

Review

Serum Biomarkers for Early Detection of Gynecologic Cancers

Yutaka Ueda, Takayuki Enomoto *, Toshihiro Kimura, Takashi Miyatake, Kiyoshi Yoshino, Masami Fujita and Tadashi Kimura

Department of Obstetrics and Gynecology, Osaka University Graduate School of Medicine, 2-2, Yamadaoka, Suita, Osaka, 565-0871, Japan; E-Mails: ZVF03563@nifty.com (Y.U.); kimbinbin@gmail.com (T.K.); tmiyatake@aim.com (T.M.); yosino-ki@mc.pref.osaka.jp (K.Y.); fujita@gyne.med.osaka-u.ac.jp (M.F.); tadashi@gyne.med.osaka-u.ac.jp (T.K.)

* Author to whom correspondence should be addressed; E-Mail: enomoto@gyne.med.osaka-u.ac.jp; Tel: +81-66879-3351; Fax: +81-66879-3359.

Received: 8 March 2010; in revised form: 31 May 2010 / Accepted: 3 June 2010 /

Published: 14 June 2010

Abstract: Ovarian, endometrial, and cervical cancers are three of the most common malignancies of the female reproductive organs. CA 125, historically the most reliable serum marker for ovarian cancer, is elevated in 50% of early-stage ovarian tumors. For endometrial cancers, there are no established serum markers. SCC, which is the best studied serum marker for squamous cell carcinomas, has been unreliable; SCC is elevated in cervical squamous cell carcinomas ranging from 28–85% of the time. Recent proteomics-based analyses show great promise for the discovery of new and more useful biomarkers. In this review, we will discuss the currently utilized serum tumor markers for gynecologic cancers and the novel biomarkers that are now under investigation.

Keywords: ovarian; endometrial; cervical; cancer; serum; tumor marker; CA 125; SCC; proteomics; biomarkers

Abbreviations: ApoA-1: apolipoprotein A-1; CA 15-3: cancer antigen-15-3; CA 19-9: cancer antigen-19-9; CA 72-4: cancer antigen-72-4; CA 125: cancer antigen-125; CEA: cancer embryonic antigen; CIN: cervical intraepithelial neoplasia; CYFRA 21-1: cytokeratin 19 fragments; HE4: (WFDC2), human epididymis-specific 4-disulfide core protein; H4: inter- α -trypsin inhibitor heavy chain fragment; IAP: immunosuppressive acidic protein; IGFII: insulin-like growth factor II; MIF: macrophage inhibitory factor; LPA: lysophosphatidic acid; MLRM: multiple logistic regression model; M-CSF: macrophage

colony-stimulating factor; Pap test: Papanicolaou's test; SAA: human serum amyloid A; SCC: squamous cell carcinoma antigen; SELDI-TOF-MS: surface-enhanced laser desorption-ionization time-of-flight mass spectrometry; sFas: serum soluble Fas; TF: transferrin; TK: thymidine kinase; TTR: transthyretin; TV-USG: transvaginal ultrasonography; USG: ultrasonography; VEGF-C: serum isoform of vascular endothelial growth factor; YKL-40: (aka CHI3L1) human cartilage glycoprotein-39

1. Introduction

Endometrial, cervical, and ovarian cancers are the three most common malignancies of the female reproductive tract. In the United States alone, roughly 12,000 women are diagnosed with uterine cervical cancer annually, and 4,000 will die from the disease [1]. The relatively low incidence of cervical cancer in the US is largely attributable to the effectiveness of Papanicolaou's cytological cervical screening test (the Pap test). The importance of screening for early tumor markers is supported by the finding that 60% of cervical cancers occur in women who have never received a Pap test, or who have not been tested in the past five years [1]. For comparison of effectiveness, the serum marker SCC, which is the best studied serum marker for squamous cell carcinomas, is elevated in only 22–60% of early-stage cervical squamous cell carcinomas [2,3].

Endometrial cancer is even more common than cervical cancer in the US, with 37,000 diagnosed with it and 7,000 deaths attributable to it in 2005, making it the fourth most common cancer in women [1]. In approximately 75% of endometrial cancer cases, the tumor remains confined to the uterus (FIGO stage I) and has a favorable prognosis. The prognosis, however, worsens dramatically as the disease progresses [4]. Despite this, screening for endometrial cancer is not currently done because of the lack of an appropriate, cost-effective, and acceptable test [5].

Intermediate in frequency between cervical and endometrial cancers, but by far the most fatal, is ovarian cancer. In 2005, 20,000 women in the US were diagnosed with ovarian cancer, but a heartbreaking 15,000 (75%) of these women died from the disease [1]. Roughly three-quarters of ovarian cancer cases present at an advanced disease stage, with the disease spread well beyond the ovaries [6]. In advanced-stage disease, patients most often have first symptoms related to the presence of an enlarging tumor and ascites fluid. However, in early- and mid-stage disease, most patients are asymptomatic for a prolonged period [5]. Serum cancer antigen-125 (CA 125) levels and transvaginal ultrasonography (TV-USG) can contribute to the early detection of ovarian cancer. Unfortunately, these tests are not currently cost-effective; they are thus not used routinely to screen for ovarian cancer [5].

For ovarian, endometrial, and cervical cancers, it is critical to detect the disease at the earliest possible stage. The discovery of useful serum biomarkers for the early detection of gynecologic cancers has thus been a high priority. Such tumor markers will be molecules arising from the presence of a tumor, which can appear in the surrounding tissue, and then within the blood and excretions.

Tumor markers can be secreted or shed by the tumor in excess of the normal tissue or cell phenotype. Sometimes, the molecule is uniquely specific to the tumor phenotype, often as embryonic, fetal (*i.e.*, AFP), undifferentiated, or stem-cell phenotypes. These can occur as re-expression of

silenced genes or as an alternative mRNA splicing expression of an already expressed gene product. Some glycoproteins produced by cancer cells have altered glycan structures, although the proteins themselves are ubiquitous [7]. Tumor markers might be unique extracellular matrix or cell adhesion molecules, receptors, growth factors, cytokines, or products of abnormal metabolism. Rarely, the marker molecules can be released by other tissues and organs in response to signals from the tumor. Even the body's own antibodies against tumor markers can be markers.

Tumor markers are secreted, released, or leaked into the interstitial fluids, and thus into the lymph, and finally (or directly) into the bloodstream, where they become detectable in serum samples. To be able to enter the bloodstream directly, larger molecules, often proteins, are cleaved into truncated forms or fragments, which are sometimes specific to the protease micro-environment of the tumor.

Tumor markers can be associated with patient diagnosis, prognosis, clinical management, and follow-up. Ideally, a serum marker would only appear in the blood of patients with a true malignancy; the marker would correlate with tumor stage and response to treatment, and it could be easily, cheaply, and reproducibly measured. The serum marker would be used for the screening of healthy populations and of specific groups with higher risk factors. The marker would enable a diagnosis for a specific type of cancer, help determine prognostic factors, and be used to monitor the course of treatment, remission, and recurrence, while receiving surgery, radiation, chemical, and immunological treatments.

