



Review The Role of miR-155 in Antitumor Immunity

Katerina Kalkusova¹, Pavla Taborska¹, Dmitry Stakheev^{1,2} and Daniel Smrz^{1,2,*}

- ¹ Department of Immunology, Second Faculty of Medicine, Charles University and Motol University Hospital, 150 06 Prague, Czech Republic
- ² Laboratory of Immunotherapy, Institute of Microbiology of the Czech Academy of Sciences, 142 20 Prague, Czech Republic
- * Correspondence: daniel.smrz@lfmotol.cuni.cz; Tel.: +420-224-43-5968; Fax: +420-224-435-962

Simple Summary: MiR-155 appears to be a significant regulator of immune responses against tumors. The aim of this review is to provide an overview of miR-155 function in distinct immune cell populations and describe how the miR-155-mediated regulation can impact the process of cancer immunoediting. Since miR-155 does not necessarily act only within a single cell, we also touch on the role of miR-155 in cellular communication. Finally, we discuss current developments in the specific targeting of this molecule in the tumor and immune cells and suggest its potential implications in developing novel therapeutic algorithms to increase the efficacy of cancer therapy.

Abstract: MicroRNAs belong to a group of short non-coding RNA molecules that are involved in the regulation of gene expression at multiple levels. Their function was described two decades ago, and, since then, microRNAs have become a rapidly developing field of research. Their participation in the regulation of cellular processes, such as proliferation, apoptosis, cell growth, and migration, made microRNAs attractive for cancer research. Moreover, as a single microRNA can simultaneously target multiple molecules, microRNAs offer a unique advantage in regulating multiple cellular processes in different cell types. Many of these cell types are tumor cells and the cells of the immune system. One of the most studied microRNAs in the context of cancer and the immune system is miR-155. MiR-155 plays a role in modulating innate and adaptive immune mechanisms in distinct immune cell types. As such, miR-155 can be part of the communication between the tumor and immune cells and thus impact the process of tumor immunoediting. Several studies have already revealed its effect on antitumor immune responses, and the targeting of this molecule is increasingly implemented in cancer immunotherapy. In this review, we discuss the current knowledge of miR-155 in the regulation of antitumor immunity and the shaping of the tumor microenvironment, and the plausible implementation of miR-155 targeting in cancer therapy.

Keywords: microRNA; immunity; tumors; cancer; immunotherapy; miR-155

1. Introduction

The immune system is responsible for maintaining the body's homeostasis. This maintenance also includes the recognition and elimination of transformed, malignant cells [1]. However, the immune system could be hijacked by the transformed cells and, on the contrary, promote their proliferation and survival in the human body. Many regulatory molecules are known to impact the balance between the pro- and antitumorigenic activities of the immune system. Although many of these molecules are proteins, there are also nonprotein molecules, such as non-coding RNAs (ncRNAs), which can substantially contribute to the balance [1–3]. One group of ncRNAs is microRNAs (miRNAs). MiRNAs were found to have a substantial regulatory impact on many distinct cellular processes, and research on these molecules has been gaining increasing interest in the last two decades. As miRNAs are often associated with the regulation of important cellular processes, such as apoptosis and proliferation [4,5], which are dysregulated during tumorigenesis, it is no surprise



Citation: Kalkusova, K.; Taborska, P.; Stakheev, D.; Smrz, D. The Role of miR-155 in Antitumor Immunity. *Cancers* 2022, *14*, 5414. https:// doi.org/10.3390/cancers14215414

Academic Editors: Antonio Curti, Alessandro Isidori, Giuseppe Lo Russo and Marina Chiara Garassino

Received: 30 September 2022 Accepted: 31 October 2022 Published: 3 November 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). that a large number of studies have been performed in the field of cancer research [6,7]. Nevertheless, despite their direct tumor suppressor or oncogenic functions, miRNAs also participate in tumorigenesis by regulating the immune system [8].

The function of miRNAs is decreasing the messenger RNA (mRNA) levels of proteincoding genes [9]. MiRNAs also participate in the post-transcriptional regulation of many targets, including other regulatory molecules or even transcription factors [10], which make miRNAs powerful players in the biology of the cell. Moreover, their distal transport in extracellular vesicles allows them to be involved not only in the regulation of the single cell in which they are transcribed but also in shaping the transcriptomes of other cells, thereby forming a mode of distal cellular communication [11]. One of the miRNAs participating in all the above-described mechanisms is miR-155, whose role in the tumor immune response is discussed in detail in this review.

2. MiRNA Biology

MiRNAs are small (approximately 22 nucleotides), single-stranded ncRNA molecules that participate in the regulation of gene expression at several levels and in cellular communication [12,13]. MiRNAs are evolutionarily conserved among species and have been identified in plants, animals, and even viruses [14,15]. These molecules were discovered almost thirty years ago in *Caenorhabditis elegans*. The first noted miRNA was lin-4, described as a negative regulator of the mRNA for the LIN-14 protein, which is essential for controlling developmental events in *C. elegans* [13,16,17]. The mechanism of its regulatory action was discovered to be based on the complementarity of lin-4 with the LIN-14 mRNA sequence and RNA–RNA interaction [16,17]. One decade after this, the function of these regulatory molecules was recognized. Initially, miRNAs were originally associated with post-transcriptional regulation [18]. However, the functions of miRNAs have been considerably expanded since then, as the mechanisms of their regulatory functions regarding gene expression were found to occur on multiple levels.

A mature miRNA creates a complex with proteins, namely the ribonucleoprotein (RNP), called the RNA-induced silencing complex (RISC) [19–21] (Figure 1). Being part of RNP, miRNA works as a sequence-specific guide that leads RNP to the target sequence. The target sequences are usually mRNAs but may also include other ncRNAs [22]. The miRNA complementarily binds at least seven nucleotides of the target sequence. Longer regions of complementarity are believed to strengthen the binding of the target sequence and miRNA [23]. The interaction of miRNA and target mRNA can result either in the mRNA degradation, its destabilization, or less efficient protein translation. These mechanisms are promoted by the proteins of the RISC complex. Degradation of mRNA is catalyzed by the argonaute protein [19,20], which is usually associated with its binding to the 3'UTR of mRNA [24]. If mRNA is not degraded, its translation can then be controlled on several levels, such as the inhibition of translation initiation, poly (A) shortening, decapping, or altered cap protein binding [25,26].

In contrast to 3'UTR mRNA targeting, binding to the 5'UTR is more often associated with the stabilization of the target mRNA, leading to increased protein translation [27,28]. In addition to mRNA binding, miRNAs can also affect DNA [29]. MiRNAs can even alter the structure of chromatin, which is another mechanism through which miRNAs regulate the expression of their targets [29,30]. The complexity of the gene expression regulation promoted by miRNAs is due to the fact that each miRNA is responsible for the regulation of multiple targets [31]. Moreover, these targets do not have to be transcribed in every cell. Thus, each miRNA can have a different impact on the protein expression in distinct cell types. Additionally, the expression levels of each miRNA differ among distinct cell populations, and they can even differ within a certain cell type depending on the stage of its differentiation. Generally, miRNA-mediated regulation is context dependent.

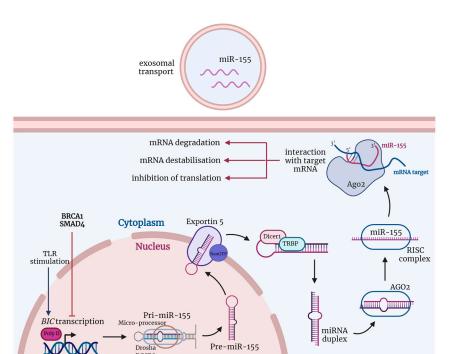


Figure 1. The biogenesis and function of miRNAs are shown via the example of miR-155. The noncoding B cell integration cluster (*BIC*), which encodes miR-155, is transcribed by polymerase II (poly II) into a double-stranded primary RNA transcript (pri-miR-155) after stimuli, such as stimulation of toll-like receptors (TLRs). Pri-miR-155 is cleaved by the ribonuclease—DROSHA and its cofactor binding protein, DGCR8—into precursor miRNA (pre-miR-155). Pre-miR-155 is transported by the GTP-dependent Exportin 5 from the nucleus to the cytoplasm, where the ribonuclease Dicer1 processes it into mature but still double-stranded miR-155. After this, miR-155 and argonaut 2 (AGO2), together with other proteins, create an RNA-induced silencing complex (RISC). RISC unwinds the double-stranded miR-155, and one of its strands is degraded while the second remains and acts as a sequence-specific guide for proteins. MiR-155 in the RISC complex promotes target mRNA degradation and destabilization or inhibits its translation. Created with BioRender.com (agreement number: DV24GT9WQS, accessed on 30 September 2022).

3. MiRNA Biogenesis

MiRNAs can be encoded in the genome as a single unit or they can be clustered. Clustered miRNAs are transcribed together as polycistronic transcripts that are processed into individual mature miRNAs that usually target mRNAs with the related function. MiRNAs are also often coded in introns and intergenic non-coding DNA sequences: "junk DNA" [25,32]. Moreover, their expression can be altered with alternative splicing. A number of miRNAs are classified into families determined by their 5′-end sequence homology [33]. The expression of miRNAs can be tissue specific, and their roles in the regulation of cell processes can differ depending on the cell type [33,34].

The biogenesis of miRNAs is a complex process that begins in the nucleus (Figure 1) [35]. MiRNAs are usually transcribed by RNA polymerase II into double-stranded primary RNA transcripts (pri-miRNAs) with a 5'cap and 3'poly (A) tail. Pri-miRNAs are significantly longer than their mature forms and need to be cleaved by a nuclear microprocessor containing ribonuclease (RNase III) Drosha and its cofactor double-stranded binding protein DiGeorge critical region 8 protein (DGCR8). In the nucleus, pri-miRNA is processed by this microprocessor complex into precursor miRNAs (pre-miRNAs) with a length of 60 to 70 nucleotides [36–38]. Thenceforth, pre-miRNAs are transported to the cytoplasm through GTP-dependent protein complex Exportin 5 (XPO5) [39,40]. In the cytoplasm, pre-miRNAs are cleaved and processed into their mature double-stranded miRNAs forms by the RNase III enzyme Dicer [41,42]. However, there are some alternative forms of this miRNA processing pathway [43,44]. The mature, but still double-stranded, miRNAs in

the cytoplasm create RISC containing AGO family proteins. One of the miRNA strands is degraded while the second strand remains within RISC [41]. When integrated into the RISC complex, miRNA interacts with complementary RNA and works as a sequence-specific guide for the proteins [45].

