



## Novel Gene Fusions in Glioblastoma Tumor Tissue and Matched Patient Plasma

Lan Wang <sup>1,+</sup>, Anudeep Yekula <sup>1,+</sup>, Koushik Muralidharan <sup>1</sup>, Julia L. Small <sup>1</sup>, Zachary S. Rosh<sup>1</sup>, Keiko M. Kang <sup>1,2</sup>, Bob S. Carter <sup>1,\*</sup> and Leonora Balaj <sup>1,\*</sup>

- <sup>1</sup> Department of Neurosurgery, Massachusetts General Hospital and Harvard Medical School, Boston, MA 02115, USA. <u>lanwang.amber@gmail.com</u> (L.W.); <u>ayekula@mgh.harvard.edu</u> (A.Y.); <u>kmuralidharan@mgh.harvard.edu</u> (K.M.); <u>jsmall6@mgh.harvard.edu</u> (J.S.); <u>Zrosh@mgh.harvard.edu</u> (Z.R.); <u>kmkang@mgh.harvard.edu</u> (K.M.K.)
- <sup>2</sup> School of Medicine, University of California San Diego, San Diego, CA 92092, USA
- + These authors contributed equally.
- \* Correspondence: <u>bcarter@mgh.harvard.edu</u> (B.S.C.); <u>Balaj.Leonora@mgh.harvard.edu</u> (L.B.)

## Supplementary



**Figure S1.** Representative Bioanalyzer profiles of RNA extracted from (A) GBM tumor, (B) patient plasma and (C) healthy control plasma.



**Figure S2.** Representative DNA Bioanalyzer profiles of libraries extracted from (A) GBM tumor, (B) patient plasma.



**Figure 3.** 1-D amplification plot of AGK-BRAF fusion droplet digital PCR validation assay of GBM tumor cDNA (left), Sample library (center) from patient P3 and No template control (right).

AKT3, ALK, AR, ARHGAP26, AXL, BRAF, BRD3, BRD4, CSF1, CSF1R, EGFR, ERG, ESR1, ESRRA ETV1, ETV4, ETV5, ETV6, EWSR1, FGFR1, FGFR2, FGFR3, FGFR3, FGR, INSR, JAZF1, MAML2, MAST1, MAST2, MET, MET, MSMB, MUSK, MYB, NOTCH1, NOTCH2, NRG1, NTRK1, NTRK2, NTRK3 NUMBL, NUTM1, PDGFRA, PDGFRB, PIK3CA, PKN1, PPARG, PRKCA, PRKCB, RAF1, RELA, RET, ROS1, RSPO2, RSPO3, TERT, TFE3, TFEB, THADA, TMPRSS2

