



Review Large B-Cell Lymphomas in the 5th Edition of the WHO-Classification of Haematolymphoid Neoplasms—Updated Classification and New Concepts

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Simple Summary: This paper provides an overview of the classification of large B-cell lymphomas in the 5th edition of the WHO classification of haematolymphoid tumors due to be published in 2023 and discusses new concepts, new entities, and the relevance of new genetic findings for the classification.

Abstract: The family/class of the large B-cell lymphomas (LBCL) in the 5th edition of the World Health Organization (WHO) classification of haematolymphoid tumors (WHO-HAEM5) features only a few major changes as compared to the 4th edition. In most entities, there are only subtle changes, many of them only representing some minor modifications in diagnostic terms. Major changes have been made in the diffuse large B-cell lymphomas (DLBCL)/high-grade B-cell lymphomas (HGBL) associated with *MYC* and *BCL2* and/or *BCL6* rearrangements. This category now consists of *MYC* and *BCL2* rearranged cases exclusively, while the *MYC/BCL6* double hit lymphomas now constitute genetic subtypes of DLBCL, not otherwise specified (NOS) or of HGBL, NOS. Other major changes are the conceptual merger of lymphomas arising in immune-privileged sites and the description of LBCL arising in the setting of immune dysregulation/deficiency. In addition, novel findings concerning underlying biological mechanisms in the pathogenesis of the different entities are provided.

Keywords: large B-cell lymphoma; diffuse large B-cell lymphoma; classification; genetics; WHO-HAEM5

1. Introduction

The family of large B-cell lymphomas (LBCL) is a heterogeneous class of tumors characterized by large lymphoid cells of the B-cell lineage that by definition form sheets or clusters. "Large cell" cytology is usually defined as a lymphoid cell with a nucleus that is larger than that of a macrophage or twice the size of a normal lymphocyte (Figure 1). Diffuse large B-cell lymphomas constitute the most common type of Non-Hodgkin lymphoma in the Western world, comprising 30-40% of cases with slight over-representation of the male gender [1–3]. The updated 5th edition of the World Health Organization (WHO) classification of haematolymphoid tumors (WHO-HAEM5), is currently available in a pre-published version [4] and online as a beta version and is expected to appear in print in summer 2023. It recognizes 17 specific entities of LBCL that can roughly be subdivided by morphology, genetic features, viral association, site of origin, and other, mostly overlapping, criteria (Table 1). The heterogeneous biology of this lymphoma family is reflected in the marked variability of clinical outcomes. In spite of the increasing complexity of the classification, displaying substantial biological differences between entities, rather monolithic chemotherapy regimens such as CHOP or related regimens combined with anti-CD20 antibodies still represent the standard of care in patients with symptomatic disease. Although a high proportion of patients achieve complete remission, therapy resistance or relapse



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still occurs in 30–40% of patients, thus evidencing an unmet need for the stratification of therapy in LBCL [2].

Figure 1. Diffuse large B-cell lymphoma, NOS (H&E \times 400). There is a diffuse effacement of the architecture by medium-sized and large blastic cells that have a moderately basophilic cytoplasm and round nuclei with light chromatin and one to several nucleoli.

 Table 1. Large B-Cell Lymphomas in WHO-HAEM5.

Diffuse large B-cell lymphoma, NOS
T-cell/histiocyte-rich large B-cell lymphoma
Diffuse large B-cell lymphoma/high-grade B-cell lymphoma with MYC and BCL2 rearrangements
ALK-positive large B-cell lymphoma
Large B-cell lymphoma with IRF4 rearrangement
High-grade B-cell lymphoma with 11q aberrations
Lymphomatoid granulomatosis
EBV-positive diffuse large B-cell lymphoma
Diffuse large B-cell lymphoma associated with chronic inflammation
Fibrin-associated large B-cell lymphoma
Fluid overload-associated large B-cell lymphoma
Plasmablastic lymphoma
Primary large B-cell lymphoma of immune-privileged sites
Primary cutaneous diffuse large B-cell lymphoma, leg type
Intravascular large B-cell lymphoma
Primary mediastinal large B-cell lymphoma
Mediastinal grey zone lymphoma
High-grade B-cell lymphoma, NOS

2. Diffuse Large B-Cell Lymphoma, Not Otherwise Specified

The etiology of DLBCL is multifaceted, and most likely several factors contribute to pathogenesis, in particular, genetic predisposition, immunodeficiency, and viral infection [5]. The large majority of diffuse large B-cell lymphoma, not otherwise specified (DLBCL, NOS), are of primary nodal origin and lack the criteria to be classified as one of the more specific entities. They most commonly involve lymph nodes but may also arise in extranodal lymphatic tissues such as in the tonsil or the Peyer patches of the gastrointestinal tract, or in extralymphatic tissues throughout the body [1,6]. Tumors with LBCL morphology may represent transformed disease from an indolent/small B-cell lymphoma, such as chronic lymphocytic leukemia, marginal zone lymphoma, or follicular lymphoma, or from nodular lymphocyte predominant Hodgkin lymphoma. However, the large majority of these cases arise de novo. WHO-HAEM5, for the first time, includes a separate chapter on "transformations of indolent lymphomas" specifically devoted to the clinical features, morphological traits, and genetic changes involved in the transformation.

The cytology of DLBCL can be variable. WHO-HAEM5 recognizes three morphological subtypes: centroblastic, immunoblastic, and anaplastic. Gene expression profiling (GEP) studies have identified two major profiles that can subdivide DLBCL, NOS into molecular subtypes named germinal center B-cell-like (GCB-like) and activated B-cell-like (ABC-like) with the former mainly expressing genes that are also expressed in cells in the reactive germinal center (GC), and the latter having a GEP of activated B-cells (DLBCL classification according to the cell of origin—COO) [7]. This concept was also retained in the proposal of the International Consensus Classification (ICC) [8]. The majority of studies revealed an association of the ABC-subtype with poorer clinical outcomes [9], while some did not observe a clear survival difference between patients with ABC-like and GCB-like DLBCL [10]. In order to enable molecular subtyping according to the COO classification independent of more sophisticated GEP methods, immunohistochemical algorithms continue to be allowed as surrogate markers, dividing DLBCL into GCB-types and non-GCB-types, for example by using the Hans algorithm [11,12]. Besides the GE-based molecular subtypes, DLBCL, NOS also comprise a genetic subtype, namely DLBCL, NOS with dual MYC and BCL6-rearrangements (DLBCL, NOS-MYC/BCL6)-see below.

3. The Molecular Pathogenesis of DLBCL

As already mentioned, DLBCL attempt to recapitulate, or rather, hijack the mechanisms of differentiation and maturation effect in GCs (Figure 2). The different molecular subtypes defined by GEP (GCB-like and ABC-like) also feature different mutational landscapes. GCB-like DLBCLs commonly show mutations in genes essential for GC development, dark zone (DZ) and light zone (LZ) transitions, and interactions with the microenvironment, such as in EZH2, GNA13, MEF2B, KMT2D, TNFRSF14, B2M, and CREBBP [13]. Moreover, 5–10% of GCB-DLBCL harbor dual rearrangements of MYC and BCL2 genes (with or without additional BCL6 translocations) [14,15] which, however, although morphologically indistinguishable from bona fide DLBCL, NOS, are separately classified as diffuse large B-cell lymphoma with *MYC* and *BCL2* rearrangements (see below). The large majority of DLBCL, NOS with an ABC-like GEP show a late GC/GC exit or post-GC origin and are characterized by constitutive activation of B-cell receptor (BCR) signaling and Nuclear Factor Kappa B (NFkB) pathway activation. In contrast to their GCB counterparts, they are usually negative for GC markers, such as CD10 and BCL6, but often express IRF4/MUM [11]. Upon mutation analysis, they often bear mutations in components of the BCR signaling pathway, such as in MYD88, CD79B, PIM1, and PRDM1/BLIMP1 [13]. The t(14;18)(q32; q21) translocation leading to IGH::BCL2 fusion is found in 20–30% of GCB-like DLBCL [16]. BCL6, on the other hand, is predominantly rearranged (30%) in ABC-like DLBCL [17]. Structural alterations of the MYC gene can be found in both GCB-like as well as ABC-like DLBCL [18], and often deregulate MYC through IG gene enhancer involvement. Around 60% of MYC translocations occur with an IGH, IGK, or IGL partner, while in 40% of cases, *MYC* is rearranged with a non-IG gene, including for example *BCL6*, *BCL11A*, *IKZF1*, or *PAX5* [19].

In *IGH::MYC* fusions, the breakpoints are usually upstream (5') of *MYC* or within exon 1 or intron 1, but often downstream of *MYC* (3') in non-IGH translocations [20]. Aggressive lymphomas with simultaneous rearrangements of *MYC* and *BCL6* are considered a genetic subtype of DLBCL, NOS, and in around 30% of these cases, *MYC* is directly translocated to *BCL6* forming a *BCL6::MYC* fusion as a result of the t(3;8)(q27;q24) [21]. The significance of this fusion and the difference between cases with dual *MYC* and *BCL6* translocations have not been elucidated so far.

More recently, more sophisticated approaches aiming at deciphering the mutational landscape of DLBCL, NOS via exome or targeted sequencing have shown that the molecular spectrum of DLBCL, NOS is highly heterogeneous with recurrent mutations found in approximately 150 coding genes or targets of copy number changes, and a mean of 8% of genes mutated per case [22]. Based on those findings, three groups have independently described molecular DLBCL subtypes that either further subdivide the COO classification or that are COO-independent (Figure 2, lower panel) [23–25]. Remarkably, in spite of the different platforms and algorithms used, these studies have provided classification trees that were overlapping, though not identical. However, since a unifying concept for the proposed clusters and their significant genetic drivers is still lacking, both WHO-HAEM5 and ICC do not yet recommend molecular subclassification of DLBCL at the present time.



Figure 2. Schematic presentation of maturating and differentiating pathways in germinal centers including some biological characteristics of GCB-like and ABC-like lymphomas derived from different compartments of the GC. The lower panel shows the more recently proposed algorithms in the molecular subclassification of DLBCL including their main defining genes.

4. T-Cell/Histiocyte-Rich Large B-Cell Lymphoma

T-cell/histiocyte-rich large B-cell lymphoma (THRLBCL) is an LBCL with <10% neoplastic large B-cells in a background rich in T-cells and histiocytes, showing some overlap with certain variants of nodular lymphocyte predominant Hodgkin lymphoma/nodular lymphocyte predominant B-cell lymphoma (NLPHL/NLPBL). No major changes have occurred as compared to the definition in WHO-HAEM4R.

THRLBCL affects lymph nodes, the spleen, the liver, and bone marrow [26]. It is a rare lymphoma usually showing a diffuse or sometimes vaguely nodular growth pattern with the previously mentioned predominating background of T-cells and histiocytic cells and randomly dispersed large B-cells that can have the appearance of peripheral B-blasts or can be Hodgkin- and Reed–Sternberg-like. There can be marked overlap with pattern E of NLPHL/NLPBL according to Fan and colleagues [27], and the distinction is usually not possible in cytologic or fine-needle preparations.

THRLBCL harbors recurrent mutations in *JUNB*, *DUSP2*, *SGK1*, *SOCS1*, and *CREBBP* genes, and these gene mutations are also encountered in NLPHL/NLPBL, which supports a possible biological relationship between the two entities [28,29]. The intriguing composition of the background is obviously due to a tolerogenic host microenvironment with a prominent pro-inflammatory and Interferon-dependent GE signature including metal-binding proteins, such as MT2A [30–33]. Among the T-cells in the background, CD4+ T-cells predominate [34,35]. Of importance, cases that contain unequivocal areas of NLPHL—and if they contain only one nodule—should be classified as NLPHL variants according to WHO-HAEM5.

5. Diffuse Large B-Cell Lymphoma/High-Grade B-Cell Lymphoma with *MYC* and *BCL2* Rearrangements

The revised 4th edition of the WHO classification of haematolymphoid tumors (WHO-HAEM4R) introduced a category of "High-grade B-cell lymphoma, unclassifiable, with rearrangements of MYC and BCL2, and/or BCL6", taking into account the inferior prognosis of neoplasms characterized by such dual or triple MYC, BCL2, and/or BCL6 rearrangements. Two important changes regarding the definition and terminology of this entity were made in WHO-HAEM5. Owing to the morphological variability of these neoplasms that can be composed of large B-cells, intermediate or blastoid cells (Figure 3), WHO-HAEM5 changed the name to diffuse large B-cell lymphoma/high-grade B-cell lymphoma (DLBCL/HGBL) so that there is no need to change the diagnostic category once FISH results have been obtained. The ICC prefers the term "high-grade B-cell lymphoma". Both in WHO-HAEM5 and the ICC, however, a fundamental change is that this entity comprises only cases with MYC and BCL2 rearrangements (DLBCL/HGBL-MYC/BCL2), with or without additional BCL6 breaks, so that aggressive lymphomas with dual rearrangements of MYC and BCL6 (without *BCL2*) are excluded. The reason for this is that aggressive lymphomas with a MYC/BCL2 double hit form a homogeneous group with respect to GEPs and the mutational profile, whereas MYC/BCL6 DH lymphomas are heterogeneous in nature [36].

DLBCL/HGBL-*MYC/BCL2* are aggressive neoplasms diagnosed in advanced disease stages in the majority of patients. Often, there is an extranodal disease, such as infiltration of the bone marrow and the central nervous system (CNS) [37,38]. *MYC* rearrangements have been reported in roughly 10% of cases with DLBCL morphology, of which 40% were DLBCL/HGBL-*MYC/BCL2* and 10–15% DLBCL/HGBL-*MYC/BCL2* with additional *BCL6* rearrangements [18,39]. Aggressive lymphomas with an *MYC* and *BCL6* double hit and either DLBCL or HGBL morphology comprise 5 bis 10% of cases [18,39–41]. With the sole exception of GCB derivation, there is no adjunct marker to reliably predict cases with *MYC* and *BCL2* rearrangements. DLBCL/HGBL-*MYC/BCL2* have a GC GEP, and/or a GCB immunophenotype in 95% of cases [11,42–44]. Accordingly, the COO of DLBCL/HGBL-*MYC/BCL2* is a mature somatically mutated GC cell. In around 30% of the cases, there is a history of preceding follicular lymphoma (FL) [37,40,45–47].



Figure 3. Diffuse large B-cell-lymphoma/High-grade-B-cell-lymphoma with MYC and BCL2 rearrangements (H&E ×400). This tumor has a DLBCL morphology. Upon FISH, it showed a "triple hit" constellation with MYC, BCL2, and BCL6 rearrangements.

In DLBCL/HGBL-MYC/BCL2, IGH::BCL2 fusions are the rule [20,24,40], and MYC is translocated to IG loci in 60% of the cases or to non-IG partners including BCL6, PAX5, RFTN1 and others [18–21,24]. In contrast to the rare precursor cell (lymphoblastic) neoplasms with MYC or dual MYC and BCL2 translocations, the MYC translocation in DLBCL/HGBL-MYC/BCL2 obviously arises on the constraints of a pre-existing BCL2 translocation and, according to the structure of the IG breakpoints, takes place in the GC. DLBCL/HGBL-MYC/BCL2 usually harbors complex karyotypic changes. The genetic landscape is characterized by mutations in genes typically altered in FL, such as BCL2, KMT2D, CREBBP, EZH2, and TNFRSF14, as well as in genes typically mutated in Burkitt lymphomas (BLs), such as MYC, CCND3, FOXO1, and others [46] and, therefore, differs from DLBCL, NOS as well as from FL and BL. As a consequence, most cases of DLBCL/HGBL-MYC/BCL2 are categorized into DLBCL molecular subgroups EZB, C3, or BCL2 [23–25,48]. More recently, two groups have independently published GE signatures that recognize DLBCL/HGBL-MYC/BCL2, termed "molecular high-grade" and "double hit" signatures. Of interest, these signatures identify not only aggressive lymphomas with this double hit status but also a significant number of lymphomas that do not harbor MYC and BCL2 rearrangements, although in some of them, retrospectively, cryptic rearrangements have been detected [14,15,49,50]. The fact that BLs also express these signatures indicates that they are not recognizing the double hit status per se, but a common biology related to the dark zone of the GC.

Rare cases of aggressive non-precursor B-cell lymphomas with either large cell or highgrade morphology and expression of Terminal Deoxynucleotidyl Transferase (TdT) have been described [51,52]. Of importance, such cases—although overall very rare—cluster within DLBCL/HGBL-MYC/BCL2, so staining for TdT is advised. Equally rare bona fide B-cell lymphoblastic leukemias/lymphomas with TdT expression and MYC translocations or, even rarer, MYC/BCL2 double hit have been mainly reported in the pediatric or adolescent setting [53,54] and are molecularly different from DLBCL/HGBL-MYC/BCL2. More recently, flow cytometry or immunohistochemistry-based classifiers have been proposed to differentiate between the two conditions [55,56].

In contrast to DLBCL/HGBL-MYC/BCL2, aggressive lymphomas with either large cell or high-grade morphology and an MYC/BCL6 double hit, but lacking BCL2 translocations, form a heterogeneous tumor category without indications of a unifying biology. They infrequently show mutations of epigenetic regulators, but instead mutations in genes that are frequently altered in ABC-DLBCL, and they often display an ABC-like GEP [24,46]. WHO-HAEM5 recommends classifying them as genetic subtypes of either DLBCL, NOS, or HGBL, NOS according to their morphological features. The ICC lists them as a provisional entity in order to allow for their more precise definition and clarification of the implication of their diagnosis in the future. Of note, both FL with dual MYC and BCL2 rearrangements as well as other specific large B-cell lymphoma entities with a dual MYC/BCL2 translocation status are excluded from this category. Figure 4 gives an overview of the algorithms for the classification of aggressive B-cell lymphomas in WHO-HAEM5 depending on the rearrangements of MYC, BCL2, and BCL6 and the complex 11q gain/loss pattern.



