

Figure S1

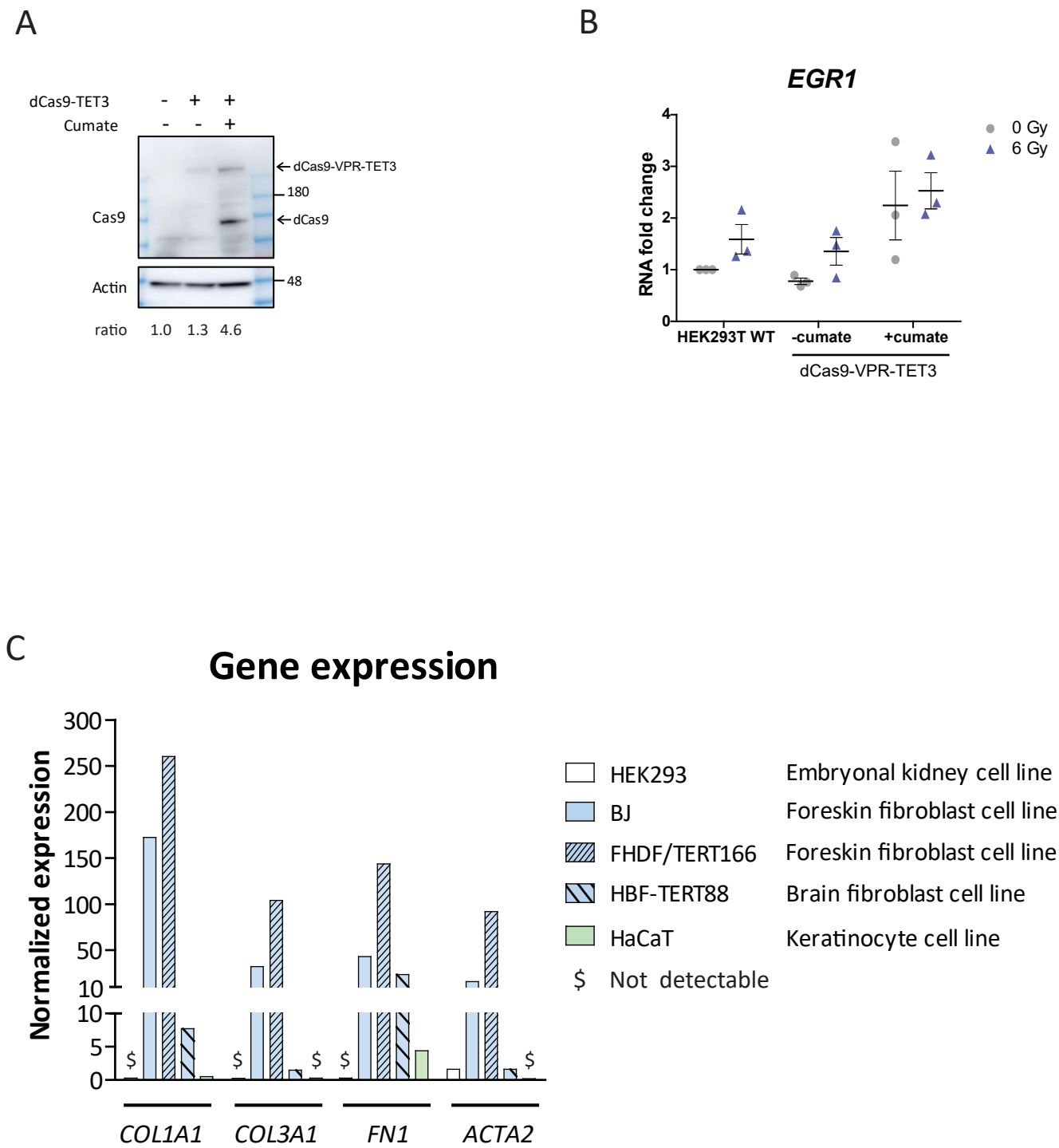
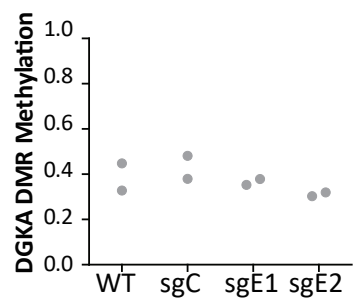


Figure S1. dCas9-VPR-TET3 expression and endogenous relative mRNA expression of indicated genes in human cell lines

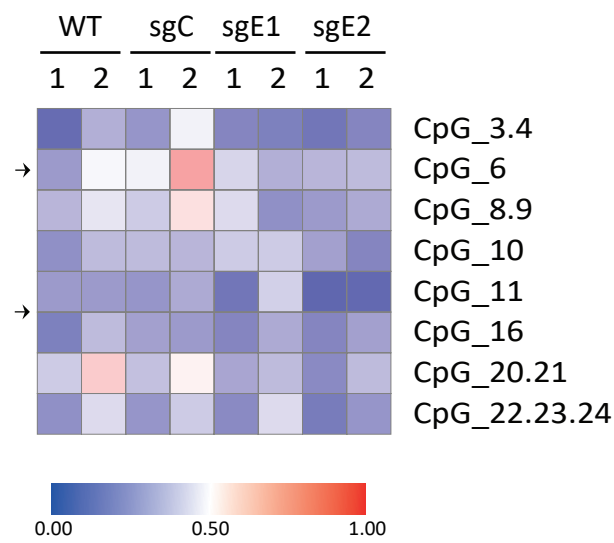
(A) Protein expression of Cas9 in dCas9-VPR-TET3-transfected HEK293T cells. Cells were treated with or without cumate (300 $\mu\text{g/ml}$) for dCas9-VPR-TET3 induction over 2 passages before harvesting. (B) *EGR1* expression in HEK293T and dCas9-VPR-TET3-transfected cells treated without or with cumate (300 $\mu\text{g/ml}$). Cells were harvested 48 hours after irradiation with 6 Gy. Data include three biological replicates and are presented as mean \pm SEM. Statistical significance was determined by one-tailed Student's t test. (C) Endogenous relative mRNA expression of indicated genes in human cell lines. Data are derived from a consensus dataset in the Human Protein Atlas (HPA), which combines data from three comprehensive databases (HPA, GTEx, and FANTOM5 [52]).

