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Acid Hydrolysis of Lignocellulosic Biomass: Sugars and Furfurals Formation

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Abstract: Hydrolysis of lignocellulosic biomass is a crucial step for the production of sugars and biobased platform chemicals. Pretreatment experiments in a semi-continuous plant with diluted sulphuric acid as catalyst were carried out to measure the time-dependent formation of sugars (glucose, xylose, mannose), furfurals, and organic acids (acetic, formic, and levulinic acid) at different hydrolysis temperatures (180, 200, 220 °C) of one representative of each basic type of lignocellulose: hardwood, softwood, and grass. The addition of the acid catalyst is followed by a sharp increase in the sugar concentration. Xylose and mannose were mainly formed in the initial stages of the process, while glucose was released slowly. Increasing the reaction temperature had a positive effect on the formation of furfurals and organic acids, especially on hydroxymethylfurfural (HMF) and levulinic acid, regardless of biomass type. In addition, large amounts of formic acid were released during the hydrolysis of miscanthus grass. Structural changes in the solid residue show a complete hydrolysis of hemicellulose at 180 °C and of cellulose at 200 °C after around 120 min reaction time. The results obtained in this study can be used for the optimisation of the hydrolysis conditions and reactor design to maximise the yields of desired products, which might be sugars or furfurals.

Keywords: spruce wood; beech wood; miscanthus; pretreatment; hydroxymethylfurfural; HMF; furfural; hydrolysate; glucose; xylose

1. Introduction

Renewable lignocellulosic feedstocks have potential for large-scale production of biofuels and chemicals in second generation biorefineries with sugars from cellulose and hemicellulose as intermediates [1]. Lignocellulosic biomass is composed mainly of cellulose, hemicellulose, and lignin, as well as some minor compounds like inorganics and extractives [2,3]. The fractionation at rather mild conditions is applied to get sugars to feed microorganism. Fermentable sugars are mainly used for bioethanol production, where the most important intermediate is glucose. Here, mild reaction conditions are employed to minimise the formation of fermentation inhibitors. This sugar can also be used for platform chemical production, for example, hydroxymethylfurfural (HMF) [4], by chemical pathways. Modification of the existing hydrolysis process would directly provide platform chemicals like HMF, furfural, and levulinic acid as primary products [5]. In this case, more severe reaction conditions can be applied, because the fact that these compounds may inhibit yeasts is of no importance. Additional research is needed to establish the conditions required for their efficient production [6,7].

The composition of lignocellulosic biomass varies in softwood, hardwood, and grasses. One representative of each group is investigated in this study, including spruce wood, beech wood, and *Miscanthus x giganteus*. *Miscanthus* is an interesting candidate for a key energy crop, because it grows rapidly and produces an annual crop without the need for replanting with very low input requirements. Reported miscanthus harvestable yield in Europe is in the range of 5 to 44 t ha⁻¹ (dry matter). However, that yield depends on the genotype, location, and harvest time [8]. It is characterised by a high energy content and low moisture content after harvest; therefore, miscanthus is expected to play an essential role in the sustainable production of chemicals in biorefineries via thermochemical conversion [9,10].

Both beech and spruce trees are some of the major forest trees in Europe and are available year-round. A representative of European softwood species is spruce wood. It produces high-quality timber, very important for the pulp and paper industry [11]. These biomasses differ from each other in the proportions of cellulose, hemicellulose, and lignin, as well as in the particular composition of the hemicellulose and lignin [12,13]. The typical composition of carbohydrate fraction is presented in Table 1. As can be seen, the dominant sugar in beech wood hemicellulose (hardwood) and miscanthus (grass) is xylose, while spruce wood (softwood) contained more mannose than other biomasses.

Table 1. Comparison of the cell composition of beech and spruce wood, and *Miscanthus x giganteus*.

	Beech Wood ¹	Spruce Wood ²	<i>Miscanthus x giganteus</i> ³
	(wt%)		
Cellulose	36.30	44.00	38.20
Hemicellulose	34.74	23.88	24.30
Arabinose	0.72	1.48	1.80
Xylose	25.80	5.98	19.00
Galactose	1.05	2.46	0.40
Glucose	1.91	3.06	0.27
Mannose	1.60	8.62	0.10
Rhamnose	0.63	0.26	0.00

¹ [14], ² [15], ³ [16,17].

The key for maximising the value of lignocellulosic biomass, owing to its structural recalcitrance, is the efficient and cost-effective transformation of these three polymeric constituents, that is, cellulose, hemicellulose, and lignin. So, the central unit operation in a lignocellulosic biorefinery is biomass fractionation whereby, for example, soluble sugars and other valuable products streams are obtained [5,18]. Various pretreatment methods have different effects on the biomass structure. In some methods, cellulose might be removed from the biomass structure, which is particularly observed in hydrothermal and acidic pretreatment methods. Meanwhile, the second type of method consists of removing of lignin, especially at alkaline conditions [5]. One of the most cost-effective methods for the recovery of sugars from lignocellulosic biomass is acid-catalysed hydrothermal fractionation. Often, sulphuric acid is used for this purpose. It is a relatively inexpensive and strong acid, and thus makes cost-intensive acid recovery unnecessary. An alternative is hydrochloric acid, which is also a low cost, strong acid and can be recovered by distillation. However, hydrochloric acid is highly corrosive, which increases the costs for the construction materials of equipment [19,20].

Hemicellulose and cellulose are broken down via hydrolysis during dilute acid pretreatment and soluble sugars are obtained. In general, the hydrolysis of hemicellulose produces sugars (glucose, xylose, mannose, and galactose) and organic acids (formic, acetic acid). Xylose is a pentose sugar, which may further be dehydrated to furfural [21,22]. In contrast to hemicellulose, cellulose has higher stability, owing to a high degree of polymerisation, its crystallinity, and the presence of hydrogen bonds. This prevents the onset of cellulose hydrolysis in milder conditions [23]. Under acidic conditions, cellulose can be hydrolysed to glucose, and then glucose can be subsequently transformed to value-added chemicals such as HMF [24–27]. During acid-catalysed pretreatment, only low amounts

of lignin are removed in forms of soluble fragments. Lignin is a complex aromatic polymer with a very random structure. It is mainly composed of phenylpropanoid units. A lot of research is currently underway to obtain valuable chemicals, for example, phenols, from lignin [28–30].

Lignocellulosic hydrolysis using dilute acid solutions has been intensively studied and is reviewed by Steinbach et al. [5] and Kang et al. [31]. Furthermore, hydrothermal treatment of beech wood was presented in the work of Nitsos et al. [32], spruce wood by Kačíková et al. [33], and *Miscanthus x giganteus* by Zhou et al. [34]. Most experimental studies were performed in batch systems [19,35,36], where intermediate compounds like sugars underwent considerable consecutive reactions. Using a semi-continuous reaction setup, the formation of intermediates can be examined in more detail, and so more insights into the hydrolysis process can be obtained [37]. Conversion of biomass in a semi-continuous reaction setup was performed with woody biomass [38] and rye straw [39]. Further, studies in a continuous reactor on the hydrolysis of seaweed and corn stover were performed by Park et al. [40] and Schell et al. [41], respectively. The results obtained strongly depend on the feedstock and operational conditions employed, like reaction time or temperature.