Figure 4. Algorithm for the classification of aggressive B-cell lymphomas in WHO-HAEM5 according to rearrangements of MYC, BCL2, and BCL6 and/or presence of the complex 11q gain/loss patterns.

FISH Analysis

6. ALK-Positive Large B-Cell Lymphoma

Anaplastic lymphoma kinase-positive (ALK+) large B-cell lymphoma (LBCL) is a rare, aggressive large cell lymphoma with immunoblastic or plasmablastic morphology of the tumor cells [57] and plasmablastic immunophenotype with reactivity for plasma cell-associated markers. No major changes have been made from WHO-HAEM4R. The ALK protein is expressed in different patterns according to the genetic aberration targeting *ALK* and can be seen in the cytoplasm only or show both cytoplasmic and nuclear reactivity. In most cases of ALK+ LBCL, there is activation and phosphorylation of *STAT3*, and MYC is also often expressed [58–61]. The *ALK* gene is involved in a variety of rearrangements including the most frequent t(2;17)(p23;q23)/*CLTC::ALK* or the t(2;5)(p23;q35)/*NPM1::ALK* fusion, the latter being the only one that leads to cytoplasmic and nuclear ALK expression [60,62–66].

7. IRF4-Rearranged Large B-Cell Lymphoma

In WHO-HAEM4R and the ICC, *IRF4*-rearranged LBCL (*IRF4*-LBCL) is listed as a variant of FL. However, owing to its predominating large cell cytology, WHO-HAEM5 categorizes *IRF4*-LBCL as a subtype of DLBCL. *IRF4*-LBCL is characterized by rearrangements involving chromosomal band 6p25.3, the locus of the *IRF4* gene, usually in the form of an *IGH::IRF4* fusion [67,68]. This neoplasm arises in children and young adolescents, but can also be seen in older adults. The most common site of origin is the tonsils, but head and neck lymph nodes are also commonly involved. Most patients are diagnosed in clinical stage I [67–72].

IRF4-LBCL can display follicular, follicular and diffuse, or diffuse growth patterns and expresses, next to pan B-cell antigens, CD10, BCL2, and BCL6. Owing to the fusion, the *IRF4* gene product MUM1 is strongly and uniformly expressed. More recently, characteristic genetic alterations have been described in *IRF4*-LBCL with mutations in *IRF4*, *CD79b*, *CARD11*, and *MYD88*, chromosomal losses at 11q and 17p, and gains of chromosome 7 [67,71,73].

8. High-Grade B-Cell Lymphoma with 11q Aberrations

WHO-HAEM4R had listed a provisional entity of "Burkitt-like lymphoma with 11q aberrations" that now has been renamed and upgraded to a definite entity in both WHO-HAEM5 and the ICC as High-grade B-cell lymphoma with 11q aberrations (HGBL-11q) or Large B-cell lymphoma with 11q aberrations, respectively. It is defined as an aggressive B-cell lymphoma harboring a characteristic 11q gain/loss pattern without *MYC* rearrangement. These tumors often present as a localized nodal or extranodal disease in the head and neck region or in the gastrointestinal tract [74,75]. Some cases have been seen in the post-transplant setting [76]. HGBL-11q is a rare lymphoma predominantly diagnosed in children or adolescents.

HGBL-11q characteristically shows diffuse infiltrates of cohesive medium-sized cells with a BL-like or intermediate/blastoid appearance and commonly a starry sky pattern [77]. Compared with BL the tumor cells are more pleomorphic, and the starry sky macrophages have ingested coarse apoptotic debris in 50–100% of cases (Figure 5) [77,78]. The immunophenotype resembles that of BL with reactivity for CD10 and BCL6 and negativity for BCL2, and there is a high proliferation index of >90%.



Figure 5. High-grade B-cell lymphoma with 11q aberrations (H&E \times 400). These tumors as a rule are composed of medium-sized blastic cells reminiscent of, but more pleomorphic than, those of Burkitt lymphoma. Note the starry sky pattern with coarse cellular debris in the macrophages.

In contrast with BL, however, HGBL-11q shows a complex aberration pattern in the long arm of chromosome 11 with a consensus region of gain at 11q23.3 and a consensus region of loss at 11q24.1-qter. Rare cases harbor only the telomeric loss or telomeric loss of heterozygosity [77]. NGS analyses have shown an absence of mutations in genes typically altered in BL, such as *ID3*, *TCF3*, *SMARCA4*, or *CCND3*, while mutations in genes typically altered in GCB-DLBCL, such as GNA13 in 50% are the rule, further substantiating the distinctiveness of HGBL-11q from BL and the rationale for changing the name [75,79].

9. Lymphomatoid Granulomatosis

Lymphomatoid granulomatosis (LYG) is an angiocentric and angiodestructive B-cell lymphoproliferative disorder involving extranodal sites, which is associated with Epstein–Barr Virus (EBV) infection (Figure 6). The definition and the diagnostic criteria of LYG have not substantially changed from WHO-HAEM4R. LYG usually involves the lung and frequently the CNS, skin, kidneys, and other organs, such as the liver. In contrast, lymph nodes and bone marrow involvement are rare [80–82].

LYG is thought to arise on a background of impaired immune surveillance of EBVinfected cells and/or an abnormal immune response to EBV [83–85], however, an overt immunodeficiency state should be absent in typical cases (see below). Monoclonal IG gene rearrangements are usually identified in grade 2 and grade 3 lesions. In the setting of inborn or acquired immunodeficiency and/or dysregulation (IDD), cases with an LYG-like picture should be diagnosed according to the guidelines given in the IDD chapter in WHO-HAEM5, such as "polymorphic lymphoproliferative disorder, LYG-type, EBV+".



(B)

Figure 6. Lymphomatoid granulomatosis. (**A**) (H&E \times 200). There is an angiocentric infiltration of several medium-sized blood vessels by a polymorphic lymphomatous infiltrate. (**B**) (\times 400) LMP staining confirms EBV association and highlights medium-sized to large blasts infected by EBV.

10. EBV-Positive Diffuse Large B-Cell Lymphoma

EBV-positive DLBCL (EBV-DLBCL), now lacking the suffix "NOS" in WHO-HAEM5, is an LBCL in which the majority of the neoplastic cells harbor EBV. Since any threshold given is arbitrary, WHO-HAEM5 avoids giving a defined threshold of the number of infected cells to put a DLBCL into this entity and states that the majority of tumor cells should be EBER positive. Of note, underlying established immune deficiency or dysregulation aside from

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immune senescence excludes the diagnosis. EBV-DLBCL has first been described in elderly patients, thus relating it to age-related immune impairment. However, the neoplasm can occur in younger patients as well and therefore, age is not a defining criterion anymore [86].

The neoplasm is more frequent in countries with a higher EBV prevalence, such as in Asia and Latin America. It is a primarily nodal disease, however, extranodal manifestations are common, with or without lymphadenopathy [87–92].

EBV-DLBCL, although by definition composed of large B-cells, can appear in variable patterns in the affected tissues. Polymorphic lesions are characterized by centroblasts, immunoblasts, and/or Hodgkin-like or Reed–Sternberg-like cells in a background infiltrate composed of activated lymphocytes, histiocytes, and plasma cells (Figure 7). In monomorphic types, the tumor cells form sheets indistinguishable from DLBCL, NOS [88,93–95]. Angioinvasive and angiodestructive growth, sometimes leading to extensive "geographical" necrosis, are often findings that lead pathologists to search for EBV infection. The immunophenotype is characterized by the expression of pan B-cell markers, and three-fourths of the cases have an ABC-like or non-GCB immunophenotype. The expression of CD30 and PD-L1/PD-L2 is common [86,88]. EBV latency patterns can be I, II, or III with a dominance of EBER reactivity +/- LMP1-positivity (types I and II), and EBNA2 coexpression in 25%, with some geographical variation [86,96].



Figure 7. EBV-positive diffuse large B-cell lymphoma (H&E \times 400). This lymphoid tumor shows numerous anaplastic Hodgkin- and Reed–Sternberg-like cells. Upon EBER in situ hybridization, practically all nuclei were positive. On the upper left, an area of necrosis is seen.

EBV is thought to play a causal role in the pathogenesis of this entity, and EBVassociated proteins have been shown to enhance or mimic B-cell receptor or CD40 signaling. In addition, the mutational burden of EBV-positive DLBCL has been found to be lower than that of EBV-negative DLBCL, and the presence of EBV may substitute for mutations in driver genes, such as *MYD88* and *CD79A* [97]. NGS analyses have revealed alterations in pathways associated with cellular activation and proliferation, such as *NFKB*, *NOTCH*, *WNT*, and *JAK/STAT*, and immune escape [97,98] Fabian 36604606. Several genes have been described as exclusively mutated in EBV-DLBCL, such as *CCR6*, *CCR7*, *DAPK1*, *TNFRSF21*, *CSNK2B*, and *YY1* [97]. PD-L1 overexpression is frequent, with younger patients presenting more frequently with high-level PD-L1+ tumors in comparison with elderly patients [86,92,99].

11. Diffuse Large B-Cell Lymphoma Associated with Chronic Inflammation

DLBCL associated with chronic inflammation (CI-DLBCL) is an EBV-associated LBCL confined to the natural body or acquired tissue spaces arising in a setting of chronic inflammation. Pyothorax-associated lymphoma (PAL) is the prototypical form. CI-DLBCL most frequently arises in the pleural cavity, bones, joints, and periarticular tissues [100]. It usually develops after a long latency period (10–35 years) after the onset of chronic infection or implant insertion, such as chronic osteomyelitis, metallic implant or surgical mesh implantation, or chronic venous ulcer of the skin [101–104]. No immunodeficiency is known in these patients. Similar to Fibrin-associated large B-cell lymphoma (FA-LBCL), CI-DLBCL is thought to be related to local immune modulation by the production of IL10 that fosters immune escape of EBV-infected B-cells [100,105–107].

Most cases are composed of centroblasts or immunoblasts, and there may be angiodestructive growth with ensuing massive necrosis. Expression of B-cell markers is typical, but plasmablastic differentiation may occur. CD30 expression and aberrant expression of T-cell-antigens have been noted. EBV type III latency is characteristic [101,102,108].

The GEP of CI-DLBCL (PAL) is characterized by EBV association and down-regulation of HLA class 1 expression [109–111]. Complex karyotypes are the rule, and recurrent mutations of *TP53* and deletions of *TNFAIP3* have been recorded [112,113].

12. Fibrin-Associated Large B-Cell Lymphoma

Fibrin-associated large B-cell lymphoma (FA-LBCL) had been listed as a variant of CI-DLBCL in WHO-HAEM4R. However, taking into account its characteristic presentation and indolent clinical course, both WHO-HAEM5 and ICC have upgraded it to a definite entity. FA-LBCL is an EBV-associated neoplasm that arises in the setting of chronic fibrin deposition in cysts and pseudocyst cavities, such as in the peri-implant space of breast implants and chronic hematomas, and in intravascular or intracardiac lesions, such as in atrial myxomas or endovascular grafts [114–118]. As in CI-DLBCL, mechanisms leading to local immune impairment are thought to lead to the escape of EBV-positive B-cells from immune surveillance [119].

The typical histological findings are aggregates of large transformed cells floating in the background of fibrin and cellular debris (Figure 8A,B), however, there may be an infiltration of the stroma in cardiac myxomas or capsular tissues as well. Of importance, there is no tumor mass formation and/or invasive growth in FA-LBCL setting this neoplasm apart from EBV-DLBCL and CI-DLBCL. As with CI-DLBCL, aberrant expression of T-cell markers may occur. EBV is present in the vast majority of cases (Figure 7B), but rare EBV-negative FA-DLBCL have been encountered [120,121]. EBV type III latency is the rule [117]. Little is known about the genetic background of FA-LBCL, although occasional cases with *BCL6* or *MYC* translocations have been described.





Figure 8. Fibrin-associated large B-cell lymphoma (**A**) (H&E \times 200). Large transformed cells are floating in the background of fibrin and cellular debris. (**B**) (\times 200) EBER in situ hybridization confirmed EBV association.

13. Fluid Overload-Associated Large B-Cell Lymphoma

Fluid overload-associated LBCL (FO-LBCL) is a large B-cell lymphoma presenting as serous effusions often in patients with fluid overload states. Of importance, it is not associated with KSHV/HHV8-infection of the tumor cells, hence the name "HHV8- and EBV- negative primary effusion-based lymphoma" in the ICC. Most cases present in the pleural cavity followed by effusions in the pericardium and peritoneum [122]. By definition, no mass formation is detected at presentation but can occur during relapse or disease progression [123].

The cytological spectrum of the tumor cells as seen in smears or cytoblocks is broad and centroblastic, immunoblastic, or anaplastic cells can be seen (Figure 9). They usually express pan B-cell markers but some of them, in particular, CD20, can be negative. A non-GCB immunophenotype is common, and CD30 expression is variable.



Figure 9. Fluid overload-associated large B-cell lymphoma (H&E \times 200) This cytoblock preparation demonstrates massive shedding of large lymphoid cells into the pleural fluid.

In roughly half of the cases, FO-LBCL is associated with a fluid overload status. KSHV/HHV8 infection is not detected, but a proportion of cases do arise in hepatitis C-positive patients [123–125]. EBV association has been reported in 10–30% of cases [123,125]. In the ICC proposal, however, EBV-associated cases are excluded from this entity. The age peak in elderly patients points to immune senescence as a pathologic factor, but overt immunodeficiency by definition is absent. Few genetic data are available pointing to complex copy number changes, recurrent deletions at 3p and 6q, and structural alterations in the *MYC* locus in 50% of cases [124,126]. Mutations have been found in chromatin-modifying genes and *MYD88*, *IRF4*, and others [126].

14. Plasmablastic Lymphoma

Plasmablastic lymphoma (PBL), similar to ALK-positive LBCL, is an aggressive lymphoma composed of immunoblastic or plasmablastic cells (Figure 10) that express a plasmablastic immunophenotype and that is negative for CD20 [127–130]. No major changes have been made from WHO-HAEM4R. PBL most often arises in settings of immune deficiency, most commonly in HIV+ patients and more rarely in patients that have undergone organ transplantation or have been diagnosed with autoimmune disease [127]. In around 30% of patients, however, there is no evidence of immune deficiency [129–133]. Most patients with PBL have an extranodal disease, most commonly, especially in HIV+ patients, in the oral cavity, nasal sinuses, the nasopharynx, and orbit but all extranodal localizations, including the bone marrow, can be involved [134,135].



Figure 10. Plasmablastic lymphoma (Giemsa \times 400). This photomicrograph shows cohesively arranged immunoblastic/plasmablastic cells with broad deeply basophilic cytoplasm. In this case, CD20 was negative, and both CD138 and MUM1 were strongly expressed.

Translocations involving *MYC*, most often as *IGH::MYC* fusion, have been detected in 50% of cases [136], and EBV infection of the tumor cells is found in 70% of cases [136]. GEP has identified a gene signature similar to plasma cell neoplasms and different from that of DLBCL [137]. More recent NGS studies have identified recurrent mutations in *KRAS* and *NRAS* genes and in genes of the *JAK-STAT* pathway [136,138].

Plasma cell myeloma/multiple myeloma (PCM/MM) with plasmablastic features are a relevant differential diagnosis of PBL because the morphology and the immunophenotype of PBL and PCM/MM are nearly identical and because both *MYC* rearrangements and EBV-

association have been noted in plasmablastic PCM/MN; therefore, the clinical presentation is often crucial in rendering the correct diagnosis [139–141].

15. Primary Large B-Cell Lymphomas of Immune-Privileged Sites

Primary LBCL of immune-privileged sites (IP-LBCL) represents a newly defined entity, or umbrella, in WHO-HAEM5 comprising primary DLBCL that arise in the central nervous system (PCNS-LBCL), the vitreoretina (PVR-LBCL), and the testis (PT-LBCL) in immunocompetent patients. In the ICC proposal, the primary DLBCL of the CNS and the testis continue to be separately listed. DLBCL arising in these localizations have similar morphological, immunophenotypic, and molecular genetic features and have been grouped into one category because of their common background of an origin in immune "sanctuaries" created by precise anatomical structures, such as the blood-brain, blood-retinal and blood-testicular barriers, and specific mechanisms of immune regulation [120,142,143].

These tumors overall are rare, occurring in middle-aged patients; their clinical features depend on the site of origin. PCNS-LBCL, PVR-LBCL, and PT-LBCL consist of medium-sized to large blasts with round to oval, sometimes pleomorphic, nuclei; light chromatin; and eosinophilic to basophilic cytoplasm. PCNS-LBCL typically infiltrate diffusely and/or in perivascular areas (Figure 11) [144], and PT-LBCL infiltrates are also mainly diffuse. In PVR-LBCL, the neoplastic cells occupy the sub-retinal space and/or the perivascular spaces of the retina with cells often also floating in the vitreous [145]. They show similar immunophenotypes with the expression of mature B-cell markers, BCL2, BCL6, and MUM1, while CD10 expression is rarely seen.



Figure 11. Primary large B-cell lymphomas (LBCL) of immune-privileged sites (H&E \times 200). This is an example of a primary central nervous system LBCL with diffuse and perivascular infiltration patterns.