Recent advances in clinical proteomics have propelled us into an exciting period of discovery of new cancer biomarkers, although the available proteomic technologies have their limitations. The principles of proteomic technology require stringent guidelines for the collection of clinical material, the application of analytical techniques, and for our interpretation of the data.

In this review, we present an overview of the serum tumor markers in current use. A lack of sensitivity and specificity has, so far, given most of the tumor markers in current use an unsatisfactory predictive value. We will discuss the novel biomarkers of the future, where there is great hope for the better detection and management of gynecologic cancers, including ovarian, endometrial, and cervical cancers.

2. Serum Markers for Cervical Cancer

Screening for cervical cancer with cervical cytology reduced the incidence of cervical cancer by more than 50% over the past 30 years in the United States [8]. However, it is estimated that 50% of the women in whom cervical cancer is diagnosed each year will have never had cervical cytology testing [8]. One approach for further reducing the incidence and the mortality of cervical cancer would be to increase the screening rates among groups of women at highest risk, who currently are not being screened. Another would be the establishment of appropriate serum testing for the early detection of cervical cancer. The squamous cell carcinoma antigen, SCC, is the most commonly used serum marker for squamous cell cervical carcinoma, which makes up 85–90% of all cervical carcinomas [4]. Elevated serum SCC levels have been detected in 28–88% of cervical squamous cell carcinomas [9–15] (Table 1). Pre-treatment levels of SCC have been shown to be related to the stage of the disease, size of the tumor, depth of the stromal invasion, the lymph-vascular space involvement, and lymph node metastasis [9,11,13,14,16–19]. Elevated SCC levels were also demonstrated to have predictive value for prognosis in some studies [16,17,19].

Table 1. Diagnostic serum markers for cervical cancer in clinical use.

Serum markers	SCC	CYFRA 21-1	CA 125	CA 19-9	CEA	IAP
Positive rates	squamous 28–85%	squamous 42–52%	adeno 27–75%	adeno 35–42%	adeno 26–48%	43–51%

Positive rates (elevated serum levels) detected for the indicated serum markers, in cases of squamous cell carcinoma (squamous), adenocarcinoma (adeno), or for all histological types.

Serum SCC levels have also been shown to parallel the responses to radiotherapy and chemotherapy [13,19–21]. Elevated pre-radiotherapy SCC levels were detected in 60% of 72 squamous cell carcinoma patients, whereas post-treatment SCC levels were above the cut-off value in only 2% of the patients whose disease was considered completely treated [13]. The serum SCC level also proved to be an independent predictor of response to neoadjuvant chemotherapy in locally advanced cervical cancer patients who received neoadjuvant chemotherapy and radical surgery [19]. SCC has also been used in the follow-up examination of cervical cancer patients. Increased serum SCC was shown to precede the clinical detection of recurrence of the disease [12,13,22].

The marker CYFRA 21-1 (serum fragments of cytokeratin 19) is also being used as a serum tumor marker for cervical cancer, especially for squamous cell carcinoma. Elevated CYFRA 21-1 levels have been detected in 42–52% of patients with squamous cell carcinoma of the uterine cervix [18,23,24] (Table 1). Similar to the usefulness of SCC, pre-treatment levels of CYFRA 21-1 are related to stage of the disease, size of the tumor, depth of the stromal invasion, the lymph-vascular space involvement, and lymph node metastasis [25–27]. However, raised CYFRA 21-1 levels were not demonstrated to be of predictive value for prognosis in some studies [18].

Serum CYFRA 21-1 levels were reported to be useful for monitoring the response to radiotherapy and chemotherapy [12,24]. The CYFRA 21-1 assay was also used in the follow-up examination of cervical cancer patients. Additionally, increased serum CYFRA 21-1 was shown to precede the clinical detection of recurrence of the disease [12].

Adenocarcinoma accounts for 10–15% of cervical cancers [4]. SCC, CYFRA 21-1, CA 125, CA 19-9 and CEA are positive in 20–25%, 33–63%, 27–75%, 34–42% and 26–48%, respectively, of such tumors [9–11,20,23,28–34] (Table 1). Raised serum CA 125 is associated with the stage of the cervical disease and is of some prognostic significance [29].

Another novel marker (that is also a target for new drug development) is immunosuppressive acidic protein (IAP), which is elevated in 43–51% of cervical carcinomas [33,34] (Table 1). Battaglia *et al.* found pre-treatment-elevation of serum IAP in 53% of squamous cell carcinoma cases, and in 40% of adenocarcinoma patients [34]. IAP level is related to disease stage and lymph node metastasis, and is of predictive value for prognosis [34].

There are some newer serum markers still under investigation (Table 2). Suzuki *et al.* showed that serum M-SCF was elevated in 27% of cervical cancer cases [35]. YKL-40, also known as CHI3L1 or human cartilage glycoprotein-39, was shown to be a potential biomarker in the detection and management of cervical cancer [28]. Elevated serum YKL-40 was found in 75% of squamous cell carcinoma patients and 78% of adenocarcinoma patients. Moreover, elevated pre-treatment-levels of YKL-40 were shown to predict an unfavorable prognosis, independent of the stage of the disease. Significantly elevated serum levels of circulating soluble Fas (sFas) were demonstrated in squamous

cell carcinomas when compared to that of healthy women ($p < 0.0001$) [36]. Vascular endothelial growth factor (VEGF), especially the VEGF-C isoform, was revealed to be elevated in the serum of patients with squamous cell carcinoma and cervical intraepithelial neoplasia (CIN) when compared to that of healthy women [37,38]. In cervical squamous cell carcinoma patients, serum VEGF levels were associated with the stage of the disease, but not with prognosis [38]. The serum level of thymidine kinase (TK) was demonstrated to be significantly higher in patients with cervical carcinoma than in normal women and patients with carcinoma in situ ($p < 0.01$ and $p < 0.05$, respectively) [39].

Table 2. Diagnostic serum markers for cervical cancer currently under investigation.

Serum markers	M-CSF	YKL-40		sFas	VEGF	TK
Positive rates	27%	squamous 75%	adeno 78%	squamous N/A	squamos N/A	N/A

Positive rates (elevated serum levels) detected for the indicated serum markers, in cases of squamous cell carcinoma (squamous), adenocarcinoma (adeno), or for all histological types.

3. Serum Markers for Endometrial Cancer

In current practice, screening for endometrial cancer is not undertaken because of the lack of an appropriate, cost-effective, and acceptable test that actually reduces mortality [5]. Routine use of an endometrial cytological test, comparable to the Pap test for cervical cancer, is too insensitive and nonspecific to be useful in screening for endometrial cancer [5]. Currently used serum markers and novel biomarkers under investigation for endometrial cancer are discussed below.

