

Review

Chasing the Target: New Phenomena of Resistance to Novel Selective RET Inhibitors in Lung Cancer. Updated Evidence and Future Perspectives

Sara Fancelli ¹, Enrico Caliman ^{1,2}, Francesca Mazzoni ¹, Marco Brugia ¹, Francesca Castiglione ³, Luca Voltolini ^{2,4}, Serena Pillozzi ¹ and Lorenzo Antonuzzo ^{1,2,*}

¹ Medical Oncology Unit, Careggi University Hospital, 50134 Florence, Italy; sara.fancelli@unifi.it (S.F.); enrico.caliman@unifi.it (E.C.); mazzonifr@aou-careggi.toscana.it (F.M.); brugiam@aou-careggi.toscana.it (M.B.); serena.pillozzi@unifi.it (S.P.)

² Department of Experimental and Clinical Medicine, University of Florence, 50134 Florence, Italy; luca.voltolini@unifi.it

³ Pathological Histology and Molecular Diagnostics Unit, Careggi University Hospital, 50134 Florence, Italy; castiglione@aou-careggi.toscana.it

⁴ Thoraco-Pulmonary Surgery Unit, Careggi University Hospital, 50134 Florence, Italy

* Correspondence: lorenzo.antonuzzo@unifi.it; Tel.: +39-055-7948406

Simple Summary: REarranged during Transfection (RET) is an emerging target for several types of cancer, including non-small cell lung cancer (NSCLC). The recent U.S. FDA approval of pralsetinib and selpercatinib raises issues regarding the emergence of secondary mutations and amplifications involved in parallel signaling pathways and receptors, liable for resistance mechanisms. The aim of this review is to explore recent knowledge on RET resistance in NSCLC in pre-clinic and in clinical settings and accordingly, the state-of-the-art in new drugs or combination of drugs development.

Abstract: The potent, RET-selective tyrosine kinase inhibitors (TKIs) pralsetinib and selpercatinib, are effective against the RET V804L/M gatekeeper mutants, however, adaptive mutations that cause resistance at the solvent front RET G810 residue have been found, pointing to the need for the development of the next-generation of RET-specific TKIs. Also, as seen in EGFR- and ALK-driven NSCLC, the rising of the co-occurring amplifications of KRAS and MET could represent other escaping mechanisms from direct inhibition. In this review, we summarize actual knowledge on RET fusions, focusing on those involved in NSCLC, the results of main clinical trials of approved RET-inhibition drugs, with particular attention on recent published results of selective TKIs, and finally, pre-clinical evidence regarding resistance mechanisms and suggestion on hypothetical and feasible drugs combinations and strategies viable in the near future.

Keywords: RET; NSCLC; selpercatinib; pralsetinib; solvent-front mutations; acquired resistances



Citation: Fancelli, S.; Caliman, E.; Mazzoni, F.; Brugia, M.; Castiglione, F.; Voltolini, L.; Pillozzi, S.; Antonuzzo, L. Chasing the Target: New Phenomena of Resistance to Novel Selective RET Inhibitors in Lung Cancer. Updated Evidence and Future Perspectives. *Cancers* **2021**, *13*, 1091. <https://doi.org/10.3390/cancers13051091>

Academic Editor: Noriaki Sunaga

Received: 13 January 2021

Accepted: 26 February 2021

Published: 4 March 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Recent evidence in non-small cell lung cancer (NSCLC) about the new highly selective REarranged during Transfection (RET) inhibitors selpercatinib and pralsetinib, despite impressive results in clinical trials, have raised the urgency to highlight the best therapeutic sequence in view of novel resistances which are able to cause selective inhibition to be useless. With this aim, and aware that the inhibition of RET has always been a challenge since its discovery in T-cell lymphoma [1], we want to summarize the available knowledge on RET inhibitors, including both ongoing trials with new drugs and pre-clinical data concerning overcoming the resistance mechanisms.

2. RET in Pills

RET is located on chromosome 10q11.2 and its expression is mediated by several DNA-binding proteins belonging to the Sp family of transcription factors (Sp1, Sp3) [2] or early growth response protein 1 (EGR1) [2], SRY-box 10 (SOX10), paired box 3 (PAX3) [3], NK2 homeobox 1 (NKX2-1) and homeobox B5 (HOXB5) [4]. *RET* encodes for a Transmembrane Tyrosine Kinase Receptor (RTK) with a unique structure composed of four cadherin-like domains, a cysteine-rich domain, a transmembrane domain and a tyrosine kinase (TK) domain, this latter has a different number of amino acids depending on the isoform transcribed (RET9, RET43 and RET51) [5]. Each isoform interacts with adaptors and signaling proteins that are able to activate different downstream pathways during embryogenesis, in homeostasis of several tissues [6]. Physiologically, beginning RET signals depend on the binding of specific ligand members of the glial cell line-derived neurotrophic factors (GDNFs) with GDNF family receptor alpha ($GFR\alpha$). The ligand family includes GDNF, neurturin (NTRN), artemin (ARTN) and persephin (PSPN) and each has a selective, although not completely specific, receptor, respectively called $GFR\alpha1$, $GFR\alpha2$, $GFR\alpha3$ and $GFR\alpha4$ [7]. The interposition of the GDNF- $GFR\alpha$ complex allows for the homodimerization between RET monomers resulting in autophosphorylation of the intracellular tyrosine residues of the main docking-site of the RET51 isoform (Y1062). RET is also able to heterodimerize with other RTKs [5]. Phosphorylated tyrosine recruits a multitude of adaptors that, in turn, mediate the activation of RAS- Mitogen-Activated Protein Kinases (MAPK) and Phosphatidylinositol-3 Kinase (PI3K)- Protein Kinase B (AKT) pathways [5]. Several docking sites (Y900, Y905, Y981, Y1015 and Y1096), able to trigger additional downstream pathways such as JAK/STAT, PKA, PKC and JNK, have been described [6]. Moreover, RET interacts with RTKs and other cell surface proteins guaranteeing a continuity and spreading of downstream signals [8] (Figure 1). During embryogenesis RET is mainly expressed in the urinary tract, nervous system and hematopoietic stem cells, justifying the pathogenesis of hereditary diseases secondary to germline mutations (loss of function). In adult life, low levels of RET expression are registered in all tissues [9] and different RET molecular alterations have been reported in tumors at either germline or somatic levels. These include gene amplification, fusion, as well as single base substitutions/small insertions/deletions.

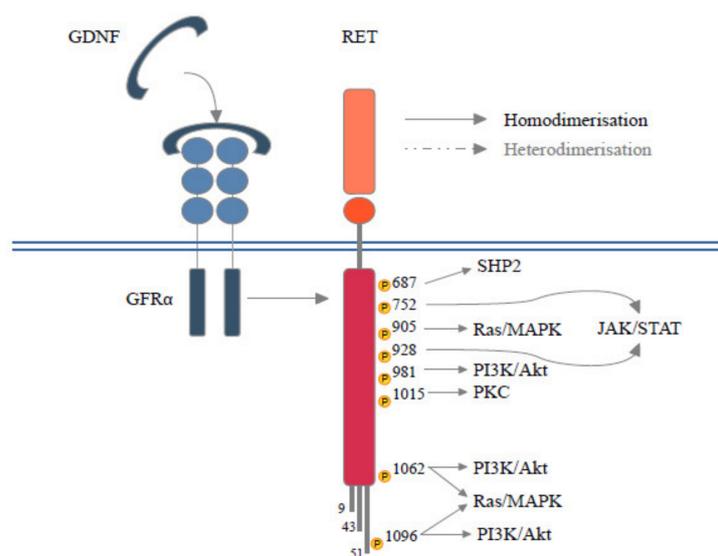


Figure 1. Schematic RET protein structure showing phosphorylation sites. RET forms a heterocomplex with $GFR\alpha$ and GFLs proteins, which in turn results in the activation of multiple signaling pathways involved in survival, differentiation, motility, proliferation, and growth.

2.1. Germline Mutations

Genitourinary and nervous system development [10,11], maturation and migration of stem cell lines and a general involvement in embryogenesis and spermatogenesis, represent the main known mechanisms in which RET's signaling is involved during embryonic development [12,13]. It's clearly understandable how RET loss of function due to germline mutations, affecting those mechanism, can lead to a variety of congenital malformations such as Hirschsprung disease (HSCR) and congenital abnormalities of the kidney and urinary tract (CAKUT), and cause numerous symptoms in patients with phenotypic variants of these syndromes [14,15]. However, a role for RET in maintenance of hematopoietic system and in development of Gut-Associated Lymphoid Tissue (GALT) has recently been recognized [16]. Germinal mutations of the proto-oncogene RET affecting cysteine-rich extracellular domains or less frequently on the intracellular domains give rise to multiple neuroendocrine neoplasia 2 (MEN2). MEN2 is classified based on clinical features in MEN2A characterized by thyroid cancer, pheochromocytoma, and hyperparathyroidism and in MEN2B with also ganglioneuromatosis and a Marfanoid habitus [17]. Similarly to MEN2, the familial medullary thyroid carcinoma (FMTC) derived from germinal point mutation that causes an increase in the effect of self-activation by increasing ATP-binding or phosphorylation activity, sustains the oncogenic and pro-proliferative stimuli [18]. Every point mutation, rarely seen outside neuroendocrine neoplasms, correlates with different prognosis and clinical outcome, suggesting the necessity to sketch out an early screening and subsequently a different therapeutic approach [19,20]. Indeed, MEN2A and FMTC, having phenotypic and clinical indolent characteristics, appear to be more of a continuum of the same disease, unlike MEN2B which has a juvenile onset and a more aggressive course [21].

2.2. Somatic Mutations and Cancer

To better understand its decisive role as a proto-oncogene in sporadic cancers we had to wait until the Chernobyl disaster in 1986 showed a correlation between the papillary thyroid carcinomas (PTC) onset and gene rearrangements in post-radiation exposed children [22]. RET/Coiled-Coil Domain Containing 6 (CCDC6) gene fusion is associated in about 80% of cases of sporadic PTC, while the Nuclear Receptor Coactivator 4 (NCOA4) gene, is mainly related to radiation exposure and younger age [23]. Countless rearrangements have been described in literature being a part of the pathogenesis of PTC [24]. Although PTCs are the most frequently associated cancers. with RET rearrangements (10–20%), many other neoplasms are associated with RET-fusion involved in creating resistances and escaping mechanisms to classical therapies. In hormone positive breast cancer (BC), RET overexpression is described in less than 0.1% of cases and is involved in resistance to anti-hormonal therapies in BC cell lines [25]. Based on preclinical evidence of crosstalk between RET and positive estrogen receptors, some clinical trials in BC patients without any convincing results in disease control explore the benefit of using multi-kinase inhibitors active on RET [26]. Recently, a single case report has been presented as part of LIBRETTO-001 trial, of a metastatic BC woman who presented a complete clinical response with Selpercatinib, suggesting a possible role of selective RET inhibitors in this field [27]. In colon cancers RET rearrangements represent 0.2% of cases [28]. Among them, 2/3 manifest in the right colon and are characterized by MSI, RAS and BRAF wild type status, and could benefit from the use of specific therapies [29,30]. Other gastrointestinal malignancies, gynecological tumors, renal and prostate cancer have a limited expression of RET fusions [31] and could benefit from treatment inside basket trials, thanks to recent gene sequencing techniques.

2.3. RET in Lung Cancer

In NSCLC the prevalence of RET alterations is estimated to be 1–2% of all cases [32]. Thanks to modern genomic sequencing methods, the first fusion gene discovery in 2011 between RET and the Kinesin Family Member 5B (KIF5B) gene has allowed to broaden the knowledge of translocations involving RET [33]. As mentioned above, rearrangements

involving chromosome 10 are intrachromosomal, leading to fusion with several genes lying on the same chromosome. In NSCLC the gene most involved in fusions is KIF5B, a gene involved in a pericentric rearrangement, followed by CCDC6 and NCOA4 which are characterized by a paracentric inversion fusion [32]. Several other inter-chromosomal rearrangements or translocations have been described, however they represent a small percentage of cases, we have summarized some in Table 1. [24,34]. Breakpoints in KIF5B are frequently found in the intron 11 at different positions and are involved in transcription of intracytoplasmic segments of RET, however, different introns are rarely involved in fostering the inclusion of the transmembrane dominion [35,36]. In addition to fusions, single amplifications or mutations with variable penetrance related to histotype and gender have been found [34,37]. KIF5B exon 15 fusion to RET exon 12 is the most frequently detected fusion in nonsmokers and young females, while CCDC6 exon 1 to RET exon 12 correlates with smoking habits and male gender [34].

Adenocarcinoma and adenosquamous carcinoma are the most frequent histologies diagnosed in rare solid subtypes as per signet ring cells or mucinous form [38]. Fusions support intracellular signaling thanks to the increase in kinase expression in those tissues normally lacking in RET expression, non-ligand mediated self-activation secondary to mutation in upstream proteins able to support coiled-coil domains interaction, and finally, the loss of self-inhibitory capacity. These modifications determine activation of the signaling pathways STAT3, JAK/STAT3 and RAS/RAF/MEK/ERK capable of supporting proliferation, differentiation, angiogenesis and metastasis as demonstrated in in vivo experiences [39–41].

Table 1. Other rearrangements and RET fusions.

