

Review

The Mechanisms Underlying the Beneficial Impact of Aerobic Training on Cancer-Related Fatigue: A Conceptual Review

Adeline Fontvieille ^{1,2,3} , Hugo Parent-Roberge ^{1,2,3}, Tamás Fülöp ^{2,4}, Michel Pavic ^{3,4}  and Eléonor Riesco ^{1,2,3,*}

¹ Faculty of Physical Activity Sciences, University of Sherbrooke, 2500 Boulevard de l'Université, Sherbrooke, QC J1K 2R1, Canada; adeline.fontvieille@usherbrooke.ca (A.F.); hugo.parent-roberge@usherbrooke.ca (H.P.-R.)

² Research Centre on Aging, 1036 Rue Belvédère Sud, Sherbrooke, QC J1H 4C4, Canada; tamas.fulop@usherbrooke.ca

³ Institut de Recherche sur le Cancer de l'Université de Sherbrooke, 12e Avenue N Porte 6, Sherbrooke, QC J1H 5N4, Canada; michel.pavic@usherbrooke.ca

⁴ Faculty of Medicine and Health Sciences, University of Sherbrooke, 3001 12e Avenue N, Sherbrooke, QC J1H 5N4, Canada

* Correspondence: e.riesco@usherbrooke.ca

Simple Summary: Cancer-related fatigue is a prevalent symptom, with a significant impact on the daily lives of those affected. While physical exercise has demonstrated effectiveness in reducing the intensity and duration of fatigue, the literature still lacks sufficient evidence on the physiological mechanisms explaining this impact. This conceptual review aimed to provide an overview of the evidence regarding the effect of acute exercise on peripheral and neuronal inflammation, immune function, and the neuroendocrine system in the context of cancer. We aim to integrate these pathways into a conceptual model that can serve as a starting point for further research into the physiological mechanisms linking exercise and cancer-related fatigue.

Abstract: Cancer-related fatigue (CRF) is a prevalent and persistent issue affecting cancer patients, with a broad impact on their quality of life even years after treatment completion. The precise mechanisms underlying CRF remain elusive, yet its multifaceted nature involves emotional, physical, and cognitive dimensions. The absence of effective medical treatments has prompted researchers to explore integrative models for potential insights. Notably, physical exercise emerges as a promising strategy for managing CRF and related symptoms, as studies showed a reduction in CRF ranging from 19% to 40%. Current recommendations highlight aerobic training at moderate intensity as beneficial, although questions about a dose–response relationship and the importance of exercise intensity persist. Despite the positive impact of exercise on CRF, the underlying mechanisms remain elusive. This review aims to provide a theoretical model explaining how aerobic exercise may alleviate CRF. Focusing on acute exercise effects, this review delves into the potential influence on peripheral and neural inflammation, immune function dysregulation, and neuroendocrine system disruptions. The objective is to enhance our understanding of the intricate relationship between exercise and CRF, ultimately paving the way for tailored interventions and potential pharmacological treatments for individuals unable to engage in physical exercise.

Keywords: immune system; hypothalamic–pituitary–adrenal axis; inflammation; neuroinflammation; aerobic exercise; acute exercise



Citation: Fontvieille, A.; Parent-Roberge, H.; Fülöp, T.; Pavic, M.; Riesco, E. The Mechanisms Underlying the Beneficial Impact of Aerobic Training on Cancer-Related Fatigue: A Conceptual Review. *Cancers* **2024**, *16*, 990. <https://doi.org/10.3390/cancers16050990>

Academic Editor: David Wong

Received: 4 February 2024

Revised: 26 February 2024

Accepted: 27 February 2024

Published: 29 February 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Cancer-related fatigue (CRF) is a significant burden for patients [1,2], affecting 14% to 99% of them, depending on various factors such as cancer stage, treatment type, and assessment methods [3,4]. While recognized as a multidimensional syndrome encompassing emotional, physical, and cognitive manifestations [5], CRF is defined by the National

Comprehensive Cancer Network as “an unusual, persistent, subjective sense of tiredness related to cancer or cancer treatment that interferes with usual functioning” [6]. CRF can persist more than 10 years after the end of cancer treatments [7,8] and has a deleterious impact on health-related quality of life [9]. Usually, this fatigue sets in within a few days of treatment, peaking 1 to 3 days post-chemotherapy, and then gradually subsides, returning to values near those before treatment around 10 days post-chemotherapy (as shown in Figure 1). There is currently no effective medical treatment for CRF, mainly because its underlying mechanisms remain unclear. In the past decade, several groups have attempted to fill this gap by suggesting integrative models that consider potential mechanisms involved in CRF [10–14]. Despite the lack of established medical treatments, these models and the systemic impact of physical exercise suggest exercise as a promising and effective strategy for managing CRF [15] and other treatment-related symptoms [16]. In fact, while we observed a reduction in CRF for up to 19% in oncogeriatric patients with exercise training [17], others have reported a greater impact in adjuvant breast cancer with a 40% reduction [16]. Currently, it is suggested that aerobic training at moderate intensity (65% heart rate max [HRmax] or 45% VO₂ max) reduced CRF, while it remains unclear if there is a dose–response relationship and if exercise intensity matters [18–20]. Nevertheless, there is a paucity of data regarding the underlying mechanisms by which exercise training reduces CRF. Unveiling the mechanisms by which exercise reduces CRF is crucial for developing personalized interventions and potential pharmacological alternatives for individuals unable to exercise. Given that chronic exercise is the outcome of accumulating acute exercise effects [21], we can infer that the decrease observed after an exercise intervention may be explained by the repetitive impact of acute exercise effects on CRF (Figure 1).



Figure 1. Kinetics of cancer-related fatigue depending on the cycle of chemotherapy treatment and exercise. Figure 1 illustrates the kinetics of CRF in relation to chemotherapy treatment cycles and their cumulative effect. The orange curve depicts a characteristic pattern of CRF intensity following each treatment cycle. A peak in CRF is observed between 3 and 5 days post-treatment, followed by a recovery period in the subsequent days. Notably, the intensity of CRF progressively increases with the accumulation of treatments. The blue curve depicts the fatigue kinetics when physical exercise is incorporated during treatment cycles, highlighting the potential for chronic effects. Here, a marked reduction in CRF intensity is observed compared to the no-exercise scenario. Additionally, the cumulative anti-cancer treatment effect appears less pronounced with exercise intervention. This observation suggests that the acute effect of exercise, practiced within a treatment cycle, might contribute to a gradual attenuation of CRF over time, potentially reflecting the cumulative impact of acute exercise. Image created with [Biorender.com](https://www.biorender.com) (accessed on 6 December 2023).

While there is no accepted mechanistic model explaining the reduction in CRF induced by exercise, it would be possible to provide such a theoretical model by using available evidence from exercise studies performed with cancer patients, or other chronic diseases

like chronic fatigue syndrome [22], as well as healthy patients. In fact, among the suggested mechanisms involved in CRF etiology, some of them may be acutely influenced by aerobic exercise, such as peripheral pro-inflammatory state [23], immune function dysregulation [14,24], as well as neuroendocrine dysregulations [25,26]. Hence, the objective of this conceptual review is to provide a theoretical model explaining how aerobic exercise may reduce CRF. Therefore, this review will mostly focus on the acute effect of exercise on peripheral and neural inflammation, as well as dysregulation of the immune function and neuroendocrine system. Nevertheless, a brief review of the current literature regarding potential mechanisms explaining CRF precedes the content section to facilitate understanding (for a more detailed review, please see [10,11,27]).