Elevated serum CA 125 levels have been detected in 11–43% of endometrial cancers [40–46] (Table 3). Pre-treatment CA 125 levels were shown to be related to the stage of the disease, the depth of myometrial invasion, peritoneal cytology, and lymph node metastasis [42–48]. Raised CA 125 levels were also demonstrated to be of predictive value for prognosis in some studies [42,47]. The serum CA 125 level usually parallels the clinical course of the disease [43,45,49]; however, the fact that serum CA 125 levels are often elevated in disease-free endometrial cancer patients who have undergone abdominal radiation should be kept in mind [50].

Table 3. Diagnostic serum markers for endometrial cancer in clinical use.

Serum markers	CA 125	CA 19-9	CA 15-3	CA 72-4	CEA	IAP
Positive rates	11–43%	22–24%	24–32%	22–32%	14–22%	55–76%

Positive rates (elevated serum levels) detected for the indicated serum marker in cases of endometrial cancer are shown.

The serum markers CA 19-9, CA 15-3, CA 72-4, and CEA levels are raised in endometrial cancer patients in 22–24%, 24–32%, 22–32% and 14–22% of the cases, respectively [11,33,40,42,45,51,52] (Table 3). Serum CA 15-3 levels were shown to be associated with prognosis [42,53]. Elevated serum IAP levels have been detected in 55–76% of endometrial cancer patients [17,33] (Table 3).

There are other serum markers for endometrial cancer now under investigation (Table 4). Raised serum M-CSF levels were detected in 25–73% of endometrial cancer cases [35,43]. HE4 was

demonstrated to provide 46% sensitivity for endometrioid adenocarcinoma of the endometrium in all stages at 95% specificity [54]. For stage I disease, HE4 yielded a 17% improvement in sensitivity compared with that of CA 125. Significantly elevated levels of serum sFas were demonstrated in endometrioid adenocarcinoma of the endometrium compared to that of healthy women ($p < 0.0001$) [36]. Human serum amyloid A (SAA) is a high-density lipoprotein that has recently been proposed as a useful biomarker for several kinds of tumors, including gastric, lung, pancreatic, and nasopharyngeal cancers [55–59]. Cocco *et al.* showed that SAA was overexpressed and actively secreted by Grade-3 endometrioid adenocarcinoma and serous papillary carcinoma of the endometrium, and that SAA was present at high concentration in the serum of these patients [55,60].

Table 4. Diagnostic serum markers for endometrial cancer under investigation.

Serum markers	M-CSF	HE4	sFas	SAA
Positive rates	25–73%	E 46%	E N/A	G3-E/S N/A

Positive rates (elevated serum levels) of each serum marker detected in cases of squamous cell carcinoma, adenocarcinoma, and for all histological types, are shown. E: endometrioid adenocarcinoma; G3-E: endometrioid adenocarcinoma Grade-3; S: serous adenocarcinoma; N/A: not assessed

Recently, panels of novel biomarkers have been developed to better detect cancers, including endometrial tumors (Table 5). Farias-Eisner *et al.* constructed a multiple logistic regression model (MLRM) with the use of the values for ApoA-1 (apolipoprotein A-1), TF (transferrin), and TTR (transthyretin) for the detection of endometrial cancer [61]. This panel distinguished normal samples from early-stage endometrial cancer with a sensitivity of 71% and a specificity of 88%. Additionally, the panel distinguished normal samples from late stage endometrial cancer with a sensitivity of 82% and a specificity of 86%.

Table 5. Novel biomarker panels for detection of endometrial cancer.

Reference	Biomarkers	Sensitivity		Specificity	
Zhu <i>et al.</i> (2006)[63]	13 proteins	93%		100%	
Farias-Eisner <i>et al.</i> (2009) [61]	3 proteins	Early 71%	Late 82%	Early 88%	Late 86%
Takano <i>et al.</i> (2009) [62]	2 proteins	82%		86%	

Shown here is the sensitivity and specificity of each serum biomarker panel used in the detection of endometrial cancer; Early: early-stage cases; Late: late-stage cases

Recent proteomic techniques, which can identify differentially expressed proteins in a large set of samples, have been applied to the discovery of new biomarkers in many diseases [62]. Zhu *et al.* has established a diagnostic system with 13 novel potential biomarkers, using surface-enhanced laser desorption-ionization time-of-flight mass spectrometry (SELDI-TOF-MS) to differentiate endometrial cancer patients from healthy women. The technique had a sensitivity of 93% and a specificity of 100% [63]. Takano *et al.* also used SELDI-TOF-MS to identify candidate markers for endometrial cancer [62]. Two of the candidates turned out to be apolipoprotein A-1 and apolipoprotein C-1. Dual-biomarker analysis for the detection of endometrial cancer yielded a sensitivity of 82% and a specificity of 86%. These studies, which analyzed test samples consisting of a significantly higher

proportion of cancer patients than would be found in a general population, and thus a significantly lower proportion of healthy controls, had methodological limitations, as previously described [64]. The prevalence of endometrial cancer was 50% in the study by Takano *et al.* [62] and 57% in the study by Zhu *et al.* [63]. Thus, these results might have overstated the sensitivity of the tests. However, because one of the candidate proteins discovered in a study by Takano *et al.* [62], apolipoprotein A-1, was also a constituent of the biomarker panel constructed by Farias-Eisner *et al.* [61], this protein may be a very promising biomarker for endometrial cancer.

4. Serum Markers for Ovarian Cancer

Establishment of an appropriate screening test for ovarian cancer has long been sought. This disease is the leading cause of death from gynecologic malignancies in the US, with its poor prognosis resulting from the lack of reliable, sensitive screening tests and our limited understanding of the mechanisms of chemoresistance and relapse. More than two-thirds of cases of ovarian cancers are diagnosed only after the disease has progressed to stage III or IV and involve the peritoneal cavity or other organs.

Symptoms that are associated with ovarian cancer are typically nonspecific and the association is often not recognized until the disease is advanced [64]. Previous studies showed that ultrasonography (USG), with or without power Doppler, provided high sensitivity. However, ultrasonography's specificity and positive predictive values were unsatisfactory [65,66]. Serum markers and novel biomarkers for early detection of ovarian cancer that are currently used or under investigation are discussed below.

Elevated serum CA 125 levels have been detected in 50% and 92% of ovarian cancers in early and late stages, respectively [67] (Table 6). According to a review by Nossow *et al.*, the positive predictive value of the CA 125 assay for the early detection of ovarian cancer is 57% [68]. Elevated CA 125 occurs in other cancers, such as in endometrial, breast, pancreatic, gastrointestinal, and lung cancers. Raised CA 125 levels are sometimes found in patients with benign gynecologic conditions, such as menstruation, pregnancy, endometriosis, and pelvic inflammatory disease, and even in non-gynecologic conditions, such as hepatitis and pancreatitis [49]. The predictive value of pre-treatment CA 125 levels for prognosis is controversial; however, changes in CA 125 levels correlate with the regression, stability, and progression of the disease in 87–94% of instances [49].

