

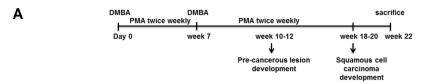


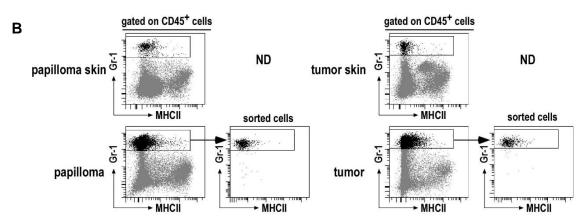
Article

Tumor-associated neutrophils dampen adaptive immunity and promote cutaneous squamous cell carcinoma development

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Supplementary Materials





Cancers 2020, 12, x S2 of S8

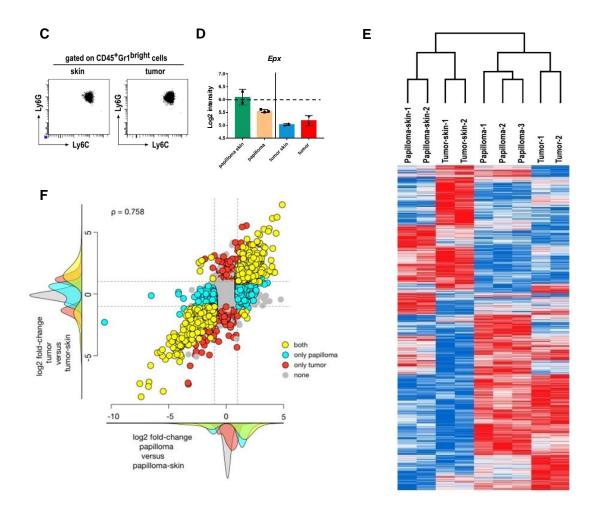
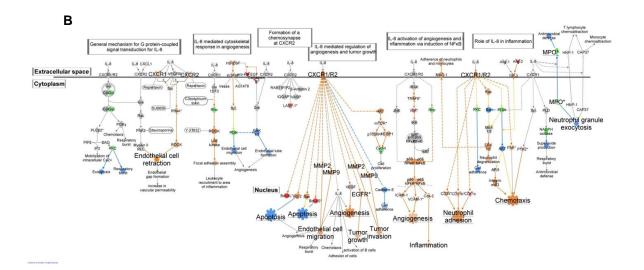


Figure S1. Gene-expression analyses of cell-sorted neutrophils purified from skin and chemically-induced cSCC, related to Figure 1. (**A**) Multi-stage chemical carcinogenesis. DMBA/PMA treatment protocol of FVB/N mice induces pre-cancerous lesions (papillomas) ~weeks 10–12, some further converting into squamous cell carcinomas (tumor) ~weeks 18–20. (**B**) Multiparameter flow cytometry cell sorting of Gr-1^{bright} cells. Representative dot plots of CD45⁺ pre-gated cells before and after Gr-1^{bright}-sorted cells (black gate), ND: not determined. (**C**) Gr-1^{bright} cells identify Ly6G⁺ neutrophils. Representative flow cytometry dot plots of CD45⁺Gr-1^{bright} gated cells. (**D**) Average intensity expression levels of *Epx* transcripts coding for eosinophil peroxidase (EPO) are shown. Dotted line delineates threshold for expression set at log2 gene expression above 6. (**E**) Hierarchical clustering. The 15% most variable probe sets with a minimum log2 average expression of 6 were analyzed within the indicated purified Gr-1^{bright} cell populations (numbers relate to experiments-see Figure 1B). Probe sets were median-centered and a clustering per rows and columns was performed with the average-linkage method. (**F**) GENAS biological correlation between tumor and papilloma modulation on all probe sets.