4. MiRNAs in Tumors and Antitumor Immunity

MiRNA-mediated regulation is important in numerous cellular processes, such as proliferation, differentiation, apoptosis, and metabolism [35]. Dysregulation of miRNAs' gene expression is involved in several diseases, including cancer. During tumor progression, miRNAs can act similarly as tumor suppressors or oncogenic molecules, depending on their targets [46]. MiRNAs with an oncogenic function are responsible for the downregulation of tumor suppressor genes while miRNAs that act as tumor suppressors are involved in the downregulation of oncogenes and their products [47]. Whereas upregulated levels of oncogenic miRNA are associated with oncogenic diseases, the tumor suppressor miRNA levels are often downregulated during tumorigenesis [48]. For this reason, miRNAs are very often considered to be potential biomarkers in cancer [49]. Moreover, they can also be valuable targets in cancer therapy [50–52].

MiRNAs are often transported from cells in vesicles called exosomes. These miRNA-transferring exosomes can be derived from cancer cells or, vice versa, from immune cells [53]. Recently, Wang et al. described exosomal crosstalk between cancer cells and tumor-associated fibroblasts and showed that exosomal miR-155, together with miR-146a, contributed to colorectal carcinoma metastasis [54]. The miR-155 exosomal crosstalk was also described between immune cells, such as dendritic cells [11]. This intercellular communication makes miRNAs a powerful tool in shaping the tumor's progression and microenvironment [53,55]. Therefore, although research on miRNAs has mostly remained within the field of oncology, these molecules have received considerable attention also in the field of immunology. MiRNAs were found to be involved in the regulation of immune cells [56,57], and since immune cells play a crucial role in antitumor defense mechanisms, a process called immunoediting [58], many miRNA-affecting immune cells are also involved in immunoediting, their involvement in which could be either pro- or antitumorigenic [7,59]. Under some circumstances, miRNAs can even act as immune checkpoint inhibitor molecules [60]. For this reason, all these characteristics make miRNAs an attractive study subject for research on immunoediting in cancer, and one of the currently most studied miRNAs in this field is miR-155.

5. MiR-155

MiR-155 is one of the most commonly studied miRNA molecules since its dysregulation is involved in many pathological processes. Its expression levels were shown to control pathways related to essential cellular processes, such as cell proliferation, apoptosis, differentiation, stemness, growth, migration, and angiogenesis [61,62]. Such broad regulatory potential in so many important cellular mechanisms makes miR-155 highly attractive for research and clinical applications [63].

As for genome localization, miR-155 is incorporated within the non-coding B cell integration cluster (*BIC*) gene, which, in humans, is located on chromosome 21. The sequence coding miR-155 is also known as the *MIR155* host gene (*MIR155HG*). The highest expression profile of miR-155 was found in the thymus and spleen, and increased levels of miR-155 were also found in CD34⁺ hematopoietic stem cells [64]. This finding led to the hypothesis that miR-155 could inhibit the differentiation of these cells [65]. The same study also suggested that a number of molecules participating in the hematopoietic differentiation are targets of miR-155 as their 3'UTR of mRNA includes the binding side for this miRNA. Similar miR-155 expression patterns were reported during erythropoiesis, where high expression levels of miR-155 were associated with lower differentiation states of erythroid cells, whereas their mature counterparts had these levels significantly decreased [66]. In addition, miR-155 plays a role in lymphocyte development [67,68]. Generally, miR-155

has been shown to play important roles in the regulation of myelopoiesis, lymphopoiesis, and erythropoiesis [65,69], and its dysregulation can negatively impact the development of cells derived from these lineages [70]. Recent studies also showed that miR-155 promotes the proliferation of tumor cells and inhibits their apoptosis through PTEN/PI3K/AKT targeting [62,71]. In addition, this mechanism was reported even upon cellular crosstalk when miR-155 was transferred in exosomes [71].

The function of miR-155 can be explained by its target genes. MiR-155 expression is regulated by several proteins that usually directly bind the promoter region of *BIC*. The tumor suppressor protein BRCA1 was described to negatively regulate the expression of miR-155 via inducing the deacetylation of histones on the miR-155 promoter [72]. Another transcriptional regulator of miR-155 is SMAD family member gene 4 (SMAD4), which can cause the repression of miR-155 expression as a result of TGF- β 1 signaling [73]. NF- κ B and TP53 were also shown to regulate miR-155 expression by directly interacting with the promoter [74,75]. Regulation of miR-155 levels does not necessarily always rely on interaction with the *BIC* promoter. The reduction of miR-155 levels can also be elicited post-transcriptionally by decreasing the stability of the miR-155 precursor and its maturation. An example of the negative regulation of miR-155 on the post-transcriptional level could be represented by the anti-inflammatory cytokine IL-10 [76,77].

MiR-155 is mostly considered an oncogenic miRNA in the field of oncology. In many studies, its upregulation was found to correlate with tumor occurrence and often with poor prognosis for many cancers [78,79]. Since miR-155 plays a role in the primary differentiation of myeloid progenitors, it is no surprise that its oncogenic property was mostly revealed in hematologic malignancies [80,81]. In addition, its oncogenic potential was also described in solid tumors, such as breast cancer [79,82,83] or gliomas [84].

6. MiR-155 in Antitumor Immunity

MiR-155 significantly impacts immune cells by targeting important regulatory molecules and transcriptional factors that regulate the immune system (Figure 2). Generally, the miR-155 expression increases after immune cell activation [85,86].

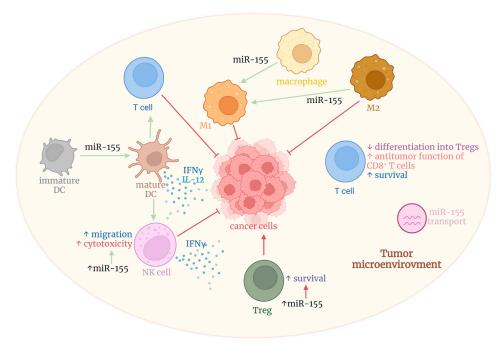


Figure 2. MiR-155-mediated shaping of the immune antitumor response. Created with BioRender. com (agreement number: RT24GT9RIE, accessed on 30 September 2022).

6.1. Danger Signals

MiR-155 levels increase in macrophages and dendritic cells after their response to danger signals: pathogen-associated molecular pattern (PAMP) or damage-associated molecular pattern (DAMP) molecules [86–89]. Several PAMP and DAMP molecules have been described in connection with miR-155 upregulation, such as lipopolysaccharide (LPS), polyinosinic–polycytidylic acid (poly I:C), and interferon beta (INF- β) [86,88,90]. This upregulation can be mediated by NF- κ B signaling [91] or the loss of Notch signaling [92]. High levels of miR-155 maintain the activated phenotypes of immune cells by inhibiting the expression of negative immune regulators, including inositol-polyphosphate-5-phosphatase-1 (SHIP-1) [93], and also contribute to the increased production of pro-inflammatory cytokines, such as TNF- α and IL-1 β [93].

6.2. Dendritic Cells

Dendritic cells (DCs) are professional antigen-presenting cells and linkers of innate and adaptive immunity, which is accomplished via their unique ability to induce naïve T cell activation [94]. Monocyte-derived DCs upregulate miR-155 during their maturation, which is suggested to be part of a negative feedback loop, since miR-155 targets the tolllike receptor and IL-1 inflammatory pathways [95]. Increased levels of miR-155 are also associated with the enhanced ability of DCs to induce T cell proliferation [96]. DCs lacking miR-155 display an antigen presentation deficiency, downregulated production of IL-12, and decreased expression of the chemokine receptor CCR7. Hence, these DCs display deficiencies in their maturation, migration, and ability to induce T cell activation [67,97–99]. MiR-155 targeting c-Fos was suggested to be the mechanism mediating DC maturation [100]. However, other miR-155 targets, such as the suppressor of cytokine signaling-1 (SOCS-1), arginase 1, and Jarid2, can contribute to these mechanisms as well [67,97–99].

DCs can amplify their functions through mutual communication, which is mainly promoted by exosomes. Functional microRNAs can be transferred in exosomes (nanovesicles) and thus influence the phenotype of the acceptor DC. Therefore, mature DCs can even affect the maturation phenotypes of other acceptor DCs [101]. Overall, the miR-155-induced maturation of DCs is associated with their antitumor phenotype, as it leads to the production of pro-inflammatory cytokines.

The most important regulatory role of miR-155 in DC functionality was further confirmed in other studies, both in vitro and in vivo. The in vitro studies showed that DCs treated with miR-155-enriched exosomes produced more IL-12 and IFN- γ , the cytokines that significantly contribute to the DCs' antitumor activities [102]. These exosome-treated DCs were found to be superior in inducing the differentiation, proliferation, and cytotoxicity of T cells [103]. Moreover, in vivo studies in a mouse model of colorectal cancer showed that adoptively transferred miR-155-treated DCs promoted increased tumor infiltration with cytotoxic and helper lymphocytes, whereas infiltration with regulatory lymphocytes was decreased [103]. These findings show that miR-155 in DCs appears to act in an antitumorigenic manner.

6.3. Macrophages

Macrophages are thought to be the most common myeloid cells present in the tumor microenvironment. Herein, the macrophages can display either a pro-tumorigenic or antitumorigenic role. Pro-tumorigenic (M2) macrophages support the growth of the primary tumor and the spread of metastases. The correlation between their abundance in the tumor and prognosis has been described for many cancers [104–106]. Antitumorigenic (M1) macrophages, on the other hand, can promote infiltration with cytotoxic lymphocytes or the antitumor activities of the tumor-infiltrating DCs [107]. MiR-155 was found to directly target and downregulate the phosphatase SHIP-1. The overexpression of miR-155 in macrophages was shown to repress SHIP-1 activity, leading to enhanced AKT signaling in stimulated cells [108]. In addition, the overexpression of miR-155 in tumor-associated macrophages (TAMs) led to macrophages' re-polarization into pro-inflammatory (antitumorigenic) M1 macrophages [109]. However, these data contrast with the role of SHIP-1 in macrophage differentiation because SHIP-1-deficient mice were found to profoundly skew macrophage differentiation into the M2 phenotype [110]. However, regardless of the role of SHIP-1, genetic miR-155 overexpression was demonstrated to promote the generation of M1-skewed TAMs [109].

