



## Editorial Electric Field Based Therapies in Cancer Treatment

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+ We dedicate this Special Issue to Dr. Justin Teissié, an outstanding scientist, extraordinary adviser, and respected friend, who lived for science, and whom we lost in September 2020. He lost his fight, but his electric-field mediated battle against cancer will indeed go on.

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Enormous progress has been made in pulsed electric field-based therapies since J. Teissié reported the occurrence of electric field-induced transient pores in phospholipid bilayer vesicles in 1981 [1]. The transient pores, which occurred upon application of short (microsecond) external electric field pulses were attributed to the dielectric breakdown of the bilayer structure and did not damage the lipid membrane [1]. While the term "electroporation" (also known as electropermeabilization or the formation of transient permeant structures) took some time to anchor within the vocabulary of the entire scientific community, experiments showed that pulsed electric field could have numerous biological consequences. Among them, the induction of ATP synthesis [2], onset of cell fusion [3], generation of cell hybrids [4], induction of cytoskeletal reorganization [5], transfection of cells [6–8], delivery of molecules including cytotoxic drugs to cells [9], and anticancer therapy [10] set the bases for therapeutic applications.

This Special Issue covers a number of hot topics in the field of electric field based therapies in cancer treatment. Original research and a review article present recent advances in calcium electroporation [11–14], the potential of expandable electrodes is presented in a porcine model undergoing open body surgery, laparoscopy and endoscopy [15], and the extension of the time of blood brain barrier disruption is highlighted after application of high frequency electroporation [16]. In addition to a review providing the state of the art in cytoskeletal alterations after electroporation [17], original research articles also present the induction of immunogenic cell death by nanosecond pulsed electric fields [18] and describe protocols for the optimization of DNA electrotransfer [19,20].

In addition to electrochemotherapy and irreversible electroporation, that already proved their efficacy to treat cancer, the combination of calcium ions with high intensity electric field pulses is emerging in clinics. As calcium ions are implicated in cell death regulation, the amplification of calcium ions uptake upon electropermeabilization results in acute and severe ATP depletion associated with cancer cell death. This approach has been used in clinics and the treatment modality is thoroughly reviewed by Frandsen et al. [14]. In addition to this comprehensive review, Agoston et al. [12] presented a Phase II Clinical trial (NCT03628417) where they compared the efficacy of calcium-based electroporation with bleomycin-based electrochemotherapy. The two approaches lead to a similar tumor response but adverse reactions, such as ulceration and hyperpigmentation, were less common after calcium-based electroporation [12]. Moreover, as highlighted by Gibot et al. [13], calcium electroporation is not genotoxic. In addition to electrochemotherapy protocols, calcium ions were shown to delay tumor growth upon combination with irreversible electroporation, as reported by Novickij et al. [11]. Their protocol was not only efficient against primary tumors in a murine model, but also destroyed the tumor microenvironment and induced anti-tumor immune response. A similar phenomenon of immunostimulation was also observed by Rossi et al. [18], who applied nanosecond pulsed electric fields in murine cancer models.

Significant advances were made in electric field-based delivery of DNA [21] to cancer cells. Indeed, protocols still need to be optimized. In this context, Sieni et al. [19] described a useful three-dimensional cellular scaffold, which is rich in extracellular matrix and appears particularly attractive for gene electro transfer studies. As electrotransfer may decrease cell viability, Wang et al. [20] show that inhibition of caspases post electrotransfer may significantly increase cell viability, without compromising the T cell receptor disruption efficiency.

High frequency electroporation was efficiently used to transiently disrupt the blood brain barrier in vivo in a healthy rat brain model, as shown by Lorenzo et al. [16]. In the mentioned study, the blood brain barrier could remain focally disrupted for 72 h following the application of high frequency electroporation, and returned to its normal 96 h following pulse exposures. This finding thus suggests a useful approach to permeate the blood brain barrier and promote drug diffusion into brain parenchyma.

