Supplementary Information

Breast-specific epigenetic regulation of deltaNp73 and its role in DNAdamage-response of *BRCA1*-mutated human mammary epithelial cells

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1.1 Figure S1

Adapted from:

Killick R, Niklison-Chirou M, Tomasini R, Bano D, Rufini A, Grespi F, Velletri T, Tucci P, Sayan BS, Conforti F, Gallagher E, Nicotera P, Mak TW, Melino G, Knight RA, Agostini M. p73: a multifunctional protein in neurobiology.
Mol Neurobiol. 2011 Apr;43(2):139-46. doi: 10.1007/s12035-011-8172-6.



Fig. S1 Schematic structure of the *TP*73 gene.

a. Genomic organization of the *TP73* gene: exons, introns and alternative splicing sites. The colors represent the specific protein domains shown below. The P1 promoter generates the TA isoforms, while the P2 promoter generates the ΔN isoforms. **b** Schematic representation of the domains encoded by the different isoforms of *p73*. TA - transactivation domain, DBD - DNA-binding domain, OD – oligomerization domain, SAM – sterile alpha motif domain, TID transactivation inhibitory domain.



Expression of ∆Np73 (RU)				
Cisplatin (μM)	0	15		
HMEC	4.2	12.3		
SAEC	0.7	3.0		
PREC	0.6	4.7		
HREC	0.7	0.4		
OVEC	0.3	0.8		

Expression of ∆Np73 (RU)					
Doxorubicin (μM)	0	0.33	1		
HMEC	2.78	52.85	74.14		
SAEC	0.16	39.61	1.38		
PREC	0.04	9.33	28.7		
HREC	0.00	0.03	0.41		

Fig. S2 Cell cytotoxicity in the various human normal primary epithelial cells following treatment with cisplatin (a) and doxorubicin (b). The cells were exposed to increasing doses of cisplatin or doxorubicin for 24 hours and cell viability was measured by XTT. HMECs – mammary, HREC – renal, SAEC – small airways, PREC – prostate, OVEC – ovary. The data represent average of 3 replicates in cells originated from 3 individuals (cisplatin) or 1 individual (doxorubicin). Error bars represent standard error of the mean (S.E.M). The tables on the right indicate that deltaNp73 induction by cisplatin or doxorubicin differed between cell types and these differences did not correlate with cell cytotoxicity.



Fig. S2c. Induction of PUMA, p21 and NOXA by DNA damage in HMECs and in other types of human epithelial cells. The cells were exposed to doxorubicin or cisplatin for 24 hours. Gene expression (mRNA) was measured by RT-qPCR and normalized to *GAPDH* (RU = $2^{\Delta CT} \times 10^4$). Epithelial cells tested: HMECs – mammary, SAEC – small airways, HREC – renal, PREC – prostate, OVEC – ovary, HCEC – colon. Error bars represent standard error of the mean (S.E.M). Number of individual tissues (n) is marked when n>1.





Fig. S3a The *deltaNp73* promoter is unmethylated in HMECs and fully methylated in white blood cells of *BRCA1* mutation carriers and noncarriers. Upper panel: DNA methylation of the *deltaNp73* promoter in primary human mammary epithelial cells (HMECs) from *BRCA1* mutation carriers (*BRCA1*-mut, n=8) and non-carriers (*BRCA1*-wt, n=14). Lower panel: DNA methylation of the *deltaNp73* promoter in white blood cells (WBCs) from *BRCA1* mutation carriers (*BRCA1*-mut, n=4) and non-carriers (*BRCA1*-wt, n=5). Methylation was measured by quantitative methylation specific PCR (qMSP). Ttest showed no significant difference between the groups. Note: The Y-axis scale is different in each panel.



Fig. S3b Cell viability in response to DNA damage in *BRCA1*-wt and *BRCA1*-mut HMECs. Dose response curves of *BRCA1*-wt (n=7 individuals) and *BRCA*-mut (n=5 individuals) HMECs show that the *BRCA1*mut cells were more sensitive to cisplatin than the *BRCA1*-wt. The data are similar to those presented in Fig.3 b-e (siControl group). The experiment was repeated twice for each individual, and XTT was done in triplicates. Error bars represent S.E.M.