Fusions	Histotype	Gender	Reference
CDC123-RET	ADC	F	[42]
CCDC6-RET	ADC, NE	M > F	[34]
CLIP1-RET	ADC	NA	[43]
CUX1-RET	ADC	M	[44]
EPHA5-RET	ADC	NA	[45]
ERC1-RET	ADC	NA	[43]
FRMD4A-RET	ADC	F	[46]
FYCO1-RET	ADC	F	[34]
ITGA8-RET	ADC	M	[34]
ITIH2-RET	AS	F	[34]
KIF13A-RET	ADC	F	[47]
KIF5B-RET	ADC, NE, NSCLC, AS	F > M	[34]
KIAA1468-RET	IMA	M	[48]
MIR3924-RET	SCC	M	[34]
MYO5C-RET	ADC	NA	[49]
NCOA4-RET	ADC	F	[50]
PICALM-RET	ADC	NA	[45]
RASSF4-RET	ADC	NA	[51]
RUFY2-RET	ADC	NA	[52]
SLC25A36-RET	ADC	F	[34]
SLC39A8-RET	ADC	F	[34]
TBC1D32-RET	ADC	F	[53]
TRIM24-RET	ADC	NA	[52]
TRIM33-RET	ADC	F	[54]
WAC-RET	ADC	F	[55]
ZBTB41-RET	ADC	M	[34]

Abbreviations: ADC adenocarcinoma; NE neuroendocrine; AS adenosquamous carcinoma; NSCLC non-small cell lung cancer; IMA invasive mucinous adenocarcinoma; F: female, M: male; NA not available.

3. Activity of MKIs in RET-Positive NSCLC

As the tyrosine kinase receptor RET shares similarities in the structure of the kinase domain with other tyrosine kinases (TK) [56], initial attempts to target RET rearrangements

focused on multikinase inhibitors (MKIs) with non-selective RET inhibitory activity have been approached. However, results obtained with MKIs suggest that RET fusions are not highly actionable. Treatment with MKIs in RET fusion-positive NSCLC demonstrated both modest clinical activity and limited response durability. Moreover, the response rates achieved in clinical experiences were lower compared with outcomes with therapies targeting other oncogenic drivers (i.e., EGFR mutations, ALK and ROS-1 fusions) [57]. Several MKIs inhibitors, that have been investigated in the treatment of RET-rearranged NSCLC, are approved for the treatment of thyroid cancers (i.e., vandetanib, cabozantinib, lenvatinib and sorafenib) or are approved for other indications (i.e., ponatinib, alectinib and sunitinib). Activity of MKIs in RET fusion-positive NSCLC has been reported in pre-clinical cancer models [31,36,58,59], in retrospective case series [45,60] and in phase I and phase II trials [43,49,54,61–64]. These agents have been developed against a variety of target-kinases other than (or in addition to) RET, such as VEGF receptors, AXL, FGFR1, EGFR, MET, c-KIT and BRAF, and unfortunately demonstrated limited potency for RET-positive cancers. Moreover, these agents have led to a variety of adverse events (AEs) which are closely related to their activity against other pathways, such as EGFR (diarrhea and dermatologic toxicities) and VEGFR (hypertension). These off-target side effects can frequently lead to discontinuation of treatment or dose reduction and, as a consequence, treatment with a dose that effectively inhibits RET would not be guaranteed. Taken together these evidence may explain the suboptimal activity and the lower clinical benefits obtained in MKIs-treated RET-positive NSCLC, compared to the outcomes of other oncogene-addicted NSCLC subtypes when treated with matched targeted therapies.

3.1. Vandetanib

Vandetanib, a multi-target TKI targeting VEGF receptors, EGFR and RET, has been investigated in a phase II trial [61]. In this study clinical antitumor activity has been reported in nine out 19 (47%) patients with RET-positive NSCLC enrolled (ORR 47%), but grade 3 or 4 AEs were common and a dose reduction was described in 53% of patients. In another phase II trial [49] vandetanib showed moderate activity in pretreated patients with NSCLC harboring RET rearrangements (ORR 18%, DCR 65%) and dose reduction was necessary in 4 of 18 (22%) patients enrolled. The efficacy outcomes reported in the above studies are comparable with the retrospective analysis of a global registry study (GLORY) with an ORR = 18% [45]. Moreover, in a retrospective analysis of four randomized phase III trials, the overall prevalence of RET rearrangements identified was 0.7% and none of the three RET-positive NSCLC patients have obtained an OR after treatment with vandetanib [65].

3.2. Cabozantinib

Cabozantinib, initially developed against AXL e MET, demonstrated activity versus a broad range of TK, such as VEGFR2, ROS1, c-KIT, TIE2 and RET. The first report of response to cabozantinib in RET-fusion positive lung adenocarcinomas was described by Drilon and colleagues [54] in three patients as preliminary data of a phase II trial. Final results of this phase II study [43] showed 28% ORR and 73% of dose reduction rate (DRR) due to AEs in the 26 patients with RET-rearrangement NSCLC, that have been treated with cabozantinib. Similarly, in the global multicenter registry in patients with RET-rearranged lung cancers, Gautschi et al. reported 32% ORR in the 21 patients treated with cabozantinib [45].

3.3. Lenvatinib

Lenvatinib is a MKI of FGFRs, VEGFRs, PDGFR-alpha, KIT and RET. This MKI has been tested as oral monotherapy in 25 RET-rearranged NSCLC patients in a phase II study [62,64]; the reported ORR and DCR were 16% and 76%, respectively. In the study, almost all patients treated with lenvatinib experienced at least one treatment-related AE: grade 3 or 4 AEs was reported in 92% of patients, 16 patients (64%) required a dose modification of the therapy and six patients (24%) discontinued treatment due to side

effects. In the GLORY database only two patients with RET-positive NSCLC received lenvatinib: one experienced partial response (PR) to treatment while a disease progression was reported for the second patient [45].

3.4. Other MKIs

Clinical data regarding the activity of other MKIs (sorafenib, sunitinib, ponatinib, alectinib, nintedanib and regorafenib) in RET fusion-positive NSCLC are lacking or have been reported in smaller experiences, case reports and in patients included in the large retrospective series already cited [45], in which MKIs were administered in various line of systemic therapy. The efficacy of sorafenib has been tested in a limited number of patients ($n = 3$) in a study by Horiike et al. [66]: one patient experienced stable disease (SD) while two showed progressive disease (PD) as best responses to treatment. Conversely, response to MKI sunitinib has been described in a case report of a patient with NSCLC harboring KIF5B-RET rearrangement [67]. Clinical activity of the anaplastic lymphoma kinase TKI alectinib in RET-rearranged NSCLC was firstly reported in two of four patients described by Lin et al. [60], successively in a case report [68] and in three among the four patients described in a case series [69]. The dose-limiting toxicity (DLT) to alectinib resulted from a phase I study, led to 1 level of dose-reduction recommended for ongoing phase II [70]: preliminary results from phase 2 showed 4% ORR (1 pt) and 52% DCR (13 pts) among in twenty-five RET inhibitor-naïve patients treated with 450 mg alectinib twice daily [71]. Moreover, among the 165 patients with RET-rearranged NSCLC accrued in the global retrospective registry (GLORY) [45], 53 patients (32%) were treated with RET MKIs. Of them, ten patients received sunitinib and two reported partial response (22% ORR), one of the two patients treated with nintedanib achieved a complete response, while none of the patients treated with sorafenib (two patients), alectinib (two patients), ponatinib (two patients) and regorafenib (one patient) experienced OR to these agents. Finally, conversely to other multi-kinase RET-inhibitors, RXDX-105 is a MKIs with a high potency against RET and BRAF while it is VEGFRs sparing [72]. Despite these factors, the overall activity of RXDX-105 in patients with RET fusion-positive NSCLC did not differ substantially from the activity of other MKIs. In a phase I/Ib trial [63] the reported ORR with RXDX-105, in the cohort of RET inhibitor-naïve patients with RET fusion positive NSCLC, was 19% (6/31). Interestingly, although KIF5B-RET is the most common RET fusion in NSCLC, RXDX-105 demonstrated activity only in non-KIF5B-RET-containing NSCLC. In this trial the response rate varied significantly from 0% in KIF5B-RET rearrangement NSCLCs to 67% in non-KIF5B-RET lung cancers. Interestingly, poor clinical outcomes have also been reported in patients with NSCLC harboring KIF5B-RET rearrangement treated with other above mentioned MKIs in several phase II trials [43,49,61,62], compared to patients with non-KIF5B-RET NSCLC. However, in the GLORY study, the reported clinical benefits in patients did not differ substantially based on different RET fusion identified [45] (Table 2).

Table 2. Antitumor activity of multikinase inhibitors (MKIs) and RET-selective inhibitors in patients with RET-positive lung cancer. Data of principal clinical trials.

Drug	Principal Kinase Targets	Type of Study	No. of pts with RET Positive—NSCLC Treated	ORR (%)	DCR (%)	DRR (%)	Grade \geq 3 AEs (%)	Common Grade 3 or 4 TEAEs (%)
Vandetanib	VEGFR, EGFR, RET	Phase II trial [61]	19	47%	90%	53%	58%	Hypertension (58%) Rash acneiform (16%) Diarrhea (11%) Prolonged QTc (11%)
		Phase II trial [49]	17	18%	65%	22%	28%	Hypertension (17%) Prolonged QTc (11%) AST/ALT elevation (6%)
		Retrospective series [45]	11	18%	45%	NA	NA	NA
		Retrospective series [65]	3	0%	33%	33%	NA	NA
Cabozantinib	VEGFR2, MET, AXL, c-KIT, FLT3, TIE-2, RET	Phase II trial [43]	26	28%	100%	73%	47%	AST/ALT elevation (16%) Lipase elevation (15%) Decreased platelet count (8%) Hypophosphatemia (8%)
		Retrospective series [45]	21	33%	57%	NA	NA	NA
Lenvatinib	VEGFR1-3, FGFR1-4, PDGFR-A, c-KIT, RET	Phase II trial [62,64]	25	16%	76%	64%	92%	Hypertension (56%) Hyponatremia (20%) Proteinuria (16%) Pneumonia (16%) Nausea (12%)
		Retrospective series [45]	2	50%	50%	NA	NA	NA
Other MKIs								
Sorafenib	VEGFR1-3, PDGFRB, c-KIT, FLT3, BRAF, c-RAF	Phase II trial [66]	3	0%	33%	33%	33%	HFS (33%) Infection (33%)
		Retrospective series [45]	2	0%	100%	NA	NA	NA

Table 2. Cont.

Other MKIs								
Sunitinib	VEGFR1-3, PDGFRB, c-KIT, FLT3, RET	Retrospective series [45]	10	22%	50%	NA	NA	NA
		Case report [67]	1	-	-	-	-	Fatigue Thrombocytopenia
Ponatinib	BCR-ABL, FLT3, SRC, c-KIT, FGFR, VEGFR, PDGFR, RET	Retrospective series [45]	2	0%	100%	NA	NA	NA
		Case series [60]	4	50%	75%	25%	25%	Hyperbilirubinemia (25%) Increased CPK (25%)
		Retrospective series [69]	4	50%	50%	0%	0%	None
Alectinib	ALK, LTK, CHEK2, FLT3, RET	Phase II trial [71]	25	4%	52%	0%	4%	Neutropenia Pneumonitis Diarrhea Hyponatremia Increased CPK Hyperbilirubinemia (percentages NA)
		Retrospective series [45]	2	0%	0%	NA	NA	NA
		Case report [68]	1	-	-	-	-	None
Nintedanib	PDGFRA-B, VEGFR1-3, FGFR1-3	Retrospective series [45]	2	50%	100%	NA	NA	NA
Regorafenib	VEGFR1-3, PDGFRB, c-KIT, FGFR, RET, c-RAF	Retrospective series [45]	1	0%	0%	NA	NA	NA
RXDX-105	RET, BRAF	Phase I/Ib trial [63]	40	15% (19% in previous untreated pts)	52%	5%	NA	Hypophosphatemia AST/ALT elevation Rash Diarrhea Fatigue (percentages NA)

Table 2. Cont.

New RET-selective inhibitors								
Selpercatinib	RET	Phase I/II trial [73]	144 (105 evaluable for response)	64% (85% in previous untreated pts)	92% (95% in previous untreated pts)	30% of the safety population (3/531 pts)	28%	AST/ALT elevation (23%) Hypertension (14%) Hyponatremia (6%) Lymphopenia (6%)
Pralsetinib	RET	Phase I/II trial [74] *	116	61% (73% in previous untreated pts)	93%	4% of the safety population (4/120)	28%	AST elevation (22%), Hypertension (18%) ALT elevation (17%) Fatigue (15%) Neutrophilia (15%)

Abbreviations: pts, patients; NSCLC, non-small cell lung cancer; ORR, objective response rate; DCR, disease control rate; DRR, dose reduction rate; AEs, adverse events; TEAEs, treatment-emergent adverse events; AST/ALT, aspartate/alanine aminotransferases; HFS, hand-foot syndrome; CPK, creatinine phosphokinase; NA, not available. VEGFR, vascular endothelial growth factor receptor; EGFR, epidermal growth factor receptor; RET, rearranged during transfection proto-oncogene; MET, MET proto-oncogene, receptor tyrosine kinase; AXL, AXL receptor tyrosine kinase; c-KIT, KIT proto-oncogene receptor tyrosine kinase; FLT3, fms related tyrosine kinase 3; TIE2, tyrosine kinase with immunoglobulin-like and EGFR-like domains 2; FGFR, fibroblast growth factor receptor; PDGFRA(B), platelet derived growth factor receptor alpha (beta); BRAF, v-raf murine sarcoma viral oncogene homolog B1; c-RAF, RAF proto-oncogene serine/threonine-protein kinase; BCR-ABL, breakpoint cluster region-Abelson murine leukemia viral oncogene homolog 1; SRC, SRC proto-oncogene, non-receptor tyrosine kinase; ALK, ALK receptor tyrosine kinase; LTK, leukocyte tyrosine kinase; CHEK2, checkpoint kinase 2. * preliminary data.