The selection of optimum pretreatment conditions for lignocellulose is crucial for maximising the yields of sugars and/or furfurals in the product fluid. Furthermore, for the industrial implementation of a process, not only the maximisation of the yields, but also the minimisation of the production costs is essential. So, an optimum has to be determined between high yields at elevated reaction conditions and, on the other side, simple processes at a short reaction time, which leads to small equipment size and investment.

Pretreatment experiments with diluted sulphuric acid were carried out for a better understanding of lignocellulosic biomass fractionation. In this study, three different feedstocks, beech wood chips, spruce wood chips, and *Miscanthus x giganteus*, were used. The experiments were performed in a semi-continuous mini plant, where water-soluble compounds, such as sugar monomers or furfurals, were quickly removed from the hot reaction zone, and thus protected as far as possible from secondary reactions. By the continuous removal of the liquid phase, the time dependence of formation of soluble organic compounds (sugars, furfurals, and organic acids) was determined. For each biomass, the influence of hydrolysis temperature on the formation of these compounds was investigated. Additionally, the yields of the main components like sugars, furfurals, and organic acids are determined for different treatment durations. In order to fully determine the effect of pretreatment conditions on the resulting products, structural changes in the solid residue were also examined.

2. Results

2.1. Influence of Temperature on Beech Wood Hydrolysis

Beech wood chips were pretreated in the presence of 0.05 mol L^{-1} sulphuric acid. A volume flow of 15 mL min^{-1} of the acid was kept constant during the experiment. The reaction temperature was varied in the fixed bed reactor and was $180 \text{ }^\circ\text{C}$, $200 \text{ }^\circ\text{C}$, or $220 \text{ }^\circ\text{C}$. Time curves of the formation of the main hydrolysis products are shown in Figures 1 and 2. The negative values of time represent the preheating period with a flow of demineralised water. The time $t = 0 \text{ min}$ corresponds to switching from demineralised water to acid solution in the feed stream.

During the preheating time with demineralised water to $180 \text{ }^\circ\text{C}$, mainly xylose was detected; see Figure 1A. The production of xylose grows rapidly, after changing from water to the acid solution. At the same point, the formation of glucose and mannose was observed. The formation of xylose reached a maximum between 10 and 15 minutes' duration. A longer reaction duration caused a substantial decrease in the concentration. The production of glucose largely increased after 10 min and remained at a steady level until the end of the measurement. The concentration of mannose was almost constant during hydrolysis and an order of magnitude lower than the concentration of other measured sugars.

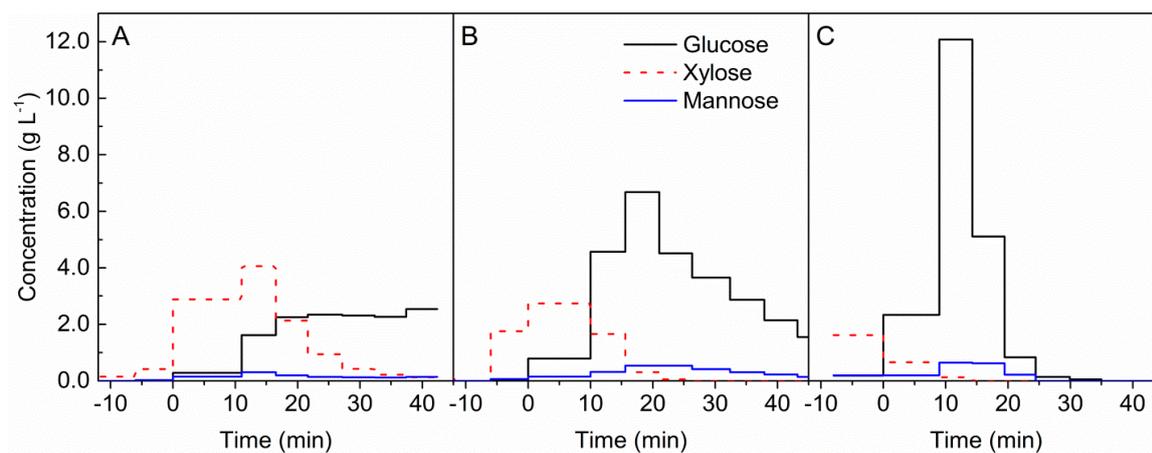


Figure 1. Formation of sugars during the hydrolysis of beech wood with 0.05 mol L^{-1} sulphuric acid at different temperatures of (A) $180 \text{ }^{\circ}\text{C}$, (B) $200 \text{ }^{\circ}\text{C}$, and (C) $220 \text{ }^{\circ}\text{C}$. $t = 0 \text{ min}$ marks the beginning of acid hydrolysis.

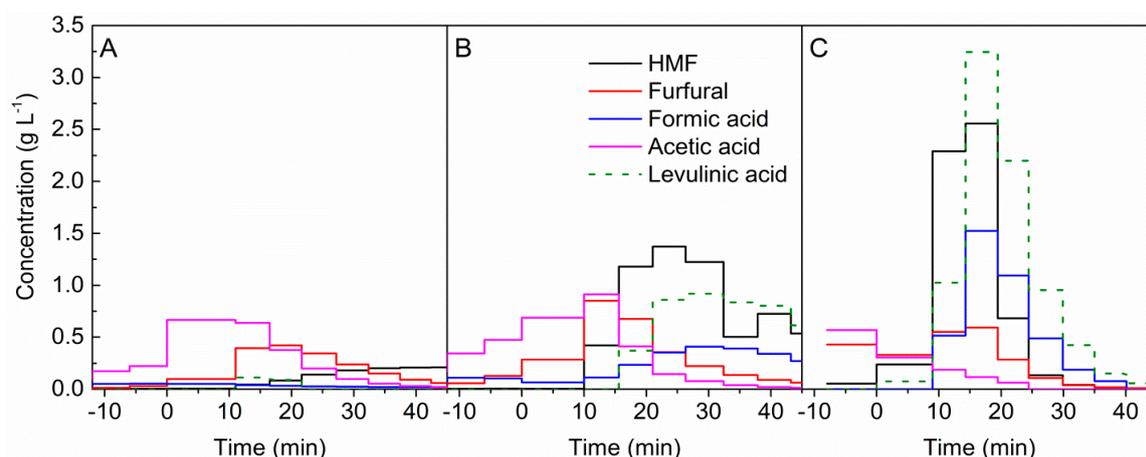


Figure 2. Formation of furfurals and organic acids during the hydrolysis of beech wood with 0.05 mol L^{-1} sulphuric acid at temperatures of (A) $180 \text{ }^{\circ}\text{C}$, (B) $200 \text{ }^{\circ}\text{C}$, and (C) $220 \text{ }^{\circ}\text{C}$. $t = 0 \text{ min}$ marks the beginning of acid hydrolysis. HMF, hydroxymehtylfurfural.