The COO of IP-LBCL is a somatically hypermutated mature GC B-cell with in part biased usage of IGHV genes (especially IGHV4-34) also found in autoimmune disorders [146–149]. Their genetic alterations are associated with immune escape and/or a

down-regulation of immune reactions, for example by genetic inactivation of MHC class I and II and *B2M* genes. *MYD88* and *CD79B* are recurrently mutated [150–156] so IP-LBCL show features of C5/MCD/MYD88 gene signatures [23–25,48,154,157,158]. In addition, IP-LBCL display enhanced BCR and Toll-like signaling, and aberrant somatic hypermutation in genes such as *BCL2*, *MYC*, *PAX5*, and *IRF4* are frequently seen [153,159]. Genomic imbalances are frequent, including gains in 18q21 and 9p24.3 and losses in 6q21 and 6p21. *CDKN2A* is frequently inactivated by bi-allelic deletions in 9p21 [154,160–162].

16. Primary Cutaneous Diffuse Large B-Cell Lymphoma, Leg Type

Primary cutaneous diffuse large B-cell lymphoma, leg type (PCLBCL-LT) is an aggressive lymphoma consisting of centroblasts and immunoblasts, usually with an ABC-like or non-GCB-like immunophenotype commonly arising on the lower leg (Figure 12). However, 10–20% of cases have been described at other sites, such as the trunk or the upper extremities [163–166]. PCLBCL-LT is a disease of typically older patients, with a median age of 75 years, and has a male/female ratio of 1:2-4 [167,168].



Figure 12. Primary cutaneous diffuse large B-cell lymphoma, leg type (H&E \times 200). The large blastic tumor cells are invading the skin and reach the overlying epidermis.

Upon histological examination, there is a usually non-epidermotropic infiltration of the dermis and subcutis, consisting of sheets of centroblasts and immunoblasts positive for B-cell markers. Characteristically, there is strong co-expression of BCL2, MUM1, FOXP1, MYC, and cytoplasmic IgM. The proliferation index is high. Aberrant immunophenotypes with the expression of CD10 may be seen in some cases [163,169–173].

PCLBCL-LT shows a high level of IgHV somatic hypermutations. Translocations involving *IGH*, *MYC*, or *BCL6* frequently occur in 50%, 5–40%, and 25–50%, respectively [166,174–178]. Some cases harbor high-level amplifications in 18q21.3, the site of the *BCL2* locus [166,179]. Recurrent losses have been demonstrated in 9p21 (*CDKN2A* and *CDKN2B*) and for *TNFAIP3* and *PRDM1* [166,179,180]. Mutations in *MYD88* frequently oc-

cur in 70% of cases, and PCLBCL-LT also frequently harbors mutations in *CD79B*, *TNFAIP3*, *CARD11*, and *PIM1* [166,175,181–184]. In addition, deletions of HLA loci and *CIITA* as well as rearrangements of *CD274* have been described [166,183]. Of note, the genetic landscape and GEP of PCDLBCL-LT are similar to IP-LBCL of immune-privileged sites, such as those seen in the CNS, the vitreoretina, and the testis.

17. Intravascular Large B-Cell Lymphoma

The definition of intravascular LBCL (IVLBCL) was not changed in WHO-HAEM5 or ICC compared to WHO-HAEM4R. IVLBCL is an aggressive extranodal B-cell lymphoma showing a proliferation of large B-cells within the lumens of blood vessels. Different subtypes are recognized according to the leading clinical presentation: the classic subtype with fever, local organ-specific symptoms, and/or multiorgan failure; in the cutaneous subtype, the disease is confined to the skin at presentation, and this subtype is described as less aggressive than the classic subtype. In the haemophagocytic subtype, symptoms related to haemophagocytic syndrome predominate. Generally, IVLBCL is a disease of elderly patients, and the haemophagocytic subtype has been predominantly seen in Asian patients [185–187].

In histologic preparations, large B-cells are found within the lumens of capillaries and small vessels with a certain size range (Figure 13). The immunophenotype is characterized by the expression of pan B-cell markers and often with reactivity for CD5 and PD-L1 [188,189]. Most cases have a non-GC B-cell immunophenotype, however, CD10 reactivity has been reported in 10% of cases [190].



Figure 13. Intravascular large B-cell lymphoma (H&E \times 200). The typical blastic tumor cells are seen within distended capillary vessels of the leptomeninx.

Pathogenetically, the lack of expression of adhesion molecules, including integrins and chemokine receptors, prevents vascular adhesion and transvascular migration of the tumor cells [191–195]. Mutations in *MYD88*, *CD79B*, *SETD1B*, and *HLA-B* have been

reported [196–199], and structural alterations in *CD274* (*PD-L1*) and *PDCD1LG2* (*PD-L2*) genes are recurrent [198]. Therefore, IVLBCL, biologically, has some similarities with IP-LBCL.

18. Primary Mediastinal Large B-Cell Lymphoma

Primary mediastinal large B-cell lymphoma (PMBCL), as the name implies, arises in the mediastinum, and its founder cell most probably resides in the thymus. PMBCL is a rare lymphoma often affecting adolescents and young adults and has a male-to-female ratio of 1:2. Secondary involvement of the mediastinum by systemic (nodal) lymphoma has to be excluded. Spread outside the mediastinum may occur at relapse [200].

The tumor cells forming the characteristic mediastinal mass show a range of cytomorphologies and can resemble centroblasts, immunoblasts, or more pleomorphic, sarcomatoid, or Hodgkin-like cells. They are often associated with sclerosis forming broad bands or leading to fine compartmentalization of the tissue (Figure 14) [201–203]. The immunophenotype is characteristic, but not specific; the tumor cells are positive for pan B-cell antigens and frequently also express CD30 and CD23 [204,205], albeit with variable intensity and number of positive cells. Expression of MAL, p63, CD200, PD-L1, and PD-L2 proteins is considered quite specific for PMBCL [205–211].



Figure 14. (H&E \times 100). In this example, large tumor cells, many of them with a clear cytoplasm, are seen in a background of in part compartmentalizing sclerosis.

The genetic landscape of PMBCL is rather distinctive, and GEP has revealed considerable overlap with classic Hodgkin lymphoma (also frequently arising in the mediastinum) [212–214]. Characteristic genetic features are gains in chromosomal band 2p16.1 involving *REL* [215] and amplifications in 9p24.1, the location of *JAK2* and *PD-L1/L2* [216]. In addition, rearrangements in 16p13, harboring *CIITA*, and *PD-L1* and *PD-L2* [217,218] are a characteristic feature of PMBCL. Characteristic point mutations are detected in *SOCS1*, *GNA13*, *STAT6*, *CD58*, *B2M*, *ITKP*, *TNFAIP3*, and *IL4R*. These observations allow the tracing back of proliferation, prevention of apoptosis, and immune evasion to the pathogenetic features of PMBCL.

19. Mediastinal Grey Zone Lymphoma

While the definition and characteristic features of PMBCL have remained largely identical in WHO-HAEM4R and WHO-HAEM5, Mediastinal grey zone lymphoma (MGZL), previously designated "B-cell-lymphoma, unclassifiable, with features intermediate between DLBCL and classic Hodgkin lymphoma" in WHO-HAEM4R, has now been renamed both in WHO-HAEM5 and ICC owing to the fact that it mainly arises in the mediastinum. There are two basic types of MGZL: one with the morphology of Hodgkin- and Reed– Sternberg (HRS)-like cells, but the expression of a preserved pan B-cell program, and the second with the morphology of LBCL, but with inappropriate reactivity for antigens typically expressed in HRS-cells, most often for CD15 [219–221]. Rare cases with morphologic and immunophenotypic features of MGZL outside of the mediastinum have been described in the literature. Such tumors, however, have been shown to harbor different gene expression profiles and DNA alterations [222]. Hence, WHO-HAEM5 recommends classifying them as DLBCL, NOS.

20. High-Grade-B-Cell-Lymphoma, NOS

WHO-HAEM4R had introduced two categories of HGBL, namely "high-grade Bcell lymphoma with *MYC* and *BCL2* and/or *BCL6* rearrangements" and "high-grade B-cell lymphoma, not otherwise specified (NOS)". As mentioned above, cases with dual rearrangements of *MYC* and *BCL2* and high-grade morphology are now listed in the new entity of DLBCL/HGBL-*MYC/BCL2*. Tumors with an *MYC* and *BCL6* double hit and highgrade or large cell morphology have been designated genetic subtypes of HGBL, NOS or DLBCL, NOS in WHO-HAEM5. According to the new classification, HGBL, NOS is viewed as a heterogeneous category of aggressive B-cell lymphomas composed of medium-sized or blastoid cells (high-grade morphology), which do not fit into other defined entities of aggressive B-cell lymphomas. HGBL, NOS is rare, representing less than 2% of lymphomas.

Usually, there is a diffuse growth pattern of moderately cohesive or non-cohesive medium-sized or blastoid cells having variously been described as "Burkitt-like", intermediate between BL and DLBCL, or blastoid (Figure 15) [223–226]. However, more recent data indicate that the reproducibility of these categories may be low. The tumor cells express pan-B-cell antigens and have a GCB-like immunophenotype with CD10 and BCL6 expression in the majority of cases and MUM1 in 60% of cases. BCL2 reactivity is seen in 60–70%, and a high proliferation index is typical [227].

In HGBL, NOS, complex karyotypes have been seen in classical cytogenetic analyses and in array-based comparative genomic hybridization (CGH). *MYC* is rearranged in up to 50% of cases, whereas *BCL2* or *BCL6* translocations are rare [36,226,228,229]. NGS analyses have elucidated frequent mutations in *ID3*, *BCL2*, *TCF3*, *CCND3*, *CREBBP*, *EP300*, and *EZH2*. More recent sequencing data have identified *KMT2D* and *TP53* as the most commonly mutated genes [36]. Using the LymphGen algorithm, HGBL, NOS are grouped into the unclassified category. Of high interest, in GEP, more than 50% of HGBL, NOS harbor the "double hit" signature that had been established from DLBCL with dual *MYC* and *BCL2* rearrangements and that points to dark zone biology [36]. One important differential diagnosis of HGBL, NOS is Burkitt lymphoma (BL), since it shares with HGBL, NOS the presence of medium-sized tumor cells and a GCB immunophenotype in the majority of cases. In addition, *MYC* rearrangements are seen in 50% of HGBL, NOS. However, as a rule, HGBL, NOS shows more pleomorphic or blastoid tumor cells different from those of BL. Of importance, in most cases, HGBL, NOS is also strongly reactive for BCL2, which excludes BL. In cases that do not express BCL2 and that harbor an isolated *MYC* rearrangement, arriving at the correct diagnosis may indeed be difficult and require additional genetic analyses such as finding an absence of an *IG::MYC* fusion or higher genetic complexity in HGBL, NOS [228].



Figure 15. High-grade B-cell lymphoma, NOS (H&E ×400). The tumor cells are small to mediumsized with scant cytoplasm and roundish nuclei. No apparent starry-sky pattern is seen. This tumor was negative for *MYC*, *BCL2*, and *BCL6* rearrangements and also negative for 11q aberrations.

21. Diffuse Large B-Cell Lymphomas Arising in Immune Deficiency/Dysregulation

DLBCL arising in the setting of immune deficiency/dysregulation (IDD) are in general histologically indistinguishable from their non-IDD counterparts (DLBCL, NOS) and usually present with diffuse sheets of large B-cells. These lymphomas may be seen in HIV infection, in post-transplant settings, in the setting of autoimmune disease, therapyrelated, or in the setting of inborn errors of immunity (IEI). In an era in which novel drugs aimed at immuno-modulation or immunotherapy, e.g., CAR-T, are continuously developed, new types of IDD lesions may emerge, and the frequency and spectrum of IDD-DLBCL may broaden.

Generally, these lymphomas can be DLBCL or plasmablastic in nature, and an ABClike GEP or non-GCB immunophenotypical constellation is more frequent, especially if associated with EBV infection. The genetic alterations encountered are related to those seen in samples from non-immunocompromised individuals, however, their frequencies may vary. For example, EBV-positive DLBCL have been reported to harbor fewer genetic alterations in the NFkB pathway than their DLBCL, NOS, counterparts [230,231]. In contrast, EBV-negative DLBCL in IDD are genetically more similar to lymphomas in immunocompetent patients and often have a GCB-like GEP [230–233]. One particular type of DLBCL commonly associated with immunodeficiency is KSHV/HHV8-positive DLBCL, the occurrence of which is usually associated with severe immune deficiency, but may also be seen in non-immunocompromised patients [234–236]. The majority of the patients are HIV-positive males.

In some instances, it is difficult, if not impossible, to decide whether a tumor arose in the setting of immunodeficiency or not. One example is lymphomas arising in elderly patients, which has led to the concept of immune senescence as a source of IDD. However, immunodeficiency related to immune senescence cannot be objectively diagnosed so far, and therefore, some tumor classes may be confounded with tumors arising in unrecognized IDD settings. If manifest immune deficiency and/or dysregulation aside from immune senescence is recognized in a patient with aggressive lymphoma, this tumor should be classified as IDD-associated neoplasm.

22. Perspective

Considerable progress has been achieved in our knowledge of the molecular pathogenesis of LBCL within the past decade. For most of the different entities, elucidation of the genetic landscape has enabled us to refine the classification of tumors and to define new subgroups. Of importance, our increasing knowledge has significantly contributed to our ability to relate distinct histomorphologic features with the underlying biology that characterizes these neoplasms. Owing to the still preliminary nature of some of these data, and pending agreement on the unification of platforms and algorithms, it would be premature to now coin an entirely molecular subclassification within and among defined entities. We will most probably see this in the upcoming 6th edition of the classification. For this task, however, and the time in between, WHO-HAEM5 provides a robust framework to incorporate these new findings.

23. Conclusions and Future Directions

There has been a vast accumulation of new molecular data in the field of the LBCL, allowing us to more precisely define entities and, at the same time, refine their diagnostic criteria. By definition, the classification laid down in WHO-HAEM5 is only a "snapshot in time" and soon, new data will challenge these new definitions and concepts. However, the classification of LBCL and the more sophisticated deciphering of the molecular basis of their origin and progression provides a solid template for future work, ultimately for the sake of patients.

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References

- 1. The Non-Hodgkin's Lymphoma Classification Project. A clinical evaluation of the International Lymphoma Study Group classification of non-Hodgkin's lymphoma. *Blood* **1997**, *89*, 3909–3918. [CrossRef]
- 2. Sehn, L.H.; Salles, G. Diffuse Large B-Cell Lymphoma. N. Engl. J. Med. 2021, 384, 842–858. [CrossRef] [PubMed]
- Yoo, K.H.; Lee, H.; Suh, C. Lymphoma epidemiology in Korea and the real clinical field including the Consortium for Improving Survival of Lymphoma (CISL) trial. *Int. J. Hematol.* 2018, 107, 395–404. [CrossRef] [PubMed]
- 4. Alaggio, R.; Amador, C.; Anagnostopoulos, I.; Attygalle, A.D.; Araujo, I.B.d.O.; Berti, E.; Bhagat, G.; Borges, A.M.; Boyer, D.; Calaminici, M.; et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: Lymphoid Neoplasms. *Leukemia* **2022**, *36*, 1720–1748. [CrossRef]
- Cerhan, J.R.; Kricker, A.; Paltiel, O.; Flowers, C.R.; Wang, S.S.; Monnereau, A.; Blair, A.; Dal Maso, L.; Kane, E.V.; Nieters, A.; et al. Medical history, lifestyle, family history, and occupational risk factors for diffuse large B-cell lymphoma: The InterLymph Non-Hodgkin Lymphoma Subtypes Project. J. Natl. Cancer Inst. Monogr. 2014, 2014, 15–25. [CrossRef]
- Anderson, J.R.; Armitage, J.O.; Weisenburger, D.D. Epidemiology of the non-Hodgkin's lymphomas: Distribution of the major subtypes differ by geographic locations. Non-Hodgkin's Lymphoma Classification Project. *Ann. Oncol.* 1998, 9, 717–720. [CrossRef]
- Rosenwald, A.; Wright, G.; Chan, W.C.; Connors, J.M.; Campo, E.; Fisher, R.I.; Gascoyne, R.D.; Muller-Hermelink, H.K.; Smeland, E.B.; Giltnane, J.M.; et al. The use of molecular profiling to predict survival after chemotherapy for diffuse large-B-cell lymphoma. *N. Engl. J. Med.* 2002, 346, 1937–1947. [CrossRef]
- Campo, E.; Jaffe, E.S.; Cook, J.R.; Quintanilla-Martinez, L.; Swerdlow, S.H.; Anderson, K.C.; Brousset, P.; Cerroni, L.; de Leval, L.; Dirnhofer, S.; et al. The International Consensus Classification of Mature Lymphoid Neoplasms: A report from the Clinical Advisory Committee. *Blood* 2022, 140, 1229–1253. [CrossRef]
- 9. Dunleavy, K.; Roschewski, M.; Wilson, W.H. Precision treatment of distinct molecular subtypes of diffuse large B-cell lymphoma: Ascribing treatment based on the molecular phenotype. *Clin. Cancer Res.* **2014**, *20*, 5182–5193. [CrossRef]
- Staiger, A.M.; Ziepert, M.; Horn, H.; Scott, D.W.; Barth, T.F.E.; Bernd, H.-W.; Feller, A.C.; Klapper, W.; Szczepanowski, M.; Hummel, M.; et al. Clinical Impact of the Cell-of-Origin Classification and the MYC/ BCL2 Dual Expresser Status in Diffuse Large B-Cell Lymphoma Treated within Prospective Clinical Trials of the German High-Grade Non-Hodgkin's Lymphoma Study Group. J. Clin. Oncol. 2017, 35, 2515–2526. [CrossRef]
- Hans, C.P.; Weisenburger, D.D.; Greiner, T.C.; Gascoyne, R.D.; Delabie, J.; Ott, G.; Muller-Hermelink, H.K.; Campo, E.; Braziel, R.M.; Jaffe, E.S.; et al. Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a tissue microarray. *Blood* 2004, 103, 275–282. [CrossRef]
- Phang, K.-C.; Akhter, A.; Tizen, N.M.S.; Rahman, F.A.; Zahratul Azma, R.; Elyamany, G.; Shabani-Rad, M.-T.; Masir, N.; Mansoor, A. Comparison of protein-based cell-of-origin classification to the Lymph2Cx RNA assay in a cohort of diffuse large B-cell lymphomas in Malaysia. J. Clin. Pathol. 2018, 71, 215–220. [CrossRef]
- 13. Pasqualucci, L.; Dalla-Favera, R. The genetic landscape of diffuse large B-cell lymphoma. *Semin. Hematol.* **2015**, 52, 67–76. [CrossRef]
- 14. Ennishi, D.; Jiang, A.; Boyle, M.; Collinge, B.; Grande, B.M.; Ben-Neriah, S.; Rushton, C.; Tang, J.; Thomas, N.; Slack, G.W.; et al. Double-Hit Gene Expression Signature Defines a Distinct Subgroup of Germinal Center B-Cell-Like Diffuse Large B-Cell Lymphoma. *J. Clin. Oncol.* **2018**, *37*, 190. [CrossRef]
- Sha, C.; Barrans, S.; Cucco, F.; Bentley, M.A.; Care, M.A.; Cummin, T.; Kennedy, H.; Thompson, J.S.; Uddin, R.; Worrillow, L.; et al. Molecular High-Grade B-Cell Lymphoma: Defining a Poor-Risk Group That Requires Different Approaches to Therapy. J. Clin. Oncol. 2019, 37, 202–212. [CrossRef]
- Ennishi, D.; Mottok, A.; Ben-Neriah, S.; Shulha, H.P.; Farinha, P.; Chan, F.C.; Meissner, B.; Boyle, M.; Hother, C.; Kridel, R.; et al. Genetic profiling of MYC and BCL2 in diffuse large B-cell lymphoma determines cell-of-origin-specific clinical impact. *Blood* 2017, 129, 2760–2770. [CrossRef]
- 17. Iqbal, J.; Greiner, T.C.; Patel, K.; Dave, B.J.; Smith, L.; Ji, J.; Wright, G.; Sanger, W.G.; Pickering, D.L.; Jain, S.; et al. Distinctive patterns of BCL6 molecular alterations and their functional consequences in different subgroups of diffuse large B-cell lymphoma. *Leukemia* **2007**, *21*, 2332–2343. [CrossRef]
- Rosenwald, A.; Bens, S.; Advani, R.; Barrans, S.; Copie-Bergman, C.; Elsensohn, M.-H.; Natkunam, Y.; Calaminici, M.; Sander, B.; Baia, M.; et al. Prognostic Significance of MYC Rearrangement and Translocation Partner in Diffuse Large B-Cell Lymphoma: A Study by the Lunenburg Lymphoma Biomarker Consortium. J. Clin. Oncol. 2019, 37, 3359–3368. [CrossRef]
- 19. Bertrand, P.; Bastard, C.; Maingonnat, C.; Jardin, F.; Maisonneuve, C.; Courel, M.-N.; Ruminy, P.; Picquenot, J.-M.; Tilly, H. Mapping of MYC breakpoints in 8q24 rearrangements involving non-immunoglobulin partners in B-cell lymphomas. *Leukemia* **2007**, *21*, 515–523. [CrossRef]
- Chong, L.C.; Ben-Neriah, S.; Slack, G.W.; Freeman, C.; Ennishi, D.; Mottok, A.; Collinge, B.; Abrisqueta, P.; Farinha, P.; Boyle, M.; et al. High-resolution architecture and partner genes of MYC rearrangements in lymphoma with DLBCL morphology. *Blood Adv.* 2018, 2, 2755–2765. [CrossRef]