Table 6. Diagnostic serum markers for ovarian cancer in clinical use.

Serum markers	CA 125		CA 19-9		CA 15-3	CA 72-4	IAP
Positive rates	Early	Late	M	non-M	50–56%	63–71%	70–93%
	50%	92%	68–83%	28–29%			

Positive rates detected for each serum marker in cases of ovarian cancer are shown.

Early: early-stage cases; Late: late-stage cases; M: mucinous adenocarcinoma; Non-M: histological types of ovarian carcinoma other than mucinous adenocarcinoma.

Serum levels of CA 19-9 (a monosialoganglioside antigen widely used in GI adenocarcinoma diagnostics) are elevated in 68–83% of mucinous ovarian cancers, and only in 28–29% of non-mucinous types. In contrast, whereas CA 125 is elevated in 80% of non-mucinous ovarian tumors [69–72].

Serum CA 15-3, CA 72-4, and CEA levels are raised in 50–56%, 24–32%, 63–71% and 70–93% of the ovarian cancer patients, respectively [71,73–78] (Table 6). According to Gadducci *et al.*, CA 19-9, CA 15-3, and CA 72-4 correlated worse than CA 125 with the clinical course of the disease. Additionally, these markers did not offer additional clinical benefit for monitoring ovarian cancer, suggesting that the serial measurement of these markers may play a role only in the management of patients with a normal CA 125 assay [49].

There are serum markers for ovarian cancer that are under active investigation (Table 7). In a review by Li *et al.*, HE4 displayed the highest sensitivity among single markers, including CA 125, in the detection of ovarian cancer in both the early (62–83%) and late (75–93%) stages, respectively [79]. Elevated serum lysophosphatidic acid (LPA) levels were found in 90% and 98% of ovarian cancer in early and late stages, respectively. However, serum levels of LPA do not correlate with the stage of the disease, and nonspecific elevation of LPA was detected in healthy and benign gynecologic conditions [68,80,81]. Significantly elevated sFas levels were detected in ovarian cancer patients when compared to that of healthy women. Additionally, the serum sFas level was demonstrated to be a statistically significant indication factor for survival, as well as for the histological grading of ovarian carcinomas [36].

Table 7. Diagnostic serum markers for ovarian cancer under investigation.

Serum markers	HE4		LPA		sFas
Positive rates	Early	Late	Early	Late	N/A
	62–83%	75–93%	90%	98%	

Positive rates detected for each serum marker in cases of ovarian cancer are shown.

Early: early-stage cases; Late: late-stage cases; N/A: not assessed

Novel biomarker panels have also been investigated for the early detection of ovarian cancers (Table 8). Zhang *et al.* identified a panel that consisted of three proteins, including ApoA-1, a truncated form of TTR, and a cleavage fragment of H4 (inter- α -trypsin inhibitor heavy chain), to detect early-stage ovarian cancer with a sensitivity of 83% and a specificity of 94% [82].

Table 8. Novel biomarker panels for the detection of ovarian cancer.

Reference	Biomarkers	Sensitivity		Specificity	
Zhang <i>et al.</i> (2004) [82]	3 proteins	early 83%		early 94%	
Su <i>et al.</i> (2007) [83]	4 proteins	early, M 95%	early, all 89%	early, M 92%	early, all 97%
Nosov <i>et al.</i> (2009) [84]	4 proteins	early, S 94%	early, E 98%	early, S 94%	early, E 98%
Visintin <i>et al.</i> (2008) [85]	6 proteins	95%		99%	

The sensitivity and specificity of each serum biomarker panel used for the detection of ovarian cancer are shown; Early: early-staged cases; Late: late-staged cases; All: cases in all stages; S: serous adenocarcinoma; M: mucinous adenocarcinoma.

Su *et al.* utilized an MLRM with values for CA 125, ApoA-I, TF, and TTR for the early detection of ovarian cancer [83]. This model provided a sensitivity of 89% and a specificity of 97% for the detection of early stage ovarian cancer. Furthermore, the sensitivity and the specificity of this panel in distinguishing between normal and mucinous ovarian cancer samples were 95% and 92%, respectively. Nosov *et al.* applied this same MLRM model and marker panel to serous and endometrioid histological types of ovarian carcinomas and demonstrated a sensitivity of 94% and a specificity of 94% for serous ovarian carcinoma in its early stage, and a sensitivity of 98% and a specificity of 98% for endometrioid ovarian carcinoma in its early stage [84].

Visintin *et al.* proposed a set of serum biomarkers that consisted of leptin, prolactin, osteopontin, insulin-like growth factor II (IGFII), macrophage inhibitory factor (MIF), and CA 125 to discriminate between ovarian cancer patients and healthy women. The panel had a sensitivity of 95% and a specificity of 99% [85]. Not surprisingly, this panel provided a significant improvement over CA 125 alone. However, these studies had the same methodological limitations, of excessive numbers of tumor cases versus matched population controls, described above in the endometrial cancer section. With that being said, novel proteomics-based investigations and bioinformatics analyses still provide the greatest promise of using existing approaches to find ever more accurate and useable biomarkers for gynecological cancers.

One important question remains: how long before the diagnosis of an ovarian cancer does the serum level of various markers begin to rise above background levels as a tumor grows? Anderson *et al.* [86] studied concentrations of CA125, HE4, and mesothelin in serum samples collected from 0–18 years before the diagnosis of a tumor, as part of an unrelated study. They found that the markers may provide some evidence of ovarian cancer up to 3 years before clinical diagnosis, but the more likely lead time for the detection of a change associated with these markers appears to be less than 1 year.

5. Conclusions

For gynecologic cervical, endometrial, and ovarian cancers, only a small handful of tumor-associated antigens, such as SCC and CA 125, have been routinely used as tumor markers. Some markers are useful, not only as diagnostic tools, but also as a predictive marker for the prognosis and the clinical course after treatment. Some recently investigated new serum markers seem to be clinically useful, such as YKL-40 for cervical cancer and HE4 for endometrial and ovarian cancers. Recent breakthroughs in proteomics and bioinformatics technology will expand our understanding of tumor-specific biomarkers. Such investigations will establish newer and more useful biomarkers for the more accurate detection and management of ovarian, endometrial, and cervical cancers.

Acknowledgements

We would like to thank Buzard, G.S., CDCP, Buzard, M.M., Kaplan University-Hagerstown, and Minekawa, R., Osaka University for their constructive critiques of our manuscript.