1.5 Figure S3 c



siCont vs sidNp73	Estimate	Error	df	t value	Pr(> t)	
HMEC BRCA1/wt	15.4426-	5.75597	103.0342	2.683-	0.0085	**
HMEC BRCA1/mut	23.9086-	6.412	62.0284	3.729-	0.000419	***

Fig. S3 c Linear mixed model analysis compared the decrease in cell viability in HMECs *BRCA1*-wt (n=11, upper panel) and in HMECs *BRCA1*-mut (n=9, lower panel), between cells transfected with siRNA control (orange) to si Δ Np73 (green), following treatment with 10-25 μ M cisplatin. Each box displays lower Q1 and upper Q3 with a black line representing the mean, and S.E.M error bars. The table below shows analysis parameters and p-values for each comparison. The data for the si Δ Np73 combines the two different si Δ Np73 sequences.



Fig. S4 DNA methylation and induction of the *deltaNp73* and the *TAp73* gene isoforms by DNA damage in MCF10A. a Methylation at the *delatNp73* promoter (P2) in MCF10A (average of 12 independent Q-MSP measures). **b** Induction of *deltaNp73* in MCF10A by various DNA damage stimuli for 24 hours. **c** Induction of *deltaNp73* and *TAp73* by doxorubicin for 24 hours in MCF10A and in BRCA1-KD MCF10A. Gene expression (mRNA) was measured by RT-qPCR and normalized to GAPDH (RU = $2^{\Delta CT} \times 10^4$).

1.7 Figure S4 d

KD cells. The cells were untreated (0) or treated with 15 μ M cisplatinum or 0.11 μ M doxorubicin for 24 hours. Gene expression (mRNA) was measured by RT-qPCR and normalized to GAPDH (RU = 2^{Δ CT} x 10⁴).

siCont vs sidNp73	Estimate	Std. Error	df	t value	Pr(> t)	
MCF10A	19.87-	8.87	18	2.2-	0.0379	*
MCF10A/shBRCA1#1	46.62-	9.18	18	5.1-	7.80E-05	***
MCF10A/shBRCA1#2	36.00-	7.49	18	4.8-	1.41E-04	***

Fig. S4 e Linear mixed model analysis compared the decrease in cell viability in MCF10A and in MCF10A/BRCAKD following deltaNp73 inhibition and treatment with cisplatin. Analysis compared cell viability in response to increasing concentrations (10-25 μM) of cisplatin, between MCF10A cells that were transfected with non-target siRNA (siControl, orange) and cells transfected with siRNA targeting deltaNp73 (si△Np73, green). Analysis compared MCF10A and MCF10A harboring shRNA for BRCA1 (shBRCA1#1, #2). Each transfection experiment was repeated 3 times and cytotoxicity measurements were done in triplicates. Each box displays lower Q1 and upper Q3 with a black line representing the mean, and S.E.M error bars. The table below shows the analysis parameters and p-value. The data for the si△Np73 represent the average of two different siRNA sequences.

Fig S4 f. Inhibition of deltaNp73 by siRNA caused a decrease in cell viability (XTT) following exposure to doxorubicin in MCF10A (left) and this effect was augmented in MCF10A/BRCA1-KD (right). **g** The difference in cell viability between siControl and si Δ Np73 was compared between MCF10A/BRCA1-KD cells and the parental MCF10A cells for data point of 0.33 μ M doxorubicin. P-values were calculated by paired student t-test. Gene expression (mRNA) was measured by RT-qPCR and normalized to GAPDH (RU = $2^{\Delta CT} \times 10^4$). Error bars represent S.E.M of 3 independent experiments.

1. 10 Figure. S5 a-b

Fig. S5 a Expression of BRCA1 in MCF10A stably transfected with shBRCA and/or stably transfected with flag-i ANp73. BRCA1 mRNA is shown in upper panel and BRCA1 protein in lower panel. Gene expression (mRNA) was measured by RT-qPCR and normalized to GAPDH (RU = $2^{\Delta CT} \times 10^4$). **b** MCF10A BRCA1-KD clones were more sensitive to cisplatin than the parental MCF10A when the cells were not treated (NT) with doxycycline (left). Following induction of deltaNp73 with doxycycline (Doxy, right), the MCF10A BRCA1-KD clones became more resistant to cisplatin than the parental MCF10A. The cells were exposed to increasing doses of cisplatin for 48 hours and viability was measured by XTT. Data represent the mean +/- S.E.M of three independent experiments. Each data point was done in triplicates. These graphs are alternative presentation of the data shown in figure 5.