- Johnson, S.M.; Umakanthan, J.M.; Yuan, J.; Fedoriw, Y.; Bociek, R.G.; Kaiser-Rogers, K.; Sanmann, J.N.; Montgomery, N.D. Lymphomas with pseudo-double-hit BCL6-MYC translocations due to t(3;8)(q27;q24) are associated with a germinal center immunophenotype, extranodal involvement, and frequent BCL2 translocations. *Hum. Pathol.* 2018, *80*, 192–200. [CrossRef] [PubMed]
- Reddy, A.; Zhang, J.; Davis, N.S.; Moffitt, A.B.; Love, C.L.; Waldrop, A.; Leppa, S.; Pasanen, A.; Meriranta, L.; Karjalainen-Lindsberg, M.-L.; et al. Genetic and Functional Drivers of Diffuse Large B Cell Lymphoma. *Cell* 2017, 171, 481–494.e15. [CrossRef] [PubMed]
- Schmitz, R.; Wright, G.W.; Huang, D.W.; Johnson, C.A.; Phelan, J.D.; Wang, J.Q.; Roulland, S.; Kasbekar, M.; Young, R.M.; Shaffer, A.L.; et al. Genetics and Pathogenesis of Diffuse Large B-Cell Lymphoma. *N. Engl. J. Med.* 2018, 378, 1396–1407. [CrossRef] [PubMed]
- Chapuy, B.; Stewart, C.; Dunford, A.J.; Kim, J.; Kamburov, A.; Redd, R.A.; Lawrence, M.S.; Roemer, M.G.M.; Li, A.J.; Ziepert, M.; et al. Molecular subtypes of diffuse large B cell lymphoma are associated with distinct pathogenic mechanisms and outcomes. *Nat. Med.* 2018, 24, 679–690. [CrossRef]
- Lacy, S.E.; Barrans, S.L.; Beer, P.A.; Painter, D.; Smith, A.G.; Roman, E.; Cooke, S.L.; Ruiz, C.; Glover, P.; van Hoppe, S.J.L.; et al. Targeted sequencing in DLBCL, molecular subtypes, and outcomes: A Haematological Malignancy Research Network report. *Blood* 2020, *135*, 1759–1771. [CrossRef]
- Achten, R.; Verhoef, G.; Vanuytsel, L.; de Wolf-Peeters, C. T-cell/histiocyte-rich large B-cell lymphoma: A distinct clinicopathologic entity. J. Clin. Oncol. 2002, 20, 1269–1277. [CrossRef]
- 27. Fan, Z.; Natkunam, Y.; Bair, E.; Tibshirani, R.; Warnke, R.A. Characterization of variant patterns of nodular lymphocyte predominant hodgkin lymphoma with immunohistologic and clinical correlation. *Am. J. Surg. Pathol.* **2003**, 27, 1346–1356. [CrossRef]
- Moore, E.M.; Swerdlow, S.H.; Gibson, S.E. J chain and myocyte enhancer factor 2B are useful in differentiating classical Hodgkin lymphoma from nodular lymphocyte predominant Hodgkin lymphoma and primary mediastinal large B-cell lymphoma. *Hum. Pathol.* 2017, 68, 47–53. [CrossRef]
- Hartmann, S.; Schuhmacher, B.; Rausch, T.; Fuller, L.; Döring, C.; Weniger, M.; Lollies, A.; Weiser, C.; Thurner, L.; Rengstl, B.; et al. Highly recurrent mutations of SGK1, DUSP2 and JUNB in nodular lymphocyte predominant Hodgkin lymphoma. *Leukemia* 2016, 30, 844–853. [CrossRef]
- Hartmann, S.; Tousseyn, T.; Doring, C.; Fluchter, P.; Hackstein, H.; Herreman, A.; Ponzoni, M.; de Wolf-Peeters, C.; Facchetti, F.; Gascoyne, R.D.; et al. Macrophages in T cell/histiocyte rich large B cell lymphoma strongly express metal-binding proteins and show a bi-activated phenotype. *Int. J. Cancer* 2013, *133*, 2609–2618. [CrossRef]
- Macon, W.R.; Cousar, J.B.; Waldron, J.A.; Hsu, S.M. Interleukin-4 may contribute to the abundant T-cell reaction and paucity of neoplastic B cells in T-cell-rich B-cell lymphomas. *Am. J. Pathol.* 1992, 141, 1031–1036.
- 32. Chetaille, B.; Bertucci, F.; Finetti, P.; Esterni, B.; Stamatoullas, A.; Picquenot, J.M.; Copin, M.C.; Morschhauser, F.; Casasnovas, O.; Petrella, T.; et al. Molecular profiling of classical Hodgkin lymphoma tissues uncovers variations in the tumor microenvironment and correlations with EBV infection and outcome. *Blood* 2009, *113*, 2765–3775. [CrossRef]
- Van Loo, P.; Tousseyn, T.; Vanhentenrijk, V.; Dierickx, D.; Malecka, A.; Vanden Bempt, I.; Verhoef, G.; Delabie, J.; Marynen, P.; Matthys, P.; et al. T-cell/histiocyte-rich large B-cell lymphoma shows transcriptional features suggestive of a tolerogenic host immune response. *Haematologica* 2010, *95*, 440–448. [CrossRef]
- Kunder, C.; Cascio, M.J.; Bakke, A.; Venkataraman, G.; O'Malley, D.P.; Ohgami, R.S. Predominance of CD4+ T Cells in T-Cell/Histiocyte-Rich Large B-Cell Lymphoma and Identification of a Subset of Patients with Peripheral B-Cell Lymphopenia. *Am. J. Clin. Pathol.* 2017, 147, 596–603. [CrossRef]
- 35. Wu, D.; Thomas, A.; Fromm, J.R. Reactive T cells by flow cytometry distinguish Hodgkin lymphomas from T cell/histiocyte-rich large B cell lymphoma. *Cytometry B Clin. Cytom.* **2016**, *90*, 424–432. [CrossRef]
- Collinge, B.; Ben-Neriah, S.; Chong, L.; Boyle, M.; Jiang, A.; Miyata-Takata, T.; Farinha, P.; Craig, J.W.; Slack, G.W.; Ennishi, D.; et al. The impact of MYC and BCL2 structural variants in tumors of DLBCL morphology and mechanisms of false-negative MYC IHC. *Blood* 2021, 137, 2196–2208. [CrossRef]
- Li, S.; Lin, P.; Fayad, L.E.; Lennon, P.A.; Miranda, R.N.; Yin, C.C.; Lin, E.; Medeiros, L.J. B-cell lymphomas with MYC/8q24 rearrangements and IGH@BCL2/t(14;18)(q32;q21): An aggressive disease with heterogeneous histology, germinal center B-cell immunophenotype and poor outcome. *Mod. Pathol.* 2012, 25, 145–156. [CrossRef]
- 38. Laude, M.-C.; Lebras, L.; Sesques, P.; Ghesquieres, H.; Favre, S.; Bouabdallah, K.; Croizier, C.; Guieze, R.; La Drieu Rochelle, L.; Gyan, E.; et al. First-line treatment of double-hit and triple-hit lymphomas: Survival and tolerance data from a retrospective multicenter French study. Am. J. Hematol. 2021, 96, 302–311. [CrossRef]
- Scott, D.W.; King, R.L.; Staiger, A.M.; Ben-Neriah, S.; Jiang, A.; Horn, H.; Mottok, A.; Farinha, P.; Slack, G.W.; Ennishi, D.; et al. High-grade B-cell lymphoma with MYC and BCL2 and/or BCL6 rearrangements with diffuse large B-cell lymphoma morphology. *Blood* 2018, 131, 2060–2064. [CrossRef]
- Aukema, S.M.; Kreuz, M.; Kohler, C.W.; Rosolowski, M.; Hasenclever, D.; Hummel, M.; Kuppers, R.; Lenze, D.; Ott, G.; Pott, C.; et al. Biological characterization of adult MYC-translocation-positive mature B-cell lymphomas other than molecular Burkitt lymphoma. *Haematologica* 2014, 99, 726–735. [CrossRef]

- Krull, J.E.; Wenzl, K.; Hartert, K.T.; Manske, M.K.; Sarangi, V.; Maurer, M.J.; Larson, M.C.; Nowakowski, G.S.; Ansell, S.M.; McPhail, E.; et al. Somatic copy number gains in MYC, BCL2, and BCL6 identifies a subset of aggressive alternative-DH/TH DLBCL patients. *Blood Cancer J.* 2020, 10, 117. [CrossRef]
- 42. Aukema, S.M.; Siebert, R.; Schuuring, E.; van Imhoff, G.W.; Kluin-Nelemans, H.C.; Boerma, E.-J.; Kluin, P.M. Double-hit B-cell lymphomas. *Blood* 2011, *117*, 2319–2331. [CrossRef] [PubMed]
- Huang, W.; Medeiros, L.J.; Lin, P.; Wang, W.; Tang, G.; Khoury, J.; Konoplev, S.; Yin, C.C.; Xu, J.; Oki, Y.; et al. MYC/BCL2/BCL6 triple hit lymphoma: A study of 40 patients with a comparison to MYC/BCL2 and MYC/BCL6 double hit lymphomas. *Mod. Pathol.* 2018, *31*, 1470–1478. [CrossRef] [PubMed]
- McPhail, E.D.; Maurer, M.J.; Macon, W.R.; Feldman, A.L.; Kurtin, P.J.; Ketterling, R.P.; Vaidya, R.; Cerhan, J.R.; Ansell, S.M.; Porrata, L.F.; et al. Inferior survival in high-grade B-cell lymphoma with MYC and BCL2 and/or BCL6 rearrangements is not associated with MYC/IG gene rearrangements. *Haematologica* 2018, 103, 1899–1907. [CrossRef] [PubMed]
- Pedersen, M.Ø.; Gang, A.O.; Clasen-Linde, E.; Breinholt, M.F.; Knudsen, H.; Nielsen, S.L.; Poulsen, T.S.; Klausen, T.W.; Høgdall, E.; Nørgaard, P. Stratification by MYC expression has prognostic impact in MYC translocated B-cell lymphoma-Identifies a subgroup of patients with poor outcome. *Eur. J. Haematol.* 2019, 102, 395–406. [CrossRef]
- Cucco, F.; Barrans, S.; Sha, C.; Clipson, A.; Crouch, S.; Dobson, R.; Chen, Z.; Thompson, J.S.; Care, M.A.; Cummin, T.; et al. Distinct genetic changes reveal evolutionary history and heterogeneous molecular grade of DLBCL with MYC/BCL2 double-hit. *Leukemia* 2020, 34, 1329–1341. [CrossRef]
- Vogelsberg, A.; Steinhilber, J.; Mankel, B.; Federmann, B.; Schmidt, J.; Montes-Mojarro, I.A.; Hüttl, K.; Rodriguez-Pinilla, M.; Baskaran, P.; Nahnsen, S.; et al. Genetic evolution of in situ follicular neoplasia to aggressive B-cell lymphoma of germinal center subtype. *Haematologica* 2021, 106, 2673–2681. [CrossRef]
- Wright, G.W.; Da Huang, W.; Phelan, J.D.; Coulibaly, Z.A.; Roulland, S.; Young, R.M.; Wang, J.Q.; Schmitz, R.; Morin, R.D.; Tang, J.; et al. A Probabilistic Classification Tool for Genetic Subtypes of Diffuse Large B Cell Lymphoma with Therapeutic Implications. *Cancer Cell* 2020, *37*, 551–568.e14. [CrossRef]
- Hilton, L.K.; Tang, J.; Ben-Neriah, S.; Alcaide, M.; Jiang, A.; Grande, B.M.; Rushton, C.K.; Boyle, M.; Meissner, B.; Scott, D.W.; et al. The double-hit signature identifies double-hit diffuse large B-cell lymphoma with genetic events cryptic to FISH. *Blood* 2019, 134, 1528–1532. [CrossRef]
- Hummel, M.; Bentink, S.; Berger, H.; Klapper, W.; Wessendorf, S.; Barth, T.F.E.; Bernd, H.-W.; Cogliatti, S.B.; Dierlamm, J.; Feller, A.C.; et al. A biologic definition of Burkitt's lymphoma from transcriptional and genomic profiling. *N. Engl. J. Med.* 2006, 354, 2419–2430. [CrossRef]
- 51. Bhavsar, S.; Liu, Y.-C.; Gibson, S.E.; Moore, E.M.; Swerdlow, S.H. Mutational Landscape of TdT+ Large B-cell Lymphomas Supports Their Distinction from B-lymphoblastic Neoplasms: A Multiparameter Study of a Rare and Aggressive Entity. *Am. J. Surg. Pathol.* **2022**, *46*, 71–82. [CrossRef]
- Qiu, L.; Xu, J.; Lin, P.; Cohen, E.N.; Tang, G.; Wang, S.A.; Khanlari, M.; Wang, W.; Khoury, J.D.; Konoplev, S.; et al. Unique pathologic features and gene expression signatures distinguish blastoid high-grade B-cell lymphoma from B acute lymphoblastic leukemia/lymphoma. *Haematologica* 2022, *3*, 895–899. [CrossRef]
- Wagener, R.; López, C.; Kleinheinz, K.; Bausinger, J.; Aukema, S.M.; Nagel, I.; Toprak, U.H.; Seufert, J.; Altmüller, J.; Thiele, H.; et al. IG-MYC+ neoplasms with precursor B-cell phenotype are molecularly distinct from Burkitt lymphomas. *Blood* 2018, 132, 2280–2285. [CrossRef]
- Liu, W.; Hu, S.; Konopleva, M.; Khoury, J.D.; Kalhor, N.; Tang, G.; Bueso-Ramos, C.E.; Jorgensen, J.L.; Lin, P.; Medeiros, L.J.; et al. De Novo MYC and BCL2 Double-hit B-Cell Precursor Acute Lymphoblastic Leukemia (BCP-ALL) in Pediatric and Young Adult Patients Associated with Poor Prognosis. *Pediatr. Hematol. Oncol.* 2015, *32*, 535–547. [CrossRef]
- 55. Khanlari, M.; Medeiros, L.J.; Lin, P.; Xu, J.; You, M.J.; Tang, G.; Yin, C.C.; Wang, W.; Qiu, L.; Miranda, R.N.; et al. Blastoid high-grade B-cell lymphoma initially presenting in bone marrow: A diagnostic challenge. *Mod. Pathol.* **2022**, *35*, 419–426. [CrossRef]
- Geyer, J.T.; Subramaniyam, S.; Jiang, Y.; Elemento, O.; Ferry, J.A.; de Leval, L.; Nakashima, M.O.; Liu, Y.-C.; Martin, P.; Mathew, S.; et al. Lymphoblastic transformation of follicular lymphoma: A clinicopathologic and molecular analysis of 7 patients. *Hum. Pathol.* 2015, 46, 260–271. [CrossRef]
- 57. Pan, Z.; Hu, S.; Li, M.; Zhou, Y.; Kim, Y.S.; Reddy, V.; Sanmann, J.N.; Smith, L.M.; Chen, M.; Gao, Z.; et al. ALK-positive Large B-cell Lymphoma: A Clinicopathologic Study of 26 Cases with Review of Additional 108 Cases in the Literature. *Am. J. Surg. Pathol.* 2017, 41, 25–38. [CrossRef]
- Laurent, C.; Do, C.; Gascoyne, R.D.; Lamant, L.; Ysebaert, L.; Laurent, G.; Delsol, G.; Brousset, P. Anaplastic lymphoma kinasepositive diffuse large B-cell lymphoma: A rare clinicopathologic entity with poor prognosis. J. Clin. Oncol. 2009, 27, 4211–4216. [CrossRef]
- Valera, A.; Colomo, L.; Martinez, A.; de Jong, D.; Balague, O.; Matheu, G.; Martinez, M.; Taddesse-Heath, L.; Jaffe, E.S.; Bacchi, C.E.; et al. ALK-positive large B-cell lymphomas express a terminal B-cell differentiation program and activated STAT3 but lack MYC rearrangements. *Mod. Pathol.* 2013, 26, 1329–1337. [CrossRef]
- Sakamoto, K.; Nakasone, H.; Togashi, Y.; Sakata, S.; Tsuyama, N.; Baba, S.; Dobashi, A.; Asaka, R.; Tsai, C.-C.; Chuang, S.-S.; et al. ALK-positive large B-cell lymphoma: Identification of EML4-ALK and a review of the literature focusing on the ALK immunohistochemical staining pattern. *Int. J. Hematol.* 2016, 103, 399–408. [CrossRef]