Cancers 2020, 12, x S3 of S8

Canonical pathways	-Lo	og10 (P value)		Activation z-score			
	papilloma versus papilloma skin	tumor versus tumor skin	tumor versus papilloma	papilloma versus papilloma skin	tumor versus tumor skin	tumor versu papilloma	
Cholecystokinin/Gastrin-mediated Signaling	3.71	2.85	0.00	2.32	2.50	NA	
IL-8 Signaling	8.31	5.52	1.97	2.92	1.26	-2.24	
HMGB1 Signaling	3.87	2.46	2.87	1.21	1.21	-2.45	
ILK Signaling	5.47	3.36	4.72	1.88	-0.41	-2.65	
Integrin Signaling	5.81	2.53	3.69	1.63	-1.46		
Signaling by Rho Family GTPases	4.97	4.94	3.88	1.22	-2.89		
RhoA Signaling	1.01	2.38	2.11	1.00			
Agrin Interactions at Neuromuscular Junction	2.58	3.67	2.29	0.71	-1.15		
Cardiac Hypertrophy Signaling	3.67	1.47	2.72	0.63	-1.28		
Regulation of Actin-based Motility by Rho	2.63	3.36	2.65	0.58	-2.32		
Tec Kinase Signaling	2.89	1.91	1.66	-0.28		NA	
Rac Signaling	0.54	1.93	Inf	-0.38		0.00	
Role of NFAT in Cardiac Hypertrophy	1.96	0.77	2.02	-0.50	-1.00	-2.24	
HGF Signaling	1.72	3.56	0.00	-0.63	-2.32	NA	
Leukocyte Extravasation Signaling	3.25	3.30	1.30	-0.77		-2.24	
LXR/RXR Activation	3.33	2.42	1.47	-1.73		0.00	
PPAR Signaling	4.66	3.20	0.21	-2.52		NA	
Cell Cycle: G1/S Checkpoint Regulation	0.64	1.52	0.87	NA	2.12	NA	
RhoGDI Signaling	4.56	3.48	3.43	-1.41	0.85	2.45	



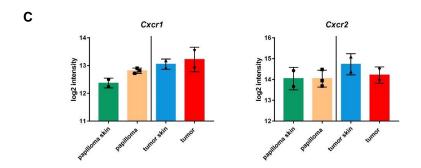


Figure S2. IPA Canonical Pathway enrichments, related to Figure 1. (**A**) Analysis of DEGs in papilloma and tumor versus their respective skin controls and their pairwise comparison. The most significant (-Log10 p-value \geq 1.39) canonical pathways are shown. A negative z-score (blue) denotes an inhibited pathway. A positive z-score (orange) stands for an activated pathway. (**B**) IL-8 Signaling Canonical Pathway. Overlay of DEGs from papilloma versus papilloma skin. Red and green colors stand for genes up and down-regulated respectively. A molecular activity predictor (MAP) was superimposed. Orange and blue stand for activation and inhibition of the gene or function. (**C**) Average intensity expression levels of Cxcr1 and Cxcr2 transcripts are shown.