## Table 2. QIAseq Targeted RNAscan Panel screens for fusions involving the above genes.

ABI1, ABL1, ABL2, ACTN4, AFF1, AFF3, AGK, AKAP9, ALK, ARHGAP26, ARHGEF12, ASPSCR1, ATF1, ATIC, AUTS2, BAG4, BAIAP2L1, BCOR, BCR, BIRC, BRAF, CARS, CASC5, CBFA2T3, CBFB, CBL, CCDC6, CCDC88C, CD74, CEP170B, CEP89, CHIC2, CIAO1, CLCN6, CLTC, CNTRL, COL1A1, COPA, CREB1, CREBBP, CRTC1, CUX1, DAZL, DCTN1, DDIT3, DDX10, DEK, DHH, ELL, ELN, EML1, EML4, EP300, EPS15, ERC1, ERG, ESRP1, ETV1, ETV4, ETV6, EWSR1, EZR, FAM131B, FCHSD1, FGFR1, FGFR1OP, FGFR2, FGFR3, FIP1L1, FLI1, FLT3, FN1, FNBP1, FOXO1, FOXO4, FOXP1, FRYL, FRYL, FUS, GAS7, GATA1, GATM, GLIS2, GNAI1, GOLGA5, GOPC, GOT1, GPHN, HACL1, HERPUD1, HIP1, HLF, HOOK3, HOXA9, HOXD13, JAK2, KAT6A, KDM5A, KIF5B, KLC1, KMT2A, KRAS, LMNA, LNP1, LNP1, LRIG3, LSM14A, MALT1, MAML2, MECOM, MKL1, MKRN1, MLF 1.00, MLLT1, MLLT1, MLLT10, MLLT11, MLLT3, MLLT6, MLLT6, MN1, MNX1, MPRIP, MRPS14, MSN, MYB, MYH11, MYO1F, MYO5A, NCOA2, NCOA4, NCOA4, NDE1, NPM1, NRG1, NSD1, NTRK1, NTRK3, NUMA1, NUP214, NUP98, P2RY8, PAPSS1, PAX3, PAX5, PAX7, PAX8, PBX1, PCM1, PDGFB, PDGFRA, PDGFRB, PHF23, PICALM, PLAG1, PML, PPARG, PPFIBP1, PRCC, PRDM16, PRKAR1A, PRKG2, PRRX1, PSIP1, PTPRR, PWWP2A, RAF1, RANBP2, RAP1GDS1, RARA, RBM15, RCSD1, RET, RHEBL1, RNF130, ROS1, RPL22, RPN1, RUNX1, RUNX1T1, SDC4, SEC31A, SEPT2, SEPT5, SEPT6, SEPT9, SET, SFPQ, SFPQ, SLC34A2, SLC45A3, SND1, SQSTM1, SRGAP3, SS18, SSX1, SSX2, SSX2B, SSX2B, STAT5B, STIL, STRN, TACC1, TACC3, TAF15, TAF15, TAL1, TCF3, TCF7L2, TET1, TFE3, TFG, TMPRSS2, TOP 1.00, TPM3, TPM4, TPR, TRIM24, TRIM27, TRIM33, TRIP11, UBE2L3, UBE3C, USP42, VCL, VTI1A, WHSC1L1, WT1, ZBTB16, ZMIZ1, ZMYM2, ZNF384, ZNF703, ZSCAN30

**Table 3.** RNA profiles indicated by Agilent Bioanalyzer 2100 Expert Software and/or Qubit 4 Fluorometer. Tumor tissue RNA concentration were determined by Qubit 4 Fluorometer. RIN, RNA fragments % > 200 bp of all RNA samples and plasma RNA concentration were calculated by 2100 Expert Software with baseline correction. RIN, RNA Integrity Number.

Commis cohort	Patient/Healthy control	DIN	Concentration	RNA	
Sample conort	ID	KIN	(ng/µL)	fragment% >200bp	
	P1	8.0	45.4	92%	
	P2	9.9	538.0	96%	
	Р3	7.7	53.9	80%	
	P4	9.3	424.0	98%	
Tumor tissue	P5	9.1	209.0	88%	
	P6	9.0	392.7	88%	
	P7	8.7	70.4	94%	
	P8	8.9	138.4	95%	
	Р9	9.6	81.2	96%	
	P1	2.0	0.074	10%	
	P2(t1)	2.6	1.820	14%	
	P2(t <sub>2</sub> )	2.7	0.981	10%	
	P2(t <sub>3</sub> )	4.8	0.153	14%	
Patient matched	P2(t4)	3.9	0.370	19%	
plasma	Р3	6.8	0.062	6%	
	P4	4.1	0.120	14%	
	P5	1.4	0.082	21%	
	P6	2.0	0.041	5%	
	P7	5.5	0.181	17%	

## Cancers 2020, 12, x FOR PEER REVIEW

-

	P8	5.3	0.135	24%
	P9	4.2	0.202	17%
	H1	2.9	1.206	11%
	H2	2.5	2.112	12%
	H3	2.6	1.869	13%
	H4	2.6	1.499	14%
Healthy control	H5	2.7	4.274	14%
plasma	H6	3.1	3.369	17%
	H7	2.8	3.187	16%
	H8	3.1	3.685	17%
	H9	2.3	2.610	14%
	H10	3.0	5.598	19%

Table S4. Summary of fusion calling and droplet digital PCR (ddPCR) validation results. Fusions identified by MGH Solid Fusion Assay and RNA-seq are listed including the fusion categories (curated, high confidence or low confidence fusions) and number of supporting molecular tags (MTs). The corresponding ddPCR validation results for the curated and high confidence fusions. (NA, Not Available).