- 61. Castillo, J.J.; Beltran, B.E.; Malpica, L.; Marques-Piubelli, M.L.; Miranda, R.N. Anaplastic lymphoma kinase-positive large B-cell lymphoma (ALK + LBCL): A systematic review of clinicopathological features and management. *Leuk. Lymphoma* **2021**, *62*, 2845–2853. [CrossRef] [PubMed]
- Onciu, M.; Behm, F.G.; Downing, J.R.; Shurtleff, S.A.; Raimondi, S.C.; Ma, Z.; Morris, S.W.; Kennedy, W.; Jones, S.C.; Sandlund, J.T. ALK-positive plasmablastic B-cell lymphoma with expression of the NPM-ALK fusion transcript: Report of 2 cases. *Blood* 2003, 102, 2642–2644. [CrossRef] [PubMed]
- 63. Rossky, P.J.; Walker, G.C. Retrospective. Paul F. Barbara (1953–2010). Science 2010, 330, 1191. [CrossRef]
- 64. Takeuchi, K.; Soda, M.; Togashi, Y.; Ota, Y.; Sekiguchi, Y.; Hatano, S.; Asaka, R.; Noguchi, M.; Mano, H. Identification of a novel fusion, SQSTM1-ALK, in ALK-positive large B-cell lymphoma. *Haematologica* **2011**, *96*, 464–467. [CrossRef]
- 65. Lee, S.E.; Kang, S.Y.; Takeuchi, K.; Ko, Y.H. Identification of RANBP2-ALK fusion in ALK positive diffuse large B-cell lymphoma. *Hematol. Oncol.* 2014, 32, 221–224. [CrossRef] [PubMed]
- 66. Ise, M.; Kageyama, H.; Araki, A.; Itami, M. Identification of a novel GORASP2-ALK fusion in an ALK-positive large B-cell lymphoma. *Leuk. Lymphoma* **2019**, *60*, 493–497. [CrossRef]
- 67. Salaverria, I.; Philipp, C.; Oschlies, I.; Kohler, C.W.; Kreuz, M.; Szczepanowski, M.; Burkhardt, B.; Trautmann, H.; Gesk, S.; Andrusiewicz, M.; et al. Translocations activating IRF4 identify a subtype of germinal center-derived B-cell lymphoma affecting predominantly children and young adults. *Blood* **2011**, *118*, 139–147. [CrossRef]
- Au-Yeung, R.K.H.; Arias Padilla, L.; Zimmermann, M.; Oschlies, I.; Siebert, R.; Woessmann, W.; Burkhardt, B.; Klapper, W. Experience with provisional WHO-entities large B-cell lymphoma with IRF4-rearrangement and Burkitt-like lymphoma with 11q aberration in paediatric patients of the NHL-BFM group. *Br. J. Haematol.* 2020, 190, 753–763. [CrossRef]
- 69. De Leval, L.; Bonnet, C.; Copie-Bergman, C.; Seidel, L.; Baia, M.; Brière, J.; Molina, T.J.; Fabiani, B.; Petrella, T.; Bosq, J.; et al. Diffuse large B-cell lymphoma of Waldeyer's ring has distinct clinicopathologic features: A GELA study. *Ann. Oncol.* **2012**, *23*, 3143–3151. [CrossRef]
- Chisholm, K.M.; Mohlman, J.; Liew, M.; Termuhlen, A.; Cairo, M.S.; Gross, T.G.; Perkins, S.L.; Miles, R.R. IRF4 translocation status in pediatric follicular and diffuse large B-cell lymphoma patients enrolled in Children's Oncology Group trials. *Pediatr. Blood Cancer* 2019, 66, e27770. [CrossRef]
- Ramis-Zaldivar, J.E.; Gonzalez-Farré, B.; Balagué, O.; Celis, V.; Nadeu, F.; Salmerón-Villalobos, J.; Andrés, M.; Martin-Guerrero, I.; Garrido-Pontnou, M.; Gaafar, A.; et al. Distinct molecular profile of IRF4-rearranged large B-cell lymphoma. *Blood* 2020, 135, 274–286. [CrossRef]
- Montes-Moreno, S.; King, R.L.; Oschlies, I.; Ponzoni, M.; Goodlad, J.R.; Dotlic, S.; Traverse-Glehen, A.; Ott, G.; Ferry, J.A.; Calaminici, M. Update on lymphoproliferative disorders of the gastrointestinal tract: Disease spectrum from indolent lymphoproliferations to aggressive lymphomas. *Virchows Arch.* 2020, 476, 667–681. [CrossRef]
- 73. Salaverria, I.; Martin-Guerrero, I.; Burkhardt, B.; Kreuz, M.; Zenz, T.; Oschlies, I.; Arnold, N.; Baudis, M.; Bens, S.; García-Orad, A.; et al. High resolution copy number analysis of IRF4 translocation-positive diffuse large B-cell and follicular lymphomas. *Genes Chromosomes. Cancer* 2013, 52, 150–155. [CrossRef]
- 74. Grygalewicz, B.; Woroniecka, R.; Rymkiewicz, G.; Rygier, J.; Borkowska, K.; Kotyl, A.; Blachnio, K.; Bystydzienski, Z.; Nowakowska, B.; Pienkowska-Grela, B. The 11q-Gain/Loss Aberration Occurs Recurrently in MYC-Negative Burkitt-like Lymphoma with 11q Aberration, as Well as MYC-Positive Burkitt Lymphoma and MYC-Positive High-Grade B-Cell Lymphoma, NOS. Am. J. Clin. Pathol. 2017, 149, 17–28. [CrossRef]
- 75. Gonzalez-Farre, B.; Ramis-Zaldivar, J.E.; Salmeron-Villalobos, J.; Balagué, O.; Celis, V.; Verdu-Amoros, J.; Nadeu, F.; Sábado, C.; Ferrández, A.; Garrido, M.; et al. Burkitt-like lymphoma with 11q aberration: A germinal center derived lymphoma genetically unrelated to Burkitt lymphoma. *Haematologica* 2019, 104, 1822. [CrossRef] [PubMed]
- Ferreiro, J.F.; Morscio, J.; Dierickx, D.; Marcelis, L.; Verhoef, G.; Vandenberghe, P.; Tousseyn, T.; Wlodarska, I. Post-transplant molecularly defined Burkitt lymphomas are frequently MYC-negative and characterized by the 11q-gain/loss pattern. *Haematologica* 2015, 100, e275–e279. [CrossRef]
- 77. Salaverria, I.; Martin-Guerrero, I.; Wagener, R.; Kreuz, M.; Kohler, C.W.; Richter, J.; Pienkowska-Grela, B.; Adam, P.; Burkhardt, B.; Claviez, A.; et al. A recurrent 11q aberration pattern characterizes a subset of MYC-negative high-grade B-cell lymphomas resembling Burkitt lymphoma. *Blood* 2014, *123*, 1187–1198. [CrossRef] [PubMed]
- 78. Horn, H.; Kalmbach, S.; Wagener, R.; Staiger, A.M.; Hüttl, K.; Mottok, A.; Bens, S.; Traverse-Glehen, A.; Fontaine, J.; Siebert, R.; et al. A Diagnostic Approach to the Identification of Burkitt-like Lymphoma with 11q Aberration in Aggressive B-Cell Lymphomas. *Am. J. Surg. Pathol.* 2021, 45, 356–364. [CrossRef]
- 79. Wagener, R.; Seufert, J.; Raimondi, F.; Bens, S.; Kleinheinz, K.; Nagel, I.; Altmüller, J.; Thiele, H.; Hübschmann, D.; Kohler, C.W.; et al. The mutational landscape of Burkitt-like lymphoma with 11q aberration is distinct from that of Burkitt lymphoma. *Blood* 2018, 133, 962–966. [CrossRef] [PubMed]
- Katzenstein, A.L.; Doxtader, E.; Narendra, S. Lymphomatoid granulomatosis: Insights gained over 4 decades. *Am. J. Surg. Pathol.* 2010, 34, e35–e48. [CrossRef]
- Song, J.Y.; Pittaluga, S.; Dunleavy, K.; Grant, N.; White, T.; Jiang, L.; Vies-Hill, T.; Raffeld, M.; Wilson, W.H.; Jaffe, E.S. Lymphomatoid granulomatosis—A single institute experience: Pathologic findings and clinical correlations. *Am. J. Surg. Pathol.* 2015, 39, 141–156. [CrossRef] [PubMed]

- Chavez, J.C.; Sandoval-Sus, J.; Horna, P.; Dalia, S.; Bello, C.; Chevernick, P.; Sotomayor, E.M.; Sokol, L.; Shah, B. Lymphomatoid Granulomatosis: A Single Institution Experience and Review of the Literature. *Clin. Lymphoma Myeloma Leuk.* 2016, 16, S170–S174. [CrossRef] [PubMed]
- 83. Sordillo, P.P.; Epremian, B.; Koziner, B.; Lacher, M.; Lieberman, P. Lymphomatoid granulomatosis: An analysis of clinical and immunologic characteristics. *Cancer* **1982**, *49*, 2070–2076. [CrossRef]
- Wilson, W.H.; Kingma, D.W.; Raffeld, M.; Wittes, R.E.; Jaffe, E.S. Association of lymphomatoid granulomatosis with Epstein-Barr viral infection of B lymphocytes and response to interferon-alpha 2b. *Blood* 1996, 87, 4531–4537. [CrossRef] [PubMed]
- Teruya-Feldstein, J.; Jaffe, E.S.; Burd, P.R.; Kanegane, H.; Kingma, D.W.; Wilson, W.H.; Longo, D.L.; Tosato, G. The role of Mig, the monokine induced by interferon-gamma, and IP-10, the interferon-gamma-inducible protein-10, in tissue necrosis and vascular damage associated with Epstein-Barr virus-positive lymphoproliferative disease. *Blood* 1997, *90*, 4099–4105. [CrossRef]
- Nicolae, A.; Pittaluga, S.; Abdullah, S.; Steinberg, S.M.; Pham, T.A.; vies-Hill, T.; Xi, L.; Raffeld, M.; Jaffe, E.S. EBV-positive large B-cell lymphomas in young patients: A nodal lymphoma with evidence for a tolerogenic immune environment. *Blood* 2015, 126, 863–872. [CrossRef]
- Park, S.; Lee, J.; Ko, Y.H.; Han, A.; Jun, H.J.; Lee, S.C.; Hwang, I.G.; Park, Y.H.; Ahn, J.S.; Jung, C.W.; et al. The impact of Epstein-Barr virus status on clinical outcome in diffuse large B-cell lymphoma. *Blood* 2007, 110, 972–978. [CrossRef]
- Dojcinov, S.D.; Venkataraman, G.; Pittaluga, S.; Wlodarska, I.; Schrager, J.A.; Raffeld, M.; Hills, R.K.; Jaffe, E.S. Age-related EBV-associated lymphoproliferative disorders in the Western population: A spectrum of reactive lymphoid hyperplasia and lymphoma. *Blood* 2011, 117, 4726–4735. [CrossRef]
- Oyama, T.; Ichimura, K.; Suzuki, R.; Suzumiya, J.; Ohshima, K.; Yatabe, Y.; Yokoi, T.; Kojima, M.; Kamiya, Y.; Taji, H.; et al. Senile EBV+ B-cell lymphoproliferative disorders: A clinicopathologic study of 22 patients. *Am. J. Surg. Pathol.* 2003, 27, 16–26. [CrossRef]
- 90. Ok, C.Y.; Papathomas, T.G.; Medeiros, L.J.; Young, K.H. EBV-positive diffuse large B-cell lymphoma of the elderly. *Blood* **2013**, *122*, 328–340. [CrossRef]
- Tracy, S.I.; Habermann, T.M.; Feldman, A.L.; Maurer, M.J.; Dogan, A.; Perepu, U.S.; Syrbu, S.; Ansell, S.M.; Thompson, C.A.; Weiner, G.J.; et al. Outcomes among North American patients with diffuse large B-cell lymphoma are independent of tumor Epstein-Barr virus positivity or immunosuppression. *Haematologica* 2018, 103, 297–303. [CrossRef] [PubMed]
- Bourbon, E.; Maucort-Boulch, D.; Fontaine, J.; Mauduit, C.; Sesques, P.; Safar, V.; Ferrant, E.; Golfier, C.; Ghergus, D.; Karlin, L.; et al. Clinicopathological features and survival in EBV-positive diffuse large B-cell lymphoma not otherwise specified. *Blood Adv.* 2021, *5*, 3227–3239. [CrossRef]
- Oyama, T.; Yamamoto, K.; Asano, N.; Oshiro, A.; Suzuki, R.; Kagami, Y.; Morishima, Y.; Takeuchi, K.; Izumo, T.; Mori, S.; et al. Age-related EBV-associated B-cell lymphoproliferative disorders constitute a distinct clinicopathologic group: A study of 96 patients. *Clin. Cancer Res.* 2007, 13, 5124–5132. [CrossRef] [PubMed]
- Asano, N.; Yamamoto, K.; Tamaru, J.-I.; Oyama, T.; Ishida, F.; Ohshima, K.; Yoshino, T.; Nakamura, N.; Mori, S.; Yoshie, O.; et al. Age-related Epstein-Barr virus (EBV)-associated B-cell lymphoproliferative disorders: Comparison with EBV-positive classic Hodgkin lymphoma in elderly patients. *Blood* 2009, 113, 2629–2636. [CrossRef]
- Montes-Moreno, S.; Odqvist, L.; Diaz-Perez, J.A.; Lopez, A.B.; de Villambrosía, S.G.; Mazorra, F.; Castillo, M.E.; Lopez, M.; Pajares, R.; García, J.F.; et al. EBV-positive diffuse large B-cell lymphoma of the elderly is an aggressive post-germinal center B-cell neoplasm characterized by prominent nuclear factor-kB activation. *Mod. Pathol.* 2012, 25, 968–982. [CrossRef]
- Chen, B.J.; Chapuy, B.; Ouyang, J.; Sun, H.H.; Roemer, M.G.; Xu, M.L.; Yu, H.; Fletcher, C.D.; Freeman, G.J.; Shipp, M.A.; et al. PD-L1 expression is characteristic of a subset of aggressive B-cell lymphomas and virus-associated malignancies. *Clin. Cancer Res.* 2013, *19*, 3462–3473. [CrossRef] [PubMed]
- Gebauer, N.; Künstner, A.; Ketzer, J.; Witte, H.M.; Rausch, T.; Benes, V.; Zimmermann, J.; Gebauer, J.; Merz, H.; Bernard, V.; et al. Genomic insights into the pathogenesis of Epstein-Barr virus-associated diffuse large B-cell lymphoma by whole-genome and targeted amplicon sequencing. *Blood Cancer J.* 2021, *11*, 102. [CrossRef]
- Zhou, Y.; Xu, Z.; Lin, W.; Duan, Y.; Lu, C.; Liu, W.; Su, W.; Yan, Y.; Liu, H.; Liu, L.; et al. Comprehensive Genomic Profiling of EBV-Positive Diffuse Large B-cell Lymphoma and the Expression and Clinicopathological Correlations of Some Related Genes. *Front. Oncol.* 2019, *9*, 683. [CrossRef]
- Takahara, T.; Satou, A.; Ishikawa, E.; Kohno, K.; Kato, S.; Suzuki, Y.; Takahashi, E.; Ohashi, A.; Asano, N.; Tsuzuki, T.; et al. Clinicopathological analysis of neoplastic PD-L1-positive EBV+ diffuse large B cell lymphoma, not otherwise specified, in a Japanese cohort. *Virchows Arch.* 2021, 478, 541–552. [CrossRef]
- Cheuk, W.; Chan, A.C.L.; Chan, J.K.C.; Lau, G.T.C.; Chan, V.N.H.; Yiu, H.H.Y. Metallic implant-associated lymphoma: A distinct subgroup of large B-cell lymphoma related to pyothorax-associated lymphoma? *Am. J. Surg. Pathol.* 2005, 29, 832–836. [CrossRef]
- 101. Petitjean, B.; Jardin, F.; Joly, B.; Martin-Garcia, N.; Tilly, H.; Picquenot, J.M.; Briere, J.; Danel, C.; Mehaut, S.; bd-Al-Samad, I.; et al. Pyothorax-associated lymphoma: A peculiar clinicopathologic entity derived from B cells at late stage of differentiation and with occasional aberrant dual B- and T-cell phenotype. *Am. J. Surg. Pathol.* 2002, *26*, 724–732. [CrossRef] [PubMed]
- 102. Nakatsuka, S.; Yao, M.; Hoshida, Y.; Yamamoto, S.; Iuchi, K.; Aozasa, K. Pyothorax-associated lymphoma: A review of 106 cases. J. Clin. Oncol. 2002, 20, 4255–4260. [CrossRef] [PubMed]