Figure S2

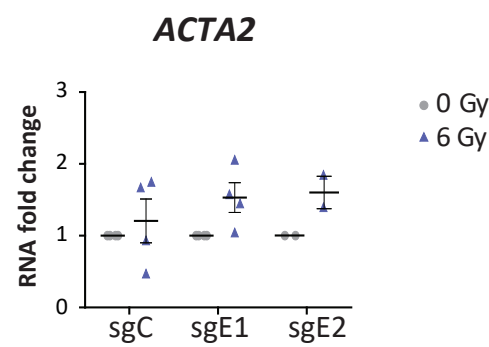
A



B



C



D

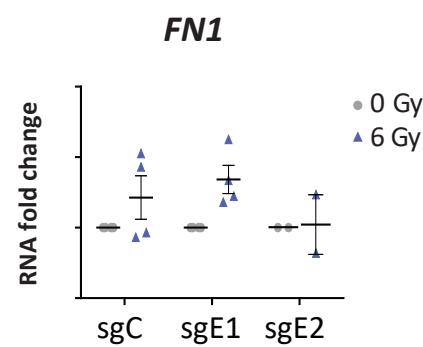


Figure S2. BJ cells with CRISPR/Cas9-edited EGR1-binding sites at the *DGKA* enhancer

(A) Methylation average across various CpGs located in the *DGKA* enhancer in parental (WT) and CRISPR/Cas9-edited (sgC, sgE1 and sgE2) BJ cells. Methylation was measured as β -values by EpiTYPER technology. (B) Heatmap of DNA methylation of all CpG sites measured in parental (WT) and CRISPR/Cas9-edited (sgC, sgE1 and sgE2) BJ cells. For each cell type, two replicates (1,2) are shown. Arrows indicate the location of the EGR1-binding motifs. The first EGR1-binding site (EGR1_1) covers CpG_6, and the second binding site (EGR1_2) covers CpG_14, which is located between CpG_11 and CpG_16 and not detected by EpiTYPER. For (A and B), data are depicted from duplicate experiments in each cell line. (C-D) Relative mRNA expression of *ACTA2* (C) and *FN1* (D) in CRISPR/Cas9-edited (sgC, sgE1 and sgE2) BJ cells. Cells were harvested 48 hours after irradiation with 6 Gy. For (C and D), data include at least two biological replicates and are presented as mean \pm SEM. Statistical significance was determined by one-tailed Student's t test.