Cancers **2020**, 12, x

	PantherPathways			Biological Process			Protein Class			Cellular Component		
	Class	Fold	-log10	Class	Fold	-log10	Class	Fold	-log10	Class	Fold	-log10
Classes 1	0.035	enrichment	(P- value)	1700.0000	enrichment > 5	(P- value) 1.50	700,002	enrichment	(P-value)		enrichment	(P- value)
Cluster 1	Cytoskeletal regulation by Rho GTPase	> 5	1,82	endocytosis cellular component organization	2,99	1,50	extracellular matrix structural protein	> 5	1,57			
Cluster 2	Cytoskeletal regulation by kno GTPase	23	1,02	cellular process	1,57	1,76						
				cellular component morphogenesis	4,48	1,71						
				anatomical structure morphogenesis	3,93	1,57						
				cellular component organization or								
				biogenesis	2,73	1,48						
	Heterotrimeric G-protein signaling			biogenesis								
	pathway-Gq alpha and Go alpha	> 5	3,39	regulation of molecular function	2,29	2,93	G-protein modulator	3,73	4,41	actin cytoskeleton	3,07	1,93
	mediated pathway						1.50			15		
Cluster3	Inflammation mediated by chemokine	2.00	1.00		2.27	2.74		4.61	2.42			
	and cytokine signaling pathway	3,69	1,69	regulation of catalytic activity	2,27	2,71	guanyl-nucleotide exchange factor	4,61	2,13			
	Pentose phosphate pathway	> 5	1,40	metabolic process	1,33	2,68	enzyme modulator	1,97	1,84			
	B cell activation	> 5	1,39									
Cluster 4				mesoderm development	4,69	5,25	receptor	2,46	2,33			
				immune system process	3,06	4,20	defense/immunity protein	3,75	2,00			
				response to stimulus	2,38	3,82	cell adhesion molecule	4,14	1,79			
				angiogenesis	> 5	2,99						
				cell communication	1,98	2,13						
				system development	2,74	2,05						
				developmental process	2,12	1,94						
Cluster 5												
Cluster 6		-	0.40		2.52	10.10			44.50			45.00
	Integrin signalling pathway	> 5	8,40	cell communication biological adhesion	2,52 > 5	10,18 10,17	extracellular matrix protein signaling molecule	> 5 3,64	11,69 7,28	extracellular region extracellular matrix	> 5 > 5	15,08 14,76
				cell-cell adhesion	>5			> 5		extracellular matrix	23	14,76
Cluster 7				cell adhesion	>5	9,15 8,99	extracellular matrix structural protein extracellular matrix linker protein	>5	5,61 4,72			
				developmental process	2,65	8,36	surfactant	>5	4,06			
				cellular process	1,75	8,10	cell adhesion molecule	4,26	3,52			
				ectoderm development	4,37	5,69	receptor	2,3	2,91			
				system process	2,8	4,98	membrane-bound signaling molecule	> 5	1,83			
				single-multicellular organism process	2,56	4,83	extracellular matrix glycoprotein	> 5	1,79			
				multicellular organismal process	2,56	4,81	nucleic acid binding	0,26	1,78			
				regulation of liquid surface tension	>5	4,03	antibacterial response protein	> 5	1,43			
				cell-matrix adhesion	>5	3,76			707			
				immune system process	2,58	3,60						
				homeostatic process	> 5	3,52						
				mesoderm development	3,49	3,48						
				system development	2,71	3,45						
				cell-cell signaling	3,51	2,70						
				macrophage activation	> 5	2,50						
				nervous system development	3	2,29						
				endocytosis	3,89	1,79						
				response to stimulus	1,88	1,77						
				receptor-mediated endocytosis	> 5	1,74						
Cluster 8	Glycolysis	> 5	1,40	glycolysis	> 5	2,61				intermediate filament cytoskeleton	> 5	1,35
Chuston 2	Glycolysis	> 5	2,47	glycolysis	>5	2,40	cysteine protease inhibitor	> 5	2,48			
Cluster 9				monosaccharide metabolic process	> 5	1,57						
Cluster 10	CCKR signaling map	> 5	2,09	cellular component biogenesis	4,2	2,39						
				metabolic process	1,32	1,57						
				nitrogen compound metabolic process	2,26	1,51						
				cellular component organization or	2,08	1,30						
				biogenesis	2,00	1,50						

Figure 3. Gene cluster GO-enrichment analyses, related to Figure 1. PantherDB (http://pantherdb.org/).