2. The Effects of Peripheral and Neural Inflammation on Cancer-Related Fatigue

2.1. Effect of Chemotherapeutic Treatments

Chemotherapy treatments, primarily used for cancer, trigger an acute response from both the innate and adaptive anti-tumor immune systems [28]. However, prolonged treatment and the accumulation of chemotherapeutic agents can lead to changes in the immune system within the tumor environment, including tumor cell death, and damage to healthy tissues due to chemicals. In response to this acute disturbance in the body's homeostatic, the immune system initiates a protective response by synthesizing cytokines. The objective is to reduce tissue damage by increasing the production of pro-inflammatory cytokines and chemokines, which help stimulate and release lymphocytes and monocytes in the affected tissue. This acute inflammatory response leads to an increase in both pro- and anti-inflammatory cytokines. However, with repeated chemotherapy treatments, this response becomes detrimental, leading to a maladaptive chronic inflammatory state. Studies have shown an increase in various inflammatory markers in the peripheral circulation during cancer [29], including IL-1, IL-6, IL-1 β , and TNF- α in different types of cancer [30–32]. Notably, this rise of inflammatory cytokines coincides with a peak in CRF levels, as shown by Raudonis et al. [33].

Progressively, the repeated elevation of pro-inflammatory cytokines after each treatment cycle triggers a rise in circulating anti-inflammatory cytokines [34,35], ultimately contributing to the maladaptive chronic inflammation mentioned above. This peripheral inflammation disrupts the blood–brain barrier (BBB) and activates the microglia, the resident macrophage population responsible for immune defense in the central nervous system (CNS) [36]. Repeated microglial activation initiates a harmful cycle, leading to the production of more pro-inflammatory cytokines and the activation of neurotoxic reactive astrocytes—both contributing to neuroinflammation [37]. Additionally, the compromised BBB allows for increased permeability [37]. It was previously shown that chemotherapeutic substances can reach and cross the BBB due to a negative impact of some pro-inflammatory cytokines (IL-1 α and IL-1 β) on tight junctions [38,39], and both IFN- γ and TNF- α can alter BBB permeability by affecting the expression and cellular distribution of junctional adhesion molecules (i.e., ICAM-1 and VCAM-1) [40]. This disruption enables an elevated number of blood-derived molecules and cells, such as activated T cells and B-cells, to enter the CNS and initiate neurodegeneration that could promote CRF. Microglia activation can also initiate an inflammatory cascade that eventually alters the metabolic pathways within CNS such as the kynurenine pathway, ultimately leading to impairments in the releasing of neurotransmitters such as serotonin and especially dopamine [41,42]. It is well established that several chemotherapeutics agents can negatively impact the hippocampal region, leading to a decreased cell proliferation or increased cell death [43]. This effect can manifest as behavioral changes, including cognitive, emotional, and spatial impairment in cancer patients. Dopamine signaling plays a crucial role in influencing motivational states [44], which, in turn, can impact hippocampal function. Dysfunction in any of these components can potentially lead to alterations in motivational behaviors and cognitive processes.

2.2. The Acute Effect of Aerobic Exercise on Peripheral and Neural Inflammation

Aerobic exercise can have a beneficial acute effect on the peripheral inflammatory profile. During exercise, a rise in pro-inflammatory cytokines occurs while a switch takes place after the end of exercise, with an increase in circulating anti-inflammatory cytokines [45]. More precisely, according to the intensity [46] and duration of aerobic exercise, many inflammatory myokines increase, especially IL-6 [47]. In response to a short bout of aerobic exercise, it was proposed that the release of myokine IL-6 could upregulate the production of anti-inflammatory cytokines, including IL-10, IL-1Ra, and TGF- β , which would dampen the inflammatory response for several hours after exercise [48,49]. When these anti-inflammatory cytokines are carried into the central nervous system (CNS), they might help reduce neural inflammation [50], potentially leading to changes in behavior. Indeed, in a large meta-analysis of 18 studies [51], evidence showed that acute moderate-intensity exercise led to an increase in anti-inflammatory cytokines, with a marked response observed after high-intensity exercise. However, it is not clear whether this increase in post-exercise anti-inflammatory cytokines leads to a decrease in CRF, as no study has directly measured this relationship. Therefore, because high peripheral levels of anti-inflammatory cytokines inhibit microglial activation by interacting with brain cytokine receptors [52], it may be possible that during the following hours after aerobic exercise, the neural activation is reduced, which could explain the lower CRF observed in response to exercise.

The BBB also benefits from aerobic exercise, notably by restoring permeability and re-establishing the expression of tight junction transmembrane proteins [53]. This effect was observed in a mouse model of multiple sclerosis, where endurance exercise led to a significant decrease in IFN- γ and IL-1 β production within the CNS, contributing to the protection of the BBB [53]. These findings imply that aerobic exercise can help restore the function of the BBB, which, in turn, reduces inflammation in the CNS. This prevents the entry of neurotoxic metabolites from the kynurenine pathway into the CNS, potentially resulting in a decrease in CRF.

3. Immune Function Dysregulation and Cancer-Related Fatigue

3.1. Immune Response after Chemotherapy

While the precise role of immune cells in the development of CRF is not yet fully understood, recent evidence indicates that immune system dysregulation, including the activity of natural killer (NK) cells and T lymphocytes, might be involved in the onset and persistence of CRF. T cells, specifically CD4⁺ and CD8⁺, are key players in adaptive immunity and have direct antitumor effects [54,55]. CD4⁺ T cells, also known as T helper, help coordinate immune responses by releasing cytokines and activating other immune cells. CD8⁺ T cells, also called cytotoxic T cells, directly kill infected or cancerous cells. Dysregulation of T lymphocytes, including a shift in the balance between different T cell subsets and altered cytokine production, has been observed in myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS) patients [56]. These dysfunctions can result in impaired immune responses against cancer cells, which can contribute to disease progression and fatigue. However, there is currently no evidence linking T-cell dysfunction in cancer patients to CRF. Moreover, research on ME/CFS patients demonstrated the reduced activity of NK cells [57,58], which has also been reported in cancer patients experiencing fatigue. Two distinct populations of NK cells can be found in peripheral circulation based on their cell surface density, the CD56^{bright} (immature) and CD56^{dim} (cytotoxic). A recent study showed a slight but significant decrease in the population of cytotoxic NK cells (CD56^{dim}) in patients with CRF compared to a non-fatigued group [24], suggesting a potential involvement of impaired NK cell function in CRF. However, the reason for reduced NK cell cytotoxic activity in cancer patients with CRF is yet to be determined.

Altogether, these dysregulations can promote the release of pro-inflammatory cytokines and other signaling molecules, which then contribute to a state of chronic inflammation. The persistent activation of the immune system and the resulting inflammatory

response can induce CRF by affecting the CNS, altering neurotransmitter levels, and disrupting normal energy metabolism.

3.2. Acute Effect of Aerobic Exercise on Immune Markers

Growing evidence suggests that aerobic exercise may improve CRF by modulating the immune system. Research has shown that exercise improves immunosurveillance by increasing the number of lymphocytes in the peripheral circulation during exercise in healthy adults [59], a key component of the immune response, although this effect diminishes after cessation. While we recently finished a proof-of-concept study investigating the relationship between acute exercise-induced immune response and CRF (clinicalTrials.gov ID: NCT04715061), it was previously reported that a single bout of moderate-intensity aerobic exercise is sufficient to enhance the mobilization of NK cells and recruitment of cytotoxic T cells (CD8⁺) [60], with both implicated in immune surveillance. Since NK cell [61] and T cell [62] dysfunction has been linked to fatigue in various populations, it may be possible that exercise-induced improvements in immune function contribute to reduced CRF.