Tumor-infiltrating macrophages are also frequent producers of exosomal miRNAs, which significantly shape the cellular crosstalk in the tumor microenvironment [111]. This mechanism is also used by tumors, as many tumor cells are also producers of miR-155, therefore impacting the immune cells in the tumor milieu [112,113].

6.4. Myeloid-Derived Suppressor Cells

MiR-155 impacts the functions of myeloid-derived suppressor cells (MDSCs), a population of immature myeloid cells with an immunosuppressive function [114]. The presence of MDSCs in the tumor microenvironment is often associated with worse antitumor immune responses and disease prognosis [115]. Loss of miR-155 in MDCSs was found to increase their migration and immunosuppressive potential [116]. This potential can even override the tumor growth inhibition mediated by the loss of miR-155 [117]. This override was demonstrated by a finding that showed that although the inhibition of miR-155 in the tumor cells suppressed their growth, the loss of miR-155 in the tumor microenvironment promoted tumor growth [117]. In contrast, the study by Li et al. suggested that the loss of miR-155 could be associated with worse MDSC proliferation by showing that the upregulation of miR-155 was linked to enhanced MDCS expansion [118]. Chen et al. also revealed that the loss of miR-155 was associated with the reduced infiltration of MDSCs into the tumor microenvironment [119]. Mechanistically, miR-155 was suggested to contribute to the MDSC suppressor activity in two ways: by inhibiting SOCS-1 and weakening the capacity of MDSCs to induce regulatory T cells (Tregs) [119]. However, the role of miR-155 in MDSCs is ambiguous, since miR-155 produced by leukemia was found to induce MDSCs and enhance their function [120].

6.5. Natural Killer Cells

Another important role in miR-155-mediated antitumor immunity is played by natural killer (NK) cells. The number of NK cells is often associated with favorable clinical outcomes of the disease [121,122]. NK cells are also regulated by miR-155. The miR-155 levels in NK cells were found to be elevated after their stimulation and associated with the production of IFN- γ , which was mechanistically promoted by the miR-155-mediated inhibition of SHIP-1 [123]. The miR-155-mediated inhibition of SHIP-1 was also found to regulate NK cell chemotaxis and migration. An miR-155 deficiency followed by SHIP-1 overexpression resulted in impaired F-actin cytoskeleton polymerization, which then negatively impacted to be associated with enhanced cytotoxicity of NK cells towards tumors [125]. Taken together, these findings indicate that miR-155 may be a positive regulator of NK cells in antitumor immunity and suggest an ability to migrate into the tumor microenvironment.

6.6. T Cells and Immune Checkpoint Inhibitors

Very early, in 2002, Haasch et al. found that miR-155 expression was significantly increased in CD4⁺ T cells after their activation [85]. Subsequent studies showed that this miRNA plays a crucial role in the proliferation and differentiation of lymphocytes [126,127]. MiR-155 was also found to promote CD4⁺ T cell differentiation into the Th17 phenotype [67,128], and this differentiation was induced by STAT3 [129]. Vice versa, CD4⁺ T cell differentiation into the Th2 phenotype is suppressed by the miR-155-mediated downregulation of c-Maf, which is a Th2 response-promoting transcription factor and an inhibitor of the Th1 response [130]. Therefore, miR-155 promotes the Th1 differentiation of CD4⁺ T cells [131,132].

MiR-155 also affects the function of Tregs. Tregs play an essential role in tumor immune escape as they promote a pro-tumorigenic environment via cytokine production and cell–cell communication [133,134]. MiR-155 is highly expressed in Tregs, and its decrease leads to a reduction in their numbers [135]. The Treg-specific transcriptional factor, FoxP3, also regulates BIC's transcription [136–138]. It was further demonstrated that miR-155 could enhance CD4⁺ T cell differentiation towards Tregs [139] and that reduced miR-155 levels in Tregs can shorten their survival [136]. However, contrasting data were obtained with an miR-155 inhibitor (antagomir), which was found to modulate the balance between Tregs and Th17 via the Jarid2/Wnt/ β -catenin pathway. The miR-155 inhibitor enhanced Tregs in vitro whereas Th17 cells were decreased [140]. Although contrasting, these findings point to the fact that miR-155 can significantly impact the functionality of Tregs and thus shape their role in antitumor immunity.

CD8⁺ T cells are critical effector cells of antitumor activities in the immune system [141]. These cells were found to be much less efficient in tumor growth control in the absence of miR-155 expression [142]. On the other hand, increased expression of miR-155 promoted their antitumor activity. The mechanism behind this regulation of CD8⁺ T cell effector function was based on the miR-155 targeting of SOCS-1 [142].

Immune checkpoint inhibitors such as cytotoxic T lymphocyte-associated antigen-4 (CTLA-4), programmed cell death protein 1 (PD-1), and its ligands PDL-1 and PDL-2 participate in the inhibition of T cell activation [143]. Inhibitors of these molecules enhance antitumor immune responses and are widely used in the current cancer immunotherapy approach, which is considered to be one of the most successful cancer immunotherapy strategies of the past decade [144]. The expression of these inhibitors can also be regulated by miR-155, as demonstrated in a study where the direct binding of miR-155 to the 3'UTR of PDL-1 mRNA downregulated the expression of this critical regulator of T cell function [145].

7. MiR-155 in Cancer Immunotherapy

Linking the above-described characteristics of miR-155 together, it is undoubtable that this molecule has potential clinical relevance in cancer therapy, especially in immunotherapy. There are many potential options for the usage of miR-155 in different immunotherapeutic modalities.

One of the most promising approaches relates to active cellular immunotherapy based on DCs. In these cells, miR-155 was shown to improve the efficacy of DC-based cancer vaccines in a mouse model. In this study, miR-155 overexpression in DCs enhanced their ability to increase CD8⁺ T cell antitumor responses [96]. In vivo, the proteasome inhibitor Bortezomib, approved for multiple myeloma treatments, was also found, in addition to other mechanisms, to induce the miR-155-mediated downregulation of SOCS-1 and SHIP-1. Suppression of these immune system regulators in the final stages led to the suppression of PD-1-mediated T cell exhaustion [146]. Another in vivo study showed that miR-155 derived from breast cancer cells was recently described to enhance the recruitment of antitumor immune cells, as the repression of SOCS-1 is associated with the upregulation of several chemokines. Moreover, miR-155 was reported to change a "cold tumor" into a "hot one" and thus sensitize the tumor for checkpoint blockage immunotherapy [147]. Bioinformatics analysis also revealed that higher levels of miR-155 in tumors were correlated with an enhanced antitumor immune profile and favorable outcomes. Moreover, the study also revealed that high miR-155 levels in serum were a good prognostic marker for breast cancer. The authors of this study suggested that increasing the levels of miR-155 in breast tumors could improve the efficacy of cancer immunotherapy by increasing tumor infiltration with immune cells [147]. Indeed, the potential of miR-155 as a predictive biomarker for immunotherapeutic efficacy has already been reported [148]. The mechanisms involved in this process are presumably based on the potential of miR-155 to modify the tumor microenvironment, namely the immune cells within. An example of this modification was demonstrated by the repolarization of the tumor-infiltrating macrophages (TAMs) [149]. This study showed that miR-155-overexpressor redox/pH dual-responsive hybrid polypeptide nanovectors that targeted TAMs repolarized immunosuppressive macrophages into M1 macrophages with antitumor properties. These properties were displayed via robust tumor regression, which was also associated with increased T and NK cell activation in the regressing tumors [149].

On the other hand, another study suggested that miR-155 should be therapeutically downregulated because high levels of miR-155 were found to be associated with cancer resistance to chemotherapy and radiotherapy [150]. Therefore, under these circumstances, miR-155 inhibitors might seem to represent a novel therapeutic approach to sensitizing tumors to chemotherapy or radiotherapy [150]. This avenue is tempting and might appear promising, but, as demonstrated by other studies discussed above, the performance of miR-155 in tumors can be ambivalent, and the flat downregulation of miR-155 can trigger multiple mechanisms with different impacts on the disease state. Downregulation of miR-155 could indeed sensitize tumors to chemotherapy or radiotherapy or radiotherapy on the one hand, but, on the other, this intervention could remodel the tumor's immune microenvironment, and the tumor could become resistant to the immune system and immunotherapy. The outcome of this flat intervention could finally be the chemo/radiotherapy-induced disappearance of large masses of the tumor while simultaneously rendering the remaining (surviving) tumor mass more resistant to the immune system and immunotherapy, which could, in the end, cause a disease relapse.

8. Future Perspectives

A large number of studies related to the field of cancer research have shown the oncogenic nature of miR-155 [83,151]. Being upregulated in many cancers and promoting the stemness of cancer cells, which is strongly associated with poor prognosis, and increasing their resistance to chemotherapy and radiotherapy [150], the systematic downregulation of miR-155 was initially a convenient therapeutic approach. However, being deeply immersed in immune system regulation and participating in the antitumor response, mainly by affecting many distinct immune cell populations, the harvesting of the regulatory powers of miR-155 for cancer therapy needs to be reconsidered based on the implementation of novel therapeutic algorithms. These algorithms should stem not only from the disease severity, including the tumor staging and grading, but also from the tumoral/paratumoral immune signatures inflicted by the disease or its treatment [152–156]. Guided by these signatures, the design of the therapeutic algorithm could then even include both miR-155 downregulation and upregulation, each corresponding to the time of treatment and the type of therapeutic modality. For instance, intratumoral miR-155 downregulation could be initially indicated as a neoadjuvant therapy before chemotherapy to sensitize the tumor to chemotherapy or radiotherapy [150]. Then, following chemotherapy or radiotherapy, miR-155 upregulation in a specific immune cell population, such as tumor-infiltrating macrophages [149], could become the basis of adjuvant immunotherapy.