Cancers 2020, 12, x S5 of S8

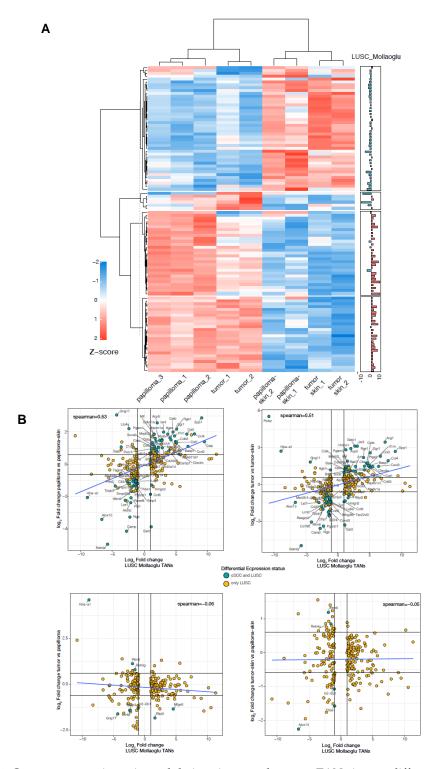
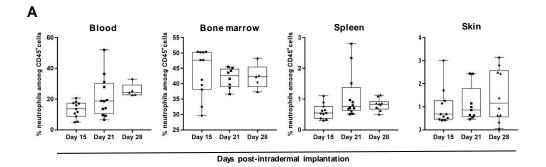


Figure S4. Common transcriptomic modulation signature between TANs in two different types of SCC, from the skin or lung, related to Figure 1. (**A**) Hierarchical clustering of differentially expressed genes from neutrophils in cSCC compared to skin control (see Figure 1) and in lung SCC compared to circulating neutrophils [35]. Only genes differentially expressed in both studies (MaxExp \geq 5, adjusted p-value \leq 0.05 and absolute log2 fold-change \geq 0.6) were considered. The log2 fold-change of each gene in the Lung SCC model between TAN and circulating neutrophils is reported as barplots on the right side of the figure. Red color of the barplot means up-regulation in the TANs, while blue stands for down-regulation. (**B**) Pairwise comparisons of the log2 fold-change modulation between [35] and our study for all genes reported as modulated in [35]. The spearman coefficient is reported for each comparison. Genes that are significantly modulated in both datasets are reported in blue, while those modulated only in the dataset [35] are reported in yellow.

Cancers 2020, 12, x S6 of S8



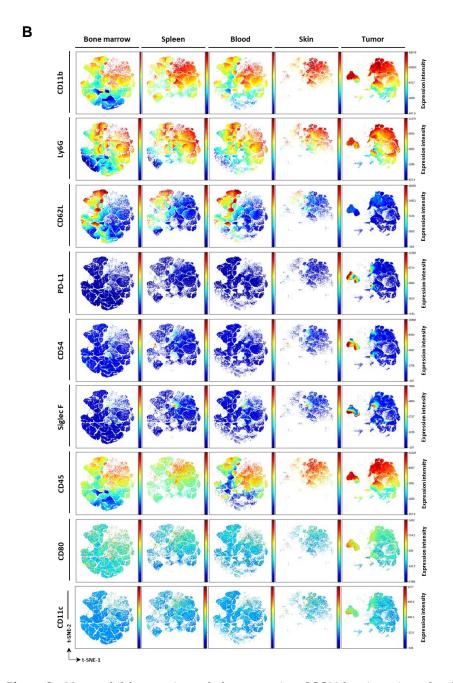


Figure S5. Neutrophil frequencies and phenotypes in mSCC38-bearing mice, related to Figure 2. (**A**) Frequencies of neutrophils overtime in the indicated tissue compartments of mSCC38-bearing mice (n = 5-11, mean \pm SEM from three independent experiments). (**B**) t-SNE analysis was performed on pre-gated CD45⁺Ly6G⁺ neutrophils from all samples. Neutrophils markers expression is presented on a rainbow heat scale in the t-SNE map of each group concatenated file.