4. New RET-Selective Inhibitors

None of the drugs described so far have been designed to preferentially bind to RET and, probably due to poor pharmacokinetic features and off-target side effects, were associated with modest clinical activity. It has been hypothesized that RET-specific antagonists could have achieved better clinical outcomes in patients harboring RET-rearranged NSCLC. Recently, two highly potent and selective RET TKIs, selpercatinib (LOXO-292) and pralsetinib (BLU-667), have been developed and their activity has been investigated in early phase trials. These agents, specifically tailored to target the activated forms of RET while sparing other kinases, offer the potential for a better clinical efficacy with a more satisfactory side effect profile. Selpercatinib has > 100-fold selectivity against VEGFR2 [75] and pralsetinib has 87-fold selectivity against VEGFR2 and 20-fold selectivity against JAK1 [74]. Furthermore, both are effective in inhibiting the RETV804L/M gatekeeper mutants and they are effective in the central nervous system [76]. Preclinically, selpercatinib (LOXO-292) demonstrated potent RET-selective antitumor activity both in *in vitro* and *in vivo* models, against both RET wild-type RET and RET alterations, with minimal activity against other kinase targets [75,77]. In a clinical setting, recent results from phase 1/2 LIBRETTO-001 trial [73] reported that selpercatinib achieved durable ORs in patients with advanced NSCLC marked by RET gene fusions. Of the first 105 enrolled patients with RET fusion-positive NSCLC, previously treated with platinum-based chemotherapy, the ORR was 64%, including two patients (2%) with a complete response and 65 patients (62%) with partial response. The median duration of response was 17.5 months and the median PFS was 16.5 months. The objective intracranial response was 91% (10/11 pts) among this cohort. Notably, in the subgroup of 39 previously untreated patients, the ORR was 85% without median PFS or OS reached at the intermediate follow-up of 9.2 months. Antitumor activity of selpercatinib was observed regardless of the specific RET fusion partner. The most common severe AEs were hypertension, hepatotoxicity, hyponatremia and lymphopenia, dose reduction was warranted in 30% of patients, but only 2% discontinued selpercatinib due to a drug-related AE. Results from LIBRETTO-001 trial led to the FDA-approval of selpercatinib for patients harboring RET-positive NSCLC, in May 2020. An ongoing phase III trial (NCT04194944) [78] is evaluating selpercatinib versus platinum-based chemotherapy (CT) with or without immunotherapy (IT) in treatment-naïve patients with advanced RET-fusion positive NSCLC. The next-generation TKI pralsetinib (BLU-667), selectively developed to target RET, demonstrated meaningful preclinical activity in a wide variety of tumors with activated RET kinase [79,80]. Preliminary data from the ongoing phase 1/2 ARROW trial (NCT03037385) demonstrated potent and durable activity and tolerability of pralsetinib in the cohort of patients with advanced RET-fusion positive NSCLC [74]. Among 116 patients with RET-positive NSCLC, 80 patients had received prior platinum treatment and 26 patients were treatment-naïve: the ORRs were 61% and 73%, respectively, with a DCR of 93% in the overall population. Furthermore, ORR was similar regardless of RET fusion partner (72% of patients had KIF5B-RET fusion NSCLC, 16% had CCDC6-RET fusion NSCLC and 12% presented other fusion) or central nervous system (CNS) involvement (56%). Most treatment-related AEs were grade 1–2 and included anemia, hepatotoxicity, constipation and hypertension. Discontinuation of treatment due to side effects was reported in 4% of patients of the safety population (all tumor types) [81]. In September 2020 pralsetinib received FDA-approval for the treatment of RET-fusion positive NSCLC patients. The international randomized phase III AcceleRET Lung study (NCT04222972) [82] is currently evaluating pralsetinib compared to standard of care as first-line in RET-positive metastatic NSCLC (Table 2).

5. Immune Checkpoint Inhibitors (ICI) and Chemotherapy (CT)

Only a few retrospective analyses based on limited amounts of patients have explored the effect of ICI as a single agent, suggesting a lack of benefits from this strategy. Particularly data from 4 retrospective analysis revealed similar characteristics among RET positive patients (i.e., young age, female gender, mainly non- or former smokers, adenocarcinoma

histotype and low expression of Programmed Death Ligand 1 (PD-L1) with disappointing PFS (2.1 to 7.6 months) and a median OS of 12.3 months. Moreover, the median OS was not reached in patients who underwent ICI earlier in their clinical history [83–86]. A correlation between different fusions in NSCLC, gender and PD-L1 or TMB expression with a poor outcome in female that frequently express the KIF5B-RET rearrangement associated with a high rate of PD-L1 had been identified [34,83]. Moreover, the role of GDNF secreted by cells in micro-environment as stimulus in PD-L1 cell expression via JAK/STAT1 in HNSCC has been demonstrated [87]. These evidence suggest, as already well-known, the lack of benefit from ICI single agent in driven-mutation NSCLC and the lack of predictive value of PD-L1. The encouraging results from the IMpower150, a phase III trial designed to evaluate a first line combination of ICI and CT also in patients with EGFR and ALK driver alterations, open to the opportunity to explore this therapeutic option also in rearranged RET patients [88]. However, the appearance of resistance mutations to MKI and TKI, the paucity of data on treatment with ICI alone, the lack of favorable data on the immunochemotherapy combination as well as the impossibility of adequate stratification of patients who could benefit from these treatments, suggests the need to try combination therapies including targeted therapies and ICI as in some recent experiences in RET positive HCC [89]. CT alone deserves a historical mention due to its role in the last decades as a therapeutic strategy in RET rearranged NSCLC. Data from GLORY global single database about the impact of this strategy in RET fusion positive NSCLC suggested a partial efficacy of the platinum combination with pemetrexed as first line (PFS 6.3 months–OS 23.6 months) [45]. Few data in the second line, and the advent of drugs with recent FDA approval (<https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-pralsetinib-lung-cancer-ret-gene-fusions> accessed on 8 January 2021) (<https://www.fda.gov/drugs/drug-approvals-and-databases/fda-approves-selpercatinib-lung-and-thyroid-cancers-ret-gene-mutations-or-fusions> accessed on 8 January 2021), relegate platinum based plus pemetrexed combination in subsequent lines, and in any case, after selective targeted therapies and available new drugs in clinical trials [86].

6. Resistance

Broadly speaking, a well-established mechanism of TKIs resistance is the development of secondary (or acquired) resistance mutations within the target kinases. These secondary somatic mutations, dynamically evolved under the selective pressure of specific TKI, enable the persistent activation of the kinases despite the presence of inhibitors. Typically, in oncogene-addicted NSCLCs, acquired mutations occur at the gatekeeper position or at the solvent front area of the kinase. These alterations confer resistance through steric interference that hinder the accessibility of drugs to the kinase ATP-binding pocket or alter the conformation of the kinase when non-contact residues are involved. Both gatekeeper and solvent front mutations have been described in different types of oncogene-driven NSCLCs. Examples of gatekeeper mutations are T790M in EGFR-mutant NSCLC, L1196M in ALK-rearranged NSCLC and L2026M in ROS-1 positive NSCLC, while typical solvent front mutations are G1202R in ALK-rearranged NSCLC and G2032R in ROS-1 positive NSCLC [90–93]. As regards RET-positive cancers, RET gatekeeper mutations at the V804 residue (V804L and V804M) primarily occur as germline mutations in sporadic medullary thyroid cancers and in about 2% of MEN2 where they act as primary driver mutations and cause intrinsic resistance to several MKIs [94]. Importantly, V804M and V804L mutations also represent the two best known secondary somatic mutations that emerge during MKIs therapies and confer resistance to treatment. Preclinically, V804M/L mutant models resulted in pan-resistance to several MKI, such as cabozantinib, vandetanib, lenvatinib and partially ponatinib, in different studies [58,80,95]. V804M/L mutations have also been reported in clinical experiences to confer resistance to vandetanib in RET-positive NSCLC patients [96,97].

In addition to gatekeeper mutations, solvent-front mutations at the G810 residue (G810A and G810R) and other mutations like S904F and I788N may be involved in sec-

ondary resistance. The G810A solvent-front mutation has been identified as a novel resistance mutation to vandetanib in cell lines expressing KIF5B-RET. However, although the RET G810A mutant cells conferred resistance to vandetanib, they acquired novel sensitivity to other MKIs, such as lenvatinib and ponatinib [58]. The missense S904F mutation occurs in the activation loop of the kinase domain, and is able to increase the autophosphorylation activity of RET kinase and confer resistance to vandetanib in vitro through an allosteric effect and has been also reported as a mechanism of acquired resistance in a patient with NSCLC harboring CCDC6-RET fusion after treatment with vandetanib [98]. Noteworthy, S904F mutation has also been described as a germline oncogene mutation with a high transforming activity, and implicated in the development of medullary thyroid cancer [99]. Moreover, in vitro analysis of KIF5B- or CCDC6-RET-rearranged cells identified I788N somatic mutation as a mechanism of acquired resistance to different MKIs, such as cabozantinib, vandetanib and AD80, but not to ponatinib [100].

Beyond the acquisition of secondary resistance mutations, another mechanism of acquired resistance in oncogene-driven NSCLC is the reactivation of different intracellular pathways, bypassing signals mediated by targeted receptor-kinase [90,101]. In RET-rearranged tumors examples of these intracellular reactivated networks include RAS/MAPK signaling, which has been reported to confer resistance to MKI AD80 in RET-rearranged cell lines and to MKI ponatinib in preclinical patient-derived models of RET-fusion positive lung adenocarcinoma [100,102]. The retained activation of EGFR and AXL signaling may contribute to the acquired resistance to MKIs, by up-regulating downstream signaling through MAPK and PI3K/AKT, respectively [102–104]. EGFR increases phosphorylation of RET in cell lines with neuroendocrine features and expression of Achaete-scute homolog 1 (ASCL1) suggesting a close cross-talking between both the RTKs and the interesting chance of combining different selective TKIs [105]. Also, the NRAS p.Q61K oncogenic mutation proved to represent another mechanism of acquired resistance to RET inhibition, again through MAPK and PI3K/AKT signaling reactivation [102]. Moreover, the increased Src activation has been reported as a further mechanism of acquired resistance to different MKIs by activating RET downstream effector ERK1/2 in RET-rearranged lung adenocarcinoma [106]. Finally, the MDM2 (a p53 antagonist) amplification has also been identified as a potential mediator of both intrinsic and acquired resistance to cabozantinib in patients with RET-rearranged lung cancers [107].

The next-generation RET-selective inhibitors selpercatinib and pralsetinib have been developed to surpass the limitations of MKIs both by sparing non-RET target kinases and by overcoming most common MKIs resistance mutations. These drugs have demonstrated equipotent and selective preclinical activity against RET rearrangements and mutations, including CCDC6-RET fusion, KIF5B-RET fusion, RET-activating mutations (C634W and M918T) and RET mutations at the gatekeeper residue (V804 L/M/E) [77,80]. Thanks to a different binding mode from MKIs, new selective RET inhibitors can avoid interference from the gatekeeper mutations, however they remain susceptible to secondary resistance from non-gatekeeper mutations. Moreover, new mechanisms of resistance have been described in preclinical models and in clinical experiences and represent an ongoing area of research. Recent studies reported that mutations at solvent front (G810R/S/C/V), hinge (Y806C/N) and β 2 strand (V738A) sites within the RET kinase domain can mediate acquired resistance to selpercatinib and pralsetinib in RET fusion-positive NSCLC and in RET-mutant medullary thyroid cancer [108,109] (Table 3). A recent multi-institutional study analyzed tumor and plasma biopsies from 18 patients with RET-rearranged NSCLC after treatment with selpercatinib and pralsetinib to characterize mechanisms of acquired resistance. The analysis detected the solvent front G810C/S mutations in two cases (10%) and identified MET amplification as recurrent mechanisms of resistance (three patients, 15%), and additionally described KRAS amplification in one resistant case [110]. In addition, MET amplification has been described in a NSCLC patient in a recently published case report [111]. These evidence are in line with the knowledge that tumor cells through

primary or secondary mechanisms of adaptations, overcome the inhibition with the re-activation of other signaling up- or downstream pathways.

Table 3. Mechanisms of resistance and IC₅₀ (μM) for each drug.

Mutation Status		Cabozantinib [112]	Vandetanib [112]	Lenvatinib [112]	Ponatinib [112]	Selpercatinib [109]	Pralsetinib [109]
Gatekeeper	V804M	4.26	5.83	5.42	0.0339	0.0559	0.0168
	V804L	3.22	6.10	10.60	0.43 [60]	0.0172	0.0018
Solvent front	G810A	0.22	2.76	0.11	0.008 [60]	-	-
	G810R	-	-	-	-	2.744	2.650
	G810S	1.05	5.47	0.67	-	0.8802	0.3906
	G810C	-	-	-	-	1.227	0.6417
	S904F	-	0.908 [98]	-	-	-	-
Other	Y806C	-	0.933 [113]	-	-	0.1744	0.2958
	Y806N	4.76	5.86	1.93	-	0.1498	0.2925
	V738A	1.20	1.05	2.35	-	0.2388	0.1775

The IC₅₀ values are mean (95% confidence interval). In red: resistant; in green: non-resistant. Values refers to BaF3 cell line, exception for Vandetanib Y806C value obtained in HEK 293.