Supporting this notion, studies by Campbell et al. [60] and others [63,64] have demonstrated that high-intensity aerobic exercise (85% peak power output) elicits a transient greater lymphocyte mobilization in the peripheral circulation compared to moderate-intensity in healthy individuals, then returning to baseline values after 60 min post-exercise. Moreover, our preliminary results (under review) further suggests that aerobic exercise, particularly high-intensity exercise in fitter individuals, can promote acute lymphocyte mobilization in the peripheral circulation in metastatic cancer patients [65]. Additionally, high-intensity exercise has been shown to induce a more pronounced increase in NK cell cytotoxic activity compared to light- or moderate-intensity aerobic exercise [66,67]. This transient mobilization of cytotoxic NK cells in peripheral blood might be transported into the tumor microenvironment. However, further research is necessary to comprehensively characterize the extent of immune modulation induced by different aerobic exercise intensities in cancer patients and to definitively establish the link between these changes and CRF.

4. Neuroendocrine Alteration and Cancer-Related Fatigue

4.1. The Influence of Chemotherapeutic Agents on the Neuroendocrine System

Neuroendocrine alterations may also contribute to CRF as reviewed by O'Higgins et al. [27]. In fact, chronic inflammation tends to reduce the synthesis and release of corticotropin-releasing hormone (CRH) [27,68], which is a central regulator of the hypothalamic–pituitary–adrenal (HPA) axis. This disrupts the HPA axis, which negatively impacts the regulation of stress hormone cortisol's synthesis and release. Furthermore, studies have observed either resistance or sensitivity to glucocorticoids [69], suggesting a potential disruption in the negative feedback loop regulating CR, ACTH, and cortisol levels. This impaired regulation may lead to the HPA system releasing cortisol at a constant level throughout the day in cancer patients. Supporting this notion, a study conducted by Bower et al. [70] reported higher daytime cortisol levels with a blunted circadian rhythm (flatter slope) compared to normal peak times (morning and evening), suggesting dysfunction of the HPA axis in patients with CRF. However, the relationship between HPA dysfunction and CRF is still controversial. It is worth noting that some cancer treatments, such as glucocorticoids and certain chemotherapy drugs, can affect the HPA axis, potentially contributing to CRF. However, this appears to be more common when glucocorticoid withdrawal coincides with adrenal insufficiency [71].

Interestingly, indoleamine-2,3-dioxygenase (IDO) is a counter-regulatory enzyme that contributes to immune suppression in the tumor microenvironment [72]. By numerous mechanisms reviewed in Johnson et al. [73], greater activation of IDO expression stimulates tryptophan catabolism, resulting in increased circulating levels of kynurenine (KYN) to the detriment of serotonin [74]. Knowing that serotonin regulates upstream CRH signaling

systems [75], a decrease in serotonin levels might reduce the activity of the HPA axis and impair cortisol production [27,76]. Studies conducted in cancer patients have shown a correlation between IDO activity and CRF severity, indicating that a low tryptophan concentration or a high ratio of kynurenine/tryptophan was correlated with high CRF or lethargic behaviors. IDO activity is closely linked to the immune system, as the peripheral mononuclear cells have been shown to be potent producers of IDO [77]. In an interesting way, KYN promotes the generation of T_{reg} cells and also inhibits the proliferation of NK cells, B cells, and $CD4^+$ and $CD8^+$ lymphocytes [78], which can be argued to play a role in the reduction of immune defense. Similarly, the hypothesis of dopaminergic imbalance appears to be akin to serotonin dysregulation. In this context, both low and excessive levels of dopamine can induce fatigue in individuals with multiple sclerosis [42]. In this scenario, T cells, especially $CD4^+$ cells, can breach the BBB in the CNS and trigger the production of $IFN-\gamma$, which, in turn, inhibits dopamine production and can even lead to the destruction of dopamine neurons. Finally, as the kynurenine metabolites reach the BBB and enter the CNS, they become metabolized into quinolinic acid by glial cells (e.g., microglia) and cause neurotoxicity, potentially leading to CRF.

4.2. The Acute Effect of Aerobic Exercise on the Neuroendocrine System

In healthy humans, the acute response to moderate-to-vigorous exercise ($>60\%$ of VO_2 max) stimulates the HPA axis by increasing adrenocorticotrophic hormone (ACTH) and cortisol levels [42]. Nevertheless, two studies on cancer survivors revealed that during moderate-intensity exercise (60% VO_2 peak), the magnitude of the increase in cortisol and ACTH blood concentration is smaller [79,80], mainly because of the higher baseline levels observed in cancer patients compared to healthy individuals. Hence, exercise-induced HPA axis activation may be somewhat limited in cancer survivors. However, to which extent aerobic training performed over several weeks in cancer patients during treatment can help restore normal baseline circulating cortisol and ACTH levels remains to be investigated.

The sensitivity of glucocorticoid receptors, principal inflammatory regulators of the HPA axis, is also affected by aerobic exercise. In fact, it was demonstrated that exercise acutely increased tissue sensitivity to glucocorticoids, which could be explained by an increasing number of glucocorticoid receptors or a shift in their isoform expression [81]. Hence, cortisol has more potential to bind to glucocorticoid receptors, which would restore appropriate negative feedback. It could then be hypothesized that exercise allows for the recovery of diurnal cortisol variations via the enhancement of glucocorticoid receptors sensitivity, thus providing a negative feedback loop.

At the central level, exercise acutely acts on dopaminergic and serotonergic pathways, which are suggested to be contributors to CRF. Acute exercise increases the bioavailability of free tryptophan in the brain, along with an increase in tryptophan hydroxylase, the enzyme that converts tryptophan into serotonin. By doing this, exercise leads to an increase in serotonin levels in the hours after exercise [82]. Even though there are still discrepancies in the literature regarding the required intensity of exercise to trigger increased cerebral serotonin levels in cancer patients, it is accepted that high-intensity exercise promotes higher serotonin and dopamine levels [83]. An adequate concentration of these two neurotransmitters ensures their optimal function and could consequently decrease CRF.

IDO activation, which was previously associated with a high level of CRF [84,85], could also be influenced by aerobic exercise. Studies investigating the acute effect of aerobic exercise showed an increase in kynurenine concentration in circulation [86,87]. This is mainly explained by the transient elevation of pro-inflammatory cytokines during exercise [88]. However, the acute anti-inflammatory impact of exercise in the following hour could also decrease IDO activity [89] and thus restore normal tryptophan metabolism. Regarding BBB permeability, acute exercise suggests the re-establishment of the BBB function and permeability and might prevent the crossing of neurotoxic metabolites from the kynurenine pathway to the CNS [36]. However, little evidence is available on this phenomenon in acute exercise in cancer patients.

5. Physiological Variability in Exercise Response

Inter-individual variability in physiological responses following exercise is observed in both healthy populations and cancer patients [90] and is influenced by a multitude of factors such as genetics, baseline fitness levels, age, and demographic variables like sex and ethnicity. Studies have demonstrated a wide range of responses to exercise, with individuals experiencing either increased fatigue or improvements in factors like cardiorespiratory fitness. Among the explaining factors, a genetic component likely contributes, as twin and family studies revealed a genetic component explaining 30 to 60% of the variation in cardiorespiratory fitness response to exercise [91]. Distinct responses observed between “low” and “high” responders in animal experiments provide evidence for genetic factors influencing training adaptations and thus possibly the impact of a single exercise session. Additionally, the type, intensity, duration, and timing of exercise play a significant role in shaping the outcome. Inconsistency in exercise prescription across studies hinders the comparison of true variability in response to exercise. Studies have employed various approaches for standardization, including fixed duration, intensity, or caloric expenditure targets. Most of the existing literature fails to report exercise interventions following CERT guidelines, often only considering adherence, defined as exercise session attendance, solely. Moreover, monitoring methods like heart rate can be affected by cardiovascular drift and pharmacological treatment, potentially leading to underestimating the actual workload. Beyond these factors, behavioral aspects like overall physical activity levels, dietary habits, and sleep quality contribute to the variability observed in exercise responses. This is particularly relevant for cancer patients due to their unique characteristics, including pre-existing comorbidities, fitness levels, and specific cancer diagnoses (e.g., initial cancer type, the presence and location of metastases, and treatment regimen).