Cancers 2020, 12, x S7 of S8

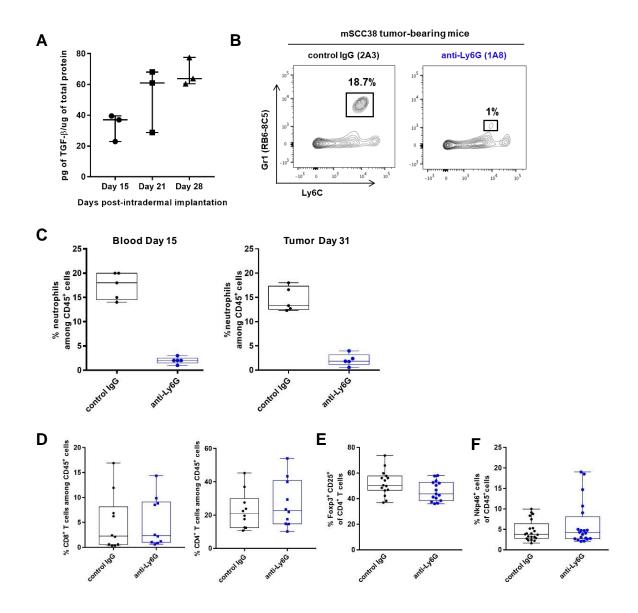


Figure S6. Analyses of tumor microenvironment of WT and neutrophil-depleted mice, related to Figure 3. (**A**) TGF-β concentration in mSCC38 tumors (n = 3, mean ± SD). (**B**) Representative contour plot showing expression of Gr1 and Ly6C in CD45⁺ cells from blood of isotype IgG control or anti-Ly6G-treated mice day 15 post-intradermal implantation. (**C**) Frequencies of neutrophils monitored in blood and tumors of isotype IgG control or anti-Ly6G-treated mice. (n = 5 mice, mean ± SD). (**D**) Frequencies of CD8⁺ and CD4⁺ T cells among CD45⁺ cells in mSCC38 tumors from isotype IgG control or anti-Ly6G-treated mice day 30 post-intradermal implantation, (n = 10, mean ± SEM from two independent experiments). (**E**) Frequencies of FoxP3⁺ CD25⁺ Treg among CD4⁺ T cells in mSCC38 tumors from isotype IgG control or anti-Ly6G-treated mice day 30, (n = 15, mean ± SEM from three independent experiments). p value ns, Mann-Whitney u test. (**F**) Frequencies of CD3⁻ NKp46⁺ NK cells among CD45⁺ cells in mSCC38 tumors from isotype IgG control or anti-Ly6G-treated mice day 30, (n = 20, mean ± SEM from three independent experiments).

Cancers 2020, 12, x S8 of S8

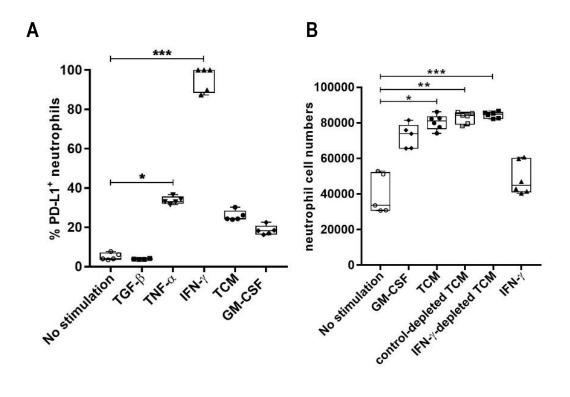


Figure S7. Modulation of PD-L1 cell surface expression and survival of neutrophils, related to Figure 4. (**A**) Frequencies of PD-L1⁺ neutrophils purified from BM and stimulated in vitro for 24 h with medium alone or in the presence of 500 ng/mL TGF- β , 500 ng/mL TNF- α , 10ng/ml IFN- γ , TCM (1/2) or 500 ng/mL GM-CSF. *p < 0.05, ***p < 0.001, Kruskal-Wallis one-way ANOVA. (**B**) neutrophil cell numbers upon culture 24 h in media alone, or supplemented with indicated TCM or cytokines. *p < 0.05, **p < 0.01, Kruskal-Wallis one-way ANOVA.



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