References

1. Centers for Disease Control and Prevention. Gynecologic Cancers. Available online: <http://www.cdc.gov/cancer/gynecologic/index.htm> (accessed on 1 March 2010).
2. Nam, J.H.; Chang, K.C.; Chambers, J.T.; Schwartz, P.E.; Cole, L.A. Urinary gonadotropin fragment, a new tumor marker. III. Use in cervical and vulvar cancers. *Gynecol. Oncol.* **1990**, *38*, 66–70.
3. Pectasides, D.; Economides, N.; Bourazanis, J.; Pozadzizou, P.; Gogou, L.; Koutsiouba, P.; Athanassiou, A. Squamous cell carcinoma antigen, tumor-associated trypsin inhibitor, and carcinoembryonic antigen for monitoring cervical cancer. *Am. J. Clin. Oncol.* **1994**, *17*, 307–312.
4. DiSaia, P.J.; Creasman, W.T. *Clinical Gynecologic Oncology*, 6th ed.; Mosby: St. Louis, MO, USA, 2002; pp. 53–112, 137–184, 259–350.
5. Berek, J.S., Ed. *Novak's Gynecology*, 13th ed.; Lippincott Williams & Wilkins: Philadelphia, PA, USA, 2002; pp. 1143–1398.
6. Scully, R.E.; Young, R.H.; Clement, P.B. Tumors of the Ovary, Maldeveloped Gonads, Fallopian Tube, and Broad Ligament. In *Atlas of Tumor Pathology*, Third Series, Fascicle 23; AFIP: Washington, DC, USA, 1998; pp. 27–50.
7. Narimatsu, H.; Sawaki, H.; Kuno, A.; Kaji, H.; Ito, H.; Ikebara, Y. A strategy for discovery of cancer glycol-biomarkers in serum using newly developed technologies for glycoproteomics. *FEBS J.* **2010**, *277*, 95–105.
8. The American College of Obstetricians and Gynecologists (ACOG). ACOG Practice Bulletin No. 109. Clinical Management Guidelines for Obstetricians and Gynecologists. Cervical Cytology Screening. *Obstet. Gynecol.* **2009**, *114*, 1409–1420.
9. Massuger, L.F.; Koper, N.P.; Thomas, C.M.; Dom, K.E.; Schijf, C.P. Improvement of clinical staging in cervical cancer with serum squamous cell carcinoma antigen and CA 125 determinations. *Gynecol. Oncol.* **1997**, *64*, 473–476.
10. Lehtovirta, P.; Viinikka, L.; Ylikorkala, O. Comparison between squamous cell carcinoma-associated antigen and CA-125 in patients with carcinoma of the cervix. *Gynecol. Oncol.* **1990**, *37*, 276–278.
11. Gadducci, A.; Ferdeghini, M.; Caenaro, G.F.; Prontera, C.; Malagnino, G.; Annichiarico, C.; Chirillo, R.; Bianchi, R.; Fioretti, P. Immunoacid protein (IAP) as marker for cervical and endometrial carcinoma: alone and in comparison with CA 125 and SCC. *Cancer J.* **1992**, *5*, 272–278.
12. Kainz, C.; Sliutz, G.; Mustafa, G.; Bieglmayr, C.; Koelbl, H.; Reinthaller, A.; Gitsch, G. Cytokeratin subunit 19 measured by CYFRA 21-1 assay in follow-up of cervical cancer. *Gynecol. Oncol.* **1995**, *56*, 402–425.
13. Micke, O.; Prott, F.J.; Schäfer, U.; Tangerding, S.; Pötter, R.; Willich, N. The impact of squamous cell carcinoma (SCC) antigen in the follow-up after radiotherapy in patients with cervical cancer. *Anticancer Res.* **2000**, *20*, 5113–5115.
14. Takeda, M.; Sakuragi, N.; Okamoto, K.; Todo, Y.; Minobe, S.; Nomura, E.; Negishi, H.; Oikawa, M.; Yamamoto, R.; Fujimoto, S. Preoperative serum SCC, CA125, and CA19-9 levels and lymph

- node status in squamous cell carcinoma of the uterine cervix. *Acta Obstet. Gynecol. Scand.* **2002**, *81*, 451–457.
- 15. Ohara, K.; Tanaka, Y.; Tsunoda, H.; Nishida, M.; Sugahara, S.; Itai, Y. Assessment of cervical cancer radioresponse by serum squamous cell carcinoma antigen and magnetic resonance imaging. *Obstet. Gynecol.* **2002**, *100*, 781–787.
 - 16. Duk, J.M.; Groenier, K.H.; de Bruijn, H.W.; Hollema, H.; ten Hoor, K.A.; van der Zee, A.G.; Aalders, J.G. Pretreatment serum squamous cell carcinoma antigen: a newly identified prognostic factor in early-stage cervical carcinoma. *J. Clin. Oncol.* **1996**, *14*, 111–118.
 - 17. Gadducci, A.; Scambia, G.; Benedetti-Panici, P.; Ferdeghini, M.; Battaglia, F.; Caenaro, G.F.; Foti, E.; Castellani, C.; Cristofani, R.; Facchini, V.; Mancuso, S. The prognostic relevance of pretreatment serum immunosuppressive acidic protein (IAP) in patients with squamous cell carcinoma of the uterine cervix: a comparison with squamous cell carcinoma antigen (SCC). *Cancer J.* **1994**, *7*, 241–247.
 - 18. Gaarenstroom, K.N.; Kenter, G.G.; Bonfrer, J.M.; Korse, C.M.; Van de Vijver, M.J.; Fleuren, G.J.; Trimbos, J.B. Can initial serum cyfra 21-1, SCC antigen, and TPA levels in squamous cell cervical cancer predict lymph node metastases or prognosis? *Gynecol. Oncol.* **2000**, *77*, 164–170.
 - 19. Scambia, G.; Benedetti, P.; Foti, E.; Ferrandina, G.; Leone, F.P.; Marciano, M.; Mancuso, S. Multiple tumour marker assays in advanced cervical cancer: relationship to chemotherapy response and clinical outcome. *Eur. J. Cancer* **1996**, *32A*, 259–263.
 - 20. Pras, E.; Willemse, P.H.; Canrinus, A.A.; de Bruijn, H.W.; Sluiter, W.J.; ten Hoor, K.A.; Aalders, J.G.; Szabo, B.G.; de Vries, E.G. Serum squamous cell carcinoma antigen and CYFRA 21-1 in cervical cancer treatment. *Int. J. Radiat. Oncol. Biol. Phys.* **2002**, *52*, 23–32.
 - 21. de Bruijn, H.W.; Duk, J.M.; van der Zee, A.G.; Pras, E.; Willemse, P.H.; Boonstra, H.; Hollema, H.; Mourits, M.J.; de Vries, E.G.; Aalders, J.G. The clinical value of squamous cell carcinoma antigen in cancer of the uterine cervix. *Tumour Biol.* **1998**, *19*, 505–516.
 - 22. Ngan, H.Y.; Cheng, G.T.; Yeung, W.S.; Wong, L.C.; Ma, H.K. The prognostic value of TPA and SCC in squamous cell carcinoma of the cervix. *Gynecol. Oncol.* **1994**, *52*, 63–68.
 - 23. Ferdeghini, M.; Gadducci, A.; Annicchiarico, C.; Prontera, C.; Malagnino, G.; Castellani, C.; Facchini, V.; Bianchi, R. Serum CYFRA 21-1 assay in squamous cell carcinoma of the cervix. *Anticancer Res.* **1993**, *13*, 1841–1844.
 - 24. Suzuki, Y.; Nakano, T.; Ohno, T.; Abe, A.; Morita, S.; Tsujii, H. Serum CYFRA 21-1 in cervical cancer patients treated with radiation therapy. *J. Cancer Res. Clin. Oncol.* **2000**, *126*, 332–336.
 - 25. Gaarenstroom, K.N.; Bonfrer, J.M.; Korse, C.M.; Kenter, G.G.; Kenemans, P. Value of Cyfra 21-1, TPA, and SCC-Ag in predicting extracervical disease and prognosis in cervical cancer. *Anticancer Res.* **1997**, *17*, 2955–2958.
 - 26. Molina, R.; Filella, X.; Augé, J.M.; Bosch, E.; Torne, A.; Pahisa, J.; Lejarcegui, J.A.; Rovirosa, A.; Mellado, B.; Ordi, J.; Biete, A. CYFRA 21.1 in patients with cervical cancer: comparison with SCC and CEA. *Anticancer Res.* **2005**, *25*, 1765–1771.
 - 27. Puthucode-Easwaran, S.; Naik, R.; Athavale, R.; Handley, G.; Lopes, A.; Godfrey, K.; Hatem, M. Comparison of pre-treatment CYFRA 21 - 1 and SCC-Antigen assay in primary cervical carcinoma - a preliminary report. *J. Obstet. Gynaecol.* **2005**, *25*, 486–488.