Fig 6. Expression of BRCA1 and deltaNp73 mRNA and protein in HMECs in response to cisplatin. a Gene expression (mRNA) of BRCA1 and BRCA2 was reduced while deltaNp73 was induced in HMECs (*BRCA*-wt) following exposure to doxorubicin or cisplatin for 24 hours. Gene expression was measured by RT-qPCR and normalized to GAPDH (RU = $2^{\Delta CT} \times 10^4$). Data points represent the average +/- S.E.M (n = HMECs from 4 individuals). **b** Western blots showing the reduction of BRCA1 and induction of deltaNp73 proteins in HMECs from 2 individuals following treatment with 20 μ M cisplatin for the indicated time points.

Fig. S7 Methylation of the *TP73* **gene in breast tumors.** TCGA analysis showing average methylation (beta values) across the entire *TP73* gene locus in breast tumors grouped according to estrogen receptor status (left) or grouped by PAM50 intrinsic tumor subtypes (right). The *TAp73* (P1) and *deltaNp73* (P2) promoter regions are marked by strips of light blue. (n= ER positive;584, ER negative;153, tumor adjacent normal tissue (TAN);97, lumA;238; LumB;267; Her2;117; basal;127, normal like;26).

Fig. S8 BRCA1 and 53BP1 did not co-precipitate with deltaNp73 in MCF10A.

a. MCF10A were induced by doxycycline to over-express exogenous flag-deltaNp73. Simultaneously, DNA damage was induced by doxorubicin or cisplatin for 24 hours. Cell protein lysates were either loaded directly on 2-14% gradient Acrylamide gel or following immunoprecipitation (IP) with anti-flag antibodies. Western blots (WB) were performed with antibodies to BRCA1, Flag-deltaNp73 and beta-Actin. The BRCA1 and the flag-deltaNp73 were detected in the total lysates whereas only flag-deltaNp73 was detected in the IP. **b**. A duplicate Similar experiment as described in a, where anti 53BP1antibodies detected the expected protein in the total lysate, but not in the anti-flag IPs.

Unprocessed western blot images

Breast-specific epigenetic regulation of deltaNp73 may facilitate cancer susceptibility in BRCA1-mutated HMECs

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General remarks

- All blots were cut to pieces horizontally or vertically before incubation with the specific antibodies according to the expected molecular weights of the corresponding proteins.
- 2. Most images contain more than one blot of more than one gel that were exposed together. These are annotated.
- 3. The relevant lanes are marked in blue.
- 4. Photographs of the original blots with M.W. colored markers are presented next to the images. The relevant lanes are framed in blue.

Figure 2c

HMEC 1

- 1. Marker
- 2. NT
- 3. NT 24h
- 4. NT 36 h
- 5. Doxorubicin 0.11µM 24h
- 6. Doxorubicin 0.33 µM 24h
- 7. Marker
- 8. NT
- 9. Cisplatin 20µM 24h
- 10. Cisplatin 20µM 36h

Figure 2c

HMEC 2

- I. Mark
- 2. NT
- 3. Cisplatin 20µM 36h
- 4. Cisplatin 20µM 48h
- 5. Marker
- 6. NT
- 7. Doxorubicin 0.33µM 24h
- 8. Doxorubicin 0.66 µM 24h
- 9. Marker
- 10. Marker

Figure 2c

HMEC 3

Figure 4c

MCF10A /shBRCA1#1

- 8. MACF10A /shBRCA1#1 Cisplatin 5μM
- 9. MACF10A /shBRCA1#1 Cisplatin 10µM
- 10. MCF10A /shBRCA1#1 Cisplatin 15µM

Figure 4c

MCF10A /shBRCA1#2

- 1. Marker
- 2. MCF10A NT
- 3. MACF10A Cisplatin 5μ M
- 4. MACF10A Cisplatin 10μM
- 5. MCF10A Cisplatin 15µM
- 6. MCF10A /shBRCA1#2 NT
- 7. MACF10A /shBRCA1#2 Cisplatin 5µM
- 8. MACF10A /shBRCA1#2 Cisplatin 10μM
- 9. MCF10A /shBRCA1#2 Cisplatin 15µM
- 10. Marker