At a reaction temperature of $200 \text{ }^{\circ}\text{C}$, formation of glucose increased after addition of the acid catalyst; see Figure 1B. The highest concentration of this sugar was achieved with an experiment duration of around 20 min. After this time, a decrease in glucose concentration was observed. Formation of xylose already started at $200 \text{ }^{\circ}\text{C}$ under hydrothermal conditions during preheating. The almost complete disappearance of xylose was achieved after 20 min. No rapid changes in mannose concentration were noticed, which is the same as in the case at $180 \text{ }^{\circ}\text{C}$.

At a reaction temperature of $220 \text{ }^{\circ}\text{C}$, glucose formation had the highest concentration (12 g L^{-1}) after 10 min; see Figure 1C. After 30 min reaction duration, glucose concentrations were very low. The concentrations of xylose were already decreasing at $220 \text{ }^{\circ}\text{C}$ during the hydrothermal heating. Mannose was mainly formed between 10 and 20 min reaction time in low concentrations.

After about 10 min of acid hydrolysis at $180 \text{ }^{\circ}\text{C}$, an abrupt increase in furfural concentration was observed; see Figure 2A. After reaching the maximum, furfural concentration decreased with a longer reaction duration. Another measured furanic compound was HMF. Its concentration gradually increased up to 40 min of hydrolysis, and then the formation of this compound stayed on an equal level. Formation of formic, acetic, and levulinic acid started under preheating conditions. The addition of the acid catalyst resulted in a substantial increase in acetic acid formation. Formic acid was produced in the same order of magnitude over the entire sampling period.

As shown in Figure 2B, the maximum of the furfural concentration at 200 °C was observed earlier than the maximum HMF concentration. In contrast to hydrolysis at 180 °C, HMF reached higher concentrations than furfural. At a higher temperature, the formation of organic acids was higher compared with at 180 °C. During the hydrolysis, acetic acid was formed up to 20 min, and then a rapid decrease was observed. In comparison with a lower temperature, the concentrations of acetic acid at 200 °C were similar. The maxima of the formic acid and levulinic acid concentrations were both in the range of 20 to 40 min of the biomass reaction duration.

At 220 °C, furfural was formed under hydrothermal conditions, and the addition of acid catalyst did not increase its formation; see Figure 2C. After 30 min, furfural was detected in low concentrations. The concentrations of HMF, as well as levulinic and formic acid, undergo distinct maxima, which are between 15 and 20 min reaction time. In contrast to hydrolysis at lower temperatures, the formation of acetic acid was decreased already during the hydrothermal heating.

The temperature influence on lignocellulose hydrolysis was also determined using softwood (spruce) and grass (miscanthus) biomass. The data for spruce and miscanthus are shown in the Supplementary Materials, Figures S2–S5. Additionally, as a reference, hydrolysis of beech wood without acid catalyst addition was performed. The biomass was hydrolysed under hydrothermal conditions at 220 °C with demineralised water. The data are presented in the Supplementary Materials, Figure S6.

2.2. Influence of Biomass Type on Hydrolysis Products Formation

The pretreatment was carried out under the same conditions for each biomass. The results with hydrolysis temperature of 200 °C are described here, whereas data for temperatures of 180 °C and 220 °C are shown in the Supplementary Materials, Figures S2–S5. *Miscanthus x giganteus*, beech, and spruce wood chips were pretreated in the presence of 0.05 mol L⁻¹ sulphuric acid. A volume flow of 15 mL min⁻¹ of the acid was kept constant during the experiment.

As shown in Figure 3 for each of the biomasses, the dominant sugar was glucose. Formation of glucose at 200 °C increased after switching water to the acid solution. The highest concentration of this sugar was achieved after 10 min of the experiment for every biomass. However, for beech wood (see Figure 3A), this concentration was two times higher than for spruce wood (see Figure 3B) and miscanthus (see Figure 3C). After reaching the maximum, a decrease in glucose production was observed for both kinds of wood. Glucose formation for miscanthus was stable up to 90 min.

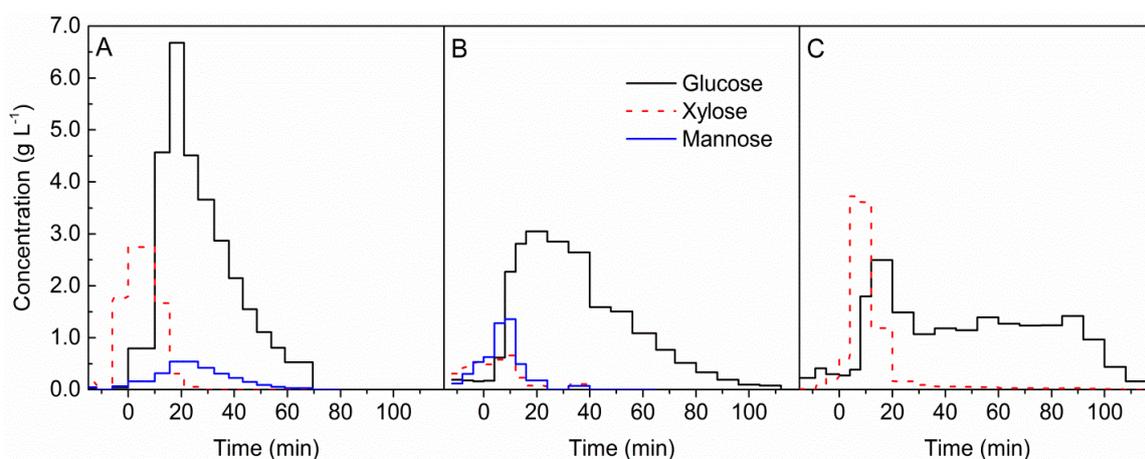


Figure 3. Formation of sugars during the hydrolysis of (A) beech wood, (B) spruce wood, and (C) *Miscanthus x giganteus* with 0.05 mol L⁻¹ sulphuric acid at a temperature of 200 °C. $t = 0$ min marks the beginning of acid hydrolysis.

For beech wood and miscanthus, a relatively high concentration of xylose was observed. Formation of xylose already started at 200 °C under hydrothermal conditions. For woods, almost complete disappearance of xylose was observed after 20 min, and for miscanthus, xylose was formed in a small amount over the entire duration of hydrolysis. At a lower temperature, xylose was the dominant sugar in beech wood and miscanthus hydrolysates, which is shown in Figure 1 and the Supplementary Materials, Figure S4.

Mannose was formed only during hydrolysis of beech and spruce woods. However, at 180 °C, for spruce wood, mannose was the sugar with the highest concentration right after glucose; see Supplementary Materials, Figure S2. Mannose was formed mainly at the initial stage of hydrolysis for spruce wood and, after 20 min, a decrease in the concentration was observed. For beech wood, the highest mannose concentration was reached at around 20 min, but it was formed over the entire duration of the hydrolysis process.