However, these algorithms will presumably require the precise targeting of miR-155 in specific cell types, namely the immune cell populations. This precision targeting could rely on novel technologies, such as nanovectors or adoptive cellular immunotherapy, where individual immune cell types could be produced ex vivo and modified accordingly. Such differential miR-155 targeting could be a promising avenue to enhance antitumor immunity and the efficacy of cancer therapy.

9. Conclusions

MiR-155 is involved in the regulation of immunoediting processes on many levels, participates in changes in cancer cells and immune cells, and plays a role in their crosstalk, and its expression levels can significantly shape the severity and therapeutic resistance of the patient's disease. These characteristics make miR-155 a potential target in cancer diagnostics, prognostics, and treatment. This review showed the promising implications of miR-155 targeting in cancer therapy. With an increasing understanding of miR-155's functions in both cancer and immune cells and its precision targeting using newly developing

technologies, miR-155 could become an immune checkpoint molecule of interest. As such, its targeting could be implemented in novel algorithms of cancer therapy in the near future.

Author Contributions: K.K. and D.S. (Daniel Smrz) conceptualized and wrote the original draft. P.T. and D.S. (Dmitry Stakheev) reviewed and edited the article. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported by the Ministry of Health, Czech Republic—Project AZV NU22-03-00300 and the Institutional IPE2 funding of the Charles University, Second Faculty of Medicine.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Abbott, M.; Ustoyev, Y. Cancer and the Immune System: The History and Background of Immunotherapy. *Semin. Oncol. Nurs.* 2019, 35, 150923. [CrossRef] [PubMed]
- Denaro, N.; Merlano, M.C.; Lo Nigro, C. Long noncoding RNAs as regulators of cancer immunity. *Mol. Oncol.* 2019, 13, 61–73. [CrossRef] [PubMed]
- 3. Papaioannou, E.; Gonzalez-Molina, M.D.P.; Prieto-Munoz, A.M.; Gamez-Reche, L.; Gonzalez-Martin, A. Regulation of Adaptive Tumor Immunity by Non-Coding RNAs. *Cancers* **2021**, *13*, 5651. [CrossRef] [PubMed]
- 4. Bueno, M.J.; Perez de Castro, I.; Malumbres, M. Control of cell proliferation pathways by microRNAs. *Cell Cycle* 2008, 7, 3143–3148. [CrossRef] [PubMed]
- 5. Yang, B.F.; Lu, Y.J.; Wang, Z.G. MicroRNAs and apoptosis: Implications in the molecular therapy of human disease. *Clin. Exp. Pharmacol. Physiol.* **2009**, *36*, 951–960. [CrossRef]
- De Paolis, V.; Lorefice, E.; Orecchini, E.; Carissimi, C.; Laudadio, I.; Fulci, V. Epitranscriptomics: A New Layer of microRNA Regulation in Cancer. *Cancers* 2021, 13, 3372. [CrossRef]
- Yi, M.; Xu, L.; Jiao, Y.; Luo, S.; Li, A.; Wu, K. The role of cancer-derived microRNAs in cancer immune escape. *J. Hematol. Oncol.* 2020, 13, 25. [CrossRef]
- Pottoo, F.H.; Iqubal, A.; Iqubal, M.K.; Salahuddin, M.; Rahman, J.U.; AlHajri, N.; Shehadeh, M. miRNAs in the Regulation of Cancer Immune Response: Effect of miRNAs on Cancer Immunotherapy. *Cancers* 2021, 13, 6145. [CrossRef]
- Rani, V.; Sengar, R.S. Biogenesis and mechanisms of microRNA-mediated gene regulation. *Biotechnol. Bioeng.* 2022, 119, 685–692. [CrossRef]
- 10. Zhang, H.M.; Kuang, S.; Xiong, X.; Gao, T.; Liu, C.; Guo, A.Y. Transcription factor and microRNA co-regulatory loops: Important regulatory motifs in biological processes and diseases. *Brief. Bioinform.* **2015**, *16*, 45–58. [CrossRef]
- Alexander, M.; Hu, R.; Runtsch, M.C.; Kagele, D.A.; Mosbruger, T.L.; Tolmachova, T.; Seabra, M.C.; Round, J.L.; Ward, D.M.; O'Connell, R.M. Exosome-delivered microRNAs modulate the inflammatory response to endotoxin. *Nat. Commun.* 2015, *6*, 7321. [CrossRef] [PubMed]
- 12. Acunzo, M.; Romano, G.; Wernicke, D.; Croce, C.M. MicroRNA and cancer—A brief overview. *Adv. Biol. Regul.* 2015, 57, 7321. [CrossRef] [PubMed]
- 13. Dexheimer, P.J.; Cochella, L. MicroRNAs: From Mechanism to Organism. Front. Cell Dev. Biol. 2020, 8, 409. [CrossRef]
- 14. Li, Z.; Xu, R.; Li, N. MicroRNAs from plants to animals, do they define a new messenger for communication? *Nutr. Metab.* **2018**, 15, 68. [CrossRef] [PubMed]
- 15. Mishra, R.; Kumar, A.; Ingle, H.; Kumar, H. The Interplay Between Viral-Derived miRNAs and Host Immunity During Infection. *Front. Immunol.* **2019**, *10*, 3079. [CrossRef]
- 16. Lee, R.C.; Feinbaum, R.L.; Ambros, V. The *C. elegans* heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14. *Cell* **1993**, *75*, 843–854. [CrossRef]
- 17. Wightman, B.; Ha, I.; Ruvkun, G. Posttranscriptional regulation of the heterochronic gene lin-14 by lin-4 mediates temporal pattern formation in *C. elegans. Cell* **1993**, *75*, 855–862. [CrossRef]
- 18. Meister, G.; Tuschl, T. Mechanisms of gene silencing by double-stranded RNA. *Nature* **2004**, 431, 343–349. [CrossRef]
- 19. Hutvagner, G.; Zamore, P.D. A microRNA in a multiple-turnover RNAi enzyme complex. Science 2002, 297, 2056–2060. [CrossRef]
- Mourelatos, Z.; Dostie, J.; Paushkin, S.; Sharma, A.; Charroux, B.; Abel, L.; Rappsilber, J.; Mann, M.; Dreyfuss, G. miRNPs: A novel class of ribonucleoproteins containing numerous microRNAs. *Genes Dev.* 2002, *16*, 720–728. [CrossRef]
- 21. Tang, G. siRNA and miRNA: An insight into RISCs. Trends Biochem. Sci. 2005, 30, 106–114. [CrossRef] [PubMed]
- Valencia-Sanchez, M.A.; Liu, J.; Hannon, G.J.; Parker, R. Control of translation and mRNA degradation by miRNAs and siRNAs. *Genes Dev.* 2006, 20, 515–524. [CrossRef] [PubMed]
- 23. Mohr, A.M.; Mott, J.L. Overview of microRNA biology. Semin. Liver Dis. 2015, 35, 3–11. [CrossRef] [PubMed]
- 24. Lai, E.C. Micro RNAs are complementary to 3' UTR sequence motifs that mediate negative post-transcriptional regulation. *Nat. Genet.* **2002**, *30*, 363–364. [CrossRef] [PubMed]
- 25. Filipowicz, W.; Bhattacharyya, S.N.; Sonenberg, N. Mechanisms of post-transcriptional regulation by microRNAs: Are the answers in sight? *Nat. Rev. Genet.* 2008, *9*, 102–114. [CrossRef]