In addition to the importance of the rising mutation or activation of alternative pathway via amplification, the moderating role of the microenvironment should be taken in consideration [114]. For instance, in an in vitro experiment in which human umbilical vein endothelial cells (HUVECs) have been used to mimic the tumor microenvironment GDNF was able to stimulate the hepatic growth factor (HGF) production and consequently the phosphorylation of its main receptor MET [115]. As GDNF is highly expressed in NSCLC [116], we could hypothesize in RET inhibitors resistant cancer cells harboring MET amplification, a cross-talking between tumor microenvironment and TK receptors. GDNF upregulates PD-L1 involved in local immune activity and leaning immune evasion [87]. With this in mind, the importance of targeting receptors or the ligand as per GDNF and understand how to modulate the cancer microenvironment is clear. To further complicate the phenomena of resistances, attention has been recently drawn to cancer cells' regulatory functions of miRNAs. In RET positive MTC, among several miRNAs identified, eight were up-regulated and one of them, miR-153-3p, was found to have tumor suppressor function, increasing the antiproliferative efficacy of cabozantinib by acting on factors involved in the mTOR pathway [117]. In NSCLC some experiences outline for miR-153-3p the function of a favorable prognostic factor when highly expressed, able to overcome of resistances to TKIs in mutated EGFR cell lines [118,119]. Mechanisms of regulation of endocytosis and RTKs cell trafficking are emerging news in pro-proliferative cancer cells behavior [120]. Several papers describe how alteration in RTKs degradation through endosomes and finally lysosomes, may affect cell proliferation, survival and migration [121]. Although it is intuitive that a lack of degradation of the RTKs could prolong the signaling, surprisingly also the intracellular accumulation inside the vesicles responsible for their removal, creates a feedback that increases the leading signals as per ERK1/2 or Akt in NSCLC [121,122]. In RET rearranged cell lines the expression of Golgi-Localized, Gamma Ear-Containing, ARF-Binding Protein 3 (GGA3), promote an everlasting recycling of the isoform RET51 on cell surfaces, promoting pro migratory functions via p-Akt [123] (Figure 2).

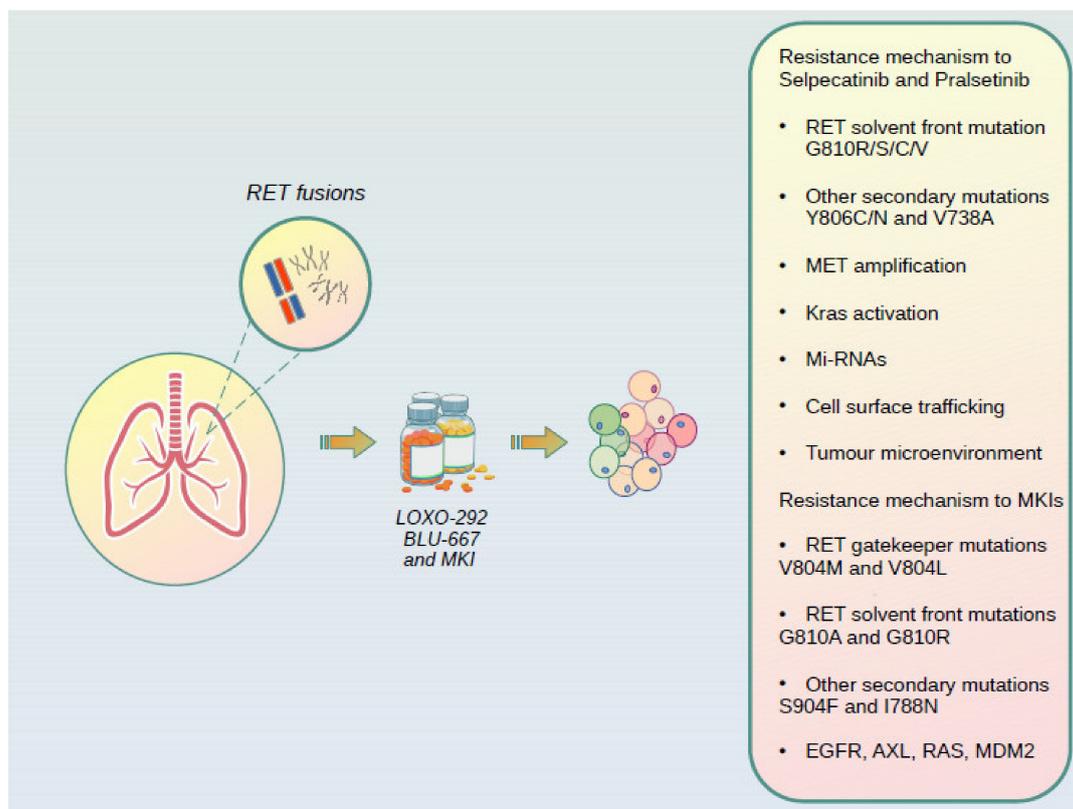


Figure 2. Most common mechanisms of resistance of multikinase inhibitors (MKIs) and new selective RET-inhibitors selpercatinib and pralsetinib.

7. Combination Strategies and Future Perspectives

According to recent experiences, resistance mechanisms to selective RET inhibitors seem to be driven mainly by off-target RET-independent mechanisms, such as MET or KRAS amplifications [110,111] while on-target resistance mutations within RET kinases after progression to selpercatinib or pralsetinib are identified less frequently in RET aberrant NSCLC. As a consequence, new treatment approaches focus on the possibility to target RET-independent resistance drivers through the combination of anti-RET therapy with other targeted agents. Preclinical studies show that the combination with the MKI vandetanib and the mTOR inhibitor everolimus is active against CCDC6-RET-positive LC-2 lung cancer cell lines and results superior to monotherapy. Everolimus targets the PI3K/AKT pathway, reactivation of which has been associated with acquired resistance to MKIs [102]. Significant antitumor activity of everolimus plus vandetanib has been demonstrated also in patients with RET rearranged NSCLC, with responses observed in all the six patients treated within the combination therapy [124]. This combination strategy showed to increase the CNS penetration and resulted particularly active against brain-metastatic RET-rearranged NSCLCs [125]. Another promising combination therapy consists in targeting both RET activated-kinase and MET amplification. In four RET-fusion positive NSCLC patients treated with selpercatinib in the LIBRETTO-001 trial, in which MET amplification has been validated as a targetable mediator of resistance to RET-directed therapy, combined therapy with selpercatinib and the MET/ALK/ROS1 inhibitor crizotinib was administered. In this case series, the combination strategy demonstrated anecdotal evidence of clinical activity and tolerability and a 10 months-lasting response was reported with the two agents [126]. Several other combined therapies have been tested in preclinical RET altered cancer models and in early phase trials in patients with RET-positive thyroid carcinomas. Treatment with RET small interfering RNA (siRNA) and irinotecan (CPT-11) has been reported to suppress RET expression and to inhibit the growth of medullary thyroid carcinoma

(MTC) xenografts via a synergic apoptotic effect [127]. The synergic antitumor activity of serine/threonine-protein kinase BRAF inhibitors (RAF256 and ZSTK474) and PI3K inhibitors (ZSTK474 and BEZ-235) on RET mediated signaling and proliferation in thyroid carcinoma cell lines harboring RET activating mutation has been described [128,129]. Combined blockade of RET and Src pathways through treatment with RPI-1 and dasatinib reduced cell proliferation in papillary thyroid carcinoma-cell lines expressing RET [130]. Moreover, association of the MKI sorafenib and a MEK inhibitor (AZD6244) demonstrated synergy in MTC cells in vitro [131], while sorafenib combined with the farnesyltransferase inhibitor tipifarnib has been evaluated in a phase I trial, showing activity in patients with RET-mutated MTC [132]. Other evidence also suggest that some repurpose drugs, such as nicotinamide, may have efficacy in RET cancer cells [133], while the use of the antibody conjugated RET-maytansine has demonstrated to be a promising strategy [134,135]. A couple of experiences in MTC and osteosarcoma cell lines has drawn the attention to the ability of piperine and ribociclib to inhibit Akt and ERK 1/2 via enhancement of activating transcription factor 4 (ATF4), suppressing RET stimuli [136,137]. In addition to combination strategies, new TKIs of different chemical scaffolds can be developed to inhibit new adaptive kinase mutants. In preclinical studies, the novel and potent RET/SRC inhibitor TPX-0046 demonstrated remarkable activity against the solvent front mutation KIF5B-RET G810R, developed as on-target resistance to selpercatinib and pralsetinib. TPX-0046 is a selective next-generation RET/SRC inhibitor, that was rationally designed with a novel macrocyclic structure and developed against various RET mutations, especially solvent front mutations [138]. BOS172738 is another novel RET inhibitor with nanomolar potency against RET and approximately 300-fold selectivity against VEGFR2 and it is currently being studied in a phase I trial (NCT03780517) [139].

Finally, further new molecules selectivity designed against RET have been tested or are currently under investigation in several preclinical trials (Table 4) [133,140–144]. In the past 10 years, several efforts have been made to discover highly selective small molecule RET inhibitors [145]. RET inhibitors based on heterocycles including benzimidazole, quinoline and pyrazolopyrimidine are reported in literature. These RET inhibitors are classified according to their hinge binder chemotypes as: pyrimidines, including the pyrazolopyrimidines, pyrimidine oxazines, quinazolines, 4-aminopyrimidines and 4-aminopyridines; indolinones; 5-aminopyrazole-4-carboxamides; 3-trifluoromethylanilines; imidazopyridines, imidazopyridazines and pyrazopyridines; nicotinonitriles; pyridones and 1,2,4-triazoles. Wang et al. [133] synthesized various nicotinamide analogs based on the scaffold of benzamide aminonaphthyridine HSN356, which was reported to inhibit RET kinase [142]. HSN608, the nicotinamide analog of HSN356 exerts strong RET inhibition and also inhibit RET(V804M/L) and RET(S905F) mutants better than alectinib, sorafenib, vandetanib and apatinib, and comparable to BLU667. Recently N-phenyl-7,8-dihydro-6H-pyrimido[5,4-b][1,4]oxazin-4-amine derivatives [143] have been reported as a new class of RET inhibitors and in particular 17d derivative, 1-(5-(tert-butyl)isoxazol-3-yl)-3-(4-((6,7,8,9-tetrahydropyrimido[5,4-b][1,4]oxazepin-4-yl)amino)phenyl)urea, potently inhibits RET and its drug resistance mutants RET-V804M and RET-V804L. Lakkaniga et al. [144] investigated a series of pyrrolo[2,3-d]pyrimidine-based derivatives and identified a lead compound, named 59, a type 2 inhibitor of RET, which shows low nanomolar potency against RET and RET V804M and additionally proposed a binding pose of 59 in RET pocket. Furthermore, new compounds targeting RET and VEGFR2 are emerging. The group of Moccia et al. [141] identified the clinical drug candidates Pz-1 and NPA101.3, who by lacking the structural liability for demethylation showed a selective inhibitory profile for both VEGFR2 and RET (WT and V804M).

Table 4. Candidates in preclinical setting.

Drugs	In Vitro			In Vivo	References
	RET IC ₅₀	VEGFR2 IC ₅₀	RET V804M IC ₅₀	Xenograft Mouse Model	
Pz-1	<0.001 µM	<0.001 µM	<0.001 µM	10 mg/kg/day per os inhibition of tumor growth	[140]
NPA-101.3	0.001 µM	0.003 µM	0.008 µM	10 mg/kg/day per os inhibition of tumor growth	[141]
HSN356	-	-	-	-	[142]
HSN608	3.16 nM	-	-	-	[133]
17d	0.01 µM	-	0.015 µM *	10–30 mg/kg/day inhibition of tumor growth	[143]
59	0.0068 µM	-	13.51 nM	-	[144]

* 0.009 µM in the V804L.

8. Conclusions

In the past decades RET oncogene has emerged as a critical tumorigenesis driver. RET mutations and rearrangements now represent a well-established mechanism that drives tumor growth across several types of neoplasms, including thyroid and lung cancer. Treatment with non-specific MKIs in RET fusion-positive NSCLC achieved modest clinical outcomes and limited response durability, especially when compared with those achieved by targeting oncogenic drivers other than RET. The two highly selective RET inhibitors, pralsetinib and selpercatinib, were specifically developed to spare non-RET target kinases and to overcome resistances to MKIs. These next-generation compounds have received FDA breakthrough designation and have been approved for clinic use based on the results of the LIBRETTO-001 and ARROW trials. Although these agents have been developed to overcome MKIs limits and have demonstrated remarkable clinical activity, new mechanisms of acquired resistance have already been reported. The emergence of off-target RET-independent mechanisms of resistance to pralsetinib and selpercatinib has highlighted the necessity to test further next-generation agents and to explore new therapeutic strategies, including concurrent inhibition of RET and parallel signaling pathways of resistance.