Figure 5a

MCF10A i∆Np73

Figure 5a

MCF10A i∆Np73 shBRCA1#1 MCF10A i∆Np73 shBRCA1#2

- 1. Marker Fluorescent
- 2. MCF10A i∆Np73 shBRCA1#1 (IP)
- 3. MCF10A i∆Np73 shBRCA1#1 +Doxycycline 0.01µg/ml (IP)
- 4. MCF10A i∆Np73 shBRCA1#2 (IP)
- 5. MCF10A i∆Np73 shBRCA1#2 +Doxycycline 0.01µg/ml (IP)
- 6. Marker
- 7. MCF10A i∆Np73 shBRCA1#1
- 8. MCF10A i∆Np73 shBRCA1#1 +Doxycycline 0.01µg/ml
- 9. MCF10A i∆Np73 shBRCA1#2
- 10. MCF10A i∆Np73 shBRCA1#2 +Doxycycline 0.01µg/ml

Figure 5c

MCF10A i∆Np73

- 8. MCF10A i∆Np73 Cisplatin 10µM
- 9. MCF10A i∆Np73 Cisplatin 15µM
- 10. Marker

Figure 6c

MCF10A i∆Np73

- 3. MCF10A i∆Np73 +Doxycycline 0.01µg/ml 24h
- 4. MCF10A i∆Np73 +Doxycycline 0.01µg/ml 72h
- 5. Marker
- 6. MCF10A i∆Np73 NT
- 7. MCF10A i∆Np73 +Doxycycline 1µg/ml 24h
- 8. MCF10A i∆Np73 +Doxycycline 1µg/ml 72h
- 9. MCF10A i∆Np73 +Doxycycline 1µg/ml 7days
- 10. Marker

- 1. Marker
- 2. MCF10A NT
- 3. MACF10A Cisplatin 5µM
- 4. MACF10A Cisplatin 10μM
- 5. MCF10A Cisplatin 15µM
- 6. Marker
- 7. MCF10A /shBRCA1#1 NT
- 8. MACF10A /shBRCA1#1 Cisplatin 5µM
- 9. MACF10A /shBRCA1#1 Cisplatin 10µM
- 10. MCF10A /shBRCA1#1 Cisplatin 15µM

Figure S5

MCF10A /shBRCA1#1 /shBRCA1#2

70 KD —

- 1. Marker Fluorescent
- 2. Marker
- 3. MCF10A
- 4. MCF10A shBRCA1#1
- 5. MCF10A shBRCA1#2
- 6. MCF10A shBRCA1#3
- 7. MCF10A shBRCA1#1+2+3
- 8. MCF10A shBRCA1#1+2+3
- 9. Marker Fluorescent
- 10. Marker

Figure S5

MCF10A i∆Np73 /shBRCA1#1 shBRCA1#2

- 8. MCF10A
- 9. Marker
- 10. Marker Fluorescent

Figure S8a

MCF10A i∆Np73

- 1. Marker Fluorescent
- 2. MCF10A i∆Np73 NT
- 3. MCF10A i∆Np73 +Doxycycline 0.1µg/ml
- 4. MCF10A i∆Np73 +Doxycycline 0.1µg/ml +Doxorubicin 0.33µM
- 5. MCF10A i∆Np73 +Doxycycline 0.1µg/ml +Cisplatin 15µM
- 6. Marker
- 7. MCF10A i∆Np73 NT (IP)
- 8. MCF10A i∆Np73 +Doxycycline 0.1µg/ml (IP)
- 9. MCF10A i∆Np73 +Doxycycline 0.1µg/ml +Doxorubicin 0.33µM(IP)
- 10. MCF10A iΔNp73 +Doxycycline 0.1µg/ml +Cisplatin 15µM (IP)

- 1. Marker
- 2. MCF10A i∆Np73 /shBRCA1#1 +Doxycycline 0.1µg/ml +Doxorubicin 0.33µM
- 3. MCF10A i∆Np73 /shBRCA1#1 +Doxycycline 0.1µg/ml
- 4. MCF10A i∆Np73 +Doxycycline 0.1µg/ml +Doxorubicin 0.33µM
- 5. MCF10A i∆Np73 +Doxycycline 0.1µg/ml
- 6. MCF10A i∆Np73 /shBRCA1#1 +Doxycycline 0.1µg/ml +Doxorubicin 0.33µM (IP)
- 7. MCF10A i∆Np73 /shBRCA1#1 +Doxycycline 0.1µg/ml (IP)
- 8. MCF10A i∆Np73 +Doxycycline 0.1µg/ml +Doxorubicin 0.33µM (IP)
- 9. MCF10A i∆Np73 +Doxycycline 0.1µg/ml (IP)
- 10. Marker+ Marker Fluorescent