Figure S3

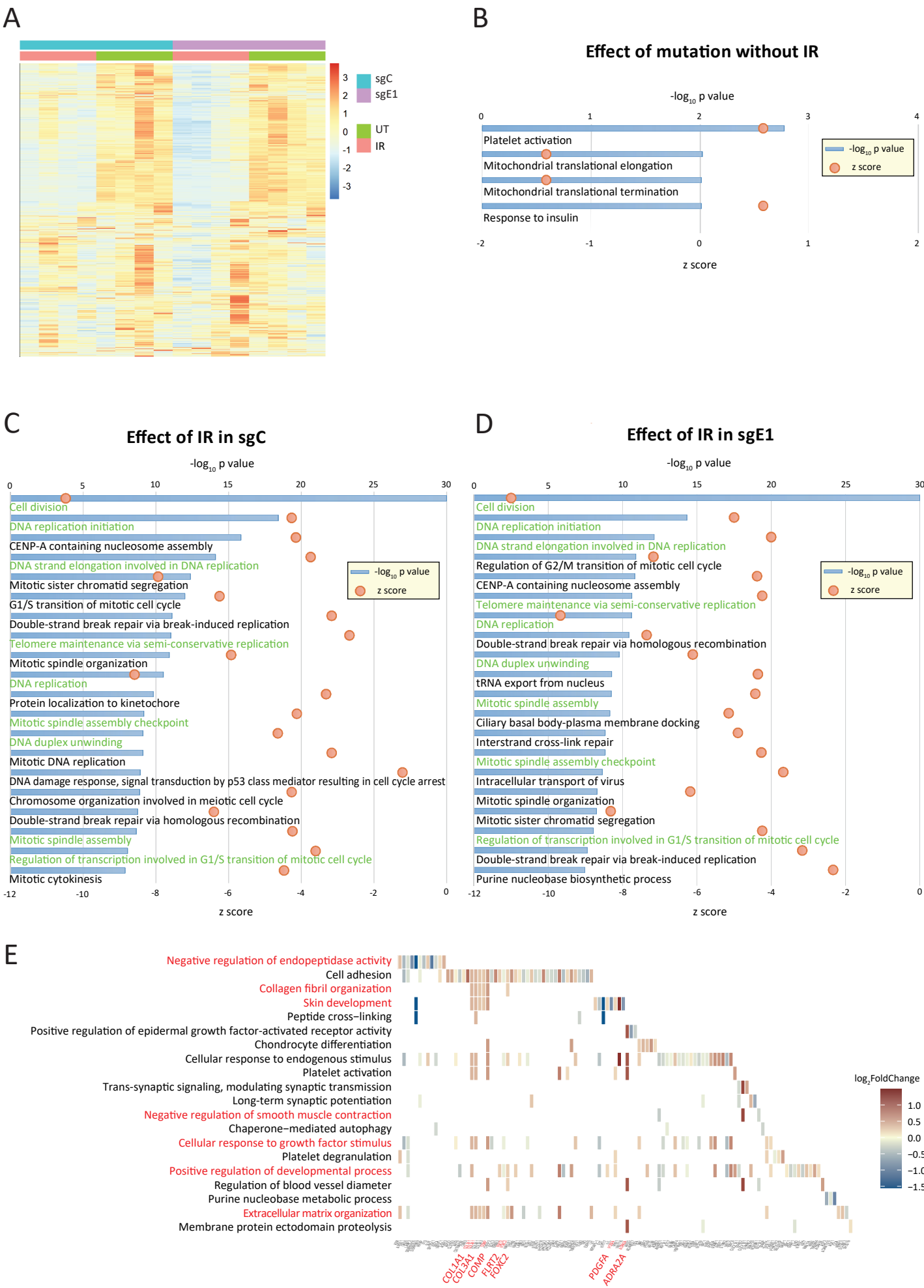


Figure S3. Genome-wide effects induced by loss of EGR1-binding at the *DGKA* enhancer

(A) Hierarchical clustering heatmap showing all differentially expressed genes (DEGs) from all comparisons (sgC versus sgE1 cells with or without 6 Gy irradiation). (B) Gene ontology (GO) analysis of the DEGs obtained from the comparison of sgC and sgE1 cells without irradiation. (C and D) GO analysis of DEGs identified after irradiation (6 Gy) of sgC (C) and sgE1 (D) cells. Significance ($-\log_{10}(\text{p value})$) of GO terms are represented by blue bars, and z scores are shown with red dots. The common biological functions and pathways existing in both sgC and sgE1 cells are highlighted in green. (E) A cluster heatmap of DEGs contributing to the top 20 GO pathways from the comparison of sgC and sgE1 cells after irradiation and adjustment for irradiation. Biological functions and pathways potentially related to fibroblast activation and fibrosis development are highlighted in red in (B) and (E).