Author Contributions: Conceptualization, S.F., E.C. and S.P.; resources, S.F. and E.C.; writing—original draft preparation, S.F., E.C., F.M. and S.P.; writing—review and editing, L.A., L.V., F.C., M.B., F.M.; supervision, L.A. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Takahashi, M.; Ritz, J.; Cooper, G.M. Activation of a novel human transforming gene, *ret*, by DNA rearrangement. *Cell* **1985**, *42*, 581–588. [\[CrossRef\]](#)
2. Andrew, S.D.; Delhanty, P.J.D.; Mulligan, L.M.; Robinson, B.G. Sp1 and Sp3 transactivate the RET proto-oncogene promoter. *Gene* **2000**, *256*, 283–291. [\[CrossRef\]](#)
3. Lang, D.; Epstein, J.A. Sox10 and Pax3 physically interact to mediate activation of a conserved c-RET enhancer. *Hum. Mol. Genet.* **2003**, *12*, 937–945. [\[CrossRef\]](#)
4. Leon, T.Y.Y.; Ngan, E.S.W.; Poon, H.-C.; So, M.-T.; Lui, V.C.H.; Tam, P.K.H.; Garcia-Barcelo, M.M. Transcriptional regulation of RET by Nkx2-1, Phox2b, Sox10, and Pax3. *J. Pediatr. Surg.* **2009**, *44*, 1904–1912. [\[CrossRef\]](#)

5. Ibáñez, C.F. Structure and Physiology of the RET Receptor Tyrosine Kinase. *Cold Spring Harb. Perspect. Biol.* **2013**, *5*, a009134. [[CrossRef](#)] [[PubMed](#)]
6. Arighi, E.; Borrello, M.G.; Sariola, H. RET tyrosine kinase signaling in development and cancer. *Cytokine Growth Factor Rev.* **2005**, *16*, 441–467. [[CrossRef](#)] [[PubMed](#)]
7. Wang, Z.; Ho, J.X.; Ruble, J.R.; Rose, J.; Rüker, F.; Ellenburg, M.; Murphy, R.; Click, J.; Soistman, E.; Wilkerson, L.; et al. Structural studies of several clinically important oncology drugs in complex with human serum albumin. *Biochim. Biophys. Acta BBA Gen. Subj.* **2013**, *1830*, 5356–5374. [[CrossRef](#)] [[PubMed](#)]
8. Li, J.; Shang, G.; Chen, Y.J.; Brautigam, C.A.; Liou, J.; Zhang, X.; Bai, X.C. Cryo-EM analyses reveal the common mechanism and diversification in the activation of RET by different ligands. *Elife* **2019**, *8*, 1–26. [[CrossRef](#)] [[PubMed](#)]
9. Su, A.I.; Cooke, M.P.; Ching, K.A.; Hakak, Y.; Walker, J.R.; Wiltshire, T.; Orth, A.P.; Vega, R.G.; Sapinoso, L.M.; Moqrich, A.; et al. Large-Scale Analysis of the Human and Mouse Transcriptomes. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 4465–4470. [[CrossRef](#)] [[PubMed](#)]
10. Avantaggiato, V.; Dathan, N.A.; Grieco, M.; Fabien, N.; Lazzaro, D.; Fusco, A.; Simeone, A.; Santoro, M. Developmental Expression of the RET Protooncogene. *Cell Growth Differ.* **1994**, *5*, 305–311. [[PubMed](#)]
11. Schuchardt, A.; Dagati, V.D.; Larsson-Blomberg, L.; Costantini, F.; Pachnis, V. Defects in the kidney and enteric nervous system of mice lacking the tyrosine kinase receptor Ret. *Nat. Cell Biol.* **1994**, *367*, 380–383. [[CrossRef](#)] [[PubMed](#)]
12. Durbec, P.L.; Larsson-Blomberg, L.B.; Schuchardt, A.; Costantini, F.; Pachnis, V. Common origin and developmental dependence on c-ret of subsets of enteric and sympathetic neuroblasts. *Development* **1996**, *122*, 349–358.
13. Meng, X.; Lindahl, M.; Hyvönen, M.E.; Parvinen, M.; De Rooij, D.G.; Hess, M.W.; Raatikainen-Ahokas, A.; Sainio, K.; Rauvala, H.; Lakso, M.; et al. Regulation of Cell Fate Decision of Undifferentiated Spermatogonia by GDNF. *Science* **2000**, *287*, 1489–1493. [[CrossRef](#)] [[PubMed](#)]
14. Mederer, T.; Schmitteckert, S.; Volz, J.; Martínez, C.; Röth, R.; Thumberger, T.; Eckstein, V.; Scheuerer, J.; Thöni, C.; Lasitschka, F.; et al. A complementary study approach unravels novel players in the pathoetiology of Hirschsprung disease. *PLoS Genet.* **2020**, *16*, e1009106. [[CrossRef](#)] [[PubMed](#)]
15. Nicolaou, N.; Renkema, K.Y.; Bongers, E.M.H.F.; Giles, R.H.; Knoers, N.V.A.M. Genetic, environmental, and epigenetic factors involved in CAKUT. *Nat. Rev. Nephrol.* **2015**, *11*, 720–731. [[CrossRef](#)] [[PubMed](#)]
16. Fonseca-Pereira, D.; Arroz-Madeira, S.; Rodrigues-Campos, M.; Barbosa, I.A.M.; Domingues, R.G.; Bento, T.; Almeida, A.R.M.; Ribeiro, H.; Potocnik, A.J.; Enomoto, H.; et al. The neurotrophic factor receptor RET drives haematopoietic stem cell survival and function. *Nat. Cell Biol.* **2014**, *514*, 98–101. [[CrossRef](#)]
17. Marx, S.J. Molecular genetics of multiple endocrine neoplasia types 1 and 2. *Nat. Rev. Cancer* **2005**, *5*, 367–375. [[CrossRef](#)]
18. Kouvaraki, M.A.; Shapiro, S.E.; Perrier, N.D.; Cote, G.J.; Gagel, R.F.; Hoff, A.O.; Sherman, S.I.; Lee, J.E.; Evans, D.B. RET Proto-Oncogene: A Review and Update of Genotype–Phenotype Correlations in Hereditary Medullary Thyroid Cancer and Associated Endocrine Tumors. *Thyroid* **2005**, *15*, 531–544. [[CrossRef](#)] [[PubMed](#)]
19. Wohllk, N.; Cote, G.J.; Evans, D.B.; Goepfert, H.; Ordóñez, N.G.; Gagel, R.F. Application of Genetic Screening Information to the Management of Medullary Thyroid Carcinoma and Multiple Endocrine Neoplasia Type 2. *Endocrinol. Metab. Clin. N. Am.* **1996**, *25*, 1–25. [[CrossRef](#)]
20. Wells, S.A. Advances in the management of MEN2: From improved surgical and medical treatment to novel kinase inhibitors. *Endocr. Relat. Cancer* **2018**, *25*, T1–T13. [[CrossRef](#)] [[PubMed](#)]
21. Kloos, R.T.; Eng, C.; Evans, D.B.; Francis, G.L.; Gagel, R.F.; Gharib, H.; Moley, J.F.; Pacini, F.; Ringel, M.D.; Schlumberger, M.; et al. Medullary Thyroid Cancer: Management Guidelines of the American Thyroid Association. *Thyroid Off. J. Am. Thyroid Assoc.* **2009**, *19*, 565–612. [[CrossRef](#)] [[PubMed](#)]
22. Suzuki, K.; Saenko, V.; Yamashita, S.; Mitsutake, N. Radiation-Induced Thyroid Cancers: Overview of Molecular Signatures. *Cancers* **2019**, *11*, 1290. [[CrossRef](#)] [[PubMed](#)]
23. Nikiforov, Y.E.; Rowland, J.M.; Bove, K.E.; Monforte-Munoz, H.; Fagin, J.A. Distinct Pattern of Ret Oncogene Rearrangements in Morphological Variants of Radiation-Induced and Sporadic Thyroid Papillary Carcinomas in Children. *Cancer Res.* **1997**, *57*, 1690–1694. [[PubMed](#)]
24. Santoro, M.; Moccia, M.; Federico, G.; Carlomagno, F. RET Gene Fusions in Malignancies of the Thyroid and Other Tissues. *Genes* **2020**, *11*, 424. [[CrossRef](#)]
25. Boulay, A.; Breuleux, M.; Stephan, C.; Fux, C.; Brisken, C.; Fiche, M.; Wartmann, M.; Stumm, M.; Lane, H.A.; Hynes, N.E. The Ret Receptor Tyrosine Kinase Pathway Functionally Interacts with the ER α Pathway in Breast Cancer. *Cancer Res.* **2008**, *68*, 3743–3751. [[CrossRef](#)] [[PubMed](#)]
26. Morandi, A.; Plaza-Menacho, I.; Isacke, C.M. RET in breast cancer: Functional and therapeutic implications. *Trends Mol. Med.* **2011**, *17*, 149–157. [[CrossRef](#)] [[PubMed](#)]
27. Takeda, M.; Watanabe, S.; Rothenberg, S.M.; Kherani, J.; French, P.P.; Olek, E. Abstract 5236: Complete response to selpercatinib (LOXO-292), a highly selective RET inhibitor, in a patient with RET fusion-positive breast cancer. *Cancer Res.* **2020**, *80*, 5236. [[CrossRef](#)]
28. Le Rolle, A.-F.; Klempner, S.J.; Garrett, C.R.; Seery, T.; Sanford, E.M.; Balasubramanian, S.; Ross, J.S.; Stephens, P.J.; Miller, V.A.; Ali, S.M.; et al. Identification and characterization of RET fusions in advanced colorectal cancer. *Oncotarget* **2015**, *6*, 28929–28937. [[CrossRef](#)] [[PubMed](#)]

29. Santos, C.; Sanz-Pamplona, R.; Salazar, R. RET-fusions: A novel paradigm in colorectal cancer. *Ann. Oncol.* **2018**, *29*, 1340–1343. [[CrossRef](#)] [[PubMed](#)]
30. Pietrantonio, F.; Di Nicolantonio, F.; Schrock, A.; Lee, J.; Morano, F.; Fucà, G.; Nikolinakos, P.; Drilon, A.; Hechtman, J.; Christiansen, J.; et al. RET fusions in a small subset of advanced colorectal cancers at risk of being neglected. *Ann. Oncol.* **2018**, *29*, 1394–1401. [[CrossRef](#)]
31. Kohno, T.; Tabata, J.; Nakaoku, T. REToma: A cancer subtype with a shared driver oncogene. *Carcinogenesis* **2019**, *41*, 123–129. [[CrossRef](#)]
32. Kohno, T.; Ichikawa, H.; Totoki, Y.; Yasuda, K.; Hiramoto, M.; Nammo, T.; Sakamoto, H.; Tsuta, K.; Furuta, K.; Shimada, Y.; et al. KIF5B-RET fusions in lung adenocarcinoma. *Nat. Med.* **2012**, *18*, 375–377. [[CrossRef](#)]
33. Ju, Y.S.; Lee, W.-C.; Shin, J.-Y.; Lee, S.; Bleazard, T.; Won, J.-K.; Kim, Y.T.; Kim, J.-I.; Kang, J.-H.; Seo, J.-S. A transforming KIF5B and RET gene fusion in lung adenocarcinoma revealed from whole-genome and transcriptome sequencing. *Genome Res.* **2011**, *22*, 436–445. [[CrossRef](#)] [[PubMed](#)]
34. Qiu, Z.; Ye, B.; Wang, K.; Zhou, P.; Zhao, S.; Li, W.; Tian, P. Unique Genetic Characteristics and Clinical Prognosis of Female Patients with Lung Cancer Harboring RET Fusion Gene. *Sci. Rep.* **2020**, *10*, 1–9. [[CrossRef](#)]
35. Mizukami, T.; Shiraishi, K.; Shimada, Y.; Ogiwara, H.; Tsuta, K.; Ichikawa, H.; Sakamoto, H.; Kato, M.; Shibata, T.; Nakano, T.; et al. Molecular Mechanisms Underlying Oncogenic RET Fusion in Lung Adenocarcinoma. *J. Thorac. Oncol.* **2014**, *9*, 622–630. [[CrossRef](#)]
36. Lipson, D.; Capelletti, M.; Yelensky, R.; Otto, G.; Parker, A.; Jarosz, M.; Curran, J.A.; Balasubramanian, S.; Bloom, T.; Brennan, K.W.; et al. Identification of new ALK and RET gene fusions from colorectal and lung cancer biopsies. *Nat. Med.* **2012**, *18*, 382–384. [[CrossRef](#)] [[PubMed](#)]
37. Kato, S.; Subbiah, V.; Marchlik, E.; Elkin, S.K.; Carter, J.L.; Kurzrock, R. RET Aberrations in Diverse Cancers: Next-Generation Sequencing of 4,871 Patients. *Clin. Cancer Res.* **2017**, *23*, 1988–1997. [[CrossRef](#)] [[PubMed](#)]
38. Tsai, T.-H.; Wu, S.-G.; Hsieh, M.-S.; Jin-Yuan, S.; Yang, J.C.-H.; Shih, J.-Y. Clinical and prognostic implications of RET rearrangements in metastatic lung adenocarcinoma patients with malignant pleural effusion. *Lung Cancer* **2015**, *88*, 208–214. [[CrossRef](#)]
39. Suzuki, M.; Makinoshima, H.; Matsumoto, S.; Suzuki, A.; Mimaki, S.; Matsushima, K.; Yoh, K.; Goto, K.; Suzuki, Y.; Ishii, G.; et al. Identification of a lung adenocarcinoma cell line with CCDC 6- RET fusion gene and the effect of RET inhibitors in vitro and in vivo. *Cancer Sci.* **2013**, *104*, 896–903. [[CrossRef](#)]
40. Melillo, R.M.; Castellone, M.D.; Guarino, V.; De Falco, V.; Cirafici, A.M.; Salvatore, G.; Caiazzo, F.; Basolo, F.; Giannini, R.; Kruhoffer, M.; et al. The RET/PTC-RAS-BRAF linear signaling cascade mediates the motile and mitogenic phenotype of thyroid cancer cells. *J. Clin. Investig.* **2016**, *126*, 1603. [[CrossRef](#)]
41. Saito, M.; Ishigame, T.; Tsuta, K.; Kumamoto, K.; Imai, T.; Kohno, T. A mouse model of KIF5B-RET fusion-dependent lung tumorigenesis. *Carcinogenesis* **2014**, *35*, 2452–2456. [[CrossRef](#)]
42. Xu, H.; Shen, J.; Xiang, J.; Li, H.; Li, B.; Zhang, T.; Zhang, L.; Mao, X.; Jian, H.; Shu, Y. Characterization of acquired receptor tyrosine-kinase fusions as mechanisms of resistance to EGFR tyrosine-kinase inhibitors. *Cancer Manag. Res.* **2019**, *11*, 6343–6351. [[CrossRef](#)]
43. Drilon, A.; Rekhman, N.; Arcila, M.; Wang, L.; Ni, A.; Albano, M.; Van Voorthuysen, M.; Somwar, R.; Smith, R.S.; Montecalvo, J.; et al. Cabozantinib in patients with advanced RET-rearranged non-small-cell lung cancer: An open-label, single-centre, phase 2, single-arm trial. *Lancet Oncol.* **2016**, *17*, 1653–1660. [[CrossRef](#)]
44. Lira, M.E.; Choi, Y.-L.; Lim, S.M.; Deng, S.; Huang, D.; Ozeck, M.; Han, J.; Jeong, J.Y.; Shim, H.S.; Cho, B.C.; et al. A Single-Tube Multiplexed Assay for Detecting ALK, ROS1, and RET Fusions in Lung Cancer. *J. Mol. Diagn.* **2014**, *16*, 229–243. [[CrossRef](#)] [[PubMed](#)]
45. Gautschi, O.; Milia, J.; Filleron, T.; Wolf, J.; Carbone, D.P.; Owen, D.; Camidge, R.; Narayanan, V.; Doebele, R.C.; Besse, B.; et al. Targeting RET in Patients With RET-Rearranged Lung Cancers: Results From the Global, Multicenter RET Registry. *J. Clin. Oncol.* **2017**, *35*, 1403–1410. [[CrossRef](#)] [[PubMed](#)]
46. Velcheti, V.; Thawani, R.; Khunger, M.; Mukhopadhyay, S.; Chute, D.J.; Schrock, A.B.; Ali, S.M. FRMD4A / RET: A Novel RET Oncogenic Fusion Variant in Non-Small Cell Lung Carcinoma. *J. Thorac. Oncol.* **2017**, *12*, e15–e16. [[CrossRef](#)]
47. Zhang, X.; Li, Y.; Liu, C.; Wang, W.; Li, M.; Lv, D.; Sun, G.; Chen, H.; Dong, X.; Miao, Z.; et al. Identification of a novel KIF13A-RET fusion in lung adenocarcinoma by next-generation sequencing. *Lung Cancer* **2018**, *118*, 27–29. [[CrossRef](#)] [[PubMed](#)]
48. Nakaoku, T.; Tsuta, K.; Ichikawa, H.; Shiraishi, K.; Sakamoto, H.; Enari, M.; Furuta, K.; Shimada, Y.; Ogiwara, H.; Watanabe, S.-I.; et al. Druggable Oncogene Fusions in Invasive Mucinous Lung Adenocarcinoma. *Clin. Cancer Res.* **2014**, *20*, 3087–3093. [[CrossRef](#)]
49. Lee, S.-H.; Lee, J.-K.; Ahn, M.-J.; Kim, D.-W.; Sun, J.-M.; Keam, B.; Kim, T.M.; Heo, D.S.; Ahn, J.S.; Choi, Y.-L.; et al. Vandetanib in pretreated patients with advanced non-small cell lung cancer-harboring RET rearrangement: A phase II clinical trial. *Ann. Oncol.* **2017**, *28*, 292–297. [[CrossRef](#)]
50. Wang, R.; Hu, H.; Pan, Y.; Li, Y.; Ye, T.; Li, C.; Luo, X.; Wang, L.; Li, H.; Zhang, Y.; et al. RET Fusions Define a Unique Molecular and Clinicopathologic Subtype of Non-Small-Cell Lung Cancer. *J. Clin. Oncol.* **2012**, *30*, 4352–4359. [[CrossRef](#)]
51. Zehir, A.; Benayed, R.; Shah, R.H.; Syed, A.; Middha, S.; Kim, H.R.; Srinivasan, P.; Gao, J.; Chakravarty, D.; Devlin, S.M.; et al. Mutational landscape of metastatic cancer revealed from prospective clinical sequencing of 10,000 patients. *Nat. Med.* **2017**, *23*, 703–713. [[CrossRef](#)] [[PubMed](#)]