HMF was the product with the highest concentration obtained from the degradation of sugars for beech and spruce woods, as shown in Figure 4A,B. An increase in HMF concentration occurred with the addition of the acid catalyst. After reaching the maximum, the concentration of HMF decreased faster for beech wood compared with spruce wood and miscanthus; see Figure 4C. For the latter, HMF was formed for a longer duration of the hydrolysis process. Another measured furanic compound was furfural. The maximum of the furfural concentration for each biomass was observed earlier than the maximum HMF concentration. In contrast to HMF, furfural was mainly formed up to 40 min of the process.

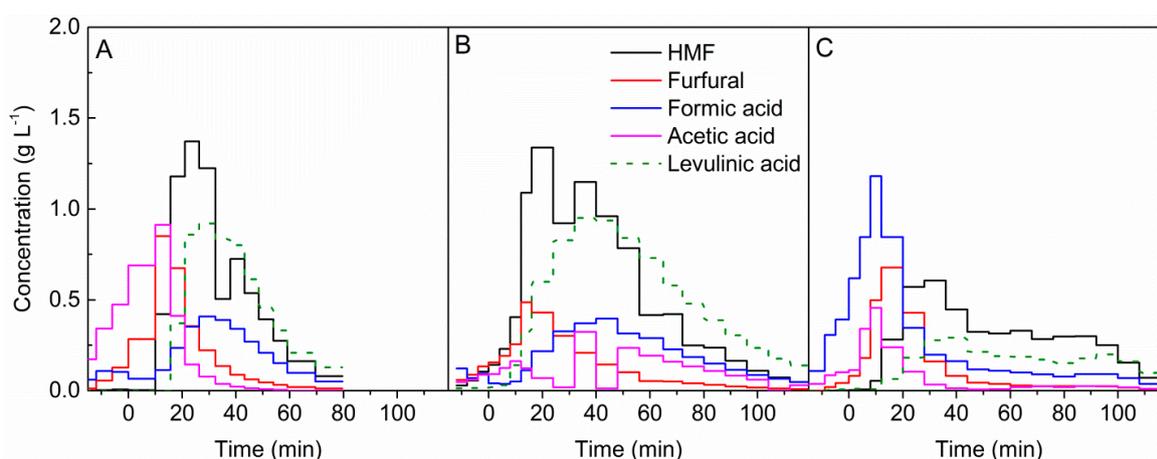


Figure 4. Formation of furfurals and organic acids during the hydrolysis of (A) beech wood, (B) spruce wood, and (C) *Miscanthus x giganteus* with 0.05 mol L⁻¹ sulphuric acid at a temperature of 200 °C. $t = 0$ min marks the beginning of acid hydrolysis.

For both woody biomasses at 200 °C, the dominant organic acid was levulinic acid. As shown in Figure 4, the formation of levulinic acid was similar to the formation of HMF for each biomass. The maximum of the levulinic acid concentration was in the range of 20 to 40 min reaction duration for beech and spruce wood. While for miscanthus formation at a constant level, similar formation to that of HMF was observed. In the same time range, the maximum concentration of formic acid was achieved for both kinds of wood. For this acid, the formation curve course was also identical that of to levulinic acid. In contrast to woody biomass, formic acid was the dominant organic acid in miscanthus hydrolysates, regardless of the temperature, as can be seen in the Supplementary Materials, Figure S5. Formation of this acid was the highest under hydrothermal preheating. For miscanthus, the maximum acetic acid concentration was obtained in the first minutes of the hydrolysis, and then only small amounts were formed. While for spruce wood, erratic concentrations were observed.

Total Products Yield

The yield of sugars, furfurals, and organic acids from the temporal concentration curves was calculated for reaction durations of 20, 40, and 60 min with acid catalyst. So, the formation of components during heat-up with water is not considered in this yield calculation. To show the yield changes depending on reaction duration and temperatures, data for spruce wood were chosen; see Table 2. The results for the three different biomasses are presented for a hydrolysis temperature of 200 °C and 40 min in Table 3, whereas data for other temperatures and reaction durations are shown in the Supplementary Materials, Tables S1 and S2.

Table 2. Main components yield after spruce wood hydrolysis with 0.05 mol L⁻¹ sulphuric acid at different temperatures. HMF, hydroxymehtylfurfural.

	Yield (mg g ⁻¹ Dry Biomass)								
	180 °C			200 °C			220 °C		
t (min)	20	40	60	20	40	60	20	40	60
Glucose	26.5	61.4	92.4	36.73	94.58	124.54	53.0	63.5	63.5
Xylose	17.6	25.7	26.5	8.31	9.49	9.49	5.1	5.8	5.8
Mannose	29.6	44.5	47.7	16.24	17.55	17.55	6.1	6.6	6.6
HMF	3.0	11.0	17.7	13.20	35.78	51.88	35.5	46.2	46.7
Furfural	2.8	7.9	10.6	6.26	12.23	14.48	9.6	14.2	14.9
Formic acid	1.0	2.4	3.6	1.89	8.53	15.58	12.3	27.9	30.6
Acetic acid	3.1	5.4	5.8	2.36	5.47	8.30	2.0	2.3	2.3
Levulinic acid	0.9	3.9	7.3	4.60	21.74	39.40	28.4	64.3	71.2

Table 3. Main components yield after 40 min hydrolysis of different biomass with 0.05 mol L⁻¹ sulphuric acid at 200 °C.

	Yield (mg g ⁻¹ Dry Biomass)		
	Beech wood	Spruce wood	Miscanthus
t (min)	40	40	40
Glucose	148.0	94.6	70.7
Xylose	42.0	9.5	60.8
Mannose	15.1	17.6	0.0
HMF	30.2	35.8	19.8
Furfural	16.8	12.2	18.5
Formic acid	10.6	8.5	29.7
Acetic acid	17.1	5.5	9.0
Levulinic acid	20.1	21.7	8.2

For glucose, an extension of the reaction duration from 20 to 40 min resulted in a more than twofold increase in yield for both 180 and 200 °C; see Table 2. Meanwhile, further extension of the reaction time to 60 min caused a smaller increase. Only for miscanthus did a longer reaction duration have a big effect on glucose yield. This is because, for this biomass, glucose was released over the entire duration of hydrolysis. Regardless of biomass type and temperature, the xylose yields nearly stayed constant from 20 to 60 min. The reason is that xylose was formed mainly at the initial stage of hydrolysis. The highest xylose yields were reported for miscanthus. While for spruce wood, the yield of xylose was nearly seven times lower. Changes in mannose yield were only observed for beech wood, where an extension of the reaction duration from 20 to 40 min resulted in a twofold yield increase. This means that, similar to xylose, mannose was formed mainly at the initial stage of the process and during the heat-up phase. Therefore, a decrease in the yield of these two sugars was observed when process temperature increased because, for this yield calculation, only the acid-catalysed reaction duration was considered, excluding the heat-up phase.