Overall, the hypothetical mechanisms by which aerobic exercise can decrease CRF are presented in Figure 2. Keeping in mind that this is a proposed theoretical model, the scientific literature on chronic fatigue syndrome provides a good starting point for future studies.

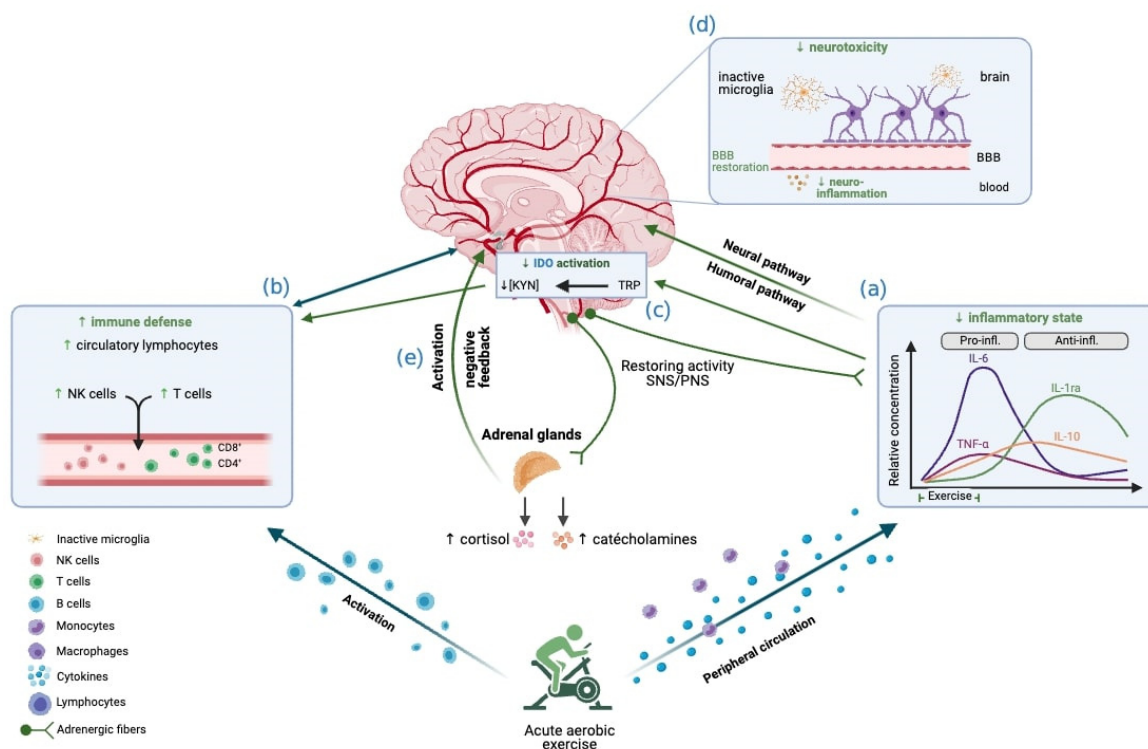


Figure 2. Effects of acute aerobic exercise on CRF, integrated into the conceptual model. The reduction in CRF observed following aerobic exercise may be explained by these mechanisms: Aerobic exercise,

particularly when performed at high intensity, is associated with an anti-inflammatory effect during the first few hours after exercise, possibly transiently alleviating chronic peripheral inflammation (a). Aerobic exercise promotes the mobilization of circulating lymphocytes, thus helping to restore immune function (b). This reduction in inflammation would lead to a reduction in the activation of IDO (c), with effects accentuated by vigorous intensity. The restoration of BBB function prevents the passage of pro-inflammatory cytokines into the CNS, which decreases microglia activation and therefore prevents neurotoxic damage (d). Aerobic exercise manages to re-regulate the activation of the HPA axis (e), notably by recovering the amplitude of cortisol release, while promoting a better sensitivity to glucocorticoids and restoring the negative feedback loop. Image created with [Biorender.com](https://www.biorender.com) (accessed on 22 September 2022).

6. Conclusions

Although the underlying mechanisms explaining how aerobic exercise impacts CRF are still under investigation, this conceptual review proposes an explanatory theoretical model of exercise based on the suggested etiology of CRF. While this review focuses on the physiological aspects and immediate impacts of chemotherapy treatment, shedding light on these aspects, the etiology of CRF is intricate and involves multiple factors, such as type of treatment, type of cancer, and chemotherapeutic agents. Additional factors mentioned in the literature may account for other aspects of CRF, such as physical deconditioning, depression, and cachexia. Understanding these biological mechanisms is crucial for developing personalized exercise interventions for cancer patients and mitigating the impact of this significant side effect on their quality of life. Further studies and a proof of concept are required to confirm or refute this model and assess its applicability based on the specifics of cancer treatment, such as the type and drugs used. Moreover, knowing that aerobic exercise is currently one of the most efficient strategies to reduce CRF, a better understanding of the underlying mechanisms would offer the opportunity to develop pharmacological treatments for individuals who cannot exercise or achieve a sufficient exercise workload to benefit from this type of intervention. By enhancing our knowledge of these mechanisms, more precise exercise prescriptions could help patients increase their survival rate after diagnosis or reduce the risk of recurrence [92], as well as alleviate treatment-related side effects such as CRF [93], initiating a virtuous circle. Establishing recommendations based on new evidence regarding the comprehension of CRF mechanisms will provide important leverage in implementing exercise as an integral part of the healthcare pathway for individuals with cancer.

Author Contributions: Conceptualization, A.F. and E.R.; methodology, A.F.; writing—original draft preparation, A.F.; writing—review and editing, A.F., H.P.-R., T.F., M.P. and E.R.; supervision, M.P. and E.R. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Fondation Sylvain Poissant and the APC was funded by the Fondation Sylvain Poissant and Faculty of Physical Activity Sciences, University of Sherbrooke. A.F. and H.P.-R. are supported by the Fonds de Recherche du Québec—Santé. E.R. is supported by the Research Chair for Healthy Aging—B. Gravel and JL Gravel Foundation.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Gerber, L.H. Cancer-Related Fatigue: Persistent, Pervasive, and Problematic. *Phys. Med. Rehabil. Clin. N. Am.* **2017**, *28*, 65–88. [[CrossRef](#)]
2. Wang, X.S.; Woodruff, J.F. Cancer-Related and Treatment-Related Fatigue. *Gynecol. Oncol.* **2015**, *136*, 446–452. [[CrossRef](#)]
3. Schmidt, M.E.; Hermann, S.; Arndt, V.; Steindorf, K. Prevalence and Severity of Long-Term Physical, Emotional, and Cognitive Fatigue across 15 Different Cancer Entities. *Cancer Med.* **2020**, *9*, 8053–8061. [[CrossRef](#)]
4. Ma, Y.; He, B.; Jiang, M.; Yang, Y.; Wang, C.; Huang, C.; Han, L. Prevalence and Risk Factors of Cancer-Related Fatigue: A Systematic Review and Meta-Analysis. *Int. J. Nurs. Stud.* **2020**, *111*, 103707. [[CrossRef](#)]
5. Jacobsen, P.B. Assessment of Fatigue in Cancer Patients. *J. Natl. Cancer Inst. Monogr.* **2004**, *2004*, 93–97. [[CrossRef](#)]
6. Mock, V.; Atkinson, A.; Barsevick, A.; Cella, D.; Cimprich, B.; Cleeland, C.; Donnelly, J.; Eisenberger, M.A.; Escalante, C.; Hinds, P.; et al. NCCN Practice Guidelines for Cancer-Related Fatigue. *Oncol. Williston Park N. Y.* **2000**, *14*, 151–161.