52. Zheng, Z.; Liebers, M.; Zhelyazkova, B.; Cao, Y.; Panditi, D.; Lynch, K.D.; Chen, J.; Robinson, H.E.; Shim, H.S.; Chmielecki, J.; et al. Anchored multiplex PCR for targeted next-generation sequencing. *Nat. Med.* **2014**, *20*, 1479–1484. [[CrossRef](#)] [[PubMed](#)]
53. Peng, P.; Zheng, Y.; Lv, J. TBC1D32-RET: A Novel RET Oncogenic Fusion in Lung Adenocarcinoma. *J. Thorac. Oncol.* **2019**, *14*, e7–e9. [[CrossRef](#)]
54. Drilon, A.; Wang, L.; Hasanovic, A.; Suehara, Y.; Lipson, D.; Stephens, P.; Ross, J.; Miller, V.; Ginsberg, M.; Zakowski, M.F.; et al. Response to Cabozantinib in Patients with RET Fusion-Positive Lung Adenocarcinomas. *Cancer Discov.* **2013**, *3*, 630–635. [[CrossRef](#)]
55. Velcheti, V.; Madison, R.; Ali, S.M.; Schrock, A.B. WAC/RET: A Novel RET Oncogenic Fusion Variant in Non-Small Cell Lung Carcinoma. *J. Thorac. Oncol.* **2018**, *13*, e122–e123. [[CrossRef](#)] [[PubMed](#)]
56. Knowles, P.P.; Murray-Rust, J.; Kjær, S.; Scott, R.P.; Hanrahan, S.; Santoro, M.; Ibáñez, C.F.; McDonald, N.Q. Structure and Chemical Inhibition of the RET Tyrosine Kinase Domain. *J. Biol. Chem.* **2006**, *281*, 33577–33587. [[CrossRef](#)]
57. Ferrara, M.G.; Di Noia, V.; D'Argento, E.; Vita, E.; Damiano, P.; Cannella, A.; Ribelli, M.; Pilotto, S.; Milella, M.; Tortora, G.; et al. Oncogene-Addicted Non-Small-Cell Lung Cancer: Treatment Opportunities and Future Perspectives. *Cancers* **2020**, *12*, 1196. [[CrossRef](#)]
58. Huang, Q.; Schneeberger, V.E.; Luetkeke, N.; Jin, C.; Afzal, R.; Budzevich, M.M.; Mankanji, R.J.; Martinez, G.V.; Shen, T.; Zhao, L.; et al. Preclinical Modeling of KIF5B-RET Fusion Lung Adenocarcinoma. *Mol. Cancer Ther.* **2016**, *15*, 2521–2529. [[CrossRef](#)] [[PubMed](#)]
59. Takeuchi, K.; Soda, M.; Togashi, Y.; Suzuki, R.; Sakata, S.; Hatano, S.; Asaka, R.; Hamanaka, W.; Ninomiya, H.; Uehara, H.; et al. RET, ROS1 and ALK fusions in lung cancer. *Nat. Med.* **2012**, *18*, 378–381. [[CrossRef](#)] [[PubMed](#)]
60. Lin, J.J.; Kennedy, E.; Sequist, L.V.; Brastianos, P.K.; Goodwin, K.E.; Stevens, S.; Wanat, A.C.; Stober, L.L.; Digumarthy, S.R.; Engelman, J.A.; et al. Clinical Activity of Alectinib in Advanced RET-Rearranged Non-Small Cell Lung Cancer. *J. Thorac. Oncol.* **2016**, *11*, 2027–2032. [[CrossRef](#)] [[PubMed](#)]
61. Yoh, K.; Seto, T.; Satouchi, M.; Nishio, M.; Yamamoto, N.; Murakami, H.; Nogami, N.; Matsumoto, S.; Kohno, T.; Tsuta, K.; et al. Vandetanib in patients with previously treated RET-rearranged advanced non-small-cell lung cancer (LURET): An open-label, multicentre phase 2 trial. *Lancet Respir. Med.* **2017**, *5*, 42–50. [[CrossRef](#)]
62. Hida, T.; Velcheti, V.; Reckamp, K.L.; Nokihara, H.; Sachdev, P.; Kubota, T.; Nakada, T.; Dutcus, C.E.; Ren, M.; Tamura, T. A phase 2 study of lenvatinib in patients with RET fusion-positive lung adenocarcinoma. *Lung Cancer* **2019**, *138*, 124–130. [[CrossRef](#)]
63. Drilon, A.; Fu, S.; Patel, M.R.; Fakih, M.; Wang, D.; Olszanski, A.J.; Morgensztern, D.; Liu, S.V.; Cho, B.C.; Bazhenova, L.; et al. A Phase I/Ib Trial of the VEGFR-Sparing Multikinase RET Inhibitor RXDX-105. *Cancer Discov.* **2018**, *9*, 384–395. [[CrossRef](#)]
64. Velcheti, V.; Hida, T.; Reckamp, K.L.; Yang, J.C.; Nokihara, H.; Sachdev, P.; Feit, K.; Kubota, T.; Nakada, T.; Dutcus, C.E.; et al. Phase 2 study of lenvatinib (LN) in patients (Pts) with RET fusion-positive adenocarcinoma of the lung. *Ann. Oncol.* **2016**, *27*, vi417. [[CrossRef](#)]
65. Platt, A.; Morten, J.; Ji, Q.; Elvin, P.; Womack, C.; Su, X.; Donald, E.; Gray, N.; Read, J.; Bigley, G.; et al. A retrospective analysis of RET translocation, gene copy number gain and expression in NSCLC patients treated with vandetanib in four randomized Phase III studies. *BMC Cancer* **2015**, *15*, 1–10. [[CrossRef](#)]
66. Horiike, A.; Takeuchi, K.; Uenami, T.; Kawano, Y.; Tanimoto, A.; Kaburaki, K.; Tambo, Y.; Kudo, K.; Yanagitani, N.; Ohyanagi, F.; et al. Sorafenib treatment for patients with RET fusion-positive non-small cell lung cancer. *Lung Cancer* **2016**, *93*, 43–46. [[CrossRef](#)]
67. Wu, H.; Shih, J.-Y.; Yang, J.C.-H. Rapid Response to Sunitinib in a Patient with Lung Adenocarcinoma Harboring KIF5B-RET Fusion Gene. *J. Thorac. Oncol.* **2015**, *10*, e95–e96. [[CrossRef](#)] [[PubMed](#)]
68. Velcheti, V.; Ahluwalia, M. Intracranial and Systemic Response to Alectinib in a Patient with RET-KIF5B Oncogenic Fusion. *J. Thorac. Oncol.* **2017**, *12*, e98–e99. [[CrossRef](#)] [[PubMed](#)]
69. Ribeiro, M.F.S.A.; Alessi, J.V.M.; Oliveira, L.J.C.; Gongora, A.B.L.; Sacardo, K.P.; Zucchetti, B.M.; Shimada, A.K.; de Galiza Barbosa, F.; Feher, O.; Katz, A. Alectinib activity in chemotherapy-refractory metastatic RET-rearranged non-small cell lung carcinomas: A case series. *Lung Cancer* **2020**, *139*, 9–12. [[CrossRef](#)] [[PubMed](#)]
70. Nosaki, K.; Takeuchi, S.; Takahara, S.; Kawakami, T.; Yoh, K.; Kono, Y.; Horiike, A.; Seto, T.; Goto, K.; Yoshimura, K.; et al. Safety of alectinib in non-small cell lung cancer patients with RET fusion gene (ALL-RET): Results from the dose-finding portion of a phase 1/2 study. *Ann. Oncol.* **2017**, *28*, x127. [[CrossRef](#)]
71. Yanagitani, N.; Takeuchi, S.; Murayama, T.; Yoshimura, K.; Imai, Y.; Takahara, S.; Kawakami, T.; Seto, T.; Hattori, Y.; Ohashi, K.; et al. OA02.01 Alectinib in Previously Treated RET-Rearranged Advanced Non-Small-Cell Lung Cancer: A Phase 1/2 Trial (ALL-RET). *J. Thorac. Oncol.* **2019**, *14*, S207. [[CrossRef](#)]
72. Li, G.G.; Somwar, R.; Joseph, J.; Smith, R.S.; Hayashi, T.; Martin, L.; Franovic, A.; Schairer, A.; Martin, E.; Riely, G.J.; et al. Antitumor Activity of RXDX-105 in Multiple Cancer Types with RET Rearrangements or Mutations. *Clin. Cancer Res.* **2017**, *23*, 2981–2990. [[CrossRef](#)] [[PubMed](#)]
73. Drilon, A.; Oxnard, G.R.; Tan, D.S.; Loong, H.H.; Johnson, M.; Gainor, J.; McCoach, C.E.; Gautschi, O.; Besse, B.; Cho, B.C.; et al. Efficacy of Selpercatinib in RET Fusion-Positive Non-Small-Cell Lung Cancer. *N. Engl. J. Med.* **2020**, *383*, 813–824. [[CrossRef](#)]
74. Gainor, J.F.; Lee, D.H.; Curigliano, G.; Doebele, R.C.; Kim, D.-W.; Baik, C.S.; Tan, D.S.-W.; Lopes, G.; Gadgeel, S.M.; Cassier, P.A.; et al. Clinical activity and tolerability of BLU-667, a highly potent and selective RET inhibitor, in patients (pts) with advanced RET-fusion+ non-small cell lung cancer (NSCLC). *J. Clin. Oncol.* **2019**, *37*, 9008. [[CrossRef](#)]