- Narimatsu, H.; Ota, Y.; Kami, M.; Takeuchi, K.; Suzuki, R.; Matsuo, K.; Matsumura, T.; Yuji, K.; Kishi, Y.; Hamaki, T.; et al. Clinicopathological features of pyothorax-associated lymphoma; a retrospective survey involving 98 patients. *Ann. Oncol.* 2007, 18, 122–128. [CrossRef] [PubMed]
- 104. Aozasa, K.; Takakuwa, T.; Nakatsuka, S. Pyothorax-associated lymphoma: A lymphoma developing in chronic inflammation. *Adv. Anat. Pathol.* **2005**, *12*, 324–331. [CrossRef]
- 105. Kanno, H.; Yasunaga, Y.; Iuchi, K.; Yamauchi, S.; Tatekawa, T.; Sugiyama, H.; Aozasa, K. Interleukin-6-mediated growth enhancement of cell lines derived from pyothorax-associated lymphoma. *Lab. Investig.* **1996**, *75*, 167–173.
- Kanno, H.; Naka, N.; Yasunaga, Y.; Aozasa, K. Role of an immunosuppressive cytokine, interleukin-10, in the development of pyothorax-associated lymphoma. *Leukemia* 1997, 11 (Suppl. 3), 525–526.
- 107. Copie-Bergman, C.; Niedobitek, G.; Mangham, D.C.; Selves, J.; Baloch, K.; Diss, T.C.; Knowles, D.N.; Delsol, G.; Isaacson, P.G. Epstein-Barr virus in B-cell lymphomas associated with chronic suppurative inflammation. *J. Pathol.* 1997, 183, 287–292. [CrossRef]
- 108. Fujimoto, M.; Haga, H.; Okamoto, M.; Obara, E.; Ishihara, M.; Mizuta, N.; Nishimura, K.; Manabe, T. EBV-associated diffuse large B-cell lymphoma arising in the chest wall with surgical mesh implant. *Pathol. Int.* 2008, 58, 668–671. [CrossRef]
- 109. Nishiu, M.; Tomita, Y.; Nakatsuka, S.; Takakuwa, T.; Iizuka, N.; Hoshida, Y.; Ikeda, J.; Iuchi, K.; Yanagawa, R.; Nakamura, Y.; et al. Distinct pattern of gene expression in pyothorax-associated lymphoma (PAL), a lymphoma developing in long-standing inflammation. *Cancer Sci.* 2004, 95, 828–834. [CrossRef]
- Kanno, H.; Ohsawa, M.; Hashimoto, M.; Iuchi, K.; Nakajima, Y.; Aozasa, K. HLA-A alleles of patients with pyothorax-associated lymphoma: Anti-Epstein-Barr virus (EBV) host immune responses during the development of EBV latent antigen-positive lymphomas. *Int. J. Cancer* 1999, 82, 630–634. [CrossRef]
- 111. Kanno, H.; Nakatsuka, S.; Iuchi, K.; Aozasa, K. Sequences of cytotoxic T-lymphocyte epitopes in the Epstein-Barr virus (EBV) nuclear antigen-3B gene in a Japanese population with or without EBV-positive lymphoid malignancies. *Int. J. Cancer* 2000, *88*, 626–632. [CrossRef] [PubMed]
- 112. Ando, M.; Sato, Y.; Takata, K.; Nomoto, J.; Nakamura, S.; Ohshima, K.; Takeuchi, T.; Orita, Y.; Kobayashi, Y.; Yoshino, T. A20 (TNFAIP3) deletion in Epstein-Barr virus-associated lymphoproliferative disorders/lymphomas. *PLoS ONE* 2013, *8*, e56741. [CrossRef]
- 113. Hongyo, T.; Kurooka, M.; Taniguchi, E.; Iuchi, K.; Nakajima, Y.; Aozasa, K.; Nomura, T. Frequent p53 mutations at dipyrimidine sites in patients with pyothorax-associated lymphoma. *Cancer Res.* **1998**, *58*, 1105–1107. [PubMed]
- 114. Boroumand, N.; Ly, T.L.; Sonstein, J.; Medeiros, L.J. Microscopic diffuse large B-cell lymphoma (DLBCL) occurring in pseudocysts: Do these tumors belong to the category of DLBCL associated with chronic inflammation? *Am. J. Surg. Pathol.* 2012, *36*, 1074–1080. [CrossRef] [PubMed]
- Rodríguez-Pinilla, S.M.; García, F.J.S.; Balagué, O.; Rodríguez-Justo, M.; Piris, M.Á. Breast implant-associated Epstein-Barr virus-positive large B-cell lymphomas: A report of three cases. *Haematologica* 2020, 105, e412–e414. [CrossRef]
- 116. Zanelli, M.; Zizzo, M.; Montanaro, M.; Gomes, V.; Martino, G.; de Marco, L.; Fraternali Orcioni, G.; Martelli, M.P.; Ascani, S. Fibrin-associated large B-cell lymphoma: First case report within a cerebral artery aneurysm and literature review. *BMC Cancer* 2019, 19, 916. [CrossRef]
- 117. Boyer, D.F.; McKelvie, P.A.; de Leval, L.; Edlefsen, K.L.; Ko, Y.-H.; Aberman, Z.A.; Kovach, A.E.; Masih, A.; Nishino, H.T.; Weiss, L.M.; et al. Fibrin-associated EBV-positive Large B-Cell Lymphoma: An Indolent Neoplasm with Features Distinct from Diffuse Large B-Cell Lymphoma Associated with Chronic Inflammation. Am. J. Surg. Pathol. 2017, 41, 299–312. [CrossRef]
- 118. Miller, D.V.; Firchau, D.J.; McClure, R.F.; Kurtin, P.J.; Feldman, A.L. Epstein-Barr virus-associated diffuse large B-cell lymphoma arising on cardiac prostheses. *Am. J Surg. Pathol.* **2010**, *34*, 377–384. [CrossRef]
- Loong, F.; Chan, A.C.L.; Ho, B.C.S.; Chau, Y.-P.; Lee, H.-Y.; Cheuk, W.; Yuen, W.-K.; Ng, W.-S.; Cheung, H.-L.; Chan, J.K.C. Diffuse large B-cell lymphoma associated with chronic inflammation as an incidental finding and new clinical scenarios. *Mod. Pathol.* 2010, 23, 493–501. [CrossRef]
- King, R.L.; Goodlad, J.R.; Calaminici, M.; Dotlic, S.; Montes-Moreno, S.; Oschlies, I.; Ponzoni, M.; Traverse-Glehen, A.; Ott, G.; Ferry, J.A. Lymphomas arising in immune-privileged sites: Insights into biology, diagnosis, and pathogenesis. *Virchows Arch.* 2020, 476, 647–665. [CrossRef]
- 121. Baugh, L.; Brown, N.; Song, J.Y.; Pandya, S.; Montoya, V.; Perry, A.M. Fibrin-Associated, EBV-Negative Diffuse Large B-Cell Lymphoma Arising in Atrial Myxoma: Expanding the Spectrum of the Entity. *Int. J. Surg. Pathol.* **2022**, *30*, 39–45. [CrossRef]
- 122. Kaji, D.; Ota, Y.; Sato, Y.; Nagafuji, K.; Ueda, Y.; Okamoto, M.; Terasaki, Y.; Tsuyama, N.; Matsue, K.; Kinoshita, T.; et al. Primary human herpesvirus 8-negative effusion-based lymphoma: A large B-cell lymphoma with favorable prognosis. *Blood Adv.* **2020**, *4*, 4442–4450. [CrossRef]
- 123. Alexanian, S.; Said, J.; Lones, M.; Pullarkat, S.T. KSHV/HHV8-negative effusion-based lymphoma, a distinct entity associated with fluid overload states. *Am. J. Surg. Pathol.* **2013**, *37*, 241–249. [CrossRef]
- 124. Wu, W.; Youm, W.; Rezk, S.A.; Zhao, X. Human herpesvirus 8-unrelated primary effusion lymphoma-like lymphoma: Report of a rare case and review of 54 cases in the literature. *Am. J. Clin. Pathol.* **2013**, *140*, 258–273. [CrossRef]
- 125. Kubota, T.; Sasaki, Y.; Shiozawa, E.; Takimoto, M.; Hishima, T.; Chong, J.-M. Age and CD20 Expression Are Significant Prognostic Factors in Human Herpes Virus-8-negative Effusion-based Lymphoma. *Am. J. Surg. Pathol.* **2018**, *42*, 1607–1616. [CrossRef]

- 126. Mendeville, M.; Roemer, M.G.M.; van den Hout, M.F.C.M.; Los-de Vries, G.T.; Bladergroen, R.; Stathi, P.; Hijmering, N.J.; Rosenwald, A.; Ylstra, B.; de Jong, D. Aggressive genomic features in clinically indolent primary HHV8-negative effusion-based lymphoma. *Blood* 2019, 133, 377–380. [CrossRef]
- Delecluse, H.J.; Anagnostopoulos, I.; Dallenbach, F.; Hummel, M.; Marafioti, T.; Schneider, U.; Huhn, D.; Schmidt-Westhausen, A.; Reichart, P.A.; Gross, U.; et al. Plasmablastic lymphomas of the oral cavity: A new entity associated with the human immunodeficiency virus infection. *Blood* 1997, *89*, 1413–1420. [CrossRef]
- 128. Montes-Moreno, S.; Gonzalez-Medina, A.R.; Rodriguez-Pinilla, S.M.; Maestre, L.; Sanchez-Verde, L.; Roncador, G.; Mollejo, M.; Garcia, J.F.; Menarguez, J.; Montalban, C.; et al. Aggressive large B-cell lymphoma with plasma cell differentiation: Immunohistochemical characterization of plasmablastic lymphoma and diffuse large B-cell lymphoma with partial plasmablastic phenotype. *Haematologica* 2010, *95*, 1342–1349. [CrossRef]
- Montes-Moreno, S.; Martinez-Magunacelaya, N.; Zecchini-Barrese, T.; de Villambrosía, S.G.; Linares, E.; Ranchal, T.; Rodriguez-Pinilla, M.; Batlle, A.; Cereceda-Company, L.; Revert-Arce, J.B.; et al. Plasmablastic lymphoma phenotype is determined by genetic alterations in MYC and PRDM1. *Mod. Pathol.* 2017, 30, 85–94. [CrossRef]
- Laurent, C.; Fabiani, B.; Do, C.; Tchernonog, E.; Cartron, G.; Gravelle, P.; Amara, N.; Malot, S.; Palisoc, M.M.; Copie-Bergman, C.; et al. Immune-checkpoint expression in Epstein-Barr virus positive and negative plasmablastic lymphoma: A clinical and pathological study in 82 patients. *Haematologica* 2016, 101, 976–984. [CrossRef]
- 131. Castillo, J.J.; Bibas, M.; Miranda, R.N. The biology and treatment of plasmablastic lymphoma. *Blood* **2015**, *125*, 2323–2330. [CrossRef] [PubMed]
- Morscio, J.; Dierickx, D.; Nijs, J.; Verhoef, G.; Bittoun, E.; Vanoeteren, X.; Wlodarska, I.; Sagaert, X.; Tousseyn, T. Clinicopathologic comparison of plasmablastic lymphoma in HIV-positive, immunocompetent, and posttransplant patients: Single-center series of 25 cases and meta-analysis of 277 reported cases. *Am. J. Surg. Pathol.* 2014, *38*, 875–886. [CrossRef] [PubMed]
- 133. Loghavi, S.; Alayed, K.; Aladily, T.N.; Zuo, Z.; Ng, S.-B.; Tang, G.; Hu, S.; Yin, C.C.; Miranda, R.N.; Medeiros, L.J.; et al. Stage, age, and EBV status impact outcomes of plasmablastic lymphoma patients: A clinicopathologic analysis of 61 patients. *J. Hematol. Oncol.* 2015, *8*, 65. [CrossRef]
- 134. Boy, S.C.; van Heerden, M.B.; Raubenheimer, E.J.; van Heerden, W.F.P. Plasmablastic lymphomas with light chain restrictionplasmablastic extramedullary plasmacytomas? *J. Oral Pathol. Med.* **2010**, *39*, 435–439. [CrossRef] [PubMed]
- 135. Fonseca, F.P.; Robinson, L.; van Heerden, M.B.; van Heerden, W.F.P. Oral plasmablastic lymphoma: A clinicopathological study of 113 cases. *J. Oral Pathol. Med.* **2021**, *50*, 594–602. [CrossRef]
- 136. Frontzek, F.; Staiger, A.M.; Zapukhlyak, M.; Xu, W.; Bonzheim, I.; Borgmann, V.; Sander, P.; Baptista, M.J.; Heming, J.-N.; Berning, P.; et al. Molecular and functional profiling identifies therapeutically targetable vulnerabilities in plasmablastic lymphoma. *Nat. Commun.* 2021, *12*, 1413. [CrossRef]
- 137. Chapman, J.; Gentles, A.J.; Sujoy, V.; Vega, F.; Dumur, C.I.; Blevins, T.L.; Bernal-Mizrachi, L.; Mosunjac, M.; Pimentel, A.; Zhu, D.; et al. Gene expression analysis of plasmablastic lymphoma identifies downregulation of B-cell receptor signaling and additional unique transcriptional programs. *Leukemia* **2015**, *29*, 2270–2273. [CrossRef]
- 138. Ramis-Zaldivar, J.E.; Gonzalez-Farre, B.; Nicolae, A.; Pack, S.; Clot, G.; Nadeu, F.; Mottok, A.; Horn, H.; Song, J.Y.; Fu, K.; et al. MAPK and JAK-STAT pathways dysregulation in plasmablastic lymphoma. *Haematologica* **2021**, *106*, 2682–2693. [CrossRef]
- Vega, F.; Chang, C.-C.; Medeiros, L.J.; Udden, M.M.; Cho-Vega, J.H.; Lau, C.-C.; Finch, C.J.; Vilchez, R.A.; McGregor, D.; Jorgensen, J.L. Plasmablastic lymphomas and plasmablastic plasma cell myelomas have nearly identical immunophenotypic profiles. *Mod. Pathol.* 2005, *18*, 806–815. [CrossRef]
- 140. Ambrosio, M.R.; de Falco, G.; Gozzetti, A.; Rocca, B.J.; Amato, T.; Mourmouras, V.; Gazaneo, S.; Mundo, L.; Candi, V.; Piccaluga, P.P.; et al. Plasmablastic transformation of a pre-existing plasmacytoma: A possible role for reactivation of Epstein Barr virus infection. *Haematologica* **2014**, *99*, e235–e237. [CrossRef]
- 141. Liu, Y.; Jelloul, F.; Zhang, Y.; Bhavsar, T.; Ho, C.; Rao, M.; Lewis, N.E.; Cimera, R.; Baik, J.; Sigler, A.; et al. Genetic Basis of Extramedullary Plasmablastic Transformation of Multiple Myeloma. *Am. J. Surg. Pathol.* **2020**, *44*, 838–848. [CrossRef]
- Riemersma, S.A.; Jordanova, E.S.; Schop, R.F.; Philippo, K.; Looijenga, L.H.; Schuuring, E.; Kluin, P.M. Extensive genetic alterations of the HLA region, including homozygous deletions of HLA class II genes in B-cell lymphomas arising in immune-privileged sites. *Blood* 2000, *96*, 3569–3577. [CrossRef]
- 143. Alame, M.; Cornillot, E.; Cacheux, V.; Rigau, V.; Costes-Martineau, V.; Lacheretz-Szablewski, V.; Colinge, J. The immune contexture of primary central nervous system diffuse large B cell lymphoma associates with patient survival and specific cell signaling. *Theranostics* **2021**, *11*, 3565–3579. [CrossRef]
- 144. Küker, W.; Nägele, T.; Korfel, A.; Heckl, S.; Thiel, E.; Bamberg, M.; Weller, M.; Herrlinger, U. Primary central nervous system lymphomas (PCNSL): MRI features at presentation in 100 patients. *J. Neurooncol.* 2005, 72, 169–177. [CrossRef]
- 145. Coupland, S.E.; Damato, B. Understanding intraocular lymphomas. Clin. Exp. Ophthalmol. 2008, 36, 564–578. [CrossRef]
- 146. Montesinos-Rongen, M.; Küppers, R.; Schlüter, D.; Spieker, T.; van Roost, D.; Schaller, C.; Reifenberger, G.; Wiestler, O.D.; Deckert-Schlüter, M. Primary central nervous system lymphomas are derived from germinal-center B cells and show a preferential usage of the V4-34 gene segment. Am. J. Pathol. 1999, 155, 2077–2086. [CrossRef]
- 147. Montesinos-Rongen, M.; Purschke, F.; Küppers, R.; Deckert, M. Immunoglobulin repertoire of primary lymphomas of the central nervous system. *J. Neuropathol. Exp. Neurol.* 2014, 73, 1116–1125. [CrossRef]