Figure S4

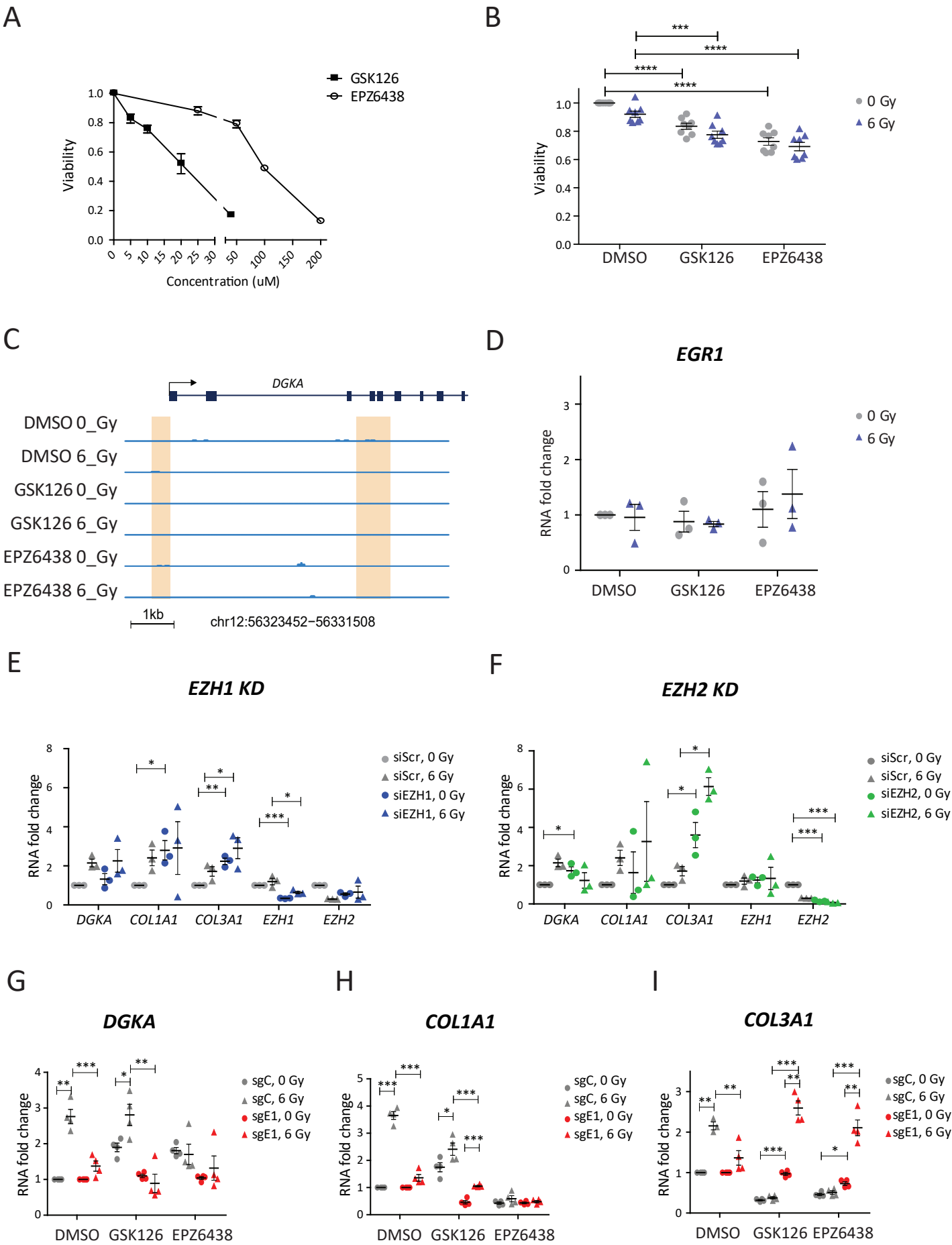


Figure S4. Effects of the inhibition of EZH1 and/or EZH2 in BJ cells

(A) Cell viability of BJ cells treated with EZH2 inhibitors. Cells were treated with DMSO, GSK126 (5 μ M) or EPZ6438 (50 μ M) at the indicated doses for 72 hours and cell viability was measured by CellTiter Blue. (B) Cell viability after EZH2 inhibitor treatment and irradiation (6 Gy) in BJ cells. Cells were pre-treated with DMSO, GSK126 (5 μ M) or EPZ6438 (50 μ M) for 48 hours, irradiated with 6 Gy, and incubated for another 48 hours with concurrent inhibitor treatment before the CellTiter Blue test. (C) Genome browser tracks comparing ACT-seq signals for H3K27me3 enrichment at the *DGKA* locus after inhibitor treatments. Promoter and enhancer regions are highlighted in orange. Data present one representative replicate from Figure 4B. (D) Relative mRNA expression of *EGR1* in BJ cells treated with EZH2 inhibitors. For (C-D), cells were pre-treated with DMSO, GSK126 (5 μ M) or EPZ6438 (50 μ M) for 48 hours, irradiated with 6 Gy, and analysed after an additional 48 hours with concurrent drug treatment. (E and F) Relative mRNA expression levels of indicated genes in BJ cells after silencing *EZH1* (E) or *EZH2* (F), respectively. Cells were pre-treated with siRNAs for 48 hours, irradiated (6 Gy), and harvested 48 hours later. (G-I) Relative mRNA expression levels of *DGKA* (G), *COL1A1* (H) and *COL3A1* (I) in CRISPR/Cas9-edited (sgC and sgE1) BJ cells. Cells were pre-treated with DMSO, GSK126 (5 μ M) or EPZ6438 (50 μ M) for 48 hours, irradiated with 6 Gy, and incubated for another 48 hours with concurrent drug treatment before harvest. Data include six (A), eight (B), three (D-F) or four (G-I) biological replicates and are presented as mean \pm SEM. Statistical significance (* $p<0.05$ ** $p<0.01$ and *** $p<0.001$) was determined by one-tailed Student's t test.