7. Husson, O.; Mols, F.; van de Poll-Franse, L.; de Vries, J.; Schep, G.; Thong, M.S.Y. Variation in Fatigue among 6011 (Long-Term) Cancer Survivors and a Normative Population: A Study from the Population-Based PROFILES Registry. *Support. Care Cancer Off. J. Multinat. Assoc. Support. Care Cancer* **2015**, *23*, 2165–2174. [[CrossRef](#)]
8. Reinertsen, K.V.; Cvancharova, M.; Loge, J.H.; Edvardsen, H.; Wist, E.; Fosså, S.D. Predictors and Course of Chronic Fatigue in Long-Term Breast Cancer Survivors. *J. Cancer Surviv. Res. Pract.* **2010**, *4*, 405–414. [[CrossRef](#)] [[PubMed](#)]
9. Arndt, V.; Koch-Gallenkamp, L.; Jansen, L.; Bertram, H.; Eberle, A.; Holleczeck, B.; Schmid-Höpfner, S.; Waldmann, A.; Zeissig, S.R.; Brenner, H. Quality of Life in Long-Term and Very Long-Term Cancer Survivors versus Population Controls in Germany. *Acta Oncol. Stockh. Swed.* **2017**, *56*, 190–197. [[CrossRef](#)] [[PubMed](#)]
10. Ryan, J.L.; Carroll, J.K.; Ryan, E.P.; Mustian, K.M.; Fiscella, K.; Morrow, G.R. Mechanisms of Cancer-Related Fatigue. *Oncologist* **2007**, *12* (Suppl. S1), 22–34. [[CrossRef](#)] [[PubMed](#)]
11. Yang, S.; Chu, S.; Gao, Y.; Ai, Q.; Liu, Y.; Li, X.; Chen, N. A Narrative Review of Cancer-Related Fatigue (CRF) and Its Possible Pathogenesis. *Cells* **2019**, *8*, 738. [[CrossRef](#)] [[PubMed](#)]
12. LaVoy, E.C.P.; Fagundes, C.P.; Dantzer, R. Exercise, Inflammation, and Fatigue in Cancer Survivors. *Exerc. Immunol. Rev.* **2016**, *22*, 82–93. [[PubMed](#)]
13. Barsevick, A.; Frost, M.; Zwinderman, A.; Hall, P.; Halyard, M.; GENEQOL Consortium. I’m so Tired: Biological and Genetic Mechanisms of Cancer-Related Fatigue. *Qual. Life Res. Int. J. Qual. Life Asp. Treat. Care Rehabil.* **2010**, *19*, 1419–1427. [[CrossRef](#)] [[PubMed](#)]
14. Bower, J.E. Cancer-Related Fatigue—Mechanisms, Risk Factors, and Treatments. *Nat. Rev. Clin. Oncol.* **2014**, *11*, 597–609. [[CrossRef](#)] [[PubMed](#)]
15. van Vulpen, J.K.; Sweegers, M.G.; Peeters, P.H.M.; Courneya, K.S.; Newton, R.U.; Aaronson, N.K.; Jacobsen, P.B.; Galvão, D.A.; Chinapaw, M.J.; Steindorf, K.; et al. Moderators of Exercise Effects on Cancer-Related Fatigue: A Meta-Analysis of Individual Patient Data. *Med. Sci. Sports Exerc.* **2020**, *52*, 303–314. [[CrossRef](#)] [[PubMed](#)]
16. Cramp, F.; Byron-Daniel, J. Exercise for the Management of Cancer-Related Fatigue in Adults. *Cochrane Database Syst. Rev.* **2012**, *11*, CD006145. [[CrossRef](#)]
17. Fontvieille, A.; Parent-Roberge, H.; Maréchal, R.; Fülöp, T.; Riesco, E.; Pavic, M. Corrigendum to “Effects of a Mixed Exercise Program on Cancer Related-Fatigue and Health-Related Quality of Life in Oncogeriatric Patients: A Feasibility Study” [Journal of Geriatric Oncology 12 Issue 6 (2021) 915–921]. *J. Geriatr. Oncol.* **2022**, *13*, 1070. [[CrossRef](#)] [[PubMed](#)]
18. Lipsett, A.; Barrett, S.; Haruna, F.; Mustian, K.; O’Donovan, A. The Impact of Exercise during Adjuvant Radiotherapy for Breast Cancer on Fatigue and Quality of Life: A Systematic Review and Meta-Analysis. *Breast Edinb. Scotl.* **2017**, *32*, 144–155. [[CrossRef](#)] [[PubMed](#)]
19. Campbell, K.L.; Winters-Stone, K.M.; Wiskemann, J.; May, A.M.; Schwartz, A.L.; Courneya, K.S.; Zucker, D.S.; Matthews, C.E.; Ligibel, J.A.; Gerber, L.H.; et al. Exercise Guidelines for Cancer Survivors: Consensus Statement from International Multidisciplinary Roundtable. *Med. Sci. Sports Exerc.* **2019**, *51*, 2375–2390. [[CrossRef](#)]
20. Mustian, K.M.; Cole, C.L.; Lin, P.J.; Asare, M.; Fung, C.; Janelins, M.C.; Kamen, C.S.; Peppone, L.J.; Magnuson, A. Exercise Recommendations for the Management of Symptoms Clusters Resulting from Cancer and Cancer Treatments. In *Seminars in Oncology Nursing*; Elsevier: Amsterdam, The Netherlands, 2016; Volume 32, pp. 383–393.
21. Farrell, C.; Turgeon, D.R. Normal Versus Chronic Adaptations to Aerobic Exercise. In *StatPearls*; StatPearls Publishing: Treasure Island, FL, USA, 2022.
22. Rusin, A.; Seymour, C.; Cocchetto, A.; Mothersill, C. Commonalities in the Features of Cancer and Chronic Fatigue Syndrome (CFS): Evidence for Stress-Induced Phenotype Instability? *Int. J. Mol. Sci.* **2022**, *23*, 691. [[CrossRef](#)]
23. Bower, J.E.; Lamkin, D.M. Inflammation and Cancer-Related Fatigue: Mechanisms, Contributing Factors, and Treatment Implications. *Brain. Behav. Immun.* **2013**, *30*, S48–S57. [[CrossRef](#)] [[PubMed](#)]
24. Regan, J.; Feng, R.; Saligan, L. 4313 Impaired Natural Killer Cell Function May Be Associated with Cancer-Related Fatigue. *J. Clin. Transl. Sci.* **2020**, *4*, 8. [[CrossRef](#)]
25. Francesco, F.; Alice, N.; Filippo, G.; Daniela, R.; Enrico, B.; Emanuela, A. Stress Axis in the Cancer Patient: Clinical Aspects and Management. *Endocrines* **2021**, *2*, 502–513. [[CrossRef](#)]
26. Simó, M.; Navarro, X.; Yuste, V.J.; Bruna, J. Autonomic Nervous System and Cancer. *Clin. Auton. Res.* **2018**, *28*, 301–314. [[CrossRef](#)] [[PubMed](#)]
27. O’Higgins, C.M.; Brady, B.; O’Connor, B.; Walsh, D.; Reilly, R.B. The Pathophysiology of Cancer-Related Fatigue: Current Controversies. *Support. Care Cancer Off. J. Multinat. Assoc. Support. Care Cancer* **2018**, *26*, 3353–3364. [[CrossRef](#)]
28. Bracci, L.; Schiavoni, G.; Sistigu, A.; Belardelli, F. Immune-Based Mechanisms of Cytotoxic Chemotherapy: Implications for the Design of Novel and Rationale-Based Combined Treatments against Cancer. *Cell Death Differ.* **2014**, *21*, 15–25. [[CrossRef](#)]
29. Mantovani, A.; Barajon, I.; Garlanda, C. IL-1 and IL-1 Regulatory Pathways in Cancer Progression and Therapy. *Immunol. Rev.* **2018**, *281*, 57–61. [[CrossRef](#)]
30. Vyas, D.; Laput, G.; Vyas, A.K. Chemotherapy-Enhanced Inflammation May Lead to the Failure of Therapy and Metastasis. *OncoTargets Ther.* **2014**, *7*, 1015–1023. [[CrossRef](#)] [[PubMed](#)]
31. Collado-Hidalgo, A.; Bower, J.E.; Ganz, P.A.; Cole, S.W.; Irwin, M.R. Inflammatory Biomarkers for Persistent Fatigue in Breast Cancer Survivors. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **2006**, *12*, 2759–2766. [[CrossRef](#)]