In general, a longer reaction time and higher temperature for each biomass had a positive effect on HMF yield. This was especially visible for miscanthus. At 200 °C, an extension of the reaction duration from 20 to 40 min resulted in an eightfold higher yield. At the considered reaction time, the highest HMF yield was for spruce wood, then for beech wood, and then miscanthus. The furfural yield did not show major changes depending on the biomass type. A reaction time extension to 40 min resulted in up to a twofold furfural yield increase compared with 20 min hydrolysis time at 200 and 220 °C.

The highest formic acid yields were obtained for miscanthus, while a longer reaction duration resulted in a 50% yield increase. Acetic acid yield changes were most pronounced for spruce wood. However, this acid did not show high yield changes along with temperature. Extension of the reaction duration from 20 to 40 min doubled the yield of this organic acid. The next extension of the reaction time from 40 to 60 min increased the yield by 50%. However, comparing yield values, the highest acetic acid yields were obtained for beech wood, followed by miscanthus. Among the discussed organic acids, reaction duration and temperature had the highest impact on levulinic acid yield. This acid yield at 200 °C after 60 min for miscanthus was even 25 times higher than that after 20 min. For beech and spruce wood, this yield was 16 and 13 times higher, respectively.

2.3. Influence of Hydrolysis Temperature on Solid Residue

It was observed that a higher temperature of hydrolysis caused the residue to become darker and more brittle when touched. The structural changes in the solid residue after pretreatment of beech wood were investigated using thermogravimetric analysis and infrared spectroscopy.

The changes in the thermogram of solid residues after sulphuric acid hydrolysis compared with starting biomass are shown in Figure 5. The DTG curve of untreated beech wood shows a small peak at 80 °C, which is owing to residual moisture. It follows a large peak at 360 °C with an upstream peak shoulder at about 300 °C. At a hydrolysis temperature of 180 °C, the peak shoulder initially disappears at 300 °C, whereas the peak at 360 °C shifts slightly forward. A sharply changed thermogram was obtained at a hydrolysis temperature of 200 °C, because the peak at 360 °C nearly disappeared. Besides, a new peak can be observed at around 415 °C. A further increase in temperature to 220 °C resulted in complete disappearance of the peak at 360 °C.

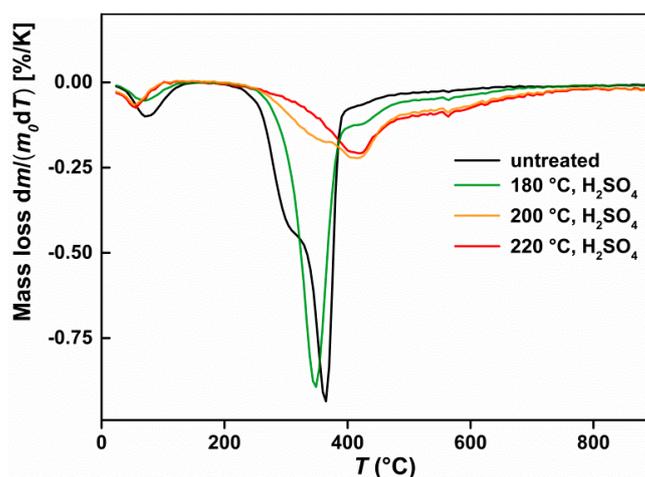


Figure 5. Thermogram (DTG curves) of untreated beech wood and the solid residue after hydrolysis with 0.05 mol L⁻¹ sulphuric acid at different temperatures.

FT-IR spectra of beech wood and solid residues from the sulphuric acid hydrolysis are shown in Figure 6. Changes in the FT-IR spectra from starting biomass to the solid residues are observed. Moreover, among the solid residues, there are differences in the IR absorption depending on the pretreatment conditions. In all solid residues, the beech wood bands disappear at 1740 and 1245 cm⁻¹. After pretreatment with sulphuric acid at higher temperatures, the bands at 1370, 1160, and 1035 cm⁻¹

become smaller. Ultimately, new bands appear after pretreatment at 1710 and 1210 cm^{-1} for a hydrolysis temperature of 200 $^{\circ}\text{C}$.

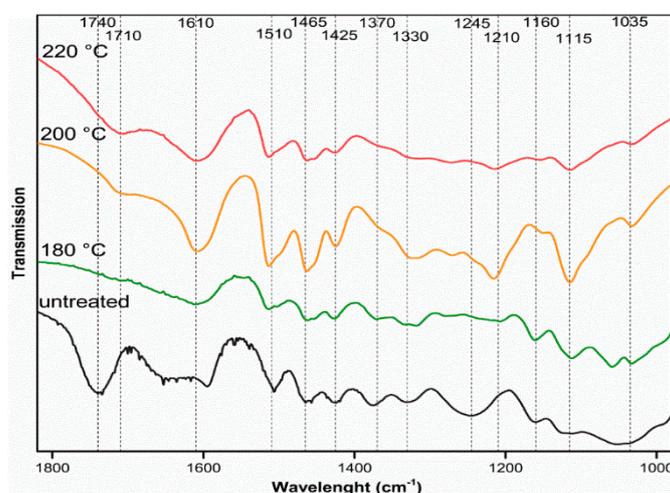


Figure 6. FT-IR spectra of untreated beech wood and solid residue after pretreatment with 0.05 mol L⁻¹ sulphuric acid at different temperatures; adsorption range from 1800 cm^{-1} to 1000 cm^{-1} .

3. Discussion

3.1. Dissolved Products Formation

In a semi-continuous process, small and soluble compounds are washed out of the reactor and measured. Here, the structure of biomass is of special importance. The cellulose fibres are stabilized by crystallinity and interconnected with hydrogen bonds. The hydrogen bonds are so strong that no swelling can occur in subcritical water. Therefore, hydrolysis occurs only at the “outer edges” of the crystalline parts or at amorphous parts [42]. Therefore, the hydrolysis of cellulose, especially with high crystallinity, is slower than that of hemicellulose. As in other publications of semi-continuous and acid-catalysed hydrolysis of biomass, the formation of oligomers is not considered, because it is known that the amount and life-time is low [43].

Glucose could originate from either the hemicellulose or cellulose fractions of lignocellulose. The glucose liberated at mild hydrolysis conditions most likely originated from hemicellulose, while the main part of glucose originated from the cellulose fraction [44,45]. The cellulose required a longer time to react at 200 $^{\circ}\text{C}$ compared with at 220 $^{\circ}\text{C}$, owing to its stable and organised structure [46].

The cellulose in beech wood was hydrolysed slowly to glucose at 180 $^{\circ}\text{C}$ with 0.05 mol L⁻¹ sulphuric acid, as indicated by the long release time of the glucose; see Figure 1. For spruce wood and miscanthus at 200 $^{\circ}\text{C}$, glucose formation was completed after 120 min; see Figure 3. This indicates that cellulose was completely reacted. For beech wood, glucose formation decreased after 70 min; see Figure 3.