Figure S5

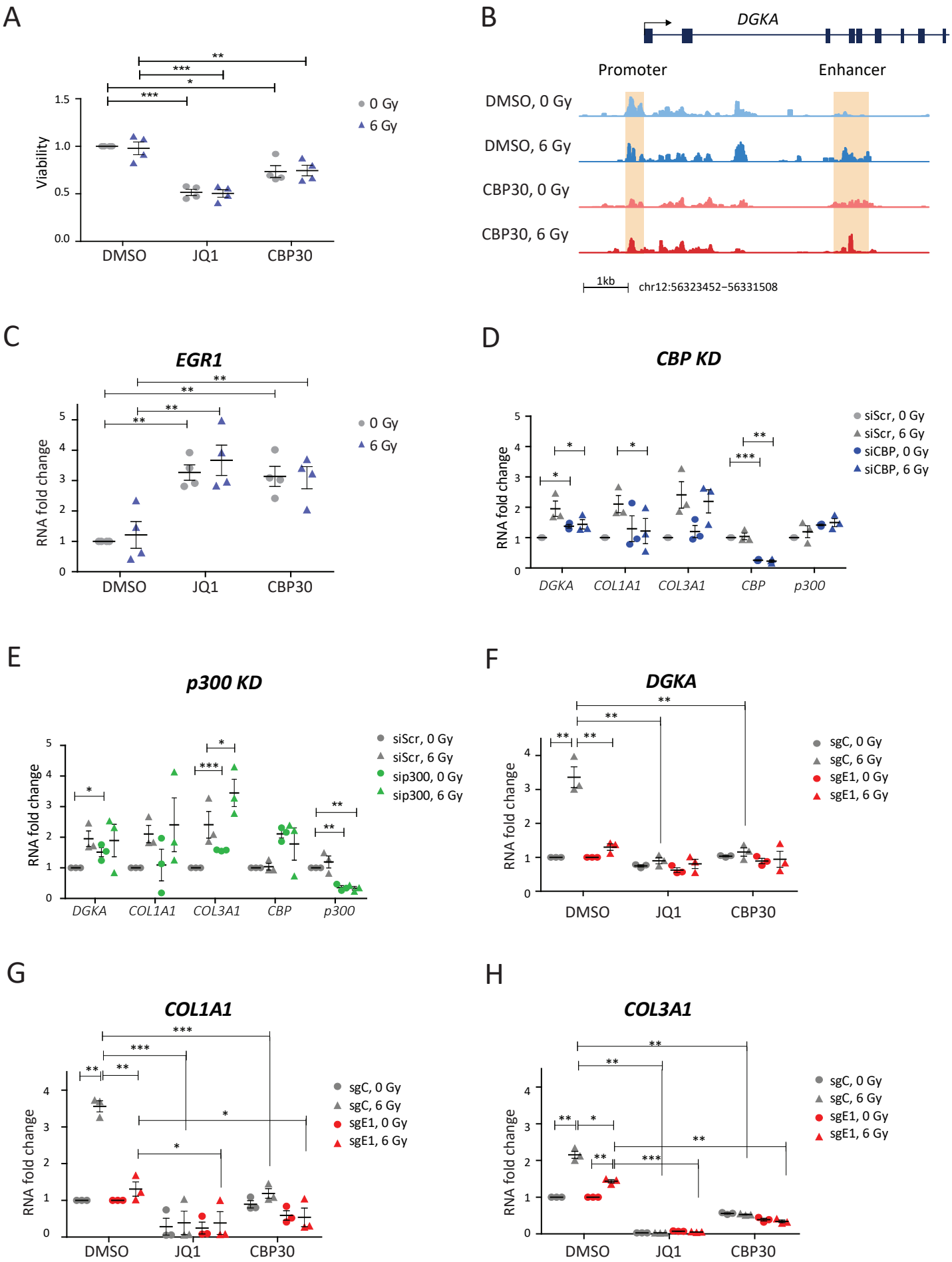


Figure S5. Effects of inhibition of bromodomain proteins in BJ cells

(A) Cell viability after bromodomain inhibitor treatment and irradiation. (B) Genome browser tracks comparing ACT-seq signals for H3K27ac at the *DGKA* locus after CBP30 treatment. Promoter and enhancer regions are highlighted in orange. Data present one representative replicate from Figure 5B. (C) Relative mRNA expression of *EGR1* in BJ cells treated with bromodomain inhibitors. For (A-C), cells were pre-treated with DMSO, CBP (10 μ M) or JQ1 (5 μ M) for 48 hours, irradiated with 6 Gy, and incubated for another 48 hours with concurrent inhibitors before harvest. (D and E) Relative mRNA expression levels of indicated genes in BJ cells after silencing CBP (D) or p300 (E), respectively. Cells were pre-treated with siRNAs for 48 hours, irradiated with 6 Gy and harvested 48 hours later. (F-H) Relative mRNA expression levels of *DGKA* (F), *COL1A1* (G) and *COL3A1* (H) in CRISPR/Cas9-edited (sgC and sgE1) BJ cells. Cells were pre-treated with DMSO, CBP (10 μ M) or JQ1 (5 μ M) for 48 hours, irradiated with 6Gy, and incubated for another 48 hours with concurrent drug treatment before harvest. Statistical data are presented as mean \pm SEM from four (A and C) or three (D-H) biological replicates. For (B), data are presented one replicate from Figure 5B. Statistical significance (* $p<0.05$ ** $p<0.01$ and *** $p<0.001$) was determined by one-tailed Student's t test.

Figure S6

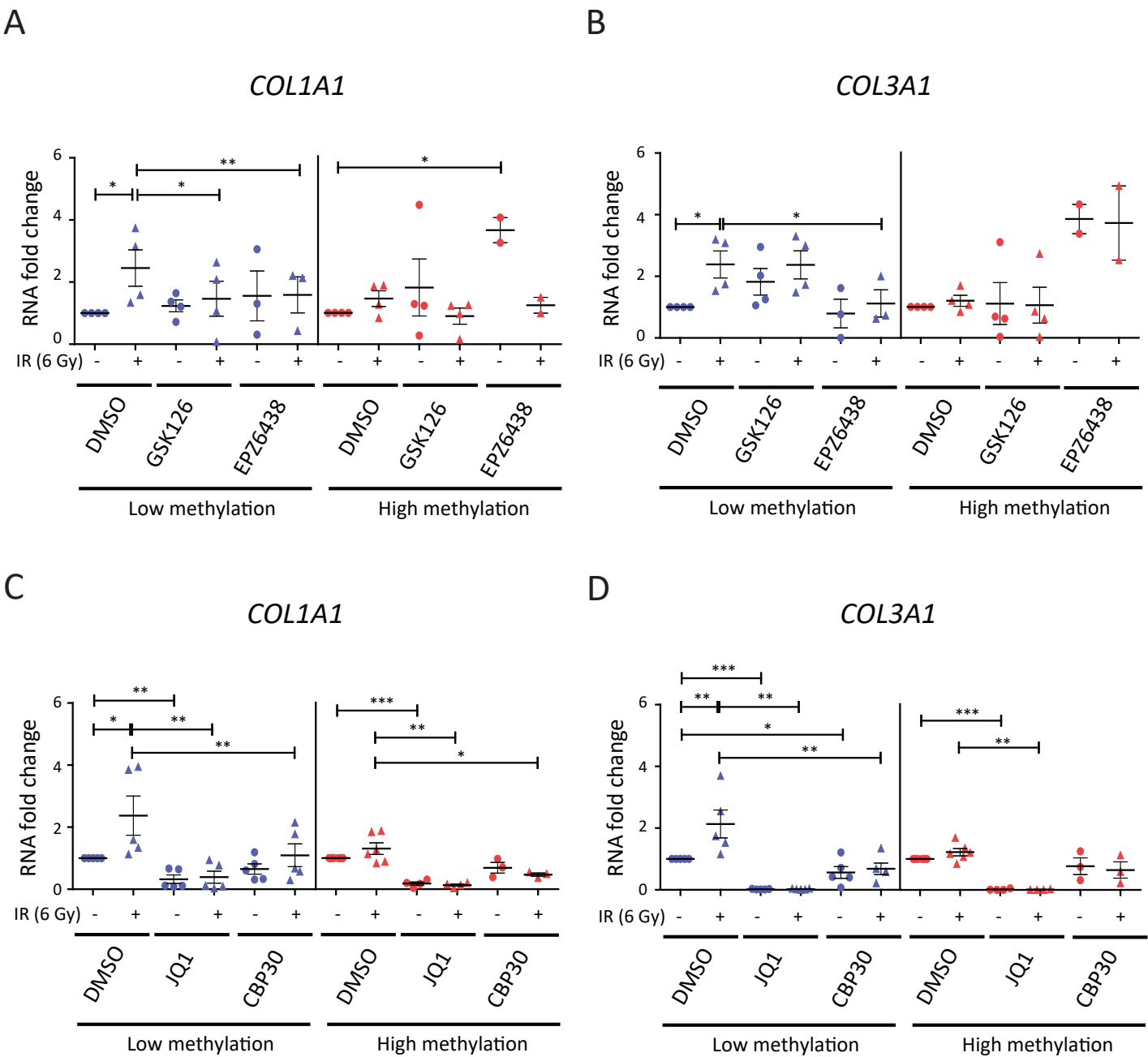


Figure S6. Effects of epigenetic inhibitors in human dermal fibroblasts with low or high DNA methylation at the *DGKA* enhancer

(A-D) Relative mRNA expression of *COL1A1* (A and C) and *COL3A1* (B and D) in NHDFs. Cells were pre-treated with EZH2 inhibitors (5 μ M GSK126 or 50 μ M GSK126) (C and D) or bromodomain inhibitors (10 μ M CBP30 or 5 μ M JQ1) (E and F), irradiated with 6 Gy, and incubated for another 48 hours with concurrent drug treatment before harvest. Data points summarize effects for both fibroblast groups and include at least duplicate experiments. Data are presented as mean \pm SEM, and statistical significance (* $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$) was determined by multiple t-test.