Last but not least, this special issue also includes a review by Graybill et al. [17] describing cytoskeletal disruption after cellular exposure to pulsed electric fields. This extensive review summarizes nearly 200 studies describing cytoskeletal disruption [22] englobing Teissié's pioneering works [23] and a series of cutting-edge papers detailing the mechanisms and outcomes of cytoskeletal disruption.

We hope that this Special Issue will be of interest to a vast number of researchers and that it will encourage new ideas and scientific discoveries. The editors are highly grateful to the editor in chief, editorial staff, reviewers, and to all contributors. We look forward to meeting you again at the forthcoming 4th World Congress in September 2021 in Copenhagen, Denmark.

Conflicts of Interest: The author declares no conflict of interest.

## References

- Teissie, J.; Tsong, T.Y. Electric field induced transient pores in phospholipid bilayer vesicles. *Biochemistry* 1981, 20, 1548–1554. [CrossRef] [PubMed]
- Teissie, J.; Knox, B.E.; Tsong, T.Y.; Wehrle, J. Synthesis of adenosine triphosphate in respiration-inhibited submitochondrial particles induced by microsecond electric pulses. *Proc. Natl. Acad. Sci. USA* 1981, 78, 7473–7477. [CrossRef] [PubMed]
- Teissie, J.; Knutson, V.; Tsong, T.; Lane, M. Electric pulse-induced fusion of 3T3 cells in monolayer culture. Science 1982, 216, 537–538. [CrossRef] [PubMed]
- 4. Finaz, C.; Lefevre, A.; Teissie, J. Electrofusion: A new, highly efficient technique for generating somatic cell hybrids. *Exp. Cell Res.* **1984**, *150*, 477–482. [CrossRef]
- Blangero, C.; Rols, M.; Teissie, J. Cytoskeletal reorganization during electric-field-induced fusion of Chinese hamster ovary cells grown in monolayers. *Biochim. Biophys. Acta (BBA)-Biomembr.* 1989, 981, 295–302. [CrossRef]
- Rols, M.P.; Coulet, D.; Teissié, J. Highly Efficient transfection of mammalian cells by electric field pulses: Application to large volumes of cell culture by using a flow system. *Eur. J. Biochem.* 1992, 206, 115–121. [CrossRef]
- 7. Wolf, H.; Rols, M.; Boldt, E.; Neumann, E.; Teissie, J. Control by pulse parameters of electric field-mediated gene transfer in mammalian cells. *Biophys. J.* **1994**, *66*, 524–531. [CrossRef]
- Golzio, M.; Teissié, J.; Rols, M.-P. Direct visualization at the single-cell level of electrically mediated gene delivery. *Proc. Natl. Acad. Sci. USA* 2002, 99, 1292–1297. [CrossRef]
- Rols, M.-P.; Golzio, M.; Delteil, C.; Teissié, J. In vitro delivery of drugs and other molecules to cells. In *Electrochemotherapy, Electrogenetherapy, and Transdermal Drug Delivery*; Springer: Berlin/Heidelberg, Germany, 2000; pp. 83–97.
- Mir, L.; Orlowski, S.; Belehradek, J., Jr.; Teissie, J.; Rols, M.; Serša, G.; Miklavčič, D.; Gilbert, R.; Heller, R. Biomedical applications of electric pulses with special emphasis on antitumor electrochemotherapy. *Bioelectrochemistry Bioenerg.* 1995, *38*, 203–207. [CrossRef]
- Novickij, V.; Česna, R.; Perminaitė, E.; Zinkevičienė, A.; Characiejus, D.; Novickij, J.; Šatkauskas, S.; Ruzgys, P.; Girkontaitė, I. Antitumor response and immunomodulatory effects of sub-microsecond irreversible electroporation and its combination with calcium electroporation. *Cancers* 2019, *11*, 1763. [CrossRef]