Hemicellulose is easily hydrolysed in water at temperatures above 180 $^{\circ}\text{C}$ [47]. This is because of a lack of repeating glycosidic bonds, and thus the random nature of the hemicellulose polymer [46]. This is probably why hydrolysates from beech, spruce wood, and miscanthus contained glucose, even at heat-up with water; see Figure 1, Figures S2 and S4. This is because all these lignocelluloses contain glucose in the hemicellulose structure, as shown in Table 1. This is in accordance with work concerning the hydrothermal treatment of hardwood and softwood [48]. Glucose released at a later stage of the process does not originate from hemicellulose. Otherwise, glucose and xylose would be hydrolysed simultaneously. Meanwhile, xylose shows an early maximum.

The hemicellulosic framework in the biomasses did not hydrolyse completely under hydrothermal preheating to 180 $^{\circ}\text{C}$. This is visible as the release of xylose and mannose (for woods) increased

rapidly after addition of the acid catalyst; see Figure 1, Figures S2 and S4. However, when heated to 180 °C, a part of the acetylated side chains was already hydrolysed, which was visible by acetic acid release, especially for beech wood. Nevertheless, by hydrothermal heating to 220 °C, the hemicellulose was largely hydrolysed, which is noticeable by declining concentrations of xylose and acetic acid during heat-up. As xylose was released during the hydrothermal heating phase to 220 °C and glucose production began in the acid catalyst environment, it is possible to separate xylose-rich and glucose-rich hydrolysates. This is of special interest when different downstream conversions for these sugars are intended.

Low concentrations of mannose in hydrolysates could be caused by the fact that hardwood contains a small amount of glucomannan in its structure [49]. However, for beech wood, mannose was released later than xylose, especially at hydrolysis temperatures of 200 °C and 220 °C; see Figure 1. Therefore, it could be assumed that the released mannose does not originate from the hemicellulose because, otherwise, mannose and xylose would be released at the same time. Mannose was released mainly when the glucose concentration was high. So, taking into account the discussed course of mannose formation, it could be assumed that glucose might epimerize to mannose.

It has to be considered that the parameter “time” here is the reaction duration for the solid material. Glucose, for example, is solved and transported out of the reactor to depress secondary reactions to furfurals, among others. This is important when the results are compared to batch experiments, wherein the reaction time for all species is the same [50]. This may explain why the differences in hydrothermal batch experiments without acids are lower [48]—here, the consecutive reaction is more slowly, but can occur, owing to the long reactions times.

HMF is formed from hexoses in acid-catalysed processes. Here, therefore, the HMF formation is observed at a shorter duration/lower temperature compared with similar experiments without acid addition [39]. Formic and levulinic acid are consecutive products formed simultaneously in equimolar yield when HMF rehydrates [51]. The yield of HMF and its degradation products gradually raised when the hydrolysis temperature was increased, regardless of the type of biomass; see Table 2 and Tables S1 and S2. However, formic acid was also released during heating to 180 °C and 200 °C under hydrothermal conditions, probably coming from the elimination of formyl groups originally linked to hemicellulose xylan [52]. This was especially noticeable for miscanthus, where the formic acid formation was similar to xylose formation, even at 220 °C. However, the formation of formic acid may also occur in different pathways, that is, sugars defragmentation, via (1) furfuryl alcohol, (2) furfural formation (3) pyruvaldehyde, and (4) D-erythrose [53].

During the pretreatment, furfural is generated by the dehydration of liberated xylose. As can be seen in Table 2 and Tables S1 and S2, only a small change in furfural yield is obtained, despite the changing xylose yields along with the temperature and type of biomass. This is because not all of the xylose consumed will end up as furfural. The reason being that two consecutive reactions of furfural—furfural resinification and furfural condensation—can take place [52].

Acetic acid is formed from hydrolysis of the acetyl groups in the hemicellulose, as a consequence of deacetylation of acetylated pentosane [54]. Furthermore, acetic acid can also originate from the defragmentation of sugars [55,56]. This is why the formation of acetic acid is dependent not only on the biomass type, but on the hydrolysis temperature too.

For the application of the hydrolysis process for a biorefinery, the yields are of special importance. When pretreatment with 0.05 mol L⁻¹ sulphuric acid was performed, the yield of glucose was maximised at 200 °C for all feedstocks. The highest glucose yield was obtained for beech wood, 148 mg g⁻¹, while for miscanthus, the yield was twofold lower. At lower temperatures, the hydrolysis of the cellulose has not progressed so far. At 220 °C, a more substantial part of the glucose undergoes subsequent reactions. This can also be seen from the sharp increase in the yields of levulinic acid and formic acid, which are consecutive products of glucose; see Table 2, Tables S1 and S2.

The yield of xylose decreased in the considered time intervals of acid-catalysed hydrolysis with higher temperatures because the hemicellulose is already hydrolysed during the heat-up of

the lignocellulose under hydrothermal conditions. Xylose yields were the highest for beech wood, 42 mg g^{-1} , and miscanthus, 61 mg g^{-1} , because they have higher xylose content in the hemicellulose structure. The same yield decrease applied to acetic acid. For beech wood, the generally larger acetic acid yields can be explained by the high degree of acetylation of xylans in hardwood [57].

For spruce wood, the decrease in mannose yields at a higher temperature can also be explained by the hydrolysis of the hemicellulose in the heating phase. In beech wood, which contains a lower amount of mannose in the hemicellulose, this argumentation is not sufficient to describe the course of the yields.

The yield of HMF increased with the hydrolysis temperature for all the biomasses, although the increase from $200 \text{ }^{\circ}\text{C}$ to $220 \text{ }^{\circ}\text{C}$ was much weaker than from $180 \text{ }^{\circ}\text{C}$ to $200 \text{ }^{\circ}\text{C}$; see Table 2 and Tables S1 and S2. This is partly because of the strongly increasing rehydration of HMF to levulinic acid and formic acid at $220 \text{ }^{\circ}\text{C}$. Consequently, a further increase in the reaction temperature would, above all, increase the yields of these two organic acids at the expense of HMF. Optimization of the hydrolysis reaction conditions for maximising HMF yields is thus not desirable, as this would also increase the yields of the HMF derivatives. Comparing biomasses, the highest HMF yield was achieved from woods, while for miscanthus, the yield was about 40% lower.

On the basis of the obtained results, it can be concluded that all three biomasses—beech, spruce wood, and miscanthus—have great potential for bio-based platform chemical production. It should be emphasized that miscanthus can compete with woody biomass. Considering the other advantages of this feedstock as well, it deserves wider interest.

3.2. Solid Residue Changes

After pretreatment with sulphuric acid at $180 \text{ }^{\circ}\text{C}$, the peak shoulder at $300 \text{ }^{\circ}\text{C}$, which can be assigned to the hemicellulose [58], firstly disappears in the thermogram; Figure 5. This indicates a complete breakdown of hemicellulose. This is also the reason the solid residue appears to be brittle. Further, in other studies, for example, Chen et al. [59], the peak shoulder of hemicellulose disappeared after pretreatment at $180 \text{ }^{\circ}\text{C}$, or even $160 \text{ }^{\circ}\text{C}$. By contrast, the pretreatment temperature of $180 \text{ }^{\circ}\text{C}$ does not appear to have an effect on the cellulose, as the cellulose peak at $360 \text{ }^{\circ}\text{C}$ remains mostly unchanged.