75. Brandhuber, B.; Haas, J.; Tuch, B.; Ebata, K.; Bouhana, K.; McFaddin, E.; Williams, L.; Winski, S.; Brown, E.; Burkhard, M.; et al. The development of a potent, KDR/VEGFR2-sparing RET kinase inhibitor for treating patients with RET-dependent cancers. *Eur. J. Cancer* **2016**, *69*, S144. [[CrossRef](#)]
76. Drilon, A.; Lin, J.J.; Filleron, T.; Ni, A.; Milia, J.; Bergagnini, I.; Hatzoglou, V.; Velcheti, V.; Offin, M.; Li, B.; et al. Frequency of Brain Metastases and Multikinase Inhibitor Outcomes in Patients With RET-Rearranged Lung Cancers. *J. Thorac. Oncol.* **2018**, *13*, 1595–1601. [[CrossRef](#)] [[PubMed](#)]
77. Subbiah, V.; Velcheti, V.; Tuch, B.; Ebata, K.; Busaidy, N.; Cabanillas, M.; Wirth, L.; Stock, S.; Smith, S.; Lauriault, V.; et al. Selective RET kinase inhibition for patients with RET-altered cancers. *Ann. Oncol.* **2018**, *29*, 1869–1876. [[CrossRef](#)]
78. Solomon, B.J.; Zhou, C.C.; Drilon, A.; Park, K.; Wolf, J.; Elamin, Y.; Davis, H.M.; Soldatenkova, V.; Sashegyi, A.; Lin, A.B.; et al. Phase III study of selpercatinib versus chemotherapy ± pembrolizumab in untreated RET positive non-small-cell lung cancer. *Future Oncol.* **2021**, *17*, 763–773. [[CrossRef](#)] [[PubMed](#)]
79. Rahal, R.; Maynard, M.; Hu, W.; Brubaker, J.; Cao, Q.; Kim, J.L.; Sheets, M.P.; Wilson, D.P.; Wilson, K.J.; DiPietro, L.; et al. Abstract B151: BLU-667 Is a Potent and Highly Selective RET Inhibitor Being Developed for RET-Driven Cancers. In *Proceedings of the AACR-NCI-EORTC International Conference: Molecular Targets and Cancer Therapeutics, Philadelphia, PA, USA, 26–30 October 2017*; American Association for Cancer Research (AACR): Philadelphia, PA, USA, 2018; Volume 17, p. B151.
80. Subbiah, V.; Gainor, J.F.; Rahal, R.; Brubaker, J.D.; Kim, J.L.; Maynard, M.; Hu, W.; Cao, Q.; Sheets, M.P.; Wilson, D.; et al. Precision Targeted Therapy with BLU-667 for RET-Driven Cancers. *Cancer Discov.* **2018**, *8*, 836–849. [[CrossRef](#)]
81. Gainor, J.F.; Curigliano, G.; Kim, D.-W.; Lee, D.H.; Besse, B.; Baik, C.S.; Doebele, R.C.; Cassier, P.A.; Lopes, G.; Tan, D.S.-W.; et al. Registrational dataset from the phase I/II ARROW trial of pralsetinib (BLU-667) in patients (pts) with advanced RET fusion+ non-small cell lung cancer (NSCLC). *J. Clin. Oncol.* **2020**, *38*, 9515. [[CrossRef](#)]
82. Besse, B.; Felip, E.; Clifford, C.; Louie-Gao, M.; Green, J.; Turner, C.D.; Papat, S. AcceleRET Lung: A phase III study of first-line pralsetinib in patients (pts) with RET-fusion+ advanced/metastatic non-small cell lung cancer (NSCLC). *J. Clin. Oncol.* **2020**, *38*, TPS9633. [[CrossRef](#)]
83. Offin, M.; Guo, R.; Wu, S.L.; Sabari, J.; Land, J.D.; Ni, A.; Montecalvo, J.; Halpenny, D.F.; Buie, L.W.; Pak, T.; et al. Immunophenotype and Response to Immunotherapy of RET-Rearranged Lung Cancers. *JCO Precis. Oncol.* **2019**, *3*, 1–8. [[CrossRef](#)]
84. Guisier, F.; Dubos-Arvis, C.; Viñas, F.; Doubre, H.; Ricordel, C.; Ropert, S.; Janicot, H.; Bernardi, M.; Fournel, P.; Lamy, R.; et al. Efficacy and Safety of Anti-PD-1 Immunotherapy in Patients with Advanced NSCLC with BRAF, HER2, or MET Mutations or RET Translocation: GFPC 01-2018. *J. Thorac. Oncol.* **2020**, *15*, 628–636. [[CrossRef](#)] [[PubMed](#)]
85. Mazieres, J.; Drilon, A.; Lusque, A.; Mhanna, L.; Cortot, A.; Mezquita, L.; Thai, A.; Mascoux, C.; Couraud, S.; Veillon, R.; et al. Immune checkpoint inhibitors for patients with advanced lung cancer and oncogenic driver alterations: Results from the IMMUNOTARGET registry. *Ann. Oncol.* **2019**, *30*, 1321–1328. [[CrossRef](#)] [[PubMed](#)]
86. Hegde, A.; Andreev-Drakhlin, A.Y.; Roszik, J.; Huang, L.; Liu, S.; Hess, K.; Cabanillas, M.; Hu, M.I.; Busaidy, N.L.; Sherman, S.I.; et al. Responsiveness to immune checkpoint inhibitors versus other systemic therapies in RET-aberrant malignancies. *ESMO Open* **2020**, *5*, e000799. [[CrossRef](#)] [[PubMed](#)]
87. Lin, C.; Cao, W.; Ren, Z.; Tang, Y.; Zhang, C.; Yang, R.; Chen, Y.; Liu, Z.; Peng, C.; Wang, L.; et al. GDNF secreted by nerves enhances PD-L1 expression via JAK2-STAT1 signaling activation in HNSCC. *Oncot Immunology* **2017**, *6*, e1353860. [[CrossRef](#)] [[PubMed](#)]
88. Socinski, M.A.; Jotte, R.M.; Cappuzzo, F.; Orlandi, F.; Stroyakovskiy, D.; Nogami, N.; Rodríguez-Abreu, D.; Moro-Sibilot, D.; Thomas, C.A.; Barlesi, F. Atezolizumab for First-Line Treatment of Metastatic Nonsquamous NSCLC. *N. Engl. J. Med.* **2018**, *378*, 2288–2301. [[CrossRef](#)]
89. Yang, X.; Shi, J.; Chen, X.; Jiang, Y.; Zhao, H. Efficacy of Cabozantinib and Nivolumab in Treating Hepatocellular Carcinoma with RET Amplification, High Tumor Mutational Burden, and PD-L1 Expression. *Oncologist* **2020**, *25*, 470–474. [[CrossRef](#)]
90. Lin, J.J.; Shaw, A.T. Resisting Resistance: Targeted Therapies in Lung Cancer. *Trends Cancer* **2016**, *2*, 350–364. [[CrossRef](#)]
91. Pailler, E.; Faugeroux, V.; Oulhen, M.; Mezquita, L.; Laporte, M.; Honoré, A.; Lecluse, Y.; Queffelec, P.; Ngo-Camus, M.; Nicotra, C.; et al. Acquired Resistance Mutations to ALK Inhibitors Identified by Single Circulating Tumor Cell Sequencing in ALK-Rearranged Non-Small-Cell Lung Cancer. *Clin. Cancer Res.* **2019**, *25*, 6671–6682. [[CrossRef](#)]
92. Nagano, T.; Tachihara, M.; Nishimura, Y. Mechanism of Resistance to Epidermal Growth Factor Receptor-Tyrosine Kinase Inhibitors and a Potential Treatment Strategy. *Cells* **2018**, *7*, 212. [[CrossRef](#)] [[PubMed](#)]
93. Drilon, A.; Ou, S.H.I.; Cho, B.C.; Kim, D.W.; Lee, J.; Lin, J.J.; Zhu, V.W.; Ahn, M.J.; Camidge, D.R.; Nguyen, J.; et al. Repotrectinib (Tpx-0005) is a Next-Generation ros1/Trk/Alk Inhibitor That Potently Inhibits ROS1/TRK/ALK Solvent-Front Mutations. *Cancer Discov.* **2018**, *8*, 1227–1236. [[CrossRef](#)] [[PubMed](#)]
94. Drilon, A.; Hu, Z.I.; Lai, G.G.Y.; Tan, D.S.W. Targeting RET-driven cancers: Lessons from evolving preclinical and clinical landscapes. *Nat. Rev. Clin. Oncol.* **2017**, *15*, 151–167. [[CrossRef](#)] [[PubMed](#)]
95. Carlomagno, F.; Guida, T.; Anaganti, S.; Vecchio, G.; Fusco, A.; Ryan, A.J.; Billaud, M.; Santoro, M. Disease associated mutations at valine 804 in the RET receptor tyrosine kinase confer resistance to selective kinase inhibitors. *Oncogene* **2004**, *23*, 6056–6063. [[CrossRef](#)] [[PubMed](#)]
96. Dagogo-Jack, I.; Stevens, S.E.; Lin, J.J.; Nagy, R.; Ferris, L.; Shaw, A.T.; Gainor, J.F. Emergence of a RET V804M Gatekeeper Mutation During Treatment With Vandetanib in RET-Rearranged NSCLC. *J. Thorac. Oncol.* **2018**, *13*, e226–e227. [[CrossRef](#)]

97. Wirth, L.J.; Kohno, T.; Udagawa, H.; Matsumoto, S.; Ishii, G.; Ebata, K.; Tuch, B.B.; Zhu, E.Y.; Nguyen, M.; Smith, S.; et al. Emergence and Targeting of Acquired and Hereditary Resistance to Multikinase RET Inhibition in Patients With RET-Altered Cancer. *JCO Precis. Oncol.* **2019**, *3*, 1–7. [[CrossRef](#)]
98. Nakaoku, T.; Kohno, T.; Araki, M.; Niho, S.; Chauhan, R.; Knowles, P.P.; Tsuchihara, K.; Matsumoto, S.; Shimada, Y.; Mimaki, S.; et al. A secondary RET mutation in the activation loop conferring resistance to vandetanib. *Nat. Commun.* **2018**, *9*, 1–9. [[CrossRef](#)]
99. Cosci, B.; Vivaldi, A.; Romei, C.; Gemignani, F.; Landi, S.; Ciampi, R.; Tacito, A.; Molinaro, E.; Agate, L.; Bottici, V.; et al. In silico and in vitro analysis of rare germline allelic variants of RET oncogene associated with medullary thyroid cancer. *Endocr. Relat. Cancer* **2011**, *18*, 603–612. [[CrossRef](#)]
100. Plenker, D.; Riedel, M.; Brägelmann, J.; Dammert, M.A.; Chauhan, R.; Knowles, P.P.; Lorenz, C.; Keul, M.; Bührmann, M.; Pagel, O.; et al. Drugging the catalytically inactive state of RET kinase in RET-rearranged tumors. *Sci. Transl. Med.* **2017**, *9*, eaah6144. [[CrossRef](#)]
101. Crystal, A.S.; Shaw, A.T.; Sequist, L.V.; Friboulet, L.; Niederst, M.J.; Lockerman, E.L.; Frias, R.L.; Gainor, J.F.; Amzallag, A.; Greninger, P.; et al. Patient-derived models of acquired resistance can identify effective drug combinations for cancer. *Science* **2014**, *346*, 1480–1486. [[CrossRef](#)]
102. Nelson-Taylor, S.K.; Le, A.T.; Yoo, M.; Schubert, L.; Mishall, K.M.; Doak, A.; Varella-Garcia, M.; Tan, A.-C.; Doebele, R.C. Resistance to RET-Inhibition in RET-Rearranged NSCLC Is Mediated By Reactivation of RAS/MAPK Signaling. *Mol. Cancer Ther.* **2017**, *16*, 1623–1633. [[CrossRef](#)] [[PubMed](#)]
103. Vaishnavi, A.; Schubert, L.; Rix, U.; Marek, L.A.; Le, A.T.; Keysar, S.B.; Glogowska, M.J.; Smith, M.A.; Kako, S.; Sumi, N.J.; et al. EGFR Mediates Responses to Small-Molecule Drugs Targeting Oncogenic Fusion Kinases. *Cancer Res.* **2017**, *77*, 3551–3563. [[CrossRef](#)]
104. Chang, H.; Sung, J.H.; Moon, S.U.; Kim, H.-S.; Kim, J.W.; Lee, J.S. EGF Induced RET Inhibitor Resistance inCCDC6-RETLung Cancer Cells. *Yonsei Med. J.* **2017**, *58*, 9–18. [[CrossRef](#)]
105. Bhinge, K.; Yang, L.; Terra, S.; Nasir, A.; Muppa, P.; Aubry, M.C.; Yi, J.; Janaki, N.; Kovtun, I.V.; Murphy, S.J.; et al. EGFR mediates activation of RET in lung adenocarcinoma with neuroendocrine differentiation characterized by ASCL1 expression. *Oncotarget* **2017**, *8*, 27155–27165. [[CrossRef](#)]
106. Kang, C.W.; Jang, K.W.; Sohn, J.; Kim, S.-M.; Pyo, K.-H.; Kim, H.; Yun, M.R.; Na Kang, H.; Kim, H.R.; Lim, S.M.; et al. Antitumor Activity and Acquired Resistance Mechanism of Dovitinib (TKI258) in RET-Rearranged Lung Adenocarcinoma. *Mol. Cancer Ther.* **2015**, *14*, 2238–2248. [[CrossRef](#)] [[PubMed](#)]
107. Somwar, R.; Smith, R.; Hayashi, T.; Ishizawa, K.; Charen, A.S.; Khodos, I.; Mattar, M.; He, J.; Balasubramanian, S.; Stephens, P.; et al. MDM2 amplification (Amp) to mediate cabozantinib resistance in patients (Pts) with advanced RET-rearranged lung cancers. *J. Clin. Oncol.* **2016**, *34*, 9068. [[CrossRef](#)]
108. Solomon, B.J.; Tan, L.; Lin, J.J.; Wong, S.Q.; Hollizeck, S.; Ebata, K.; Tuch, B.B.; Yoda, S.; Gainor, J.F.; Sequist, L.V.; et al. RET Solvent Front Mutations Mediate Acquired Resistance to Selective RET Inhibition in RET-Driven Malignancies. *J. Thorac. Oncol.* **2020**, *15*, 541–549. [[CrossRef](#)]
109. Subbiah, V.; Shen, T.; Terzyan, S.; Liu, X.; Hu, X.; Patel, K.; Hu, M.; Cabanillas, M.; Behrang, A.; Meric-Bernstam, F.; et al. Structural basis of acquired resistance to selpercatinib and pralsetinib mediated by non-gatekeeper RET mutations. *Ann. Oncol.* **2021**, *32*, 261–268. [[CrossRef](#)]
110. Lin, J.J.; Liu, S.V.; McCoach, C.E.; Zhu, V.W.; Tan, A.C.; Yoda, S.; Peterson, J.; Do, A.; Prutisto-Chang, K.; Dagogo-Jack, I.; et al. Mechanisms of resistance to selective RET tyrosine kinase inhibitors in RET fusion-positive non-small-cell lung cancer. *Ann. Oncol.* **2020**, *31*, 1725–1733. [[CrossRef](#)]
111. Zhu, V.W.; Madison, R.; Schrock, A.B.; Ou, S.-H.I. Emergence of High Level of MET Amplification as Off-Target Resistance to Selpercatinib Treatment in KIF5B-RET NSCLC. *J. Thorac. Oncol.* **2020**, *15*, e124–e127. [[CrossRef](#)]
112. Liu, X.; Shen, T.; Mooers, B.H.M.; Hilberg, F.; Wu, J. Drug resistance profiles of mutations in the RET kinase domain. *Br. J. Pharmacol.* **2018**, *175*, 3504–3515. [[CrossRef](#)]
113. Carlomagno, F.; Guida, T.; Anaganti, S.; Provitera, L.; Kjaer, S.; McDonald, N.Q.; Ryan, A.J.; Santoro, M. Identification of tyrosine 806 as a molecular determinant of RET kinase sensitivity to ZD6474. *Endocr. Relat. Cancer* **2009**, *16*, 233–241. [[CrossRef](#)] [[PubMed](#)]
114. Mulligan, L.M. GDNF and the RET Receptor in Cancer: New Insights and Therapeutic Potential. *Front. Physiol.* **2019**, *9*, 1873. [[CrossRef](#)]
115. Nakasatomi, M.; Takahashi, S.; Sakairi, T.; Ikeuchi, H.; Kaneko, Y.; Hiromura, K.; Nojima, Y.; Maeshima, A. Enhancement of HGF-induced tubulogenesis by endothelial cell-derived GDNF. *PLoS ONE* **2019**, *14*, e0212991. [[CrossRef](#)] [[PubMed](#)]
116. Garnis, C.; Campbell, J.; Davies, J.J.; Macaulay, C.; Lam, S.; Lam, W.L. Involvement of multiple developmental genes on chromosome 1p in lung tumorigenesis. *Hum. Mol. Genet.* **2004**, *14*, 475–482. [[CrossRef](#)]
117. Joo, L.J.S.; Weiss, J.; Gill, A.J.; Clifton-Bligh, R.; Brahmhatt, H.; MacDiarmid, J.A.; Gild, M.L.; Robinson, B.G.; Zhao, J.T.; Sidhu, S.B. RET Kinase-Regulated MicroRNA-153-3p Improves Therapeutic Efficacy in Medullary Thyroid Carcinoma. *Thyroid.* **2019**, *29*, 830–844. [[CrossRef](#)]
118. Zhang, W.; Dong, Y.-Z.; Du, X.; Peng, X.-N.; Shen, Q.-M. MiRNA-153-3p promotes gefitinib-sensitivity in non-small cell lung cancer by inhibiting ATG5 expression and autophagy. *Eur. Rev. Med. Pharmacol. Sci.* **2019**, *23*, 2444–2452.