- Guo, H.; Ingolia, N.T.; Weissman, J.S.; Bartel, D.P. Mammalian microRNAs predominantly act to decrease target mRNA levels. *Nature* 2010, 466, 835–840. [CrossRef]
- Henke, J.I.; Goergen, D.; Zheng, J.; Song, Y.; Schuttler, C.G.; Fehr, C.; Junemann, C.; Niepmann, M. microRNA-122 stimulates translation of hepatitis C virus RNA. *EMBO J.* 2008, 27, 3300–3310. [CrossRef]
- Moretti, F.; Thermann, R.; Hentze, M.W. Mechanism of translational regulation by miR-2 from sites in the 5' untranslated region or the open reading frame. RNA 2010, 16, 2493–2502. [CrossRef]
- 29. Catalanotto, C.; Cogoni, C.; Zardo, G. MicroRNA in Control of Gene Expression: An Overview of Nuclear Functions. *Int. J. Mol. Sci.* 2016, 17, 1712. [CrossRef]
- Salmanidis, M.; Pillman, K.; Goodall, G.; Bracken, C. Direct transcriptional regulation by nuclear microRNAs. Int. J. Biochem. Cell Biol. 2014, 54, 304–311. [CrossRef]
- Lim, L.P.; Lau, N.C.; Garrett-Engele, P.; Grimson, A.; Schelter, J.M.; Castle, J.; Bartel, D.P.; Linsley, P.S.; Johnson, J.M. Microarray analysis shows that some microRNAs downregulate large numbers of target mRNAs. *Nature* 2005, 433, 769–773. [CrossRef] [PubMed]
- 32. Slack, F.J. Regulatory RNAs and the demise of 'junk' DNA. Genome Biol. 2006, 7, 328. [CrossRef] [PubMed]
- Lagos-Quintana, M.; Rauhut, R.; Yalcin, A.; Meyer, J.; Lendeckel, W.; Tuschl, T. Identification of tissue-specific microRNAs from mouse. *Curr. Biol.* 2002, 12, 735–739. [CrossRef]
- Londin, E.; Loher, P.; Telonis, A.G.; Quann, K.; Clark, P.; Jing, Y.; Hatzimichael, E.; Kirino, Y.; Honda, S.; Lally, M.; et al. Analysis of 13 cell types reveals evidence for the expression of numerous novel primate- and tissue-specific microRNAs. *Proc. Natl. Acad. Sci. USA* 2015, *112*, E1106–E1115. [CrossRef]
- 35. Bartel, D.P. MicroRNAs: Genomics, biogenesis, mechanism, and function. Cell 2004, 116, 281–297. [CrossRef]
- Chen, C.Z.; Li, L.; Lodish, H.F.; Bartel, D.P. MicroRNAs modulate hematopoietic lineage differentiation. *Science* 2004, 303, 83–86. [CrossRef]
- Lee, Y.; Jeon, K.; Lee, J.T.; Kim, S.; Kim, V.N. MicroRNA maturation: Stepwise processing and subcellular localization. *EMBO J.* 2002, 21, 4663–4670. [CrossRef]
- Lee, Y.; Ahn, C.; Han, J.; Choi, H.; Kim, J.; Yim, J.; Lee, J.; Provost, P.; Radmark, O.; Kim, S.; et al. The nuclear RNase III Drosha initiates microRNA processing. *Nature* 2003, 425, 415–419. [CrossRef]
- Denli, A.M.; Tops, B.B.; Plasterk, R.H.; Ketting, R.F.; Hannon, G.J. Processing of primary microRNAs by the Microprocessor complex. *Nature* 2004, 432, 231–235. [CrossRef]
- 40. Yi, R.; Qin, Y.; Macara, I.G.; Cullen, B.R. Exportin-5 mediates the nuclear export of pre-microRNAs and short hairpin RNAs. *Genes Dev.* **2003**, *17*, 3011–3016. [CrossRef]
- 41. Hammond, S.M.; Boettcher, S.; Caudy, A.A.; Kobayashi, R.; Hannon, G.J. Argonaute2, a link between genetic and biochemical analyses of RNAi. *Science* 2001, 293, 1146–1150. [CrossRef] [PubMed]
- 42. Saito, K.; Ishizuka, A.; Siomi, H.; Siomi, M.C. Processing of pre-microRNAs by the Dicer-1-Loquacious complex in Drosophila cells. *PLoS Biol.* 2005, *3*, e235. [CrossRef] [PubMed]
- Cheloufi, S.; Dos Santos, C.O.; Chong, M.M.; Hannon, G.J. A dicer-independent miRNA biogenesis pathway that requires Ago catalysis. *Nature* 2010, 465, 584–589. [CrossRef] [PubMed]
- Cifuentes, D.; Xue, H.; Taylor, D.W.; Patnode, H.; Mishima, Y.; Cheloufi, S.; Ma, E.; Mane, S.; Hannon, G.J.; Lawson, N.D.; et al. A novel miRNA processing pathway independent of Dicer requires Argonaute2 catalytic activity. *Science* 2010, 328, 1694–1698. [CrossRef]
- 45. Kertesz, M.; Iovino, N.; Unnerstall, U.; Gaul, U.; Segal, E. The role of site accessibility in microRNA target recognition. *Nat. Genet.* **2007**, *39*, 1278–1284. [CrossRef]
- Wu, W.; Sun, M.; Zou, G.M.; Chen, J. MicroRNA and cancer: Current status and prospective. *Int. J. Cancer* 2007, 120, 953–960. [CrossRef]
- 47. Shenouda, S.K.; Alahari, S.K. MicroRNA function in cancer: Oncogene or a tumor suppressor? *Cancer Metastasis Rev.* 2009, 28, 369–378. [CrossRef]
- 48. Oliveto, S.; Mancino, M.; Manfrini, N.; Biffo, S. Role of microRNAs in translation regulation and cancer. *World J. Biol. Chem.* 2017, *8*, 45–56. [CrossRef]
- 49. Condrat, C.E.; Thompson, D.C.; Barbu, M.G.; Bugnar, O.L.; Boboc, A.; Cretoiu, D.; Suciu, N.; Cretoiu, S.M.; Voinea, S.C. miRNAs as Biomarkers in Disease: Latest Findings Regarding Their Role in Diagnosis and Prognosis. *Cells* **2020**, *9*, 276. [CrossRef]
- Karimzadeh, M.R.; Pourdavoud, P.; Ehtesham, N.; Qadbeigi, M.; Asl, M.M.; Alani, B.; Mosallaei, M.; Pakzad, B. Regulation of DNA methylation machinery by epi-miRNAs in human cancer: Emerging new targets in cancer therapy. *Cancer Gene Ther.* 2021, 28, 157–174. [CrossRef]
- Shah, V.; Shah, J. Recent trends in targeting miRNAs for cancer therapy. J. Pharm. Pharmacol. 2020, 72, 1732–1749. [CrossRef] [PubMed]
- Sohrabi, B.; Dayeri, B.; Zahedi, E.; Khoshbakht, S.; Nezamabadi Pour, N.; Ranjbar, H.; Davari Nejad, A.; Noureddini, M.; Alani, B. Mesenchymal stem cell (MSC)-derived exosomes as novel vehicles for delivery of miRNAs in cancer therapy. *Cancer Gene Ther.* 2022, 29, 1105–1116. [CrossRef] [PubMed]
- Bauer, K.M.; Round, J.L.; O'Connell, R.M. No small matter: Emerging roles for exosomal miRNAs in the immune system. *FEBS J.* 2022, 289, 4021–4037. [CrossRef] [PubMed]

- Wang, D.; Wang, X.; Song, Y.; Si, M.; Sun, Y.; Liu, X.; Cui, S.; Qu, X.; Yu, X. Exosomal miR-146a-5p and miR-155-5p promote CXCL12/CXCR7-induced metastasis of colorectal cancer by crosstalk with cancer-associated fibroblasts. *Cell Death Dis.* 2022, 13, 380. [CrossRef]
- 55. Di Martino, M.T.; Riillo, C.; Scionti, F.; Grillone, K.; Polera, N.; Caracciolo, D.; Arbitrio, M.; Tagliaferri, P.; Tassone, P. miRNAs and lncRNAs as Novel Therapeutic Targets to Improve Cancer Immunotherapy. *Cancers* 2021, 13, 1587. [CrossRef]
- O'Connell, R.M.; Kahn, D.; Gibson, W.S.; Round, J.L.; Scholz, R.L.; Chaudhuri, A.A.; Kahn, M.E.; Rao, D.S.; Baltimore, D. MicroRNA-155 promotes autoimmune inflammation by enhancing inflammatory T cell development. *Immunity* 2010, 33, 607–619. [CrossRef]
- 57. Xiao, C.; Rajewsky, K. MicroRNA control in the immune system: Basic principles. Cell 2009, 136, 26–36. [CrossRef]
- Dunn, G.P.; Old, L.J.; Schreiber, R.D. The three Es of cancer immunoediting. *Annu. Rev. Immunol.* 2004, 22, 329–360. [CrossRef]
 Cortez, M.A.; Anfossi, S.; Ramapriyan, R.; Menon, H.; Atalar, S.C.; Aliru, M.; Welsh, J.; Calin, G.A. Role of miRNAs in immune
- responses and immunotherapy in cancer. *Genes Chromosomes Cancer* **2019**, *58*, 244–253. [CrossRef]
- Nguyen, H.T.; Phung, C.D.; Tran, T.H.; Pham, T.T.; Pham, L.M.; Nguyen, T.T.; Jeong, J.H.; Choi, H.G.; Ku, S.K.; Yong, C.S.; et al. Manipulating immune system using nanoparticles for an effective cancer treatment: Combination of targeted therapy and checkpoint blockage miRNA. J. Control. Release Off. J. Control. Release Soc. 2021, 329, 524–537. [CrossRef]
- 61. Catela Ivkovic, T.; Voss, G.; Cornella, H.; Ceder, Y. microRNAs as cancer therapeutics: A step closer to clinical application. *Cancer Lett.* **2017**, 407, 113–122. [CrossRef]
- Zuo, W.N.; Zhu, H.; Li, L.P.; Jin, A.Y.; Wang, H.Q. MiR-155 promotes proliferation and inhibits apoptosis of nasopharyngeal carcinoma cells through targeting PTEN-PI3K/AKT pathway. *Eur. Rev. Med. Pharmacol. Sci.* 2019, 23, 7935–7942. [CrossRef] [PubMed]
- 63. Witten, L.; Slack, F.J. miR-155 as a novel clinical target for hematological malignancies. *Carcinogenesis* **2020**, *41*, 2–7. [CrossRef] [PubMed]
- 64. Tam, W. Identification and characterization of human BIC, a gene on chromosome 21 that encodes a noncoding RNA. *Gene* **2001**, 274, 157–167. [CrossRef]
- Georgantas, R.W., 3rd; Hildreth, R.; Morisot, S.; Alder, J.; Liu, C.G.; Heimfeld, S.; Calin, G.A.; Croce, C.M.; Civin, C.I. CD34+ hematopoietic stem-progenitor cell microRNA expression and function: A circuit diagram of differentiation control. *Proc. Natl. Acad. Sci. USA* 2007, 104, 2750–2755. [CrossRef] [PubMed]
- 66. Masaki, S.; Ohtsuka, R.; Abe, Y.; Muta, K.; Umemura, T. Expression patterns of microRNAs 155 and 451 during normal human erythropoiesis. *Biochem. Biophys. Res. Commun.* 2007, 364, 509–514. [CrossRef]
- 67. Rodriguez, A.; Vigorito, E.; Clare, S.; Warren, M.V.; Couttet, P.; Soond, D.R.; van Dongen, S.; Grocock, R.J.; Das, P.P.; Miska, E.A.; et al. Requirement of bic/microRNA-155 for normal immune function. *Science* **2007**, *316*, 608–611. [CrossRef] [PubMed]
- Vigorito, E.; Perks, K.L.; Abreu-Goodger, C.; Bunting, S.; Xiang, Z.; Kohlhaas, S.; Das, P.P.; Miska, E.A.; Rodriguez, A.; Bradley, A.; et al. microRNA-155 regulates the generation of immunoglobulin class-switched plasma cells. *Immunity* 2007, 27, 847–859. [CrossRef]
- 69. Johanson, T.M.; Skinner, J.P.; Kumar, A.; Zhan, Y.; Lew, A.M.; Chong, M.M. The role of microRNAs in lymphopoiesis. *Int. J. Hematol.* **2014**, 100, 246–253. [CrossRef]
- 70. Ranganath, P. MicroRNA-155 and Its Role in Malignant Hematopoiesis. Biomark. Insights 2015, 10, 95–102. [CrossRef]
- Sun, J.F.; Zhang, D.; Gao, C.J.; Zhang, Y.W.; Dai, Q.S. Exosome-Mediated MiR-155 Transfer Contributes to Hepatocellular Carcinoma Cell Proliferation by Targeting PTEN. *Med. Sci. Monit. Basic Res.* 2019, 25, 218–228. [CrossRef] [PubMed]
- Chang, S.; Wang, R.H.; Akagi, K.; Kim, K.A.; Martin, B.K.; Cavallone, L.; Kathleen Cuningham Foundation Consortium for Research into Familial Breast Cancer (kConFab); Haines, D.C.; Basik, M.; Mai, P.; et al. Tumor suppressor BRCA1 epigenetically controls oncogenic microRNA-155. *Nat. Med.* 2011, *17*, 1275–1282. [CrossRef] [PubMed]
- Zhao, H.; Zhang, J.; Shao, H.; Liu, J.; Jin, M.; Chen, J.; Huang, Y. Transforming Growth Factor beta1/Smad4 Signaling Affects Osteoclast Differentiation via Regulation of miR-155 Expression. *Mol. Cells* 2017, 40, 211–221. [CrossRef] [PubMed]
- Gerloff, D.; Grundler, R.; Wurm, A.A.; Brauer-Hartmann, D.; Katzerke, C.; Hartmann, J.U.; Madan, V.; Muller-Tidow, C.; Duyster, J.; Tenen, D.G.; et al. NF-kappaB/STAT5/miR-155 network targets PU.1 in FLT3-ITD-driven acute myeloid leukemia. *Leuk. Off. J. Leuk. Soc. Am. Leuk. Res. Fund* 2015, 29, 535–547. [CrossRef]
- Van Roosbroeck, K.; Fanini, F.; Setoyama, T.; Ivan, C.; Rodriguez-Aguayo, C.; Fuentes-Mattei, E.; Xiao, L.; Vannini, I.; Redis, R.S.; D'Abundo, L.; et al. Combining Anti-Mir-155 with Chemotherapy for the Treatment of Lung Cancers. *Clin. Cancer Res.* 2017, 23, 2891–2904. [CrossRef]
- Cheung, S.T.; So, E.Y.; Chang, D.; Ming-Lum, A.; Mui, A.L. Interleukin-10 inhibits lipopolysaccharide induced miR-155 precursor stability and maturation. *PLoS ONE* 2013, 8, e71336. [CrossRef]
- McCoy, C.E.; Sheedy, F.J.; Qualls, J.E.; Doyle, S.L.; Quinn, S.R.; Murray, P.J.; O'Neill, L.A. IL-10 inhibits miR-155 induction by toll-like receptors. J. Biol. Chem. 2010, 285, 20492–20498. [CrossRef]
- 78. Yang, M.; Shen, H.; Qiu, C.; Ni, Y.; Wang, L.; Dong, W.; Liao, Y.; Du, J. High expression of miR-21 and miR-155 predicts recurrence and unfavourable survival in non-small cell lung cancer. *Eur. J. Cancer* **2013**, *49*, 604–615. [CrossRef]
- 79. Kong, W.; He, L.; Richards, E.J.; Challa, S.; Xu, C.X.; Permuth-Wey, J.; Lancaster, J.M.; Coppola, D.; Sellers, T.A.; Djeu, J.Y.; et al. Upregulation of miRNA-155 promotes tumour angiogenesis by targeting VHL and is associated with poor prognosis and triple-negative breast cancer. *Oncogene* 2014, *33*, 679–689. [CrossRef]