Sample Cohort	Patient/ Healthy Control ID	MGH Solid Fusion Assay Fusions	RNA-seq Fusions	Fusion Category	Supporting MTs	ddPCR Positive Droplet Concentration (copies/µL)
			FGFR3-TACC3	Curated	751	65550
	P1	FGFR3-	RANBP2-RGPD8	Low confidence	150	-
	11	TACC3	PICALM-SYTL2	Low confidence	12	-
			COL1A1-COL1A2	Low confidence	11	-
			VTI1A-TCF7L2	Curated	1	151.5
	P2	None	COL1A1-COL1A2	Low confidence	136	-
			COL1A1-COL2A1	Low confidence	27	-
			AGK-BRAF	Curated	625	51650
			SND1-TMEM178B	High confidence	425	29250
			CLU-TOP1	High confidence	7	-
	P3	AGK-BRAF	PTPRZ1-ELN	High confidence	8	NA
			RANBP2-RGPD8	Low confidence	269	-
GBM			EWSR1-FUS	Low confidence	10	-
Tumor			PICALM-SYTL2	Low confidence	7	FAIL
Tissue			RANBP2-RGPD8	Low confidence	212	-
			MAP1B-TOP1	Low confidence	6	-
	P4	None	CD74-CLU	Low confidence	5	-
			CLU-TOP1	Low confidence	5	-
			EWSR1-FUS	Low confidence	6	-
	P5	None	VTI1A-TCF7L2	Curated	1	128.5
	P6	None	COL1A1-COL1A2	Low confidence	17	-
	P7	None	None	NA	0	-
			FGFR3-TACC3	Curated	2	-
	Do	None	RANBP2-RGPD8	Low confidence	171	-
	Fo		SYN2-PPARG	Low confidence	4	-
			ATRNL1-MLLT10	Low confidence	3	-
	Р9	None	RANBP2-RGPD8	Low confidence	248	-
	P1	Not tested	None	NA	0	-
	P2 (t1)	Not tested	None	NA	0	-
	P2 (t2)	Not tested	None	NA	0	-
			FGFR3-TACC3	Curated	2	507600
	P2 (t3)	Not tested	VTI1A-TCF7L2	Curated	2	16615
Patient			UBE2L3-VPS39	High confidence	55	18500
Matched	P2 (t4)	Not tested	None	NA	0	-
Plasma	P3	Not tested	CD74-GID8	Filtered	57	-
	<b>D</b> (	Not tested	TMEM91-TAL1	High confidence	12	33600
	1'4		CRTC1-ABHD12	High confidence	49	8800
	P5	Not tested	None	NA	0	-
	P6	Not tested	None	NA	0	-
	P7	Not tested	RUNX1-RUNX3	Low confidence	4	-

	DO	NT 1	TMEM91-TAL1	High confidence	7	7725
	P8	Not tested	FIP1L1-SCFD2	Low confidence	6	-
			RAB7A-FOXP1	High confidence	8	8065
	P9	Not tested	RANBP2-RGPD8	Filtered	13	-
			NPM1-FBXO38	Filtered	4	-
			TMEM91-TAL1	High confidence	22	49400
	<b>U</b> 1	Not tostad	CDCA7L-MLLT3	High confidence	12	50400
	111	Not tested	UBA5-FOXP1	High confidence	14	17350
			FIP1L1-SCFD2	Low confidence	6	-
	H2	Not tested	None	NA	0	-
Healthy H3 Control H5 Plasma H6 H7 H8 H9 H10	H3	Not tested	FIP1L1-SCFD2	High confidence	8	34850
	H4	Not tested	ACTN1-ACTN4	Low confidence	8	-
	H5	Not tested	ELL-TAL1	High confidence	4	NA
	H6	Not tested	None	NA	0	-
	H7	Not tested	FIP1L1-ATP8A1	High confidence	64	NA
	ЦQ	Not tostad	GLB1-TAL1	High confidence	6	NA
	110	Not tested	RASA3-TAL1	High confidence	6	NA
	H9	Not tested	None	NA	0	-
	H10	Not tested	None	NA	0	-

**Table S5.** Primers design for fusion transcript droplet digital PCR validation. F and R primers for fusion transcript sequences used for droplet digital PCR target amplification. F: forward primers; R: reverse primers.