Supplementary Methods

2.1 Quantitative Methylation-Specific PCR (Q-MSP)

Purification of genomic DNA from fresh tissues or cells used standard methodologies of TNES-Proteinase K and phenol-chloroform extraction followed by ethanol precipitation. Purification of DNA from FFPE Tissues used QIAamp DNA isolation Kit (Qiagen, Germantown, MD USA) according to kit instructions. DNA quantification and purity analysis used NanoDrop 2000 spectrophotometer (ThermoFisher). 0.1-1 mg of purified genomic DNA was treated with sodium bisulfite using the EZ methylation kit (Zymo research, Orange, CA, USA). Two sets of specific primers, for the un-methylated and methylated DNA forms were designed for each of the TP73 gene promoters. Primer sequences are provided below. The PCR mix contained 5 ng of sodium bisulfite-treated DNA, 200 nM of each primer and SYBR Premix Ex Taq (Takara Bio Inc, Shiga, Japan) in 10 µl final reaction volume. PCR amplification included cycles of 1 second at 95°C followed by 30 seconds at 60°C (Rotor Gene, Corbett, Australia). Human sperm DNA was used as non-methylated contro and DNA treated with SssI CpG methyltransferase (NEB, Ipswich, MA, USA) was used as methylated positive control. In addition, DNA from MCF10A cell line was used as both unmethylated and methylated controls. True amplification products were recognized by their complete overlap with the control melting points (T_M). The C_T (cycle of threshold) of the methylated and the unmethylated products for each promoter were used to calculate deltaCT (Δ CT (U-M) = CT ummethylated – CT methylated). Methylation percentage for each gene were calculated accordingly: % methylation = $1/(1 + 2^{\Delta}CT^{(U-M)})^*100$.

Illumina Probe	Illumina Probe Sequence (gene strand +)	CpG Island location*
TAp73(P1)		
cg005656884	TGTTTGGAAGGAGCTCGCGGGCGTCGGTGCGCTTGGAGTTGTTCTCTGCG(-)	1:3556965- 3559547
cg043911115	CGGGCCCCATAAGCATCAGACCATAAGCAGCGCCGCCACTGAGAGCCGCT	1:3556965- 3559547
cg059245836	GAAGCTGAGGCCTCGGGATTGGCACAGCCCTGCAGGTCGGAGGGAG	1:3556965- 3559547
cg167417103	CGCCAAGTCCCAGGGGCCGATCCAACTCCGAGGGAGCCCCTGTGTCGCCT (-)	1:3556965- 3559547
cg197831141	GGCGGGGAGGCAGGGCGGGCTGCCCGGCCCCTAGGCGGGTTATATGGGCG (-)	1:3556965- 3559547
cg258851082	CGGCCCATCTTTCCTGACACCCGGGTCTCTCCTGGCCGCCGCCACTAGCG (-)	1:3556965- 3559547
DeltaNp73(P2)		
cg038467673	CGGCCCGCATGTTCCCCAGCATCCTCGGCTCCTGCCTCACTAGCTGCGGA(+)	1:3596889- 3597535
cg166070652	TCCCATCTCCCTTAGTTCTGTCAACTGGCTGAATCCAACAACAAAACCCG(-)	1:3596889- 3597535
cg251154601	CGCCGGTCAGCGCCGGCTCCATAATTAAACCCACCAGGGCTCCTTCCGAG(-)	1:3596889- 3597535
cg262089304	CGGGACACCAGTTCCCTGGCGTGTGCAGACCCCCCGGCGCCTACCATGCT(+)	1:3596889- 3597535

List of Illumina methylation array (Methyl-27K) probes of the TP73 gene promoters (P1 & P2)