Figure S7

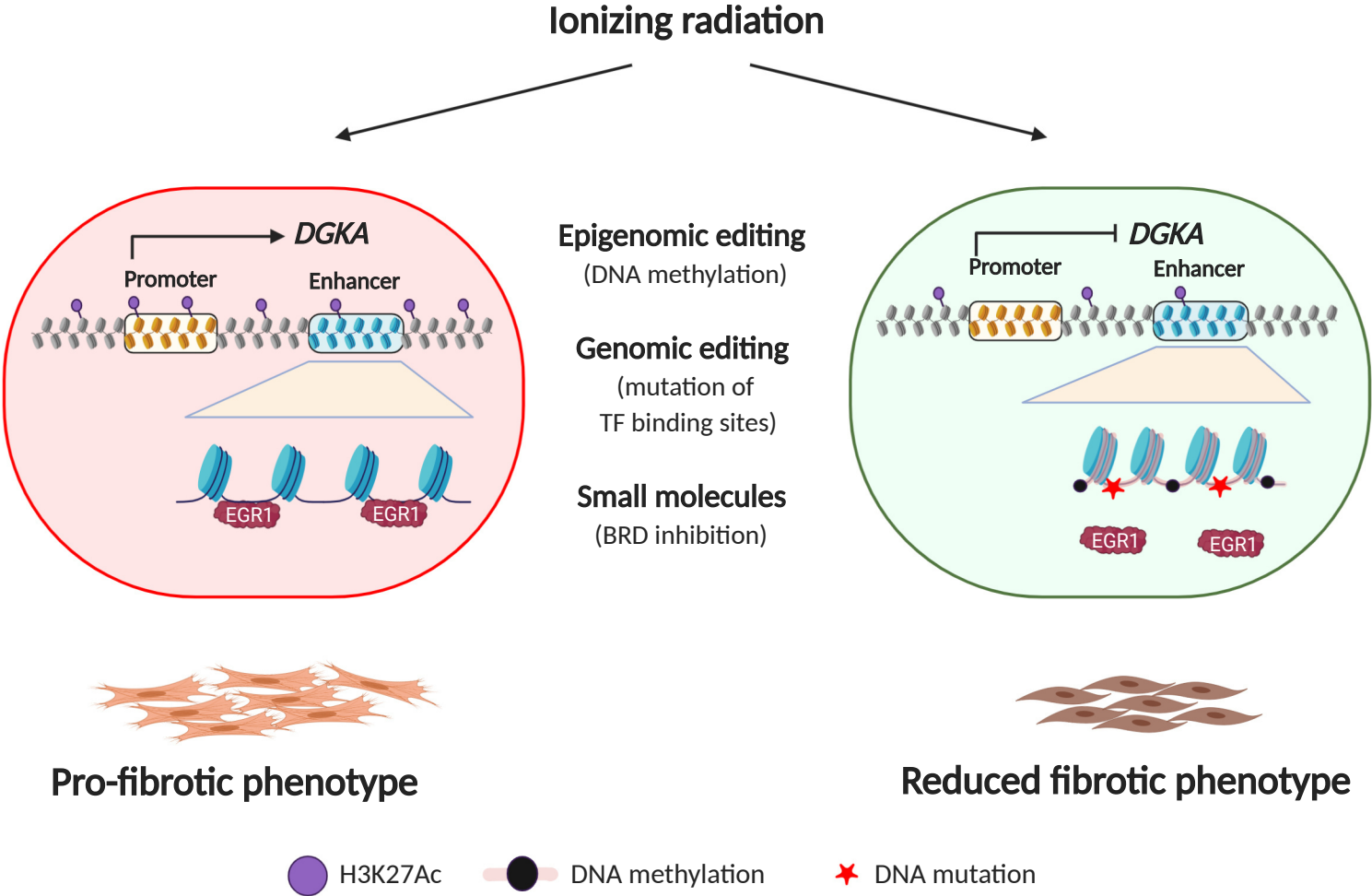


Figure S7. Schematic diagram of epigenetic modulation of *DGKA* expression

After ionizing radiation, unmethylated *DGKA* enhancer region enables the binding of the radiation-inducible transcription factor EGR1, thus upregulate *DGKA* pro-fibrotic marker expression. In this study, we were able to modulate radiation-induced *DGKA* expression by epigenomic (CRISPR/dCas9-VPR-TET3) and genomic (CRISPR-Cas9) editing of the *DGKA* enhancer and administering epigenetic drugs (JQ1, CBP30).

Figure S8. Original EMSA images.