Fusion name		Breakpoint Location	Primers	Sequences
	5'	TMEM 91 exon 1, chr19:41,351,202 (+)	F	CGTGCGAGACGAGACACAT
TMEM91-TAL1	3'	TAL1 exon 2, chr1:47,229,351 (-)	R	CTGAGAGGCCTGCAGTTACG
ODTO1 ADUD10	5'	CRTC1 exon 1, chr19:18,683,828 (+)	F	ACAATCAGAAGCAGGCGGAG
CKTCI-ABHD12	3'	ABHD12 exon 3, chr20:25,323,430 (-)	R	TGATTCAAACCTTGATCCTGTGG
	5'	FIP 1L1 exon 12, chr4:53,428,183 (+)	F	GCACTGCTCCACCTCTGATT
FIP1LI-SCFD2	3'	SCFD2 exon 5, chr4:53,145,582 (-)	R	GCTGCTTTAACACAACGGACA
UBE2L3-VPS39	5'	UBE2L3 exon 3, chr22:21,611,043 (+)	F	CCAAACATCGACGAAAAGGGG
	3'	VPS39 exon 2, chr15:42,199,961 (-)	R	TGGTTCCCACAAGAAGCCATT
DARTA EOVDI	5'	RAB7A exon 1, chr3:128,726,359 (+)	F	GTTTAGTCTCCTCCGGCG
RAB7A-FOXP1	3'	ROXP1 exon 12, chr3:71,015,653 (-)	R	TGGCCACTTGCATACACCAT
CD74 CID9	5'	CD74 exon 9, chr5:150,402,152 (-)	F	CAGTCCCCATGTGAGAGCAG
CD/4-GID6	3'	GID8 exon 5, chr20:62,944,739 (+)	R	TCCGTGGTAGACCGAATCCT
CDCA7L MLLT2	5'	CDCA7L exon 1, chr7:21,945,781 (-)	F	GCCCGGTTAGGAAGAATGGA
CDCA7L-MLLT3	3'	MLLT3 exon 9, chr9:20,354,879 (-)	R	TGCTTATCTGATTTGCTTTGCTT
LIBAE EOVDI	5'	UBA5 exon 1, chr3:132,660,698 (+)	F	GAGCTCAGAGGTGGTGGATTC
UDA5-FOAT I	3'	FOXP1 exon 12, chr3:71,015,653 (-)	R	CAGCCTGGCCACTTGCATA
EID111 SCED2	5'	FIP1L1 exon 9, chr4:53,399,839 (+)	F	TTGTTCAAGACTGGGCTTCCA
FII ILI-SCID2	3'	SCFD2 exon 3, chr4:53,313,763 (-)	R	GCAGGTTTTCTCTGCTTGCC
VTI1A TOE7I 2	5'	VTI1A exon 3, chr10:112,464,657 (+)	F	TCCGAGAGATACCACCCCAA
V111A1CF/L2	3'	TCF7L2 exon 5, chr10:113,141,184 (+)	R	GCACCACTGGCACTTTGTTAG
FGFR3-TACC3	5'	FGFR3 exon 16, chr4:1,806,934 (+)	F	GACCTGGACCGTGTCCTTACC
	3'	TACC3 exon 9, chr4:1,737,598 (+)	R	TGGAGCAGGTCCACTATAGGTC
AGK-BRAF	5'	AGK exon 2, chr7:141,555,567 (+)	F	ACGCTTCGAAATCACTGGAAG
	3'	BRAF exon 7, chr7:140,794,467 (-)	R	TGAAGGAGACGGACTGGTGA
SND1-TMEM178B	5'	SND1 exon 10, chr7:127,721,400 (+)	F	ACGATTCACCTGTCCAGCAT
	3'	TMEM178B exon 1, chr7:141,212,591 (+)	R	TAGGGCATGCCACTCATCCT
DTDD 71 EI N	5'	PTRPZ1 exon 21, chr7:122,040,979 (+)	F	GCCTGACATGGGAGTACCAG
I II KZI-ELN	3'	ELN exon 3, chr7:74,037,707 (+)	R	GGCTTAAGAGGTTTGCCTCCA
DICALM SVTL2	5'	PICALM exon 19, chr11:85,974,708 (-)	F	AAACCCCTTTGGCCCTGTATC
TICALM-51112	3'	SYTL2 exon 1, chr11:85,758,114 (-)	R	CAGCACACTCACTCTCTGGT
ELL TAL1	5'	ELL exon 1, chr19:18,521,921 (-)	F	GCGCTGAAGGAGGATAGGAG
ELL-IALI	3'	TAL1 exon 2, chr1:47,229,351 (-)	R	CTGAGAGGCCTGCAGTTACG
<b>FIP1I 1_A TP8A 1</b>	5'	FIP1L1 exon 12, chr4:53,428,183 (+)	F	GCACTGCTCCACCTCTGATT
FIP1L1-ATP8A1	3'	ATP8A1 exon 5, chr4:42,616,078 (-)	R	GCGTTTGTTTCTTGTTCACTGC
CI B1-TAI 1	5'	GLB1 exon 1, chr3:33,097,011 (-)	F	CGGGGTTCCTGGTTCGCAT
GLD1-IALI	3'	TAL1 exon 2, chr1:47,229,351 (-)	R	TTACGCTGCGGTGTGGTCC
RASA3-TAI1	5'	RASA exon 1, chr13:114,132,435 (-)	F	GTCTTCCAGAGCGTGAAGATCA
KASA3-IALI	3'	TAL1 exon 2, chr1:47,229,351 (-)	R	CTGAGAGGCCTGCAGTTACG