119. Chen, W.-J.; Zhang, E.-N.; Zhong, Z.-K.; Jiang, M.-Z.; Yang, X.-F.; Zhou, D.-M.; Wang, X.-W. MicroRNA-153 Expression and Prognosis in Non-Small Cell Lung Cancer. *Int. J. Clin. Exp. Pathol.* **2015**, *8*, 8671–8675.
120. Zhang, M.; Liu, L.; Lin, X. Article A Translocation Pathway for Vesicle-Mediated Article A Translocation Pathway for Vesicle-Mediated Unconventional Protein Secretion. *Cell* **2020**, 1–16. [[CrossRef](#)]
121. Xiao, G.-Y.; Schmid, S. Abstract 118: FCHSD2 controls oncogenic ERK1/2 signaling by regulating endocytic trafficking in non-small-cell lung cancer. *Immunology* **2020**, *80*, 118. [[CrossRef](#)]
122. Batrouni, A.G.; Baskin, J.M. A MAP for PI3K Activation on Endosomes. *Nat. Cell Biol.* **2020**, *22*, 1292–1294. [[CrossRef](#)]
123. Crupi, M.J.F.; Maritan, S.M.; Reyes-Alvarez, E.; Lian, E.Y.; Hyndman, B.D.; Rekab, A.N.; Moodley, S.; Antonescu, C.N.; Mulligan, L.M. GGA3-mediated recycling of the RET receptor tyrosine kinase contributes to cell migration and invasion. *Oncogene* **2020**, *39*, 1361–1377. [[CrossRef](#)] [[PubMed](#)]
124. Cascone, T.; Subbiah, V.; Hess, K.R.; Nelson, S.; Nilsson, M.B.; Subbiah, I.M.; Ali, S.M.; Carbone, D.P.; Salgia, R.; Owonikoko, T.K.; et al. Significant systemic and CNS activity of RET inhibitor vandetanib combined with mTOR inhibitor everolimus in patients with advanced NSCLC with RET fusion. *J. Clin. Oncol.* **2016**, *34*, 9069. [[CrossRef](#)]
125. Subbiah, V.; Berry, J.; Roxas, M.; Guha-Thakurta, N.; Subbiah, I.M.; Ali, S.M.; McMahon, C.; Miller, V.; Cascone, T.; Pai, S.; et al. Systemic and CNS activity of the RET inhibitor vandetanib combined with the mTOR inhibitor everolimus in KIF5B-RET re-arranged non-small cell lung cancer with brain metastases. *Lung Cancer* **2015**, *89*, 76–79. [[CrossRef](#)] [[PubMed](#)]
126. Rosen, E.Y.; Johnson, M.L.; Clifford, S.E.; Somwar, R.; Kherani, J.F.; Son, J.; Bertram, A.A.; Davare, M.A.; Gladstone, E.G.; Ivanova, E.V.; et al. Overcoming MET-Dependent Resistance to Selective RET Inhibition in Patients with RET Fusion-Positive Lung Cancer by Combining Selpercatinib with Crizotinib. *Clin. Cancer Res.* **2021**, *27*, 34–42. [[CrossRef](#)] [[PubMed](#)]
127. Koga, K.; Hattori, Y.; Komori, M.; Narishima, R.; Yamasaki, M.; Hakoshima, M.; Fukui, T.; Maitani, Y. Combination of RET siRNA and irinotecan inhibited the growth of medullary thyroid carcinoma TT cells and xenografts via apoptosis. *Cancer Sci.* **2010**, *101*, 941–947. [[CrossRef](#)]
128. Bertazza, L.; Barollo, S.; Radu, C.M.; Cavedon, E.; Simioni, P.; Faggian, D.; Plebani, M.; Pelizzo, M.R.; Rubin, B.; Boscaro, M.; et al. Synergistic antitumor activity of RAF265 and ZSTK474 on human TT medullary thyroid cancer cells. *J. Cell. Mol. Med.* **2015**, *19*, 2244–2252. [[CrossRef](#)] [[PubMed](#)]
129. Jin, N.; Jiang, T.; Rosen, D.M.; Nelkin, B.D.; Ball, D.W. Synergistic Action of a RAF Inhibitor and a Dual PI3K/mTOR Inhibitor in Thyroid Cancer. *Clin. Cancer Res.* **2011**, *17*, 6482–6489. [[CrossRef](#)]
130. Caccia, D.; Miccichè, F.; Cassinelli, G.; Mondellini, P.; Casalini, P.; Bongarzone, I. Dasatinib reduces FAK phosphorylation increasing the effects of RPI-1 inhibition in a RET/PTC1-expressing cell line. *Mol. Cancer* **2010**, *9*, 278. [[CrossRef](#)] [[PubMed](#)]
131. Koh, Y.W.; Shah, M.H.; Agarwal, K.; Mccarty, S.K.; Koo, B.S.; Brendel, V.J.; Wang, C.; Porter, K.; Jarjoura, D.; Saji, M.; et al. Sorafenib and Mek inhibition is synergistic in medullary thyroid carcinoma in vitro. *Endocr. Relat. Cancer* **2012**, *19*, 29–38. [[CrossRef](#)] [[PubMed](#)]
132. Hong, D.S.; Sebti, S.M.; Newman, R.A.; Blaskovich, M.A.; Ye, L.; Gagel, R.F.; Moulder, S.; Wheler, J.J.; Naing, A.; Tannir, N.M.; et al. Phase I Trial of a Combination of the Multikinase Inhibitor Sorafenib and the Farnesyltransferase Inhibitor Tipifarnib in Advanced Malignancies. *Clin. Cancer Res.* **2009**, *15*, 7061–7068. [[CrossRef](#)]
133. Wang, M.; Naganna, N.; Sintim, H.O. Identification of nicotinamide aminonaphthyridine compounds as potent RET kinase inhibitors and antitumor activities against RET rearranged lung adenocarcinoma. *Bioorg. Chem.* **2019**, *90*, 103052. [[CrossRef](#)]
134. Nguyen, M.; Miyakawa, S.; Kato, J.; Mori, T.; Arai, T.; Armanini, M.; A Gelmon, K.; Yerushalmi, R.; Leung, S.; Gao, D.; et al. Preclinical Efficacy and Safety Assessment of an Antibody–Drug Conjugate Targeting the c-RET Proto-Oncogene for Breast Carcinoma. *Clin. Cancer Res.* **2015**, *21*, 5552–5562. [[CrossRef](#)] [[PubMed](#)]
135. Bosco, E.E.; Christie, R.J.; Carrasco, R.; Sabol, D.; Zha, J.; Dacosta, K.; Brown, L.; Kennedy, M.; Meekin, J.; Phipps, S.; et al. Preclinical evaluation of a GFRA1 targeted antibody-drug conjugate in breast cancer. *Oncotarget* **2018**, *9*, 22960–22975. [[CrossRef](#)]
136. Bagheri-Yarmand, R.; Williams, M.D.; Grubbs, E.G.; Gagel, R.F. ATF4 Targets RET for Degradation and is a Candidate Tumor Suppressor Gene in Medullary Thyroid Cancer. *J. Clin. Endocrinol. Metab.* **2016**, *102*, 933–941. [[CrossRef](#)]
137. Luo, J.; Xia, Y.; Yin, Y.; Luo, J.; Liu, M.; Zhang, H.; Zhang, C.; Zhao, Y.; Yang, L.; Kong, L. ATF4 destabilizes RET through nonclassical GRP78 inhibition to enhance chemosensitivity to bortezomib in human osteosarcoma. *Theranostics* **2019**, *9*, 6334–6353. [[CrossRef](#)]
138. Drilon, A.; Rogers, E.; Zhai, D.; Deng, W.; Zhang, X.; Lee, D.; Ung, J.; Whitten, J.; Zhuang, H.; Liu, J.; et al. TPX-0046 is a novel and potent RET/SRC inhibitor for RET-driven cancers. *Ann. Oncol.* **2019**, *30*, v190–v191. [[CrossRef](#)]
139. Schoffski, P.; Aftimos, P.G.; Massard, C.; Italiano, A.; Jungels, C.; Andreas, K.; Keegan, M.; Ho, P.T.C. A phase I study of BOS172738 in patients with advanced solid tumors with RET gene alterations including non-small cell lung cancer and medullary thyroid cancer. *J. Clin. Oncol.* **2019**, *37*, TPS3162. [[CrossRef](#)]
140. Frett, B.; Carlomagno, F.; Moccia, M.L.; Brescia, A.; Federico, G.; De Falco, V.; Admire, B.; Chen, Z.; Qi, W.; Santoro, M.; et al. Fragment-Based Discovery of a Dual pan-RET/VEGFR2 Kinase Inhibitor Optimized for Single-Agent Polypharmacology. *Angew. Chem. Int. Ed.* **2015**, *54*, 8717–8721. [[CrossRef](#)] [[PubMed](#)]
141. Moccia, M.; Frett, B.; Zhang, L.; Lakkaniga, N.R.; Briggs, D.C.; Chauhan, R.; Brescia, A.; Federico, G.; Yan, W.; Santoro, M.; et al. Bioisosteric Discovery of NPA101.3, a Second-Generation RET/VEGFR2 Inhibitor Optimized for Single-Agent Polypharmacology. *J. Med. Chem.* **2020**, *63*, 4506–4516. [[CrossRef](#)] [[PubMed](#)]

142. Larocque, E.; Naganna, N.; Ma, X.; Opoku-Temeng, C.; Carter-Cooper, B.; Chopra, G.; Lapidus, R.G.; Sintim, H.O. Aminoisoquinoline benzamides, FLT3 and Src-family kinase inhibitors, potently inhibit proliferation of acute myeloid leukemia cell lines. *Future Med. Chem.* **2017**, *9*, 1213–1225. [[CrossRef](#)] [[PubMed](#)]
143. Yang, J.; Chen, K.; Zhang, G.; Yang, Q.-Y.; Li, Y.-S.; Huang, S.-Z.; Wang, Y.-L.; Yang, W.; Jiang, X.-J.; Yan, H.-X.; et al. Structural optimization and structure-activity relationship studies of N-phenyl-7,8-dihydro-6H-pyrimido[5,4-b][1,4]oxazin-4-amine derivatives as a new class of inhibitors of RET and its drug resistance mutants. *Eur. J. Med. Chem.* **2018**, *143*, 1148–1164. [[CrossRef](#)] [[PubMed](#)]
144. Lakkaniga, N.R.; Gunaganti, N.; Zhang, L.; Belachew, B.; Frett, B.; Leung, Y.-K.; Li, H. Pyrrolo[2,3-d]pyrimidine derivatives as inhibitors of RET: Design, synthesis and biological evaluation. *Eur. J. Med. Chem.* **2020**, *206*, 112691. [[CrossRef](#)] [[PubMed](#)]
145. Jia, C.-C.; Chen, W.; Feng, Z.-L.; Liu, Z.-P. Recent developments of RET protein kinase inhibitors with diverse scaffolds as hinge binders. *Future Med. Chem.* **2021**, *13*, 45–62. [[CrossRef](#)] [[PubMed](#)]