Figure 1B

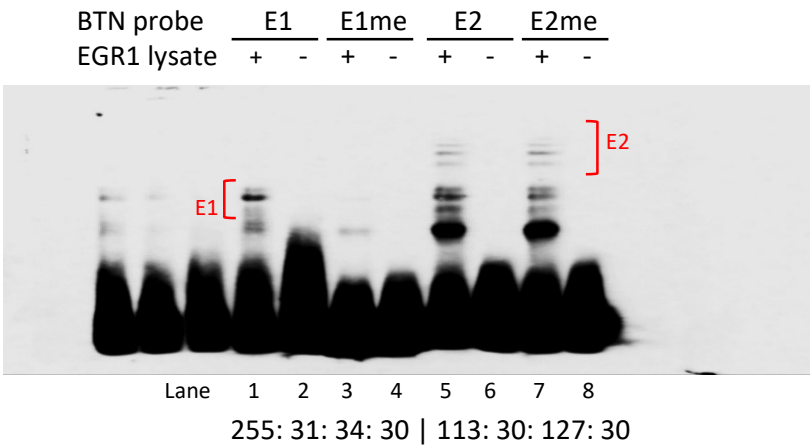


Figure 2B

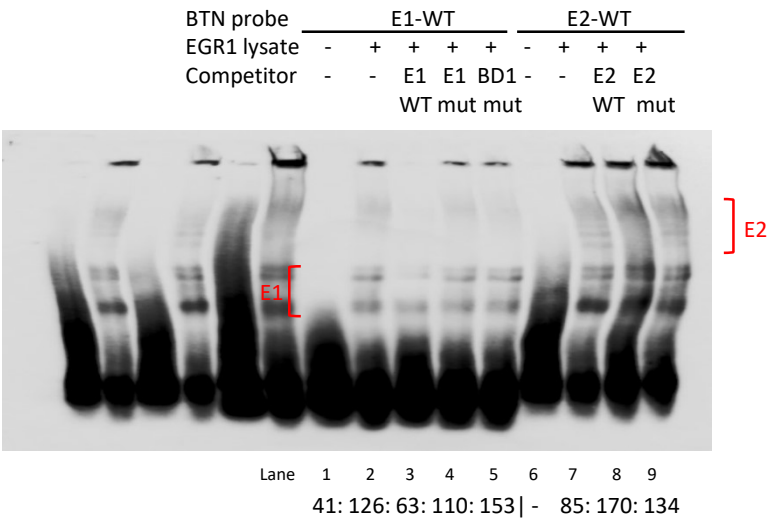


Figure S9. Original Western blot images for H3K27me3 and corresponding H3.

Figure 4A

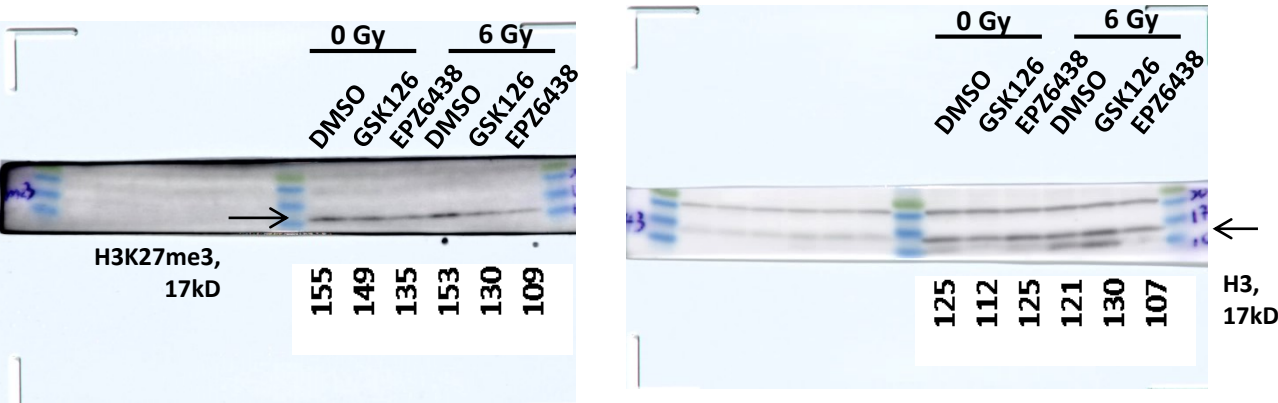


Figure 4A, image for statistic

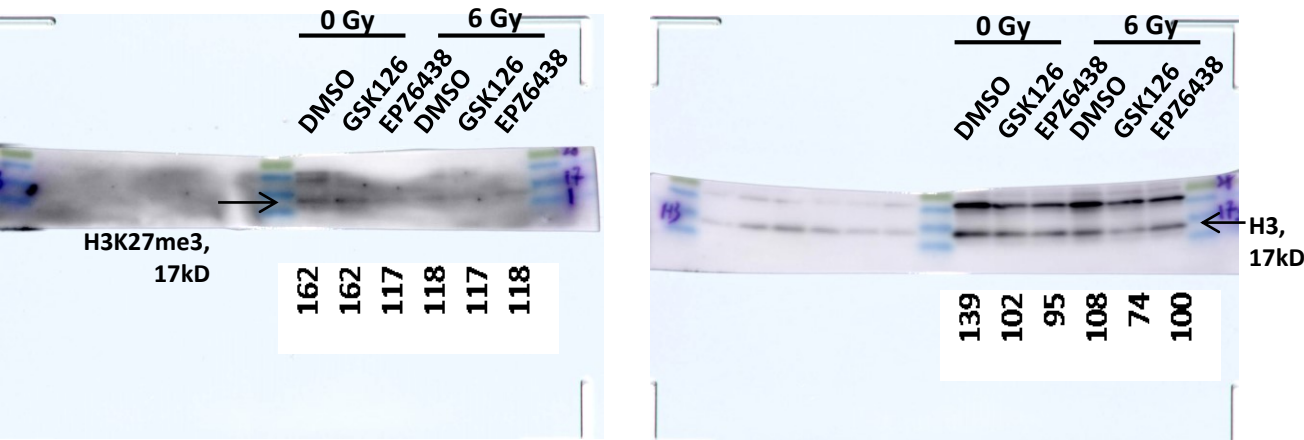


Figure S10. Original Western blot images for H3K27ac and corresponding H3.