- Cui, B.; Chen, L.; Zhang, S.; Mraz, M.; Fecteau, J.F.; Yu, J.; Ghia, E.M.; Zhang, L.; Bao, L.; Rassenti, L.Z.; et al. MicroRNA-155 influences B-cell receptor signaling and associates with aggressive disease in chronic lymphocytic leukemia. *Blood* 2014, 124, 546–554. [CrossRef]
- 81. Eis, P.S.; Tam, W.; Sun, L.; Chadburn, A.; Li, Z.; Gomez, M.F.; Lund, E.; Dahlberg, J.E. Accumulation of miR-155 and BIC RNA in human B cell lymphomas. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 3627–3632. [CrossRef] [PubMed]
- Jiang, S.; Zhang, H.W.; Lu, M.H.; He, X.H.; Li, Y.; Gu, H.; Liu, M.F.; Wang, E.D. MicroRNA-155 functions as an OncomiR in breast cancer by targeting the suppressor of cytokine signaling 1 gene. *Cancer Res* 2010, 70, 3119–3127. [CrossRef]
- Mattiske, S.; Suetani, R.J.; Neilsen, P.M.; Callen, D.F. The oncogenic role of miR-155 in breast cancer. *Cancer Epidemiol. Biomark.* Prev. 2012, 21, 1236–1243. [CrossRef] [PubMed]
- Zhou, Y.; Wang, X.; Liu, Z.; Huang, X.; Li, X.; Cheng, K.; Jiang, X. Prognostic role of microRNA-155 expression in gliomas: A meta-analysis. *Clin. Neurol. Neurosurg.* 2019, 176, 103–109. [CrossRef] [PubMed]
- Haasch, D.; Chen, Y.W.; Reilly, R.M.; Chiou, X.G.; Koterski, S.; Smith, M.L.; Kroeger, P.; McWeeny, K.; Halbert, D.N.; Mollison, K.W.; et al. T cell activation induces a noncoding RNA transcript sensitive to inhibition by immunosuppressant drugs and encoded by the proto-oncogene, BIC. *Cell. Immunol.* 2002, 217, 78–86. [CrossRef]
- O'Connell, R.M.; Taganov, K.D.; Boldin, M.P.; Cheng, G.; Baltimore, D. MicroRNA-155 is induced during the macrophage inflammatory response. *Proc. Natl. Acad. Sci. USA* 2007, 104, 1604–1609. [CrossRef]
- Ruggiero, T.; Trabucchi, M.; De Santa, F.; Zupo, S.; Harfe, B.D.; McManus, M.T.; Rosenfeld, M.G.; Briata, P.; Gherzi, R. LPS induces KH-type splicing regulatory protein-dependent processing of microRNA-155 precursors in macrophages. *FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol.* 2009, 23, 2898–2908. [CrossRef]
- Taganov, K.D.; Boldin, M.P.; Chang, K.J.; Baltimore, D. NF-kappaB-dependent induction of microRNA miR-146, an inhibitor targeted to signaling proteins of innate immune responses. *Proc. Natl. Acad. Sci. USA* 2006, 103, 12481–12486. [CrossRef]
- Tili, E.; Michaille, J.J.; Cimino, A.; Costinean, S.; Dumitru, C.D.; Adair, B.; Fabbri, M.; Alder, H.; Liu, C.G.; Calin, G.A.; et al. Modulation of miR-155 and miR-125b levels following lipopolysaccharide/TNF-alpha stimulation and their possible roles in regulating the response to endotoxin shock. *J. Immunol.* 2007, 179, 5082–5089. [CrossRef]
- Martinez-Nunez, R.T.; Louafi, F.; Friedmann, P.S.; Sanchez-Elsner, T. MicroRNA-155 modulates the pathogen binding ability of dendritic cells (DCs) by down-regulation of DC-specific intercellular adhesion molecule-3 grabbing non-integrin (DC-SIGN). J. Biol. Chem. 2009, 284, 16334–16342. [CrossRef]
- Koch, M.; Mollenkopf, H.J.; Klemm, U.; Meyer, T.F. Induction of microRNA-155 is TLR- and type IV secretion system-dependent in macrophages and inhibits DNA-damage induced apoptosis. *Proc. Natl. Acad. Sci. USA* 2012, 109, E1153–E1162. [CrossRef] [PubMed]
- Wang, L.; Zhang, H.; Rodriguez, S.; Cao, L.; Parish, J.; Mumaw, C.; Zollman, A.; Kamoka, M.M.; Mu, J.; Chen, D.Z.; et al. Notch-dependent repression of miR-155 in the bone marrow niche regulates hematopoiesis in an NF-kappaB-dependent manner. *Cell Stem Cell* 2014, 15, 51–65. [CrossRef] [PubMed]
- Jin, H.M.; Kim, T.J.; Choi, J.H.; Kim, M.J.; Cho, Y.N.; Nam, K.I.; Kee, S.J.; Moon, J.B.; Choi, S.Y.; Park, D.J.; et al. MicroRNA-155 as a proinflammatory regulator via SHIP-1 down-regulation in acute gouty arthritis. *Arthritis Res. Ther.* 2014, 16, R88. [CrossRef] [PubMed]
- 94. Sallusto, F.; Lanzavecchia, A. The instructive role of dendritic cells on T-cell responses. Arthritis Res. 2002, 4, S127–S132. [CrossRef]
- Ceppi, M.; Pereira, P.M.; Dunand-Sauthier, I.; Barras, E.; Reith, W.; Santos, M.A.; Pierre, P. MicroRNA-155 modulates the interleukin-1 signaling pathway in activated human monocyte-derived dendritic cells. *Proc. Natl. Acad. Sci. USA* 2009, 106, 2735–2740. [CrossRef]
- Hodge, J.; Wang, F.; Wang, J.; Liu, Q.; Saaoud, F.; Wang, Y.; Singh, U.P.; Chen, H.; Luo, M.; Ai, W.; et al. Overexpression of microRNA-155 enhances the efficacy of dendritic cell vaccine against breast cancer. *Oncoimmunology* 2020, 9, 1724761. [CrossRef]
- Lu, C.; Huang, X.; Zhang, X.; Roensch, K.; Cao, Q.; Nakayama, K.I.; Blazar, B.R.; Zeng, Y.; Zhou, X. miR-221 and miR-155 regulate human dendritic cell development, apoptosis, and IL-12 production through targeting of p27kip1, KPC1, and SOCS-1. *Blood* 2011, 117, 4293–4303. [CrossRef]
- Thai, T.H.; Calado, D.P.; Casola, S.; Ansel, K.M.; Xiao, C.; Xue, Y.; Murphy, A.; Frendewey, D.; Valenzuela, D.; Kutok, J.L.; et al. Regulation of the germinal center response by microRNA-155. *Science* 2007, *316*, 604–608. [CrossRef]
- Wang, J.; Iwanowycz, S.; Yu, F.; Jia, X.; Leng, S.; Wang, Y.; Li, W.; Huang, S.; Ai, W.; Fan, D. microRNA-155 deficiency impairs dendritic cell function in breast cancer. *Oncoimmunology* 2016, *5*, e1232223. [CrossRef]
- Dunand-Sauthier, I.; Santiago-Raber, M.L.; Capponi, L.; Vejnar, C.E.; Schaad, O.; Irla, M.; Seguin-Estevez, Q.; Descombes, P.; Zdobnov, E.M.; Acha-Orbea, H.; et al. Silencing of c-Fos expression by microRNA-155 is critical for dendritic cell maturation and function. *Blood* 2011, 117, 4490–4500. [CrossRef]
- Montecalvo, A.; Larregina, A.T.; Shufesky, W.J.; Stolz, D.B.; Sullivan, M.L.; Karlsson, J.M.; Baty, C.J.; Gibson, G.A.; Erdos, G.; Wang, Z.; et al. Mechanism of transfer of functional microRNAs between mouse dendritic cells via exosomes. *Blood* 2012, 119, 756–766. [CrossRef] [PubMed]
- Hiramoto, J.S.; Tsung, K.; Bedolli, M.; Norton, J.A.; Hirose, R. Antitumor immunity induced by dendritic cell-based vaccination is dependent on interferon-gamma and interleukin-12. *J. Surg. Res.* 2004, 116, 64–69. [CrossRef] [PubMed]