- Belhouachi, N.; Xochelli, A.; Boudjoghra, M.; Lesty, C.; Cassoux, N.; Fardeau, C.; Tran, T.H.C.; Choquet, S.; Sarker, B.; Houillier, C.; et al. Primary vitreoretinal lymphomas display a remarkably restricted immunoglobulin gene repertoire. *Blood Adv.* 2020, 4, 1357–1366. [CrossRef] [PubMed]
- 149. Coupland, S.E.; Hummel, M.; Müller, H.-H.; Stein, H. Molecular analysis of immunoglobulin genes in primary intraocular lymphoma. *Investig. Ophthalmol. Vis. Sci.* 2005, *46*, 3507–3514. [CrossRef]
- Montesinos-Rongen, M.; Godlewska, E.; Brunn, A.; Wiestler, O.D.; Siebert, R.; Deckert, M. Activating L265P mutations of the MYD88 gene are common in primary central nervous system lymphoma. *Acta Neuropathol.* 2011, 122, 791–792. [CrossRef]
- 151. Montesinos-Rongen, M.; Schafer, E.; Siebert, R.; Deckert, M. Genes regulating the B cell receptor pathway are recurrently mutated in primary central nervous system lymphoma. *Acta Neuropathol.* **2012**, *124*, 905–906. [CrossRef]
- 152. Cheah, C.Y.; Wirth, A.; Seymour, J.F. Primary testicular lymphoma. Blood 2014, 123, 486–493. [CrossRef] [PubMed]
- 153. Vater, I.; Montesinos-Rongen, M.; Schlesner, M.; Haake, A.; Purschke, F.; Sprute, R.; Mettenmeyer, N.; Nazzal, I.; Nagel, I.; Gutwein, J.; et al. The mutational pattern of primary lymphoma of the central nervous system determined by whole-exome sequencing. *Leukemia* **2015**, *29*, 677–685. [CrossRef]
- 154. Chapuy, B.; Roemer, M.G.M.; Stewart, C.; Tan, Y.; Abo, R.P.; Zhang, L.; Dunford, A.J.; Meredith, D.M.; Thorner, A.R.; Jordanova, E.S.; et al. Targetable genetic features of primary testicular and primary central nervous system lymphomas. *Blood* 2016, 127, 869–881. [CrossRef]
- 155. Fontanilles, M.; Marguet, F.; Bohers, É.; Viailly, P.-J.; Dubois, S.; Bertrand, P.; Camus, V.; Mareschal, S.; Ruminy, P.; Maingonnat, C.; et al. Non-invasive detection of somatic mutations using next-generation sequencing in primary central nervous system lymphoma. *Oncotarget* 2017, *8*, 48157–48168. [CrossRef]
- 156. Bödör, C.; Alpár, D.; Marosvári, D.; Galik, B.; Rajnai, H.; Bátai, B.; Nagy, Á.; Kajtár, B.; Burján, A.; Deák, B.; et al. Molecular Subtypes and Genomic Profile of Primary Central Nervous System Lymphoma. J. Neuropathol. Exp. Neurol. 2020, 79, 176–183. [CrossRef]
- 157. Bonzheim, I.; Giese, S.; Deuter, C.; Süsskind, D.; Zierhut, M.; Waizel, M.; Szurman, P.; Federmann, B.; Schmidt, J.; Quintanilla-Martinez, L.; et al. High frequency of MYD88 mutations in vitreoretinal B-cell lymphoma: A valuable tool to improve diagnostic yield of vitreous aspirates. *Blood* 2015, 126, 76–79. [CrossRef] [PubMed]
- 158. Hübschmann, D.; Kleinheinz, K.; Wagener, R.; Bernhart, S.H.; López, C.; Toprak, U.H.; Sungalee, S.; Ishaque, N.; Kretzmer, H.; Kreuz, M.; et al. Mutational mechanisms shaping the coding and noncoding genome of germinal center derived B-cell lymphomas. *Leukemia* 2021, 35, 2002–2016. [CrossRef]
- 159. Montesinos-Rongen, M.; van Roost, D.; Schaller, C.; Wiestler, O.D.; Deckert, M. Primary diffuse large B-cell lymphomas of the central nervous system are targeted by aberrant somatic hypermutation. *Blood* **2004**, *103*, 1869–1875. [CrossRef]
- 160. Cobbers, J.M.; Wolter, M.; Reifenberger, J.; Ring, G.U.; Jessen, F.; An, H.X.; Niederacher, D.; Schmidt, E.E.; Ichimura, K.; Floeth, F.; et al. Frequent inactivation of CDKN2A and rare mutation of TP53 in PCNSL. *Brain Pathol.* **1998**, *8*, 263–276. [CrossRef] [PubMed]
- Schwindt, H.; Vater, I.; Kreuz, M.; Montesinos-Rongen, M.; Brunn, A.; Richter, J.; Gesk, S.; Ammerpohl, O.; Wiestler, O.D.; Hasenclever, D.; et al. Chromosomal imbalances and partial uniparental disomies in primary central nervous system lymphoma. *Leukemia* 2009, 23, 1875–1884. [CrossRef] [PubMed]
- 162. Bonzheim, I.; Sander, P.; Salmerón-Villalobos, J.; Süsskind, D.; Szurman, P.; Gekeler, F.; Spitzer, M.S.; Steinhilber, J.; Kohler, E.; Büssgen, M.; et al. The molecular hallmarks of primary and secondary vitreoretinal lymphoma. *Blood Adv.* 2022, *6*, 1598–1607. [CrossRef] [PubMed]
- Kodama, K.; Massone, C.; Chott, A.; Metze, D.; Kerl, H.; Cerroni, L. Primary cutaneous large B-cell lymphomas: Clinicopathologic features, classification, and prognostic factors in a large series of patients. *Blood* 2005, 106, 2491–2497. [CrossRef]
- 164. Zinzani, P.L.; Quaglino, P.; Pimpinelli, N.; Berti, E.; Baliva, G.; Rupoli, S.; Martelli, M.; Alaibac, M.; Borroni, G.; Chimenti, S.; et al. Prognostic factors in primary cutaneous B-cell lymphoma: The Italian Study Group for Cutaneous Lymphomas. *J. Clin. Oncol.* 2006, 24, 1376–1382. [CrossRef]
- 165. Senff, N.J.; Hoefnagel, J.J.; Jansen, P.M.; Vermeer, M.H.; van Baarlen, J.; Blokx, W.A.; Canninga-van Dijk, M.R.; Geerts, M.-L.; Hebeda, K.M.; Kluin, P.M.; et al. Reclassification of 300 primary cutaneous B-Cell lymphomas according to the new WHO-EORTC classification for cutaneous lymphomas: Comparison with previous classifications and identification of prognostic markers. J. Clin. Oncol. 2007, 25, 1581–1587. [CrossRef] [PubMed]
- 166. Mareschal, S.; Pham-Ledard, A.; Viailly, P.J.; Dubois, S.; Bertrand, P.; Maingonnat, C.; Fontanilles, M.; Bohers, E.; Ruminy, P.; Tournier, I.; et al. Identification of Somatic Mutations in Primary Cutaneous Diffuse Large B-Cell Lymphoma, Leg Type by Massive Parallel Sequencing. *J. Investig. Dermatol.* 2017, 137, 1984–1994. [CrossRef]
- 167. Vermeer, M.H.; Geelen, F.A.; van Haselen, C.W.; Van, V.; Geerts, M.L.; van Vloten, W.A.; Willemze, R. Primary cutaneous large B-cell lymphomas of the legs. A distinct type of cutaneous B-cell lymphoma with an intermediate prognosis. Dutch Cutaneous Lymphoma Working Group. Arch Dermatol. 1996, 132, 1304–1308. [CrossRef]
- 168. Dobos, G.; de Masson, A.; Ram-Wolff, C.; Beylot-Barry, M.; Pham-Ledard, A.; Ortonne, N.; Ingen-Housz-Oro, S.; Battistella, M.; d'Incan, M.; Rouanet, J.; et al. Epidemiological changes in cutaneous lymphomas: An analysis of 8593 patients from the French Cutaneous Lymphoma Registry. Br. J. Dermatol. 2021, 184, 1059–1067. [CrossRef]
- Goodlad, J.R.; Krajewski, A.S.; Batstone, P.J.; McKay, P.; White, J.M.; Benton, E.C.; Kavanagh, G.M.; Lucraft, H.H. Primary cutaneous diffuse large B-cell lymphoma: Prognostic significance of clinicopathological subtypes. *Am. J. Surg. Pathol.* 2003, 27, 1538–1545. [CrossRef]

- Hoefnagel, J.J.; Vermeer, M.H.; Jansen, P.M.; Fleuren, G.J.; Meijer, C.J.L.M.; Willemze, R. Bcl-2, Bcl-6 and CD10 expression in cutaneous B-cell lymphoma: Further support for a follicle centre cell origin and differential diagnostic significance. *Br. J. Dermatol.* 2003, 149, 1183–1191. [CrossRef]
- 171. Grange, F.; Petrella, T.; Beylot-Barry, M.; Joly, P.; D'Incan, M.; Delaunay, M.; Machet, L.; Avril, M.-F.; Dalac, S.; Bernard, P.; et al. Bcl-2 protein expression is the strongest independent prognostic factor of survival in primary cutaneous large B-cell lymphomas. *Blood* 2004, 103, 3662–3668. [CrossRef]
- 172. Koens, L.; Vermeer, M.H.; Willemze, R.; Jansen, P.M. IgM expression on paraffin sections distinguishes primary cutaneous large B-cell lymphoma, leg type from primary cutaneous follicle center lymphoma. *Am. J. Surg. Pathol.* **2010**, *34*, 1043–1048. [CrossRef]
- 173. Demirkesen, C.; Tüzüner, N.; Esen, T.; Lebe, B.; Ozkal, S. The expression of IgM is helpful in the differentiation of primary cutaneous diffuse large B cell lymphoma and follicle center lymphoma. *Leuk. Res.* **2011**, *35*, 1269–1272. [CrossRef]
- 174. Hallermann, C.; Kaune, K.M.; Gesk, S.; Martin-Subero, J.I.; Gunawan, B.; Griesinger, F.; Vermeer, M.H.; Santucci, M.; Pimpinelli, N.; Willemze, R.; et al. Molecular cytogenetic analysis of chromosomal breakpoints in the IGH, MYC, BCL6, and MALT1 gene loci in primary cutaneous B-cell lymphomas. *J. Investig. Dermatol.* 2004, 123, 213–219. [CrossRef]
- 175. Pham-Ledard, A.; Prochazkova-Carlotti, M.; Andrique, L.; Cappellen, D.; Vergier, B.; Martinez, F.; Grange, F.; Petrella, T.; Beylot-Barry, M.; Merlio, J.P. Multiple genetic alterations in primary cutaneous large B-cell lymphoma, leg type support a common lymphomagenesis with activated B-cell-like diffuse large B-cell lymphoma. *Mod. Pathol.* **2014**, 27, 402–411. [CrossRef]
- 176. Schrader, A.M.R.; Jansen, P.M.; Vermeer, M.H.; Kleiverda, J.K.; Vermaat, J.S.P.; Willemze, R. High Incidence and Clinical Significance of MYC Rearrangements in Primary Cutaneous Diffuse Large B-Cell Lymphoma, Leg Type. Am. J. Surg. Pathol. 2018, 42, 1488–1494. [CrossRef]
- 177. Menguy, S.; Laharanne, E.; Prochazkova-Carlotti, M.; Gros, A.; Vergier, B.; Parrens, M.; Beylot-Barry, M.; Pham-Ledard, A.; Merlio, J.-P. Challenges in Assessing MYC Rearrangement in Primary Cutaneous Diffuse Large B-Cell Lymphoma, Leg-Type. Am. J. Surg. Pathol. 2020, 44, 424–427. [CrossRef]
- 178. Lucioni, M.; Pescia, C.; Bonometti, A.; Fraticelli, S.; Moltrasio, C.; Ramponi, A.; Riboni, R.; Roccio, S.; Ferrario, G.; Arcaini, L.; et al. Double expressor and double/triple hit status among primary cutaneous diffuse large B-cell lymphoma: A comparison between leg type and not otherwise specified subtypes. *Hum. Pathol.* **2021**, *111*, 1–9. [CrossRef]
- 179. Dijkman, R.; Tensen, C.P.; Jordanova, E.S.; Knijnenburg, J.; Hoefnagel, J.J.; Mulder, A.A.; Rosenberg, C.; Raap, A.K.; Willemze, R.; Szuhai, K.; et al. Array-based comparative genomic hybridization analysis reveals recurrent chromosomal alterations and prognostic parameters in primary cutaneous large B-cell lymphoma. J. Clin. Oncol. 2006, 24, 296–305. [CrossRef]
- Senff, N.J.; Zoutman, W.H.; Vermeer, M.H.; Assaf, C.; Berti, E.; Cerroni, L.; Espinet, B.; de Misa Cabrera, R.F.; Geerts, M.-L.; Kempf, W.; et al. Fine-mapping chromosomal loss at 9p21: Correlation with prognosis in primary cutaneous diffuse large B-cell lymphoma, leg type. J. Investig. Dermatol. 2009, 129, 1149–1155. [CrossRef]
- Koens, L.; Zoutman, W.H.; Ngarmlertsirichai, P.; Przybylski, G.K.; Grabarczyk, P.; Vermeer, M.H.; Willemze, R.; Jansen, P.M.; Schmidt, C.A.; Tensen, C.P. Nuclear factor-kB pathway-activating gene aberrancies in primary cutaneous large B-cell lymphoma, leg type. J. Investig. Dermatol. 2014, 134, 290–292. [CrossRef] [PubMed]
- Pham-Ledard, A.; Beylot-Barry, M.; Barbe, C.; Leduc, M.; Petrella, T.; Vergier, B.; Martinez, F.; Cappellen, D.; Merlio, J.-P.; Grange, F. High frequency and clinical prognostic value of MYD88 L265P mutation in primary cutaneous diffuse large B-cell lymphoma, leg-type. *JAMA Dermatol.* 2014, 150, 1173–1179. [CrossRef]
- 183. Zhou, X.A.; Louissaint, A.; Wenzel, A.; Yang, J.; Martinez-Escala, M.E.; Moy, A.P.; Morgan, E.A.; Paxton, C.N.; Hong, B.; Andersen, E.F.; et al. Genomic Analyses Identify Recurrent Alterations in Immune Evasion Genes in Diffuse Large B-Cell Lymphoma, Leg Type. J. Investig. Dermatol. 2018, 138, 2365–2376. [CrossRef] [PubMed]
- 184. Ducharme, O.; Beylot-Barry, M.; Pham-Ledard, A.; Bohers, E.; Viailly, P.-J.; Bandres, T.; Faur, N.; Frison, E.; Vergier, B.; Jardin, F.; et al. Mutations of the B-Cell Receptor Pathway Confer Chemoresistance in Primary Cutaneous Diffuse Large B-Cell Lymphoma Leg Type. J. Investig. Dermatol. 2019, 139, 2334–2342.e8. [CrossRef] [PubMed]
- 185. Ferreri, A.J.M.; Dognini, G.P.; Campo, E.; Willemze, R.; Seymour, J.F.; Bairey, O.; Martelli, M.; de Renz, A.O.; Doglioni, C.; Montalban, C.; et al. Variations in clinical presentation, frequency of hemophagocytosis and clinical behavior of intravascular lymphoma diagnosed in different geographical regions. *Haematologica* 2007, 92, 486–492. [CrossRef]
- Ponzoni, M.; Campo, E.; Nakamura, S. Intravascular large B-cell lymphoma: A chameleon with multiple faces and many masks. Blood 2018, 132, 1561–1567. [CrossRef]
- Liu, Z.; Zhang, Y.; Zhu, Y.; Zhang, W. Prognosis of Intravascular Large B Cell Lymphoma (IVLBCL): Analysis of 182 Patients from Global Case Series. *Cancer Manag. Res.* 2020, 12, 10531–10540. [CrossRef]
- 188. Sakakibara, A.; Inagaki, Y.; Imaoka, E.; Sakai, Y.; Ito, M.; Ishikawa, E.; Shimada, S.; Shimada, K.; Suzuki, Y.; Nakamura, S.; et al. Divergence and heterogeneity of neoplastic PD-L1 expression: Two autopsy case reports of intravascular large B-cell lymphoma. *Pathol. Int.* 2019, 69, 148–154. [CrossRef]
- Gupta, G.K.; Jaffe, E.S.; Pittaluga, S. A study of PD-L1 expression in intravascular large B cell lymphoma: Correlation with clinical and pathological features. *Histopathology* 2019, 75, 282–286. [CrossRef]
- Murase, T.; Yamaguchi, M.; Suzuki, R.; Okamoto, M.; Sato, Y.; Tamaru, J.-I.; Kojima, M.; Miura, I.; Mori, N.; Yoshino, T.; et al. Intravascular large B-cell lymphoma (IVLBCL): A clinicopathologic study of 96 cases with special reference to the immunophenotypic heterogeneity of CD5. *Blood* 2007, 109, 478–485. [CrossRef]