- Ágoston, D.; Baltás, E.; Ócsai, H.; Rátkai, S.; Lázár, P.G.; Korom, I.; Varga, E.; Németh, I.B.; Dósa-Rácz Viharosné, É.; Gehl, J. Evaluation of calcium electroporation for the treatment of cutaneous metastases: A double blinded randomised controlled phase II trial. *Cancers* 2020, *12*, 179. [CrossRef] [PubMed]
- Gibot, L.; Montigny, A.; Baaziz, H.; Fourquaux, I.; Audebert, M.; Rols, M.-P. Calcium Delivery by Electroporation Induces In Vitro Cell Death through Mitochondrial Dysfunction without DNA Damages. *Cancers* 2020, *12*, 425. [CrossRef] [PubMed]
- 14. Frandsen, S.K.; Vissing, M.; Gehl, J. A comprehensive review of calcium electroporation—A novel cancer treatment modality. *Cancers* 2020, 12, 290. [CrossRef] [PubMed]
- 15. Izzo, F.; Ionna, F.; Granata, V.; Albino, V.; Patrone, R.; Longo, F.; Guida, A.; Delrio, P.; Rega, D.; Scala, D. New Deployable Expandable Electrodes in the Electroporation Treatment in a Pig Model: A Feasibility and Usability Preliminary Study. *Cancers* **2020**, *12*, 515. [CrossRef] [PubMed]
- 16. Lorenzo, M.F.; Thomas, S.C.; Kani, Y.; Hinckley, J.; Lee, M.; Adler, J.; Verbridge, S.S.; Hsu, F.-C.; Robertson, J.L.; Davalos, R.V. Temporal characterization of blood–brain barrier disruption with high-frequency electroporation. *Cancers* **2019**, *11*, 1850. [CrossRef] [PubMed]
- 17. Graybill, P.M.; Davalos, R.V. Cytoskeletal Disruption after Electroporation and Its Significance to Pulsed Electric Field Therapies. *Cancers* **2020**, *12*, 1132. [CrossRef] [PubMed]
- Rossi, A.; N Pakhomova, O.; Mollica, P.A.; Casciola, M.; Mangalanathan, U.; G Pakhomov, A.; Muratori, C. Nanosecond Pulsed Electric Fields Induce Endoplasmic Reticulum Stress Accompanied by Immunogenic Cell Death in Murine Models of Lymphoma and Colorectal Cancer. *Cancers* 2019, *11*, 2034. [CrossRef]
- Sieni, E.; Dettin, M.; De Robertis, M.; Bazzolo, B.; Conconi, M.T.; Zamuner, A.; Marino, R.; Keller, F.; Campana, L.G.; Signori, E. The Efficiency of Gene Electrotransfer in Breast-Cancer Cell Lines Cultured on a Novel Collagen-Free 3D Scaffold. *Cancers* 2020, *12*, 1043. [CrossRef]
- 20. Wang, C.; Chang, C.-C.; Wang, L.; Yuan, F. Inhibition of Caspases Improves Non-Viral T Cell Receptor Editing. *Cancers* **2020**, *12*, 2603. [CrossRef]
- 21. Rols, M.-P.; Delteil, C.; Golzio, M.; Dumond, P.; Cros, S.; Teissie, J. In vivo electrically mediated protein and gene transfer in murine melanoma. *Nat. Biotechnol.* **1998**, *16*, 168–171. [CrossRef]
- 22. Rols, M.-P.; Teissié, J. Experimental evidence for the involvement of the cytoskeleton in mammalian cell electropermeabilization. *Biochim. Biophys. Acta* (*BBA*)-*Biomembr.* **1992**, *1111*, 45–50. [CrossRef]
- 23. Teissie, J.; Rols, M.P. Manipulation of cell cytoskeleton affects the lifetime of cell membrane electropermeabilization. *Ann. N. Y. Acad. Sci.* **1994**, 720, 98–110. [CrossRef] [PubMed]

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