- 103. Asadirad, A.; Baghaei, K.; Hashemi, S.M.; Dehnavi, S.; Ghanbarian, H.; Mortaz, E.; Anissian, A.; Asadzadeh Aghdaei, H.; Amani, D. Dendritic cell immunotherapy with miR-155 enriched tumor-derived exosome suppressed cancer growth and induced antitumor immune responses in murine model of colorectal cancer induced by CT26 cell line. *Int. Immunopharmacol.* 2022, 104, 108493. [CrossRef] [PubMed]
- Larionova, I.; Tuguzbaeva, G.; Ponomaryova, A.; Stakheyeva, M.; Cherdyntseva, N.; Pavlov, V.; Choinzonov, E.; Kzhyshkowska, J. Tumor-Associated Macrophages in Human Breast, Colorectal, Lung, Ovarian and Prostate Cancers. *Front. Oncol.* 2020, 10, 566511. [CrossRef]
- 105. Leek, R.D.; Lewis, C.E.; Whitehouse, R.; Greenall, M.; Clarke, J.; Harris, A.L. Association of macrophage infiltration with angiogenesis and prognosis in invasive breast carcinoma. *Cancer Res.* **1996**, *56*, 4625–4629.
- 106. Steidl, C.; Lee, T.; Shah, S.P.; Farinha, P.; Han, G.; Nayar, T.; Delaney, A.; Jones, S.J.; Iqbal, J.; Weisenburger, D.D.; et al. Tumor-associated macrophages and survival in classic Hodgkin's lymphoma. *N. Engl. J. Med.* **2010**, *362*, 875–885. [CrossRef]
- 107. DeNardo, D.G.; Ruffell, B. Macrophages as regulators of tumour immunity and immunotherapy. *Nat. Rev. Immunol.* **2019**, 19, 369–382. [CrossRef]
- 108. O'Connell, R.M.; Chaudhuri, A.A.; Rao, D.S.; Baltimore, D. Inositol phosphatase SHIP1 is a primary target of miR-155. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 7113–7118. [CrossRef]
- Cai, X.; Yin, Y.; Li, N.; Zhu, D.; Zhang, J.; Zhang, C.Y.; Zen, K. Re-polarization of tumor-associated macrophages to proinflammatory M1 macrophages by microRNA-155. J. Mol. Cell Biol. 2012, 4, 341–343. [CrossRef]
- Rauh, M.J.; Ho, V.; Pereira, C.; Sham, A.; Sly, L.M.; Lam, V.; Huxham, L.; Minchinton, A.I.; Mui, A.; Krystal, G. SHIP represses the generation of alternatively activated macrophages. *Immunity* 2005, 23, 361–374. [CrossRef]
- 111. Zheng, P.; Chen, L.; Yuan, X.; Luo, Q.; Liu, Y.; Xie, G.; Ma, Y.; Shen, L. Exosomal transfer of tumor-associated macrophage-derived miR-21 confers cisplatin resistance in gastric cancer cells. *J. Exp. Clin. Cancer Res.* **2017**, *36*, 53. [CrossRef]
- 112. Wu, Q.; Sun, S.; Li, Z.; Yang, Q.; Li, B.; Zhu, S.; Wang, L.; Wu, J.; Yuan, J.; Yang, C.; et al. Tumour-originated exosomal miR-155 triggers cancer-associated cachexia to promote tumour progression. *Mol. Cancer* **2018**, *17*, 155. [CrossRef] [PubMed]
- 113. Zhou, X.; Yan, T.; Huang, C.; Xu, Z.; Wang, L.; Jiang, E.; Wang, H.; Chen, Y.; Liu, K.; Shao, Z.; et al. Melanoma cell-secreted exosomal miR-155-5p induce proangiogenic switch of cancer-associated fibroblasts via SOCS1/JAK2/STAT3 signaling pathway. J. Exp. Clin. Cancer Res. 2018, 37, 242. [CrossRef] [PubMed]
- 114. Veglia, F.; Sanseviero, E.; Gabrilovich, D.I. Myeloid-derived suppressor cells in the era of increasing myeloid cell diversity. *Nat. Rev. Immunol.* **2021**, *21*, 485–498. [CrossRef] [PubMed]
- 115. Ai, L.; Mu, S.; Wang, Y.; Wang, H.; Cai, L.; Li, W.; Hu, Y. Prognostic role of myeloid-derived suppressor cells in cancers: A systematic review and meta-analysis. *BMC Cancer* **2018**, *18*, 1220. [CrossRef]
- Wang, J.; Yu, F.; Jia, X.; Iwanowycz, S.; Wang, Y.; Huang, S.; Ai, W.; Fan, D. MicroRNA-155 deficiency enhances the recruitment and functions of myeloid-derived suppressor cells in tumor microenvironment and promotes solid tumor growth. *Int. J. Cancer* 2015, 136, E602–E613. [CrossRef] [PubMed]
- 117. Kim, S.; Song, J.H.; Kim, S.; Qu, P.; Martin, B.K.; Sehareen, W.S.; Haines, D.C.; Lin, P.C.; Sharan, S.K.; Chang, S. Loss of oncogenic miR-155 in tumor cells promotes tumor growth by enhancing C/EBP-beta-mediated MDSC infiltration. *Oncotarget* 2016, 7, 11094–11112. [CrossRef]
- Li, L.; Zhang, J.; Diao, W.; Wang, D.; Wei, Y.; Zhang, C.Y.; Zen, K. MicroRNA-155 and MicroRNA-21 promote the expansion of functional myeloid-derived suppressor cells. *J. Immunol.* 2014, 192, 1034–1043. [CrossRef]
- Chen, S.; Wang, L.; Fan, J.; Ye, C.; Dominguez, D.; Zhang, Y.; Curiel, T.J.; Fang, D.; Kuzel, T.M.; Zhang, B. Host miR155 promotes tumor growth through a myeloid-derived suppressor cell-dependent mechanism. *Cancer Res.* 2015, 75, 519–531. [CrossRef]
- 120. Bruns, H.; Bottcher, M.; Qorraj, M.; Fabri, M.; Jitschin, S.; Dindorf, J.; Busch, L.; Jitschin, R.; Mackensen, A.; Mougiakakos, D. CLL-cell-mediated MDSC induction by exosomal miR-155 transfer is disrupted by vitamin D. *Leuk. Off. J. Leuk. Soc. Am. Leuk. Res. Fund* 2017, *31*, 985–988. [CrossRef]
- 121. Miller, J.S.; Soignier, Y.; Panoskaltsis-Mortari, A.; McNearney, S.A.; Yun, G.H.; Fautsch, S.K.; McKenna, D.; Le, C.; Defor, T.E.; Burns, L.J.; et al. Successful adoptive transfer and in vivo expansion of human haploidentical NK cells in patients with cancer. *Blood* 2005, 105, 3051–3057. [CrossRef] [PubMed]
- 122. Waldhauer, I.; Steinle, A. NK cells and cancer immunosurveillance. Oncogene 2008, 27, 5932–5943. [CrossRef] [PubMed]
- 123. Trotta, R.; Chen, L.; Ciarlariello, D.; Josyula, S.; Mao, C.; Costinean, S.; Yu, L.; Butchar, J.P.; Tridandapani, S.; Croce, C.M.; et al. miR-155 regulates IFN-gamma production in natural killer cells. *Blood* 2012, 119, 3478–3485. [CrossRef]
- 124. Kandell, W.M.; Donatelli, S.S.; Trinh, T.L.; Calescibetta, A.R.; So, T.; Tu, N.; Gilvary, D.L.; Chen, X.; Cheng, P.; Adams, W.A.; et al. MicroRNA-155 governs SHIP-1 expression and localization in NK cells and regulates subsequent infiltration into murine AT3 mammary carcinoma. *PLoS ONE* **2020**, *15*, e0225820. [CrossRef] [PubMed]
- 125. Jovanovic, M.Z.; Geller, D.A.; Gajovic, N.M.; Jurisevic, M.M.; Arsenijevic, N.N.; Jovanovic, M.M.; Supic, G.M.; Vojvodic, D.V.; Jovanovic, I.P. Dual blockage of PD-L/PD-1 and IL33/ST2 axes slows tumor growth and improves antitumor immunity by boosting NK cells. *Life Sci.* 2022, 289, 120214. [CrossRef] [PubMed]
- 126. O'Connell, R.M.; Rao, D.S.; Chaudhuri, A.A.; Boldin, M.P.; Taganov, K.D.; Nicoll, J.; Paquette, R.L.; Baltimore, D. Sustained expression of microRNA-155 in hematopoietic stem cells causes a myeloproliferative disorder. *J. Exp. Med.* 2008, 205, 585–594. [CrossRef] [PubMed]