At a hydrolysis temperature of $200 \text{ }^{\circ}\text{C}$, the cellulose has already mainly been converted, which can be confirmed by the disappearance of the cellulose peak at $360 \text{ }^{\circ}\text{C}$ [60]. There appears to be a temperature threshold above which the crystalline regions of cellulose either undergo structural change and/or are degraded by hydrolysis [61]. This temperature threshold is in the pretreatment of beech with 0.05 mol L^{-1} sulphuric acid between $180 \text{ }^{\circ}\text{C}$ and $200 \text{ }^{\circ}\text{C}$. The process is not well understood yet. The temperature still is too low to enable a transformation of crystalline cellulose to amorphous, but may change the structure of lignin [62]. This may influence the hydrolysis behaviour, as the lignin may cover the cellulose.

After pretreatment with sulphuric acid, a new, broad peak at approximately $415 \text{ }^{\circ}\text{C}$ is generated in the thermogram. The peak is still weak at a pretreatment temperature of $180 \text{ }^{\circ}\text{C}$, but becomes more abundant at a pretreatment temperature of $200 \text{ }^{\circ}\text{C}$. The emergence of a new peak in the region of $410\text{--}420 \text{ }^{\circ}\text{C}$ was also observed in the sulphuric acid-catalysed pretreatment of miscanthus, as well as in the hydrothermal carbonisation of various lignocelluloses [63–65]. It can be assumed that the $415 \text{ }^{\circ}\text{C}$ peak is an inert pseudo lignin formed by solid-to-solid transformations of the polysaccharides [66].

IR spectroscopy in Figure 4 shows that, after pretreatment of beech wood, the $\text{C}=\text{O}$ valence vibration at 1740 cm^{-1} has disappeared, which can be attributed to the acetylated side chains of the hemicellulose [67]. Thus, the acetylated side chains of hemicellulose are fully hydrolysed at all pretreatment conditions tested. This is related to the work of Xiao et al. [67], which investigated hydrothermal pretreatment at lower temperatures of $100\text{--}200 \text{ }^{\circ}\text{C}$. They found that the $\text{C}=\text{O}$ valence vibration at 1740 cm^{-1} decreases with the increasing hydrolysis temperature. Further, the skeleton of the hemicellulose is completely hydrolysed, which can be visualized with the disappearance of the $\text{C}-\text{O}$ stretching vibration at 1245 cm^{-1} [68]. Moreover, in the studies of Chen et al. [59] and

Sannigrahi et al. [69] after pretreatment, the C–O valence vibration of hemicellulose disappeared at 1245 cm^{-1} .

The hydrolysis of the cellulose was mostly completed when the sulphuric acid hydrolysis was carried out at 200 °C. Only then did the C–O stretching vibration of the cellulose disappear completely at 1370 cm^{-1} . This is consistent with the decrease in C–O–C valence vibration of glycosidic bonds at 1160 cm^{-1} [68].

The newly occurring C=O stretching vibration at 1710 cm^{-1} after sulphuric acid pretreatment can be attributed to pseudo lignin structures. This newly formed band was also found after the pretreatment of lignin-removed poplar [69]. The newly formed band at 1210 cm^{-1} can also be derived from pseudo lignin.

The enhancement of the aromatic C=C valence vibration at 1610 and 1510 cm^{-1} was also reported by Chen et al. and Xiao et al. and can be justified by two hypotheses [59,67]. First, condensation reactions of reactive intermediates can form new aromatics. On the other hand, the relative proportion of the aromatic structures in the solid residue increases because lignin is enriched by the hydrolysis of the polysaccharides. The latter also explains why the C–H valence vibration of methoxy groups at 2840 cm^{-1} becomes visible at higher temperatures.

4. Materials and Methods

4.1. Feedstocks

In this study, three different lignocellulosic feedstocks were used in chip size. Wood chips from pre-dried and bark-free beech wood were obtained from Joh. Sinnerbrink GmbH & Co (Verl, Germany). The chips had a length of about 30 mm, a width of about 15 mm, and a thickness of 1–2 mm, and were used without further drying. Spruce wood chips came from the Hermann Keller GmbH sawmill (Achern, Germany) with a length of about 35 mm, a width of 6–20 mm, and a thickness of about 4 mm. Originally spruce wood chips contained 48 wt% of water and were gently air-dried at 40 °C for six days.

Miscanthus x giganteus was cultivated at the fields of the University of Hohenheim (Stuttgart, Germany), and harvested in the fall of 2017 with subsequent sun-drying on the field. Then, samples were chopped with a cutting mill with a 4 mm sieve diameter. The results of the proximate and ultimate analysis of the feedstocks are shown in Table 4. Elemental analysis of the feedstocks was performed on a CHNS analyser EuroEA, 3000 Series (HEKAtech GmbH, Wegberg, Germany) according to the standard (DIN-51732). The moisture and ash content were analysed according to industrial standard proximate analysis (ASTM D1762-84). The measurements were made in duplicates, and the average data were reported.

Table 4. Proximate and ultimate analysis of the feedstocks.

Biomass	Parameter (wt%)					
	C ¹	H ¹	N ¹	S ¹	Ash ¹	Moisture ²
Beech wood	50.2	6.0	0.2	0.2	0.5	8.0
Spruce wood	48.3	6.6	0.1	0.1	0.3	3.0
<i>Miscanthus x giganteus</i>	53.7	6.7	0.5	-	3.2	12.0

¹ based on dry material, ² based on pre-dried material.

4.2. Diluted Acid Hydrothermal Fractionation

The pretreatment experiments of the lignocellulosic biomass with dilute sulphuric acid were performed using a semi-continuous mini plant. A process flow diagram of the plant is shown in the Supplementary Materials, Figure S1. A self-constructed 1.4571-stainless steel reactor with an internal volume of 100 mL was used. The reactor was clamped during the experiment in metallic heating

blocks. For temperature measurement, two thermocouples with accuracy ± 1 °C were placed in the reactor—one at the bottom and one at the middle. The reactor (R-1-SM, Figure S1) was loaded before the experiment with pre-dried biomass in chips size as a fixed bed. The biomass mass was 15.0 g of spruce and beech wood and 12.0 g of miscanthus. From a feed tank, the demineralised water (T-1) or dilute acid solution (T-2) was continuously pumped with a flow of 15 mL min^{-1} into the reactor by a preparative high-performance liquid chromatography (HPLC) pump (P-1). The liquid stream was not pre-heated before entering the reactor. The average residence time of the liquid in the reactor was 7 min, which was determined by a residence time distribution experiment using the step response method. During the heating phase (ca. 40 min), demineralised water was pumped through the reactor (R-1). When the required temperature in the reactor was reached, the feed stream was switched to acid solution. The liquid left the reactor continuously and was cooled in a heat exchanger (HE-1), and then collected in interval samples of 5–8 min duration. The samples were stored at 4 °C. After the experiment, demineralised water was pumped through the reactor for about 60 min to cool down the solid residue. Then, this residue was manually removed from the reactor. The total residence time of solid residue in the reactor was about 240 min including heat-up and cool down.