32. Parent-Roberge, H.; Fontvieille, A.; Maréchal, R.; Wagner, R.; Fülöp, T.; Pavic, M.; Riesco, E. Effects of Combined Exercise Training on the Inflammatory Profile of Older Cancer Patients Treated with Systemic Therapy. *Brain Behav. Immun. Health* **2019**, *2*, 100016. [[CrossRef](#)] [[PubMed](#)]
33. Raudonis, B.M.; Kelley, I.H.; Rowe, N.; Ellis, J. A Pilot Study of Proinflammatory Cytokines and Fatigue in Women with Breast Cancer During Chemotherapy. *Cancer Nurs.* **2017**, *40*, 323–331. [[CrossRef](#)] [[PubMed](#)]
34. Cruz, F.M.; Munhoz, B.A.; Alves, B.C.; Gehrke, F.S.; Fonseca, F.L.; Kuniyoshi, R.K.; Cubero, D.; Peppone, L.J.; del Giglio, A. Biomarkers of Fatigue Related to Adjuvant Chemotherapy for Breast Cancer: Evaluation of Plasma and Lymphocyte Expression. *Clin. Transl. Med.* **2015**, *4*, 4. [[CrossRef](#)] [[PubMed](#)]
35. Paulsen, Ø.; Laird, B.; Aass, N.; Lea, T.; Fayers, P.; Kaasa, S.; Klepstad, P. The Relationship between Pro-Inflammatory Cytokines and Pain, Appetite and Fatigue in Patients with Advanced Cancer. *PLoS ONE* **2017**, *12*, e0177620. [[CrossRef](#)]
36. Małkiewicz, M.A.; Szarmach, A.; Sabisz, A.; Cubala, W.J.; Szurowska, E.; Winklewski, P.J. Blood-Brain Barrier Permeability and Physical Exercise. *J. Neuroinflamm.* **2019**, *16*, 15. [[CrossRef](#)]
37. Michael, B.D.; Griffiths, M.J.; Granerod, J.; Brown, D.; Keir, G.; Wnek, M.; Cox, D.J.; Vidyasagar, R.; Borrow, R.; Parkes, L.M.; et al. The Interleukin-1 Balance During Encephalitis Is Associated with Clinical Severity, Blood-Brain Barrier Permeability, Neuroimaging Changes, and Disease Outcome. *J. Infect. Dis.* **2016**, *213*, 1651–1660. [[CrossRef](#)] [[PubMed](#)]
38. Roerink, M.E.; van der Schaaf, M.E.; Dinarello, C.A.; Knoop, H.; van der Meer, J.W.M. Interleukin-1 as a Mediator of Fatigue in Disease: A Narrative Review. *J. Neuroinflamm.* **2017**, *14*, 16. [[CrossRef](#)]
39. Wardill, H.R.; Mander, K.A.; Van Seville, Y.Z.A.; Gibson, R.J.; Logan, R.M.; Bowen, J.M.; Sonis, S.T. Cytokine-Mediated Blood Brain Barrier Disruption as a Conduit for Cancer/Chemotherapy-Associated Neurotoxicity and Cognitive Dysfunction. *Int. J. Cancer* **2016**, *139*, 2635–2645. [[CrossRef](#)]
40. Weiss, N.; Miller, F.; Cazaubon, S.; Couraud, P.-O. The Blood-Brain Barrier in Brain Homeostasis and Neurological Diseases. *Biochim. Biophys. Acta* **2009**, *1788*, 842–857. [[CrossRef](#)]
41. Capuron, L.; Miller, A.H. Immune System to Brain Signaling: Neuropsychopharmacological Implications. *Pharmacol. Ther.* **2011**, *130*, 226–238. [[CrossRef](#)]
42. Dobryakova, E.; Genova, H.M.; DeLuca, J.; Wylie, G.R. The Dopamine Imbalance Hypothesis of Fatigue in Multiple Sclerosis and Other Neurological Disorders. *Front. Neurol.* **2015**, *6*, 52. [[CrossRef](#)]
43. Dietrich, J.; Prust, M.; Kaiser, J. Chemotherapy, Cognitive Impairment and Hippocampal Toxicity. *Neuroscience* **2015**, *309*, 224–232. [[CrossRef](#)]
44. Hamid, A.A.; Pettibone, J.R.; Mabrouk, O.S.; Hetrick, V.L.; Schmidt, R.; Vander Weele, C.M.; Kennedy, R.T.; Aragona, B.J.; Berke, J.D. Mesolimbic Dopamine Signals the Value of Work. *Nat. Neurosci.* **2016**, *19*, 117–126. [[CrossRef](#)] [[PubMed](#)]
45. Petersen, A.M.W.; Pedersen, B.K. The Anti-Inflammatory Effect of Exercise. *J. Appl. Physiol.* **2005**, *98*, 1154–1162. [[CrossRef](#)] [[PubMed](#)]
46. Windsor, M.T.; Bailey, T.G.; Perissiou, M.; Meital, L.; Golledge, J.; Russell, F.D.; Askew, C.D. Cytokine Responses to Acute Exercise in Healthy Older Adults: The Effect of Cardiorespiratory Fitness. *Front. Physiol.* **2018**, *9*, 203. [[CrossRef](#)] [[PubMed](#)]
47. Fischer, C.P. Interleukin-6 in Acute Exercise and Training: What Is the Biological Relevance? *Exerc. Immunol. Rev.* **2006**, *12*, 6–33.
48. Nash, D.; Hughes, M.G.; Butcher, L.; Aicheler, R.; Smith, P.; Cullen, T.; Webb, R. IL-6 Signaling in Acute Exercise and Chronic Training: Potential Consequences for Health and Athletic Performance. *Scand. J. Med. Sci. Sports* **2023**, *33*, 4–19. [[CrossRef](#)]
49. Antunes, B.M.; Campos, E.Z.; Dos Santos, R.V.T.; Rosa-Neto, J.C.; Franchini, E.; Bishop, N.C.; Lira, F.S. Anti-Inflammatory Response to Acute Exercise Is Related with Intensity and Physical Fitness. *J. Cell. Biochem.* **2019**, *120*, 5333–5342. [[CrossRef](#)]
50. Mee-inta, O.; Zhao, Z.-W.; Kuo, Y.-M. Physical Exercise Inhibits Inflammation and Microglial Activation. *Cells* **2019**, *8*, 691. [[CrossRef](#)]
51. Cerqueira, É.; Marinho, D.A.; Neiva, H.P.; Lourenço, O. Inflammatory Effects of High and Moderate Intensity Exercise—A Systematic Review. *Front. Physiol.* **2020**, *10*, 1550. [[CrossRef](#)]
52. Cianciulli, A.; Dragone, T.; Calvello, R.; Porro, C.; Trotta, T.; Lofrumento, D.D.; Panaro, M.A. Lofrumento IL-10 Plays a Pivotal Role in Anti-Inflammatory Effects of Resveratrol in Activated Microglia Cells. *Int. Immunopharmacol.* **2015**, *24*, 369–376. [[CrossRef](#)]
53. Souza, P.S.; Gonçalves, E.D.; Pedroso, G.S.; Farias, H.R.; Junqueira, S.C.; Marcon, R.; Tuon, T.; Cola, M.; Silveira, P.C.L.; Santos, A.R.; et al. Physical Exercise Attenuates Experimental Autoimmune Encephalomyelitis by Inhibiting Peripheral Immune Response and Blood-Brain Barrier Disruption. *Mol. Neurobiol.* **2017**, *54*, 4723–4737. [[CrossRef](#)]
54. Miggelbrink, A.M.; Jackson, J.D.; Lorrey, S.J.; Srinivasan, E.S.; Waibl-Polania, J.; Wilkinson, D.S.; Fecci, P.E. CD4 T-Cell Exhaustion: Does It Exist and What Are Its Roles in Cancer? *Clin. Cancer Res.* **2021**, *27*, 5742–5752. [[CrossRef](#)]
55. Dolina, J.S.; Van Braeckel-Budimir, N.; Thomas, G.D.; Salek-Ardakani, S. CD8+ T Cell Exhaustion in Cancer. *Front. Immunol.* **2021**, *12*, 715234. [[CrossRef](#)]
56. Mandarano, A.H.; Maya, J.; Giloteaux, L.; Peterson, D.L.; Maynard, M.; Gottschalk, C.G.; Hanson, M.R. Myalgic Encephalomyelitis/Chronic Fatigue Syndrome Patients Exhibit Altered T Cell Metabolism and Cytokine Associations. *J. Clin. Investig.* **2020**, *130*, 1491–1505. [[CrossRef](#)] [[PubMed](#)]
57. Brenu, E.W.; van Driel, M.L.; Staines, D.R.; Ashton, K.J.; Hardcastle, S.L.; Keane, J.; Tajouri, L.; Peterson, D.; Ramos, S.B.; Marshall-Gradisnik, S.M. Longitudinal Investigation of Natural Killer Cells and Cytokines in Chronic Fatigue Syndrome/Myalgic Encephalomyelitis. *J. Transl. Med.* **2012**, *10*, 88. [[CrossRef](#)] [[PubMed](#)]