Figure 5A

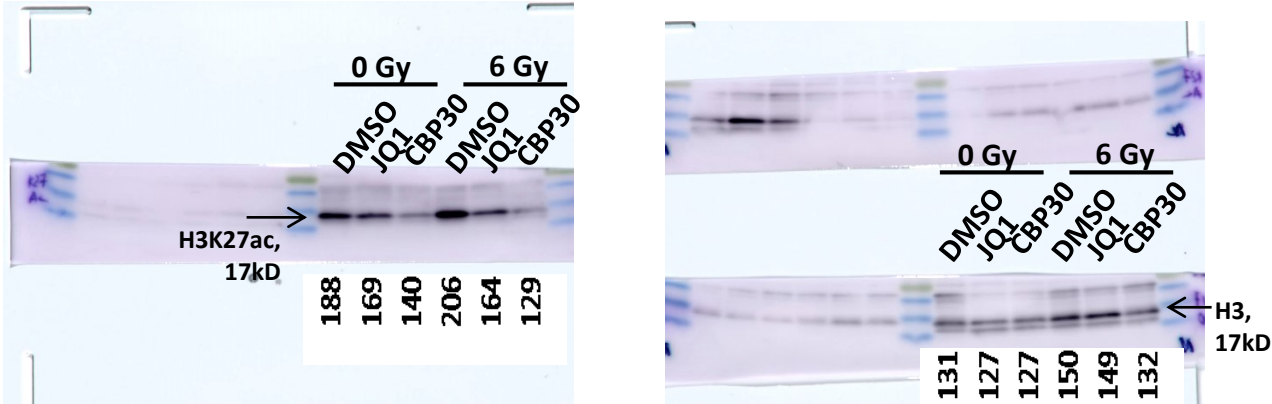


Figure 5A, image for statistic

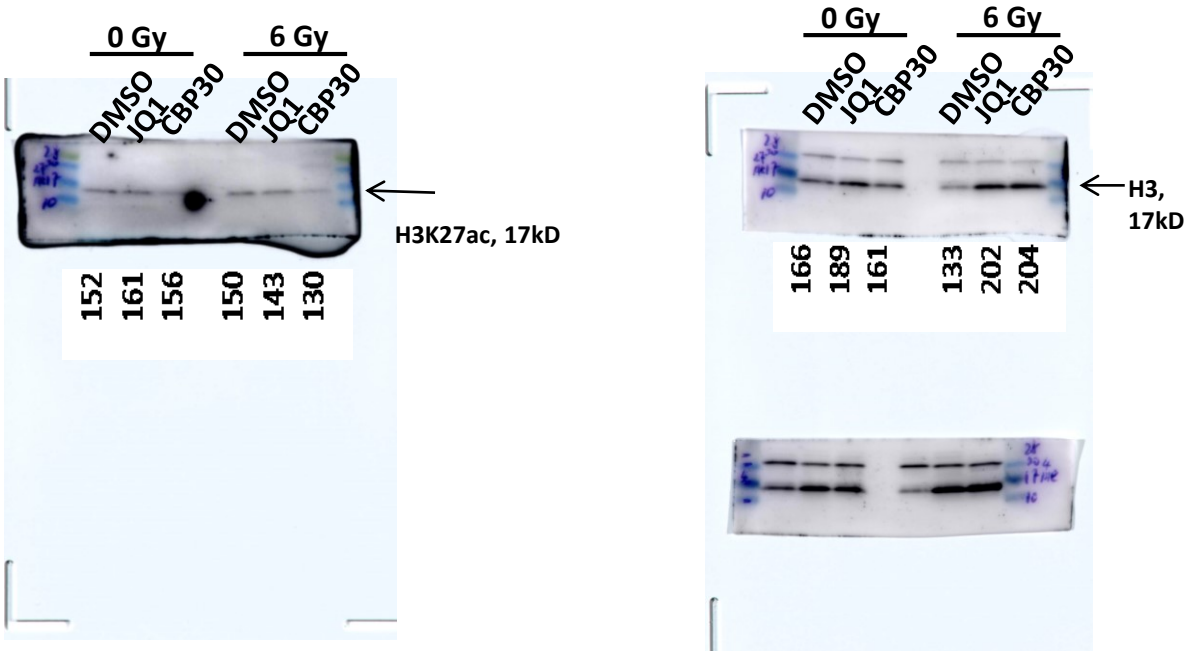


Figure 5A, image for statistic

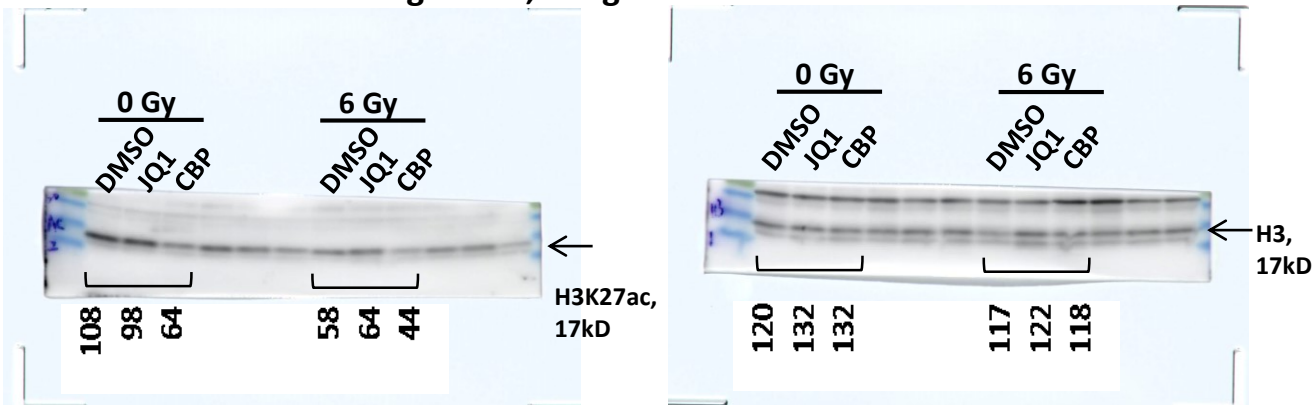


Figure S11. Original Western blot images for Figure S1A.