- 127. Turner, M.; Vigorito, E. Regulation of B- and T-cell differentiation by a single microRNA. *Biochem. Soc. Trans.* **2008**, *36*, 531–533. [CrossRef]
- O'Connell, R.M.; Rao, D.S.; Chaudhuri, A.A.; Baltimore, D. Physiological and pathological roles for microRNAs in the immune system. *Nat. Rev. Immunol.* 2010, 10, 111–122. [CrossRef]
- 129. Escobar, T.; Yu, C.R.; Muljo, S.A.; Egwuagu, C.E. STAT3 activates miR-155 in Th17 cells and acts in concert to promote experimental autoimmune uveitis. *Investig. Ophthalmol. Vis. Sci.* 2013, 54, 4017–4025. [CrossRef]
- 130. Ho, I.C.; Lo, D.; Glimcher, L.H. c-maf promotes T helper cell type 2 (Th2) and attenuates Th1 differentiation by both interleukin 4-dependent and -independent mechanisms. *J. Exp. Med.* **1998**, *188*, 1859–1866. [CrossRef]
- Banerjee, A.; Schambach, F.; DeJong, C.S.; Hammond, S.M.; Reiner, S.L. Micro-RNA-155 inhibits IFN-gamma signaling in CD4+ T cells. *Eur. J. Immunol.* 2010, 40, 225–231. [CrossRef] [PubMed]
- Goncalves-Alves, E.; Saferding, V.; Schliehe, C.; Benson, R.; Kurowska-Stolarska, M.; Brunner, J.S.; Puchner, A.; Podesser, B.K.; Smolen, J.S.; Redlich, K.; et al. MicroRNA-155 Controls T Helper Cell Activation During Viral Infection. *Front. Immunol.* 2019, 10, 1367. [CrossRef]
- Li, C.; Jiang, P.; Wei, S.; Xu, X.; Wang, J. Regulatory T cells in tumor microenvironment: New mechanisms, potential therapeutic strategies and future prospects. *Mol. Cancer* 2020, 19, 116. [CrossRef] [PubMed]
- 134. Laine, A.; Labiad, O.; Hernandez-Vargas, H.; This, S.; Sanlaville, A.; Leon, S.; Dalle, S.; Sheppard, D.; Travis, M.A.; Paidassi, H.; et al. Regulatory T cells promote cancer immune-escape through integrin alphavbeta8-mediated TGF-beta activation. *Nat. Commun.* 2021, 12, 6228. [CrossRef] [PubMed]
- 135. Kohlhaas, S.; Garden, O.A.; Scudamore, C.; Turner, M.; Okkenhaug, K.; Vigorito, E. Cutting edge: The Foxp3 target miR-155 contributes to the development of regulatory T cells. *J. Immunol.* **2009**, *182*, 2578–2582. [CrossRef]
- 136. Lu, L.F.; Thai, T.H.; Calado, D.P.; Chaudhry, A.; Kubo, M.; Tanaka, K.; Loeb, G.B.; Lee, H.; Yoshimura, A.; Rajewsky, K.; et al. Foxp3-dependent microRNA155 confers competitive fitness to regulatory T cells by targeting SOCS1 protein. *Immunity* 2009, 30, 80–91. [CrossRef]
- Marson, A.; Kretschmer, K.; Frampton, G.M.; Jacobsen, E.S.; Polansky, J.K.; MacIsaac, K.D.; Levine, S.S.; Fraenkel, E.; von Boehmer, H.; Young, R.A. Foxp3 occupancy and regulation of key target genes during T-cell stimulation. *Nature* 2007, 445, 931–935. [CrossRef]
- Zheng, Y.; Josefowicz, S.Z.; Kas, A.; Chu, T.T.; Gavin, M.A.; Rudensky, A.Y. Genome-wide analysis of Foxp3 target genes in developing and mature regulatory T cells. *Nature* 2007, 445, 936–940. [CrossRef]
- Heyn, J.; Luchting, B.; Hinske, L.C.; Hubner, M.; Azad, S.C.; Kreth, S. miR-124a and miR-155 enhance differentiation of regulatory T cells in patients with neuropathic pain. *J. Neuroinflamm.* 2016, 13, 248. [CrossRef]
- 140. Zhu, F.; Li, H.; Liu, Y.; Tan, C.; Liu, X.; Fan, H.; Wu, H.; Dong, Y.; Yu, T.; Chu, S.; et al. miR-155 antagomir protect against DSS-induced colitis in mice through regulating Th17/Treg cell balance by Jarid2/Wnt/beta-catenin. *Biomed. Pharmacother. Biomed. Pharmacother.* 2020, 126, 109909. [CrossRef]
- Raskov, H.; Orhan, A.; Christensen, J.P.; Gogenur, I. Cytotoxic CD8(+) T cells in cancer and cancer immunotherapy. *Br. J. Cancer* 2021, 124, 359–367. [CrossRef] [PubMed]
- 142. Dudda, J.C.; Salaun, B.; Ji, Y.; Palmer, D.C.; Monnot, G.C.; Merck, E.; Boudousquie, C.; Utzschneider, D.T.; Escobar, T.M.; Perret, R.; et al. MicroRNA-155 is required for effector CD8+ T cell responses to virus infection and cancer. *Immunity* 2013, 38, 742–753. [CrossRef] [PubMed]
- 143. Shiravand, Y.; Khodadadi, F.; Kashani, S.M.A.; Hosseini-Fard, S.R.; Hosseini, S.; Sadeghirad, H.; Ladwa, R.; O'Byrne, K.; Kulasinghe, A. Immune Checkpoint Inhibitors in Cancer Therapy. *Curr. Oncol.* **2022**, *29*, 3044–3060. [CrossRef]
- 144. Hargadon, K.M.; Johnson, C.E.; Williams, C.J. Immune checkpoint blockade therapy for cancer: An overview of FDA-approved immune checkpoint inhibitors. *Int. Immunopharmacol.* **2018**, *62*, 29–39. [CrossRef]
- 145. Yee, D.; Shah, K.M.; Coles, M.C.; Sharp, T.V.; Lagos, D. MicroRNA-155 induction via TNF-alpha and IFN-gamma suppresses expression of programmed death ligand-1 (PD-L1) in human primary cells. *J. Biol. Chem.* **2017**, 292, 20683–20693. [CrossRef]
- 146. Renrick, A.N.; Thounaojam, M.C.; de Aquino, M.T.P.; Chaudhuri, E.; Pandhare, J.; Dash, C.; Shanker, A. Bortezomib Sustains T Cell Function by Inducing miR-155-Mediated Downregulation of SOCS1 and SHIP1. Front. Immunol. 2021, 12, 607044. [CrossRef]
- 147. Wang, J.; Wang, Q.; Guan, Y.; Sun, Y.; Wang, X.; Lively, K.; Wang, Y.; Luo, M.; Kim, J.A.; Murphy, E.A.; et al. Breast cancer cell-derived microRNA-155 suppresses tumor progression via enhancing immune cell recruitment and anti-tumor function. *J. Clin. Investig.* 2022, 132, e157248. [CrossRef] [PubMed]
- 148. Nguyen, M.T.; Luo, Y.H.; Li, A.L.; Tsai, J.C.; Wu, K.L.; Chung, P.J.; Ma, N. miRNA as a Modulator of Immunotherapy and Immune Response in Melanoma. *Biomolecules* **2021**, *11*, 1648. [CrossRef]
- 149. Liu, L.; Yi, H.; He, H.; Pan, H.; Cai, L.; Ma, Y. Tumor associated macrophage-targeted microRNA delivery with dual-responsive polypeptide nanovectors for anti-cancer therapy. *Biomaterials* **2017**, *134*, 166–179. [CrossRef]
- 150. Bayraktar, R.; Van Roosbroeck, K. miR-155 in cancer drug resistance and as target for miRNA-based therapeutics. *Cancer Metastasis Rev.* 2018, *37*, 33–44. [CrossRef]
- Kopp, K.L.; Ralfkiaer, U.; Gjerdrum, L.M.; Helvad, R.; Pedersen, I.H.; Litman, T.; Jonson, L.; Hagedorn, P.H.; Krejsgaard, T.; Gniadecki, R.; et al. STAT5-mediated expression of oncogenic miR-155 in cutaneous T-cell lymphoma. *Cell Cycle* 2013, 12, 1939–1947. [CrossRef] [PubMed]

- 152. Strizova, Z.; Snajdauf, M.; Stakheev, D.; Taborska, P.; Vachtenheim, J., Jr.; Biskup, J.; Lischke, R.; Bartunkova, J.; Smrz, D. The paratumoral immune cell signature reveals the potential for the implementation of immunotherapy in esophageal carcinoma patients. *J. Cancer Res. Clin. Oncol.* **2020**, *146*, 1979–1992. [CrossRef] [PubMed]
- Strizova, Z.; Taborska, P.; Stakheev, D.; Partlova, S.; Havlova, K.; Vesely, S.; Bartunkova, J.; Smrz, D. NK and T cells with a cytotoxic/migratory phenotype accumulate in peritumoral tissue of patients with clear cell renal carcinoma. *Urol. Oncol.* 2019, 37, 503–509. [CrossRef] [PubMed]
- 154. Kuchar, M.; Strizova, Z.; Votava, M.; Plzak, J. The relevance of Fas/Fas ligand axis in the tumor microenvironment of salivary gland adenoid cystic carcinoma. *Oral Oncol.* 2019, *97*, 135–136. [CrossRef] [PubMed]
- 155. Strizova, Z.; Bartunkova, J.; Smrz, D. The challenges of adoptive cell transfer in the treatment of human renal cell carcinoma. *Cancer Immunol. Immunother.* **2019**, *68*, 1831–1838. [CrossRef] [PubMed]
- 156. Das, S.; Camphausen, K.; Shankavaram, U. Cancer-Specific Immune Prognostic Signature in Solid Tumors and Its Relation to Immune Checkpoint Therapies. *Cancers* 2020, *12*, 2476. [CrossRef]