28. Mitsuhashi, A.; Matsui, H.; Usui, H.; Nagai, Y.; Tate, S.; Unno, Y.; Hirashiki, K.; Seki, K.; Shozu, M. Serum YKL-40 as a marker for cervical adenocarcinoma. *Ann. Oncol.* **2009**, *20*, 71–77.
29. Ngan, H.Y.; Cheung, A.N.; Lauder, I.J.; Cheng, D.K.; Wong, L.C.; Ma, H.K. Tumour markers and their prognostic value in adenocarcinoma of the cervix. *Tumour Biol.* **1998**, *19*, 439–444.
30. Leminen, A. Tumor markers CA 125, carcinoembryonic antigen and tumor-associated trypsin inhibitor in patients with cervical adenocarcinoma. *Gynecol. Oncol.* **1990**, *39*, 358–363.
31. Duk, J.M.; Aalders, J.G.; Fleuren, G.J.; Krans, M.; De Bruijn, H.W. Tumor markers CA 125, squamous cell carcinoma antigen, and carcinoembryonic antigen in patients with adenocarcinoma of the uterine cervix. *Obstet. Gynecol.* **1989**, *73*, 661–668.
32. Borras, G.; Molina, R.; Xercavins, J.; Ballesta, A.; Iglesias, J. Tumor antigens CA 19.9, CA 125, and CEA in carcinoma of the uterine cervix. *Gynecol Oncol.* **1995**, *57*, 205–211.
33. Sawada, M.; Okudaira, Y.; Matsui, Y.; Shimizu, Y. Immunosuppressive acidic protein in patients with gynecologic cancer. *Cancer* **1984**, *54*, 652–656.
34. Battaglia, F.; Scambia, G.; Panici, P.B.; Castelli, M.; Ferrandina, G.; Foti, E.; Amoroso, M.; D'Andrea, G.; Mancuso, S. Immunosuppressive acidic protein (IAP) and squamous cell carcinoma antigen (SCC) in patients with cervical cancer. *Gynecol. Oncol.* **1994**, *53*, 176–182.
35. Suzuki, M.; Ohwada, M.; Sato, I.; Nagatomo, M. Serum level of macrophage colony-stimulating factor as a marker for gynecologic malignancies. *Oncology* **1995**, *52*, 128–133.
36. Konno, R.; Takano, T.; Sato, S.; Yajima, A. Serum soluble Fas level as a prognostic factor in patients with gynecological malignancies. *Clin. Cancer Res.* **2000**, *6*, 3576–3580.
37. Mathur, S.P.; Mathur, R.S.; Gray, E.A.; Lane, D.; Underwood, P.G.; Kohler, M.; Creasman, W.T. Serum vascular endothelial growth factor C (VEGF-C) as a specific biomarker for advanced cervical cancer: Relationship to insulin-like growth factor II (IGF-II), IGF binding protein 3 (IGF-BP3) and VEGF-A. *Gynecol. Oncol.* **2005**, *98*, 467–483.
38. Lebrecht, A.; Ludwig, E.; Huber, A.; Klein, M.; Schneeberger, C.; Tempfer, C.; Koelbl, H.; Hefler, L. Serum vascular endothelial growth factor and serum leptin in patients with cervical cancer. *Gynecol. Oncol.* **2002**, *85*, 32–35.
39. Fujiwaki, R.; Hata, K.; Moriyama, M.; Iwanari, O.; Katabuchi, H.; Okamura, H.; Miyazaki, K. Clinical value of thymidine kinase in patients with cervical carcinoma. *Oncology* **2001**, *61*, 47–54.
40. Takeshima, N.; Shimizu, Y.; Umezawa, S.; Hirai, Y.; Chen, J.T.; Fujimoto, I.; Yamauchi, K.; Hasumi, K. Combined assay of serum levels of CA125 and CA19-9 in endometrial carcinoma. *Gynecol. Oncol.* **1994**, *54*, 321–326.
41. Panici, P.B.; Scambia, G.; Baiocchi, G.; Perrone, L.; Greggi, S.; Battaglia, F.; Mancuso, S. Multiple serum markers in patients with endometrial cancer. *Gynecol. Obstet. Invest.* **1989**, *27*, 208–212.
42. Scambia, G.; Gadducci, A.; Panici, P.B.; Foti, E.; Ferdeghini, M.; Ferrandina, G.; Amoroso, M.; Castellani, C.; Facchini, V.; Mancuso, S. Combined use of CA 125 and CA 15-3 in patients with endometrial carcinoma. *Gynecol. Oncol.* **1994**, *54*, 292–297.
43. Hakala, A.; Kacinski, B.M.; Stanley, E.R.; Kohorn, E.I.; Puistola, U.; Risteli, J.; Risteli, L.; Tomás, C.; Kauppila, A. Macrophage colony-stimulating factor 1, a clinically useful tumor marker in endometrial adenocarcinoma: comparison with CA 125 and the aminoterminal propeptide of type III procollagen. *Am. J. Obstet. Gynecol.* **1995**, *173*, 112–119.