Sulphuric acid was selected as the hydrolysis catalyst with a concentration of 0.05 mol L^{-1} , corresponding to a 0.49 wt% solution. This is a typical concentration range that is used in industrial pretreatment units [70]. The hydrolysis process was carried out at a pressure of 25 bar and at three temperatures of 180, 200, and 220 °C.

4.3. Analytical Methods

4.3.1. Analysis of Liquid Products

Characterisation of the liquid samples was performed with different high-performance liquid chromatography (HPLC) systems. Preliminary filtration with $0.45 \mu\text{m}$ GHP syringe filters (Pall, New York, NY, USA) was performed to remove solid particles.

The first HPLC system for analysing monosaccharides (glucose, xylose, and mannose) consisted of an amperometric detector with a gold working electrode in pulsed measuring mode. Chromatographic separation was achieved using a Metrosep Carb 2 column (Metrohm, Filderstadt, Germany), which was maintained at 35 °C by the heater. The mixture of 0.1 mol L^{-1} sodium hydroxide aqueous solution and 0.01 mol L^{-1} sodium acetate as an eluent passed through the column at a flow rate of 0.5 mL min^{-1} . The accuracy of peak identification and sugar concentration was crossed checked with another HPLC method using MetaCarb 87C column from Agilent (Santa Clara, CA, USA) at 85 °C with 0.6 mL min^{-1} of water eluent and RI-detector. The crossed check result confirmed the qualification correctness.

The second HPLC system for determination of HMF and furfural was equipped with a UV detector at a wavelength of 290 nm. Isocratic elution was performed with 9:1 (volume ratio) water–acetonitrile mobile phase with a flow rate of 1.4 mL min^{-1} and a LiChrospher 100 RP-18 column (Merck, Darmstadt, Germany) at a column temperature of 20 °C.

Short-chain organic acids were quantified by a third HPLC system. The eluent used is a 0.004 mol L^{-1} sulphuric acid solution at a flow rate of 0.65 mL min^{-1} . The separation in the column is carried out either at 60 °C, for levulinic acid determination, or at 25 °C, for formic and acetic acid. The Aminex HPX—87H (Biorad, Hercules, CA, USA) separation column was used. Detection takes place with two detectors connected in series—RI and DAD.

The yield of the sugars, furfurals, and organic acids from the temporal concentration curves was calculated according to Equation (1).

$$Y = \frac{1}{m_{input}} \int_{0 \text{ min}}^{t_1} \dot{V} \cdot c(t) dt \quad (1)$$

where Y ($\text{mg}\cdot\text{g}_{\text{biomass}}^{-1}$), m_{input} (g), t_1 (min), \dot{V} ($\text{L}\cdot\text{min}^{-1}$), and $c(t)$ ($\text{mg}\cdot\text{L}^{-1}$) correspond to the component yield, dry mass of the used feedstock, reaction duration, diluted sulphuric acid solution flow, and sample interval concentration of a component, respectively. For comparison of yields of the main components, moderate reaction durations (relative to the lignocellulosic biomass residue) of 20, 40, and 60 min were selected.

4.3.2. Analysis of Solid Residue

The analysis of the solids after pretreatment of beech wood was carried out by thermal and spectroscopic measurement methods. Before the analysis, solid residues from the hydrolysis were dried at $105\text{ }^{\circ}\text{C}$ for 16 h.

Thermogravimetric analysis (TGA) was carried out on a thermobalance TGA851 from Mettler-Toledo (Columbus, OH, USA) at a heating rate of $10\text{ K}\cdot\text{min}^{-1}$ to $900\text{ }^{\circ}\text{C}$ under a nitrogen atmosphere. Around 10 mg of the dried and ground sample was weighed.

The IR-spectra were recorded with an FT-IR spectrometer 660-IR from Varian (Palo Alto, CA, USA) in transmittance mode for wavenumbers of $4000\text{--}400\text{ cm}^{-1}$. The IR spectroscopy of the dried and ground samples was carried out using KBr pellets and the obtained spectra were normalized.

5. Conclusions

Hydrolysis of lignocellulosic materials is a crucial step for sugars as well as biobased platform chemicals production (HMF, furfural, levulinic acid). A semi-continuous reactor type was used because it allows to combine rather a long reaction time for the low-reactive components of biomass and a short reaction time for the reactive hydrolysis products.

This study focused on dilute acid hydrolysis at different temperatures of three lignocellulosic feedstocks. At the same reaction conditions, cellulose in beech wood required nearly a twofold shorter time to react than for spruce wood and miscanthus. A temperature increase from $200\text{ }^{\circ}\text{C}$ to $220\text{ }^{\circ}\text{C}$ led to a considerable faster hydrolysis reaction and, at the same time, the yields of the obtained sugars were reduced. A concomitant shortening of the reaction time of the solid would generally have a positive effect on the profitability of a possible process because high product concentrations facilitate the purification and, at the same time, less wastewater is obtained. As consequence of the faster hydrolysis of hemicellulose compared with cellulose, most of the difference between the three feedstocks reflects the different composition and content of the hemicellulose. The hydrolysates from beech wood and miscanthus contained more xylose than spruce wood hydrolysates. Furthermore, the release of glucose already started at hydrothermal conditions.

At higher temperatures, sugar derivative products such as organic acids increase, which causes reduced glucose yields. From the time course of the main components in the pretreatment of lignocellulose, it can be concluded that HMF, levulinic acid, and formic acid are secondary products of hexoses, whereas furfural is a secondary product of xylose. However, the formic and acetic acid formation also depends on the biomass type.

Structural changes in the solid residue show that a complete hydrolysis was obtained after around 120 min reaction time (1) at $180\text{ }^{\circ}\text{C}$ in the case of hemicellulose and (2) at $200\text{ }^{\circ}\text{C}$ in the case of cellulose. At higher pretreatment temperatures, larger amounts of an inert pseudo lignin are formed.

The results obtained in this study show that both temperature and time have a strong impact on the formation of hydrolysis products. The optimization of the hydrolysis conditions to maximise the desired products of both sugars and furfurals is crucial for the efficient use of biomass.

Supplementary Materials: The following are available online at <http://www.mdpi.com/2073-4344/10/4/437/s1>, Figure S1: Process flow diagram of the mini plant for the fractionation of biomass; Figure S2: Formation of sugars in the hydrolysis of spruce wood; Figure S3: Formation of furfurals and organic acids in the hydrolysis of spruce wood; Figure S4: Formation of sugars in the hydrolysis of miscanthus; Figure S5: Formation of furfurals and organic acids in the hydrolysis of miscanthus; Figure S6: Formation of sugars and furfurals in the hydrolysis of beech wood without acid catalyst. Table S1: Main components yield after beech wood hydrolysis; Table S2: Main components yield after miscanthus hydrolysis.

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