular dimeric neopullulanase on cyclodextrin, acarbose and maltose. *Biotechnol. Appl. Biochem.* **2002**, *35*, 27–34, doi:10.1042/BA20010052.
128. Balolong, M.P.; Chae, J.P.; Kang, D.-K. Expression and characterisation of neopullulanase from *Lactobacillus mucosae*. *Biotechnol. Lett.* **2016**, *38*, 1753–1760.
129. Rajdevi, K.P.; Yogeeswaran, G. Cooperativity and substrate specificity of an alkaline amylase and neopullulanase complex of *Micrococcus halobius* OR-1. *Appl. Biochem. Biotechnol.* **2001**, *90*, 233–249.

130. Kim, H.J.; Park, J.N.; Kim, H.O.; Shin, D.J.; Chin, J.E.; Lee, H.B.; Chun, S.B.; Bai, S. Cloning and expression of a *Paenibacillus* sp. neopullulanase gene in *Saccharomyces cerevisiae* producing *Schwanniomyces occidentalis* glucoamylase. *J. Microbiol. Biotechnol.* **2002**, *12*, 340–344.
131. Sakano, Y.; Hiraiwa, S.I.; Fukushima, J.; Kobayashi, T. Enzymatic properties and action patterns of *Thermoactinomyces vulgaris*  $\alpha$ -amylase. *Agr. Biol. Chem.* **1982**, *46*, 1121–1129.
132. Shimizu, M.; Kanno, M.; Tamura, M.; Suekane, M. Purification and some properties of a novel  $\alpha$ -amylase produced by a strain of *Thermoactinomyces vulgaris*. *Agr. Biol. Chem.* **1978**, *42*, 1681–1688.
133. Tonozuka, T.; Ohtsuka, M.; Sakano, Y.; Mogi, S.I.; Sakai, H.; Ohta, T. A neopullulanase-type  $\alpha$ -amylase gene from *Thermoactinomyces vulgaris* R-47. *Biosci. Biotechnol. Biochem.* **1993**, *57*, 395–401.
134. Aoki, H.; Sakano, Y.Y. Molecular cloning and heterologous expression of the isopullulanase gene from *Aspergillus niger* A.T.C.C. 9642. *Biochem. J.* **1997**, *323*, 757–764.
135. Mizuno, M.; Koide, A.; Yamamura, A.; Akeboshi, H.; Yoshida, H.; Kamitori, S.; Sakano, Y.; Nishikawa, A.; Tonozuka, T. Crystal structure of *Aspergillus niger* isopullulanase, a member of glycoside hydrolase family 49. *J. Mol. Biol.* **2008**, *376*, 210–220.
136. Aoki, H.; Yopi; Padmajanti, A.; Sakano, Y. Two components of cell-bound isopullulanase from *Aspergillus niger* ATCC 9642-their purification and enzymatic properties. *Biosci. Biotechnol. Biochem.* **1996**, *60*, 1795–1798.
137. Roy, A.; Messaoud, E.B.; Bejar, S. Isolation and purification of an acidic pullulanase type II from newly isolated *Bacillus* sp. US149. *Enzyme Microb. Technol.* **2003**, *33*, 720–724.
138. Niehaus, F.; Peters, A.; Groudieva, T.; Antranikian, G. Cloning, expression and biochemical characterisation of a unique thermostable pullulan-hydrolysing enzyme from the hyperthermophilic archaeon *Thermococcus aggregans*. *FEMS Microbiol. Lett.* **2000**, *190*, 223–229.
139. Ahmad, N.; Rashid, N.; Haider, M.S.; Akram, M.; Akhtar, M. Novel maltotriose-hydrolyzing thermoacidophilic type III pullulan hydrolase from *Thermococcus kodakarensis*. *Appl. Environ. Microbiol.* **2014**, *80*, 1108–1115.