* CpG island position on chromosome 1 by Illumina map

Primers for sodium bisulfite-QMSP (quantitative methylation spesific PCR)
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Primers for sodium bisulfite-QMSP (quantitative methylation spesific PCR)				
Name	Direction	Sequence		
TAp73_P1 _FUM	Unmethylated;forward	GGTATTTGGGTTTGTAGTTTT		
TAp73_P1_RUM	Unmethylated;reverse	ATCCCAACTCATCCCCAATCA		
TAp73_P1_FM	Methylated;forward	TAGGTATTTGGGTTCGTAGTTTC		
TAp73_P1 _RM	Methylated; reverse	CCAACTCGTCCCCGATCG		
DeltaNp73_P2_FUM3	Unmethylated;forward	TTTTGGTGTGGTTTAATATATTATT		
DeltaNp73_P2_FUM3	Unmethylated; reverse	СТСАТАААТАТТСАТССАААТССА		
DeltaNp73_P2_FM2	Methylated; forward	TT CGGCGTTTATTATGTTGTAC		
DeltaNp73_P2_FM2	Methylated; reverse	CACATCACACCTACCGTAACG		
Primers for q-PCR				
Name	Direction	Sequence		
TAp73(V1) ex1-2_GEXF	Forward	ACG TTTGAGCACCTCTGGA		
TAp73(V1) ex1-2_GEXR	Reverse	TTCCGCCCACCACCTCAT		
DeltaNp73(V2)ex3-4_GEXF	Forward	CATGCTGTACGTCGGTGA		
DeltaNp73(V2)ex3-4_GEXF	Reverse	CTGCTCATCTGGTCCATGGT		
BRCA1_GEX_F	Forward	ACCAACATGCCCACAGATCA		
BRCA1_GEX_R	Reverse	CCTGTGCCAAGGGTGAATGA		
BRCA2_GEX_F	Forward	GCTCAAATCAT TCCTGGTACA		
BRCA2_GEX_R	Reverse	CCATACAAAGTGATAAAGGACTT		
p53 _GEX_F	Forward	CGGGTCACTGCCATGGA		
p53 _GEX_R	Reverse	GGTCTGAAAATGTTTCTTGACTCA		
p21 _GEX_F	Forward	TGGAGACTCTCAGGGTCGAAA		
p21 _GEX_R	Reverse	GGCGTTTGGAGTGGTAGAAATC		
PUMA_GEX_F	Forward	GACGACCTCAACGCACAGTA		
PUMA_GEX_R	Reverse	CCATGATGAGATTGTACAGGA		
NOXA _GEX_F	Forward	GCGCAAGAACGCTCAA		
NOXA _GEX_R	Reverse	GTTTGGATATCAGATTCAGAAG		
CK7_GEX_F	Forward	GCTCCTGAAGGCTTATTCCA		
CK7_GEX_R	Reverse	TCTTGTGATTGTGGGTGGTG		
GAPDH_GEX_F	Forward	TGCACCACCAACTGTTAGC		
GAPDH_GEX_R	Reverse	GGCATGGACTGTGGCATGAG		

Sequences of siRNA and shRNA

Name	Direction	Sequence(5' to 3')	Target
si∆Np73(1)	Sense	UGACAGAACUAAGGGAGAUGGGAAA	DeltaNp73
	Antisense	UUUCCCAUCUCCCUUAGUUCUGUCA	DeltaNp73
si∆Np73(2)	Sense	GCGCCUACCAUGCUGUACGUCGGUG	DeltaNp73
	Antisense	CACCGACGUACAGCAUGGUAGGCGC	DeltaNp73
siControl	Sense	UGAAGCAAUGAGGGAUAGGGACAAA	no target
	Antisense	UUUGUCCCUAUCCCUCAUUGCUUCA	no target
shBRCA1(1)	Sense	GAGTATGCAAACAGCTATAAT	BRCA1
shBRCA1(2)	Sense	TTGCAACCTGAGGTCTATAAA	BRCA1

SiRNA was designed by BLOCK-iT[™] RNAi Designer software engine (<u>http://rnaidesigner.thermofisher.com</u>)

Design referred to Homo sapiens tumor protein p73 (TP73), transcript variant 2, mRNA NCBI Reference Sequence: NM_001126240.2

GGATTCAGCCAGTTGACAGAACTAAGGGAGATGGGAAAAGCGAAAATGCCAACAAACGGCCCGCATGTTCCCC AGCATCCTCGGCTCCTGCCTCACTAGCTGCGGAGCCTCTCCCGGTCCACGCTGCCGGGCGGCCACGACC GTGACCCTTCCCCTCGGGCCGCCCAGATCCATGCCTCGTCCCACGGGACACCAGTTCCCTGGCGTGTGCAGAC CCCCCGGCGCCTACCATGCTGTACGTCGGTGACCCCGCACGGCACCTCGCCACG

shBRCA1 was designed using Vectorbuilder software design https://en.vectorbuilder.com/design.html according to the BRCA1 mRNA sequence NM_007294.3

2.4