44. Peters-Engl, C.; Buxbaum, P.; Ogris, E.; Sevelda, P.; Medl, M. TATI (tumor associated trypsin inhibitor) and cancer antigen 125 (CA 125) in patients with early-stage endometrial cancer. *Anticancer Res.* **1998**, *18*, 4635–4639.
45. Cherchi, P.L.; Dessole, S.; Ruiu, G.A.; Ambrosini, G.; Farina, M.; Capobianco, G.; Ambrosini, A. The value of serum CA 125 and association CA 125/CA 19-9 in endometrial carcinoma. *Eur. J. Gynaecol. Oncol.* **1999**, *20*, 315–317.
46. Ginath, S.; Menczer, J.; Fintsi, Y.; Ben-Shem, E.; Glezerman, M.; Avinoach, I. Tissue and serum CA125 expression in endometrial cancer. *Int. J. Gynecol. Cancer* **2002**, *12*, 372–375.
47. Sood, A.K.; Buller, R.E.; Burger, R.A.; Dawson, J.D.; Sorosky, J.I.; Berman, M. Value of preoperative CA 125 level in the management of uterine cancer and prediction of clinical outcome. *Obstet. Gynecol.* **1997**, *90*, 441–447.
48. Hsieh, C.H.; ChangChien, C.C.; Lin, H.; Huang, E.Y.; Huang, C.C.; Lan, K.C.; Chang, S.Y. Can a preoperative CA 125 level be a criterion for full pelvic lymphadenectomy in surgical staging of endometrial cancer? *Gynecol. Oncol.* **2002**, *86*, 28–33.
49. Gadducci, A.; Cosio, S.; Carpi, A.; Nicolini, A.; Genazzani, A.R. Serum tumor markers in the management of ovarian, endometrial and cervical cancer. *Biomed. Pharmacother.* **2004**, *58*, 24–38.
50. Carpenter, P.M.; Gamboa, G.P.; Dorion, G.E.; Ramsinghani, N.S.; Assi, A.M.; Manetta, A. Radiation-induced CA 125 production by mesothelial cells. *Gynecol. Oncol.* **1996**, *63*, 328–332.
51. Hareyama, H.; Sakuragi, N.; Makinoda, S.; Fujimoto, S. Serum and tissue measurements of CA72-4 in patients with endometrial carcinoma. *J. Clin. Pathol.* **1996**, *49*, 967–970.
52. Gadducci, A.; Ferdeghini, M.; Prontera, C.; Giordano, P.; Cristofani, R.; Bianchi, R.; Fioretti, P. A comparison of pretreatment serum levels of four tumor markers in patients with endometrial and cervical carcinoma. *Eur. J. Gynaecol. Oncol.* **1990**, *11*, 283–288.
53. Lo, S.S.; Cheng, D.K.; Ng, T.Y.; Wong, L.C.; Ngan, H.Y. Prognostic significance of tumour markers in endometrial cancer. *Tumour Biol.* **1997**, *18*, 241–249.
54. Moore, R.G.; Brown, A.K.; Miller, M.C.; Badgwell, D.; Lu, Z.; Allard, W.J.; Granai, C.O.; Bast, R.C., Jr.; Lu, K. Utility of a novel serum tumor biomarker HE4 in patients with endometrioid adenocarcinoma of the uterus. *Gynecol. Oncol.* **2008**, *110*, 196–201.
55. Cocco, E.; Bellone, S.; El-Sahwi, K.; Cargnelutti, M.; Buza, N.; Tavassoli, F.A.; Schwartz, P.E.; Rutherford, T.J.; Pecorelli, S.; Santin, A.D. Serum amyloid A: a novel biomarker for endometrial cancer. *Cancer* **2010**, *116*, 843–851.
56. Chan, D.C.; Chen, C.J.; Chu, H.C.; Chang, W.K.; Yu, J.C.; Chen, Y.J.; Wen, L.L.; Huang, S.C.; Ku, C.H.; Liu, Y.C.; Chen, J.H. Evaluation of serum amyloid A as a biomarker for gastric cancer. *Ann. Surg. Oncol.* **2007**, *14*, 84–93.
57. Yokoi, K.; Shih, L.C.; Kobayashi, R.; Koomen, J.; Hawke, D.; Li, D.; Hamilton, S.R.; Abbruzzese, J.L.; Coombes, K.R.; Fidler, I.J. Serum amyloid A as a tumor marker in sera of nude mice with orthotopic human pancreatic cancer and in plasma of patients with pancreatic cancer. *Int. J. Oncol.* **2005**, *27*, 1361–1369.
58. Cho, W.C.; Yip, T.T.; Yip, C.; Yip, V.; Thulasiraman, V.; Ngan, R.K.; Yip, T.T.; Lau, W.H.; Au, J.S.; Law, S.C.; Cheng, W.W.; Ma, V.W.; Lim, C.K. Identification of serum amyloid a protein as a

- potentially useful biomarker to monitor relapse of nasopharyngeal cancer by serum proteomic profiling. *Clin. Cancer Res.* **2004**, *10*, 43–52.
59. Khan, N.; Cromer, C.J.; Campa, M.; Patz, E.F., Jr. Clinical utility of serum amyloid A and macrophage migration inhibitory factor as serum biomarkers for the detection of nonsmall cell lung carcinoma. *Cancer* **2004**, *101*, 379–384.
60. Cocco, E.; Bellone, S.; El-Sahwi, K.; Cargnelutti, M.; Casagrande, F.; Buza, N.; Tavassoli, F.A.; Siegel, E.R.; Visintin, I.; Ratner, E.; Silasi, D.A.; Azodi, M.; Schwartz, P.E.; Rutherford, T.J.; Pecorelli, S.; Santin, A.D. Serum amyloid A (SAA): a novel biomarker for uterine serous papillary cancer. *Br. J. Cancer* **2009**, *101*, 335–341.
61. Farias-Eisner, G.; Su, F.; Robbins, T.; Kotlerman, J.; Reddy, S.; Farias-Eisner, R. Validation of serum biomarkers for detection of early- and late-stage endometrial cancer. *Am. J. Obstet. Gynecol.* **2010**, *202*, 73.e1–e5.
62. Takano, M.; Kikuchi, Y.; Asakawa, T.; Goto, T.; Kita, T.; Kudoh, K.; Kigawa, J.; Sakuragi, N.; Sakamoto, M.; Sugiyama, T.; Yaegashi, N.; Tsuda, H.; Seto, H.; Shiwa, M. Identification of potential serum markers for endometrial cancer using protein expression profiling. *J. Cancer Res. Clin. Oncol.* **2010**, *136*, 475–481.
63. Zhu, L.R.; Zhang, W.Y.; Yu, L.; Zheng, Y.H.; Zhang, J.Z.; Liao, Q.P. Serum proteomic features for detection of endometrial cancer. *Int. J. Gynecol. Cancer* **2006**, *16*, 1374–1378.
64. Clarke-Pearson, D.L. Clinical practice. Screening for ovarian cancer. *N. Engl. J. Med.* **2009**, *361*, 170–177.
65. Lerner, J.P.; Timor-Tritsch, I.E.; Federman, A.; Abramovich, G. Transvaginal ultrasonographic characterization of ovarian masses with an improved, weighted scoring system. *Am. J. Obstet. Gynecol.* **1994**, *170*, 81–85.
66. Kupesic, S.; Plavsic, B.M. Early ovarian cancer: 3-D power Doppler. *Abdom. Imaging* **2006**, *31*, 613–619.
67. Jacobs, I.; Bast, R.C., Jr. The CA 125 tumour-associated antigen: a review of the literature. *Hum. Reprod.* **1989**, *4*, 1–12.
68. Nossov, V.; Amneus, M.; Su, F.; Lang, J.; Janco, J.M.; Reddy, S.T.; Farias-Eisner, R. The early detection of ovarian cancer: from traditional methods to proteomics. Can we really do better than serum CA-125? *Am. J. Obstet. Gynecol.* **2008**, *199*, 215–223.
69. Kudoh, K.; Kikuchi, Y.; Kita, T.; Tode, T.; Takano, M.; Hirata, J.; Mano, Y.; Yamamoto, K.; Nagata, I. Preoperative determination of several serum tumor markers in patients with primary epithelial ovarian carcinoma. *Gynecol. Obstet. Invest.* **1999**, *47*, 52–57.
70. Negishi, Y.; Furukawa, T.; Oka, T.; Sakamoto, M.; Hirata, T.; Okabe, K.; Matayoshi, K.; Akiya, K.; Soma, H. Clinical use of CA 125 and its combination assay with other tumor marker in patients with ovarian carcinoma. *Gynecol. Obstet. Invest.* **1987**, *23*, 200–207.
71. Gadducci, A.; Ferdeghini, M.; Prontera, C.; Moretti, L.; Mariani, G.; Bianchi, R.; Fioretti, P. The concomitant determination of different tumor markers in patients with epithelial ovarian cancer and benign ovarian masses: relevance for differential diagnosis. *Gynecol. Oncol.* **1992**, *44*, 147–154.
72. Koelma, I.A.; Nap, M.; Rodenburg, C.J.; Fleuren, G.J. The value of tumour marker CA 125 in surgical pathology. *Histopathology* **1987**, *11*, 287–294.