**Table S6.** Overview of RNA-seq mapping metrics generated by GeneGlobe Data Analysis Center. RNA Control Primers refer to a set of RNA control primers targeting housekeeping genes that can be used as an indicator of RNA diversity (QIAseq<sup>™</sup> Targeted RNAscan Data Analysis Handbook, Qiagen).

Sample Cohort	Patient/ Healthy Control ID	Raw Reads/ Sample	Mapped Reads/ Sample	Average RNA Control Primers	Reads% > 200 bn
	P1	3 254 559	3 084 254	4 712 00	31.04%
	P2	3 251 846	3 177 744	4 291 80	33 15%
	P3	3 739 661	3 488 775	2 857 80	23.65%
	P4	2,854,488	2,729,137	5.061.00	36.34%
GBM Tumor	P5	2,120,682	2.025.337	4,000 80	35.87%
Tissue	P6	2.891.115	2,795,359	3,230.20	33.38%
	P7	2.563.322	2,432,636	5.364.20	35.42%
	P8	2,982,206	2,825,331	6,289.80	32.92%
	Р9	3,109,352	2,970,338	6,059.50	43.20%
	P1	2,394,700	2,250,364	82.5	17.40%
	P2 (t1)	2,741,135	2,600,817	116.2	20.31%
	P2 (t2)	2,404,068	2,277,630	129.2	18.60%
	P2 (t3)	3,227,719	3,060,547	184.8	9.90%
	P2 (t4)	3,227,967	3,044,622	157.5	21.12%
Matched Patient	P3	2,742,402	2,536,385	174.5	21.15%
Plasma	P4	2,345,261	2,231,584	152.2	18.83%
	P5	2,745,203	2,608,936	153.5	21.52%
	P6	1,910,362	1,813,593	136.5	19.80%
	P7	1,429,178	1,352,839	115.5	19.30%
	P8	2,694,475	2,546,389	163.2	18.60%
	Р9	3,157,529	3,005,320	202.2	20.50%
	H1	6,352,456	5,887,078	195.5	20.08%
	H2	5,955,852	5,607,881	290.8	19.90%
	H3	4,668,412	4,385,014	227.5	19.50%
	H4	5,681,289	5,211,746	232.8	17.67%
Healthy Control	H5	2,519,716	2,377,286	233	17.70%
Plasma	H6	2,016,223	1,906,711	127.2	18.30%
	H7	2,753,480	2,614,837	183.8	19.84%
	H8	2,809,201	2,663,076	255.8	20.53%
	H9	2,543,422	2,420,411	182.2	20.12%
	H10	2,617,655	2,487,716	180.2	19.80%