58. Ojo-Amaize, E.A.; Conley, E.J.; Peter, J.B. Decreased Natural Killer Cell Activity Is Associated with Severity of Chronic Fatigue Immune Dysfunction Syndrome. *Clin. Infect. Dis. Off. Publ. Infect. Dis. Soc. Am.* **1994**, *18* (Suppl. S1), S157–S159. [\[CrossRef\]](#) [\[PubMed\]](#)
59. Sand, K.L.; Flatebo, T.; Andersen, M.B.; Maghazachi, A.A. Effects of Exercise on Leukocytosis and Blood Hemostasis in 800 Healthy Young Females and Males. *World J. Exp. Med.* **2013**, *3*, 11–20. [\[CrossRef\]](#)
60. Campbell, J.P.; Riddell, N.E.; Burns, V.E.; Turner, M.; van Zanten, J.J.C.S.V.; Drayson, M.T.; Bosch, J.A. Acute Exercise Mobilises CD8+ T Lymphocytes Exhibiting an Effector-Memory Phenotype. *Brain. Behav. Immun.* **2009**, *23*, 767–775. [\[CrossRef\]](#)
61. Brenu, E.W.; Hardcastle, S.L.; Atkinson, G.M.; van Driel, M.L.; Kreijkamp-Kaspers, S.; Ashton, K.J.; Staines, D.R.; Marshall-Gradisnik, S.M. Natural Killer Cells in Patients with Severe Chronic Fatigue Syndrome. *Autoimmun. Highlights* **2013**, *4*, 69–80. [\[CrossRef\]](#)
62. Zhang, Z.; Liu, S.; Zhang, B.; Qiao, L.; Zhang, Y.; Zhang, Y. T Cell Dysfunction and Exhaustion in Cancer. *Front. Cell Dev. Biol.* **2020**, *8*, 17. [\[CrossRef\]](#)
63. Mathes, S.; Mester, J.; Bloch, W.; Wahl, P. Impact of High-Intensity and High-Volume Exercise on Short-Term Perturbations in the Circulating Fraction of Different Cell Types. *J. Sports Med. Phys. Fitness* **2017**, *57*, 130–137. [\[CrossRef\]](#)
64. Dorneles, G.P.; Haddad, D.O.; Fagundes, V.O.; Vargas, B.K.; Kloecker, A.; Romão, P.R.T.; Peres, A. High Intensity Interval Exercise Decreases IL-8 and Enhances the Immunomodulatory Cytokine Interleukin-10 in Lean and Overweight-Obese Individuals. *Cytokine* **2016**, *77*, 1–9. [\[CrossRef\]](#)
65. Parent-Roberge, H.; Fontvieille, A.; Tai, L.H.; Pavic, M.; Fülöp, T.; Riesco, E. Acute Nk Cells Response to Different Aerobic Exercise Modalities in Metastatic Cancer Patients Undergoing Chemotherapy. *Med. Sci. Sports Exerc.* **2023**, *55*, 479. [\[CrossRef\]](#)
66. Bigley, A.B.; Rezvani, K.; Chew, C.; Sekine, T.; Pistillo, M.; Crucian, B.; Bollard, C.M.; Simpson, R.J. Acute Exercise Preferentially Redeploys NK-Cells with a Highly-Differentiated Phenotype and Augments Cytotoxicity against Lymphoma and Multiple Myeloma Target Cells. *Brain. Behav. Immun.* **2014**, *39*, 160–171. [\[CrossRef\]](#)
67. Coletta, A.M.; Agha, N.H.; Baker, F.L.; Niemiro, G.M.; Mylabathula, P.L.; Brewster, A.M.; Bevers, T.B.; Fuentes-Mattei, E.; Basen-Engquist, K.; Gilchrist, S.C.; et al. The Impact of High-Intensity Interval Exercise Training on NK-Cell Function and Circulating Myokines for Breast Cancer Prevention among Women at High Risk for Breast Cancer. *Breast Cancer Res. Treat.* **2021**, *187*, 407–416. [\[CrossRef\]](#) [\[PubMed\]](#)
68. Saligan, L.N.; Olson, K.; Filler, K.; Larkin, D.; Cramp, F.; Yennurajalingam, S.; Sriram, Y.; Escalante, C.P.; del Giglio, A.; Kober, K.M.; et al. The Biology of Cancer-Related Fatigue: A Review of the Literature. *Support. Care Cancer Off. J. Multinat. Assoc. Support. Care Cancer* **2015**, *23*, 2461–2478. [\[CrossRef\]](#)
69. Olivas-Aguirre, M.; Torres-López, L.; Pottosin, I.; Dobrovinskaya, O. Overcoming Glucocorticoid Resistance in Acute Lymphoblastic Leukemia: Repurposed Drugs Can Improve the Protocol. *Front. Oncol.* **2021**, *11*, 617937. [\[CrossRef\]](#)
70. Bower, J.E.; Ganz, P.A.; Dickerson, S.S.; Petersen, L.; Aziz, N.; Fahey, J.L. Diurnal Cortisol Rhythm and Fatigue in Breast Cancer Survivors. *Psychoneuroendocrinology* **2005**, *30*, 92–100. [\[CrossRef\]](#)
71. Dinsen, S.; Baslund, B.; Klose, M.; Rasmussen, A.K.; Friis-Hansen, L.; Hilsted, L.; Feldt-Rasmussen, U. Why Glucocorticoid Withdrawal May Sometimes Be as Dangerous as the Treatment Itself. *Eur. J. Intern. Med.* **2013**, *24*, 714–720. [\[CrossRef\]](#) [\[PubMed\]](#)
72. Kim, S.; Miller, B.J.; Stefanek, M.E.; Miller, A.H. Inflammation-Induced Activation of the Indoleamine 2,3-Dioxygenase Pathway: Relevance to Cancer-Related Fatigue. *Cancer* **2015**, *121*, 2129–2136. [\[CrossRef\]](#)
73. Johnson, T.S.; McGaha, T.; Munn, D.H. Chemo-Immunotherapy: Impact of Indoleamine 2,3-Dioxygenase in Defining Immuno-genic versus Tolerogenic Cell Death in the Tumor Microenvironment. *Adv. Exp. Med. Biol.* **2017**, *1036*, 91–104. [\[CrossRef\]](#)
74. Lanser, L.; Kink, P.; Egger, E.M.; Willenbacher, W.; Fuchs, D.; Weiss, G.; Kurz, K. Inflammation-Induced Tryptophan Breakdown Is Related with Anemia, Fatigue, and Depression in Cancer. *Front. Immunol.* **2020**, *11*, 249. [\[CrossRef\]](#)
75. Heisler, L.K.; Pronchuk, N.; Nonogaki, K.; Zhou, L.; Raber, J.; Tung, L.; Yeo, G.S.H.; O’Rahilly, S.; Colmers, W.F.; Elmquist, J.K.; et al. Serotonin Activates the Hypothalamic-Pituitary-Adrenal Axis via Serotonin 2C Receptor Stimulation. *J. Neurosci. Off. J. Soc. Neurosci.* **2007**, *27*, 6956–6964. [\[CrossRef\]](#)
76. Neefjes, E.C.W.; van der Vorst, M.J.D.L.; Blauwhoff-Buskermolen, S.; Verheul, H.M.W. Aiming for a Better Understanding and Management of Cancer-Related Fatigue. *Oncologist* **2013**, *18*, 1135–1143. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Jones, S.P.; Franco, N.F.; Varney, B.; Sundaram, G.; Brown, D.A.; de Bie, J.; Lim, C.K.; Guillemin, G.J.; Brew, B.J. Expression of the Kynurenine Pathway in Human Peripheral Blood Mononuclear Cells: Implications for Inflammatory and Neurodegenerative Disease. *PLoS ONE* **2015**, *10*, e0131389. [\[CrossRef\]](#)
78. Frumento, G.; Rotondo, R.; Tonetti, M.; Damonte, G.; Benatti, U.; Ferrara, G.B. Tryptophan-Derived Catabolites Are Responsible for Inhibition of T and Natural Killer Cell Proliferation Induced by Indoleamine 2,3-Dioxygenase. *J. Exp. Med.* **2002**, *196*, 459–468. [\[CrossRef\]](#) [\[PubMed\]](#)
79. Evans, E.S.; Hackney, A.C.; Pebole, M.M.; McMurray, R.G.; Muss, H.B.; Deal, A.M.; Battaglini, C.L. Adrenal Hormone and Metabolic Biomarker Responses to 30 Min of Intermittent Cycling Exercise in Breast Cancer Survivors. *Int. J. Sports Med.* **2016**, *37*, 921–929. [\[CrossRef\]](#) [\[PubMed\]](#)
80. Evans, E.S.; Little, J.; McNeill, K.T.; Poole, C.; Bailey, S.P. Hypothalamic-Pituitary-Adrenal Axis Responses to an Acute Bout of Moderate Intensity Aerobic Exercise in Breast Cancer Survivors. *FASEB J.* **2019**, *33*, 534.3. [\[CrossRef\]](#)
81. Duclos, M.; Gouarne, C.; Bonnemaïson, D. Acute and Chronic Effects of Exercise on Tissue Sensitivity to Glucocorticoids. *J. Appl. Physiol.* **2003**, *94*, 869–875. [\[CrossRef\]](#)