73. Yedema, C.; Massuger, L.; Hilgers, J.; Servaas, J.; Poels, L.; Thomas, C.; Kenemans, P. Pre-operative discrimination between benign and malignant ovarian tumors using a combination of CA125 and CA15.3 serum assays. *Int. J. Cancer* **1988**, *3*, 61–67.
74. Fioretti, P.; Gadducci, A.; Ferdeghini, M.; Prontera, C.; Malagnino, G.; Facchini, V.; Mariani, G.; Bianchi, R. The concomitant determination of different serum tumor markers in epithelial ovarian cancer: relevance for monitoring the response to chemotherapy and follow-up of patients. *Gynecol. Oncol.* **1992**, *44*, 155–160.
75. Negishi, Y.; Iwabuchi, H.; Sakunaga, H.; Sakamoto, M.; Okabe, K.; Sato, H.; Asano, G. Serum and tissue measurements of CA72-4 in ovarian cancer patients. *Gynecol. Oncol.* **1993**, *48*, 148–154.
76. Sawada, M.; Okudaira, Y.; Matsui, Y.; Shimizu, Y. Immunosuppressive acidic protein in patients with ovarian cancer. *Cancer* **1983**, *52*, 2081–2085.
77. Castelli, M.; Romano, P.; Atlante, G.; Pozzi, M.; Ferrini, U. Immunosuppressive acidic protein (IAP) and CA 125 assays in detection of human ovarian cancer: preliminary results. *Int. J. Biol. Markers* **1987**, *2*, 187–190.
78. Castelli, M.; Battaglia, F.; Scambia, G.; Panici, P.B.; Ferrandina, G.; Mileo, A.M.; Mancuso, S.; Ferrini, U. Immunosuppressive acidic protein and CA 125 levels in patients with ovarian cancer. *Oncology* **1991**, *48*, 13–17.
79. Li, J.; Dowdy, S.; Tipton, T.; Podratz, K.; Lu, W.G.; Xie, X.; Jiang, S.W. HE4 as a biomarker for ovarian and endometrial cancer management. *Expert Rev. Mol. Diagn.* **2009**, *9*, 555–566.
80. Urban, N. Specific keynote: Ovarian cancer risk assessment and the potential for early detection. *Gynecol. Oncol.* **2003**, *88*, S75–S79.
81. Xu, Y.; Shen, Z.; Wiper, D.W.; Wu, M.; Morton, R.E.; Elson, P.; Kennedy, A.W.; Belinson, J.; Markman, M.; Casey, G. Lysophosphatidic acid as a potential biomarker for ovarian and other gynecologic cancers. *JAMA* **1998**, *280*, 719–723.
82. Zhang, Z.; Bast, R.C., Jr.; Yu, Y.; Li, J.; Sokoll, L.J.; Rai, A.J.; Rosenzweig, J.M.; Cameron, B.; Wang, Y.Y.; Meng, X.Y.; Berchuck, A.; Van Haaften-Day, C.; Hacker, N.F.; de Bruijn, H.W.; van der Zee, A.G.; Jacobs, I.J.; Fung, E.T.; Chan, D.W. Three biomarkers identified from serum proteomic analysis for the detection of early stage ovarian cancer. *Cancer Res.* **2004**, *64*, 5882–5890.
83. Su, F.; Lang, J.; Kumar, A.; Ng, C.; Hsieh, B.; Suchard, M.A.; Reddy, S.T.; Farias-Eisner, R. Validation of candidate serum ovarian cancer biomarkers for early detection. *Biomark Insights* **2007**, *2*, 369–375.
84. Nosov, V.; Su, F.; Amneus, M.; Birrer, M.; Robins, T.; Kotlerman, J.; Reddy, S.; Farias-Eisner, R. Validation of serum biomarkers for detection of early-stage ovarian cancer. *Am. J. Obstet. Gynecol.* **2009**, *200*, 639.e1–e5.
85. Visintin, I.; Feng, Z.; Longton, G.; Ward, D.C.; Alvero, A.B.; Lai, Y.; Tenthorey, J.; Leiser, A.; Flores-Saaib, R.; Yu, H.; Azori, M.; Rutherford, T.; Schwartz, P.E.; Mor, G. Diagnostic markers for early detection of ovarian cancer. *Clin. Cancer Res.* **2008**, *14*, 1065–1072.

86. Anderson, G.L.; McIntosh, M.; Wu, L.; Barnett, M.; Goodman, G.; Thorpe, J.D.; Bergan, L.; Thornquist, M.D.; Scholler, N.; Kim, N.; O'Briant, K.; Drescher, C.; Urban, N. Assessing lead time of selected ovarian cancer biomarkers: a nested case-control study. *J. Natl. Cancer Inst.* **2010**, *102*, 26–38.

© 2010 by the authors; licensee MDPI, Basel, Switzerland. This article is an Open Access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/3.0/>).