- Ponzoni, M.; Arrigoni, G.; Gould, V.E.; Del Curto, B.; Maggioni, M.; Scapinello, A.; Paolino, S.; Cassisa, A.; Patriarca, C. Lack of CD 29 (beta1 integrin) and CD 54 (ICAM-1) adhesion molecules in intravascular lymphomatosis. *Hum. Pathol.* 2000, *31*, 220–226. [CrossRef]
- 192. Kinoshita, M.; Izumoto, S.; Hashimoto, N.; Kishima, H.; Kagawa, N.; Hashiba, T.; Chiba, Y.; Yoshimine, T. Immunohistochemical analysis of adhesion molecules and matrix metalloproteinases in malignant CNS lymphomas: A study comparing primary CNS malignant and CNS intravascular lymphomas. *Brain Tumor Pathol.* 2008, 25, 73–78. [CrossRef]
- 193. Alon, R.; Shulman, Z. Chemokine triggered integrin activation and actin remodeling events guiding lymphocyte migration across vascular barriers. *Exp. Cell Res.* 2011, 317, 632–641. [CrossRef]
- Kasuya, A.; Fujiyama, T.; Shirahama, S.; Hashizume, H.; Tokura, Y. Decreased expression of homeostatic chemokine receptors in intravascular large B-cell lymphoma. *Eur. J. Dermatol.* 2012, 22, 272–273. [CrossRef]
- 195. Shimada, K.; Shimada, S.; Sugimoto, K.; Nakatochi, M.; Suguro, M.; Hirakawa, A.; Hocking, T.D.; Takeuchi, I.; Tokunaga, T.; Takagi, Y.; et al. Development and analysis of patient-derived xenograft mouse models in intravascular large B-cell lymphoma. *Leukemia* 2016, 30, 1568–1579. [CrossRef]
- 196. Schrader, A.M.R.; Jansen, P.M.; Willemze, R.; Vermeer, M.H.; Cleton-Jansen, A.-M.; Somers, S.F.; Veelken, H.; van Eijk, R.; Kraan, W.; Kersten, M.J.; et al. High prevalence of MYD88 and CD79B mutations in intravascular large B-cell lymphoma. *Blood* 2018, 131, 2086–2089. [CrossRef]
- 197. Suehara, Y.; Sakata-Yanagimoto, M.; Hattori, K.; Nanmoku, T.; Itoh, T.; Kaji, D.; Yamamoto, G.; Abe, Y.; Narita, K.; Takeuchi, M.; et al. Liquid biopsy for the identification of intravascular large B-cell lymphoma. *Haematologica* **2018**, *103*, e241–e244. [CrossRef]
- 198. Shimada, K.; Yoshida, K.; Suzuki, Y.; Iriyama, C.; Inoue, Y.; Sanada, M.; Kataoka, K.; Yuge, M.; Takagi, Y.; Kusumoto, S.; et al. Frequent genetic alterations in immune checkpoint-related genes in intravascular large B-cell lymphoma. *Blood* 2021, 137, 1491–1502. [CrossRef]
- 199. Gonzalez-Farre, B.; Ramis-Zaldivar, J.E.; Castrejón de Anta, N.; Rivas-Delgado, A.; Nadeu, F.; Salmeron-Villalobos, J.; Enjuanes, A.; Karube, K.; Balagué, O.; Cobo, F.; et al. Intravascular Large B-Cell Lymphoma Genomic Profile Is Characterized by Alterations in Genes Regulating NF-κB and Immune Checkpoints. *Am. J. Surg. Pathol.* 2022, 47, 202–211. [CrossRef]
- 200. Savage, K.J. Primary mediastinal large B-cell lymphoma. Blood 2022, 140, 955–970. [CrossRef]
- Yousem, S.A.; Weiss, L.M.; Warnke, R.A. Primary mediastinal non-Hodgkin's lymphomas: A morphologic and immunologic study of 19 cases. Am. J. Clin. Pathol. 1985, 83, 676–680. [CrossRef] [PubMed]
- Möller, P.; Lämmler, B.; Herrmann, B.; Otto, H.F.; Moldenhauer, G.; Momburg, F. The primary mediastinal clear cell lymphoma of B-cell type has variable defects in MHC antigen expression. *Immunology* 1986, 59, 411–417. [PubMed]
- 203. Paulli, M.; Strater, J.; Gianelli, U.; Rousset, M.T.; Gambacorta, M.; Orlandi, E.; Klersy, C.; Lavabre-Bertrand, T.; Morra, E.; Manegold, C.; et al. Mediastinal B-cell lymphoma: A study of its histomorphologic spectrum based on 109 cases. *Hum. Pathol.* 1999, 30, 178–187. [CrossRef] [PubMed]
- 204. Higgins, J.P.; Warnke, R.A. CD30 expression is common in mediastinal large B-cell lymphoma. *Am. J. Clin. Pathol.* **1999**, 112, 241–247. [CrossRef]
- 205. Pileri, S.A.; Gaidano, G.; Zinzani, P.L.; Falini, B.; Gaulard, P.; Zucca, E.; Pieri, F.; Berra, E.; Sabattini, E.; Ascani, S.; et al. Primary mediastinal B-cell lymphoma: High frequency of BCL-6 mutations and consistent expression of the transcription factors OCT-2, BOB.1, and PU.1 in the absence of immunoglobulins. *Am. J. Pathol.* 2003, *162*, 243–253. [CrossRef]
- 206. Kim, H.-J.; Kim, H.K.; Park, G.; Min, S.K.; Cha, H.J.; Lee, H.; Choi, S.J.; Na, H.Y.; Choe, J.-Y.; Kim, J.E. Comparative pathologic analysis of mediastinal B-cell lymphomas: Selective expression of p63 but no GATA3 optimally differentiates primary mediastinal large B-cell lymphoma from classic Hodgkin lymphoma. *Diagn. Pathol.* 2019, 14, 133. [CrossRef]
- 207. Copie-Bergman, C.; Gaulard, P.; Maouche-Chrétien, L.; Brière, J.; Haioun, C.; Alonso, M.A.; Roméo, P.H.; Leroy, K. The MAL gene is expressed in primary mediastinal large B-cell lymphoma. *Blood* **1999**, *94*, 3567–3575. [CrossRef]
- Calaminici, M.; Piper, K.; Lee, A.M.; Norton, A.J. CD23 expression in mediastinal large B-cell lymphomas. *Histopathology* 2004, 45, 619–624. [CrossRef]
- Dorfman, D.M.; Shahsafaei, A.; Alonso, M.A. Utility of CD200 immunostaining in the diagnosis of primary mediastinal large B cell lymphoma: Comparison with MAL, CD23, and other markers. *Mod. Pathol.* 2012, 25, 1637–1643. [CrossRef]
- 210. Shi, M.; Roemer, M.G.M.; Chapuy, B.; Liao, X.; Sun, H.; Pinkus, G.S.; Shipp, M.A.; Freeman, G.J.; Rodig, S.J. Expression of programmed cell death 1 ligand 2 (PD-L2) is a distinguishing feature of primary mediastinal (thymic) large B-cell lymphoma and associated with PDCD1LG2 copy gain. Am. J. Surg. Pathol. 2014, 38, 1715–1723. [CrossRef]
- Gentry, M.; Bodo, J.; Durkin, L.; Hsi, E.D. Performance of a Commercially Available MAL Antibody in the Diagnosis of Primary Mediastinal Large B-Cell Lymphoma. *Am. J. Surg. Pathol.* 2017, *41*, 189–194. [CrossRef]
- 212. Rosenwald, A.; Wright, G.; Leroy, K.; Yu, X.; Gaulard, P.; Gascoyne, R.D.; Chan, W.C.; Zhao, T.; Haioun, C.; Greiner, T.C.; et al. Molecular diagnosis of primary mediastinal B cell lymphoma identifies a clinically favorable subgroup of diffuse large B cell lymphoma related to Hodgkin lymphoma. *J. Exp. Med.* 2003, *198*, 851–862. [CrossRef]
- 213. Savage, K.J.; Monti, S.; Kutok, J.L.; Cattoretti, G.; Neuberg, D.; de Leval, L.; Kurtin, P.; Dal Cin, P.; Ladd, C.; Feuerhake, F.; et al. The molecular signature of mediastinal large B-cell lymphoma differs from that of other diffuse large B-cell lymphomas and shares features with classical Hodgkin lymphoma. *Blood* **2003**, *102*, 3871–3879. [CrossRef]

- 214. Mottok, A.; Wright, G.; Rosenwald, A.; Ott, G.; Ramsower, C.; Campo, E.; Braziel, R.M.; Delabie, J.; Weisenburger, D.D.; Song, J.Y.; et al. Molecular classification of primary mediastinal large B-cell lymphoma using routinely available tissue specimens. *Blood* 2018, 132, 2401–2405. [CrossRef]
- 215. Weniger, M.A.; Gesk, S.; Ehrlich, S.; Martin-Subero, J.I.; Dyer, M.J.; Siebert, R.; Moller, P.; Barth, T.F. Gains of REL in primary mediastinal B-cell lymphoma coincide with nuclear accumulation of REL protein. *Genes Chromosomes Cancer* 2007, 46, 406–415. [CrossRef]
- 216. Green, M.R.; Monti, S.; Rodig, S.J.; Juszczynski, P.; Currie, T.; O'Donnell, E.; Chapuy, B.; Takeyama, K.; Neuberg, D.; Golub, T.R.; et al. Integrative analysis reveals selective 9p24.1 amplification, increased PD-1 ligand expression, and further induction via JAK2 in nodular sclerosing Hodgkin lymphoma and primary mediastinal large B-cell lymphoma. *Blood* 2010, *116*, 3268–3277. [CrossRef]
- 217. Twa, D.D.; Chan, F.C.; Ben-Neriah, S.; Woolcock, B.W.; Mottok, A.; Tan, K.L.; Slack, G.W.; Gunawardana, J.; Lim, R.S.; McPherson, A.W.; et al. Genomic rearrangements involving programmed death ligands are recurrent in primary mediastinal large B-cell lymphoma. *Blood* 2014, 123, 2062–2065. [CrossRef]
- 218. Steidl, C.; Shah, S.P.; Woolcock, B.W.; Rui, L.; Kawahara, M.; Farinha, P.; Johnson, N.A.; Zhao, Y.; Telenius, A.; Neriah, S.B.; et al. MHC class II transactivator CIITA is a recurrent gene fusion partner in lymphoid cancers. *Nature* 2011, 471, 377–381. [CrossRef]
- 219. Traverse-Glehen, A.; Pittaluga, S.; Gaulard, P.; Sorbara, L.; Alonso, M.A.; Raffeld, M.; Jaffe, E.S. Mediastinal gray zone lymphoma: The missing link between classic Hodgkin's lymphoma and mediastinal large B-cell lymphoma. *Am. J. Surg. Pathol.* 2005, 29, 1411–1421. [CrossRef]
- 220. Pilichowska, M.; Pittaluga, S.; Ferry, J.A.; Hemminger, J.; Chang, H.; Kanakry, J.A.; Sehn, L.H.; Feldman, T.; Abramson, J.S.; Kritharis, A.; et al. Clinicopathologic consensus study of gray zone lymphoma with features intermediate between DLBCL and classical HL. *Blood Adv.* 2017, 1, 2600–2609. [CrossRef]
- 221. Sarkozy, C.; Copie-Bergman, C.; Damotte, D.; Ben-Neriah, S.; Burroni, B.; Cornillon, J.; Lemal, R.; Golfier, C.; Fabiani, B.; Chassagne-Clément, C.; et al. Gray-zone Lymphoma Between cHL and Large B-Cell Lymphoma: A Histopathologic Series from the LYSA. Am. J. Surg. Pathol. 2019, 43, 341–351. [CrossRef] [PubMed]
- 222. Sarkozy, C.; Hung, S.S.; Chavez, E.A.; Duns, G.; Takata, K.; Chong, L.C.; Aoki, T.; Jiang, A.; Miyata-Takata, T.; Telenius, A.; et al. Mutational landscape of gray zone lymphoma. *Blood* **2021**, *137*, 1765–1776. [CrossRef] [PubMed]
- 223. Rimsza, L.; Pittaluga, S.; Dirnhofer, S.; Copie-Bergman, C.; de Leval, L.; Facchetti, F.; Pileri, S.; Rosenwald, A.; Wotherspoon, A.; Fend, F. The clinicopathologic spectrum of mature aggressive B cell lymphomas. *Virchows Arch.* 2017, 471, 453–466. [CrossRef] [PubMed]
- Li, J.; Liu, X.; Yao, Z.; Zhang, M. High-Grade B-Cell Lymphomas, Not Otherwise Specified: A Study of 41 Cases. Cancer Manag. Res. 2020, 12, 1903–1912. [CrossRef]
- 225. Kanagal-Shamanna, R.; Medeiros, L.J.; Lu, G.; Wang, S.A.; Manning, J.T.; Lin, P.; Penn, G.M.; Young, K.H.; You, M.J.; Vega, F.; et al. High-grade B cell lymphoma, unclassifiable, with blastoid features: An unusual morphological subgroup associated frequently with BCL2 and/or MYC gene rearrangements and a poor prognosis. *Histopathology* 2012, *61*, 945–954. [CrossRef]
- Moore, E.M.; Aggarwal, N.; Surti, U.; Swerdlow, S.H. Further Exploration of the Complexities of Large B-Cell Lymphomas with MYC Abnormalities and the Importance of a Blastoid Morphology. *Am. J. Surg. Pathol.* 2017, *41*, 1155–1166. [CrossRef]
- 227. Ott, G. Aggressive B-cell lymphomas in the update of the 4th edition of the World Health Organization classification of haematopoietic and lymphatic tissues: Refinements of the classification, new entities and genetic findings. *Br. J. Haematol.* 2017, 178, 871–887. [CrossRef]
- 228. Hüttl, K.S.; Staiger, A.M.; Richter, J.; Ott, M.M.; Kalmbach, S.; Klapper, W.; Biesdorf, A.-S.; Trümper, L.; Rosenwald, A.; Ziepert, M.; et al. The "Burkitt-like" immunophenotype and genotype is rarely encountered in diffuse large B cell lymphoma and high-grade B cell lymphoma, NOS. *Virchows Arch.* 2021, 479, 575–583. [CrossRef]
- 229. Gonzalez de Villambrosia, S.; Bastos, M.; Palanca, J.M.; Cruz, J.G.; Navarro, J.-T.; Tapia, G.; Alonso, S.A.; Martin, A.; Blanco, O.; Abrisqueta, P.; et al. BCL2 translocation in high grade B cell lymphoma (NOS, DH/TH) is associated with reduced progression free survival. *Leuk. Lymphoma* 2022, 63, 101–108. [CrossRef]
- 230. Menter, T.; Juskevicius, D.; Alikian, M.; Steiger, J.; Dirnhofer, S.; Tzankov, A.; Naresh, K.N. Mutational landscape of B-cell post-transplant lymphoproliferative disorders. *Br. J. Haematol.* **2017**, *178*, 48–56. [CrossRef]
- 231. Chapman, J.R.; Bouska, A.C.; Zhang, W.; Alderuccio, J.P.; Lossos, I.S.; Rimsza, L.M.; Maguire, A.; Yi, S.; Chan, W.C.; Vega, F.; et al. EBV-positive HIV-associated diffuse large B cell lymphomas are characterized by JAK/STAT (STAT3) pathway mutations and unique clinicopathologic features. *Br. J. Haematol.* 2021, 194, 870–878. [CrossRef]
- Morscio, J.; Dierickx, D.; Ferreiro, J.F.; Herreman, A.; van Loo, P.; Bittoun, E.; Verhoef, G.; Matthys, P.; Cools, J.; Wlodarska, I.; et al. Gene expression profiling reveals clear differences between EBV-positive and EBV-negative posttransplant lymphoproliferative disorders. *Am. J. Transplant.* 2013, 13, 1305–1316. [CrossRef]
- 233. Ferreiro, J.F.; Morscio, J.; Dierickx, D.; Vandenberghe, P.; Gheysens, O.; Verhoef, G.; Zamani, M.; Tousseyn, T.; Wlodarska, I. EBV-Positive and EBV-Negative Posttransplant Diffuse Large B Cell Lymphomas Have Distinct Genomic and Transcriptomic Features. Am. J. Transplant. 2016, 16, 414–425. [CrossRef]
- 234. Engels, E.A.; Pittaluga, S.; Whitby, D.; Rabkin, C.; Aoki, Y.; Jaffe, E.S.; Goedert, J.J. Immunoblastic lymphoma in persons with AIDS-associated Kaposi's sarcoma: A role for Kaposi's sarcoma-associated herpesvirus. *Mod. Pathol.* 2003, *16*, 424–429. [CrossRef]

- 235. Courville, E.L.; Sohani, A.R.; Hasserjian, R.P.; Zukerberg, L.R.; Harris, N.L.; Ferry, J.A. Diverse clinicopathologic features in human herpesvirus 8-associated lymphomas lead to diagnostic problems. *Am. J. Clin. Pathol.* **2014**, *142*, 816–829. [CrossRef]
- 236. Gonzalez-Farre, B.; Martinez, D.; Lopez-Guerra, M.; Xipell, M.; Monclus, E.; Rovira, J.; Garcia, F.; Lopez-Guillermo, A.; Colomo, L.; Campo, E.; et al. HHV8-related lymphoid proliferations: A broad spectrum of lesions from reactive lymphoid hyperplasia to overt lymphoma. *Mod. Pathol.* 2017, *30*, 745–760. [CrossRef]

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