82. Chaouloff, F. Effects of Acute Physical Exercise on Central Serotonergic Systems. *Med. Sci. Sports Exerc.* **1997**, *29*, 58–62. [[CrossRef](#)]
83. Basso, J.C.; Suzuki, W.A. The Effects of Acute Exercise on Mood, Cognition, Neurophysiology, and Neurochemical Pathways: A Review. *Brain Plast.* **2017**, *2*, 127–152. [[CrossRef](#)]
84. Kurz, K.; Fiegl, M.; Holzner, B.; Giesinger, J.; Pircher, M.; Weiss, G.; Denz, H.A.; Fuchs, D. Fatigue in Patients with Lung Cancer Is Related with Accelerated Tryptophan Breakdown. *PLoS ONE* **2012**, *7*, e36956. [[CrossRef](#)]
85. Lyon, D.E.; Walter, J.M.; Starkweather, A.R.; Schubert, C.M.; McCain, N.L. Tryptophan Degradation in Women with Breast Cancer: A Pilot Study. *BMC Res. Notes* **2011**, *4*, 156. [[CrossRef](#)]
86. Metcalfe, A.J.; Koliamitra, C.; Javelle, F.; Bloch, W.; Zimmer, P. Acute and Chronic Effects of Exercise on the Kynurenine Pathway in Humans—A Brief Review and Future Perspectives. *Physiol. Behav.* **2018**, *194*, 583–587. [[CrossRef](#)]
87. Joisten, N.; Walzik, D.; Metcalfe, A.J.; Bloch, W.; Zimmer, P. Physical Exercise as Kynurenine Pathway Modulator in Chronic Diseases: Implications for Immune and Energy Homeostasis. *Int. J. Tryptophan Res.* **2020**, *13*, 1178646920938688. [[CrossRef](#)] [[PubMed](#)]
88. Agudelo, L.Z.; Femenía, T.; Orhan, F.; Porsmyr-Palmertz, M.; Goiny, M.; Martinez-Redondo, V.; Correia, J.C.; Izadi, M.; Bhat, M.; Schuppe-Koistinen, I.; et al. Skeletal Muscle PGC-1 α Modulates Kynurenine Metabolism and Mediates Resilience to Stress-Induced Depression. *Cell* **2014**, *159*, 33–45. [[CrossRef](#)] [[PubMed](#)]
89. Joisten, N.; Kummerhoff, F.; Koliamitra, C.; Schenk, A.; Walzik, D.; Hardt, L.; Knoop, A.; Thevis, M.; Kiesl, D.; Metcalfe, A.J.; et al. Exercise and the Kynurenine Pathway: Current State of Knowledge and Results from a Randomized Cross-over Study Comparing Acute Effects of Endurance and Resistance Training. *Exerc. Immunol. Rev.* **2020**, *26*, 24–42. [[PubMed](#)]
90. Cormie, P.; Lamb, S.; Newton, R.; Valentine, L.; McKiernan, S.; Spry, N.; Joseph, D.; Taaffe, D.; Galvao, D. One Size Doesn't Fit All: Responders and Nonresponders to Exercise Interventions for Cancer Patients and Survivors. *Asia-Pac. J. Clin. Oncol.* **2017**, *13*, 85–86.
91. Bouchard, C.; Rankinen, T. Individual Differences in Response to Regular Physical Activity. *Med. Sci. Sports Exerc.* **2001**, *33*, S446–S451; discussion S452–S453. [[CrossRef](#)]
92. Morishita, S.; Hamaue, Y.; Fukushima, T.; Tanaka, T.; Fu, J.B.; Nakano, J. Effect of Exercise on Mortality and Recurrence in Patients with Cancer: A Systematic Review and Meta-Analysis. *Integr. Cancer Ther.* **2020**, *19*, 1534735420917462. [[CrossRef](#)]
93. Cormie, P.; Zopf, E.M.; Zhang, X.; Schmitz, K.H. The Impact of Exercise on Cancer Mortality, Recurrence, and Treatment-Related Adverse Effects. *Epidemiol. Rev.* **2017**, *39*, 71–92. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.