

Supplementary Information: Novel Platform for Regulation of Extracellular Vesicles and Metabolites Secretion from Cells Using a Multi-Linkable Horizontal Co-Culture Plate

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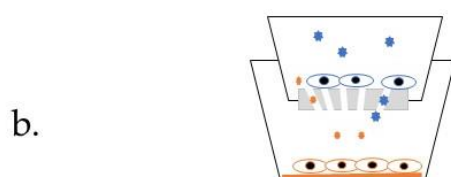
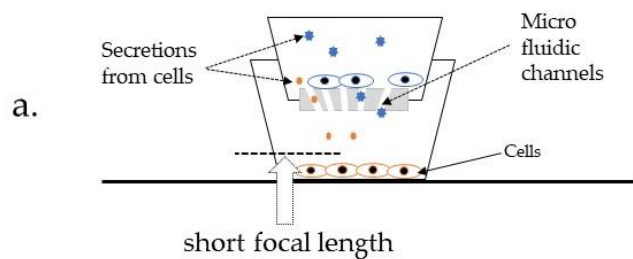
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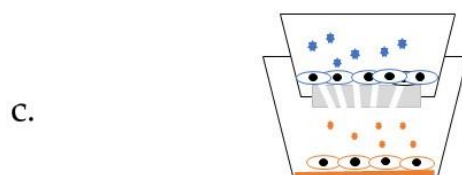
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Supplementary information



Because the material of the surface to which the cells adhere is different, the cells respond differently.



High density of the cells may decrease the exchange of materials

Figure S1. The three main problems associated with a vertical-type co-culture plate (VTCP). (a) The upper container cannot be observed because of the short focal length of the microscope. (b) The surface material to which the cells adhere differs, meaning that the cells may respond differently. (c) The high density of cells may prevent the exchange of materials.

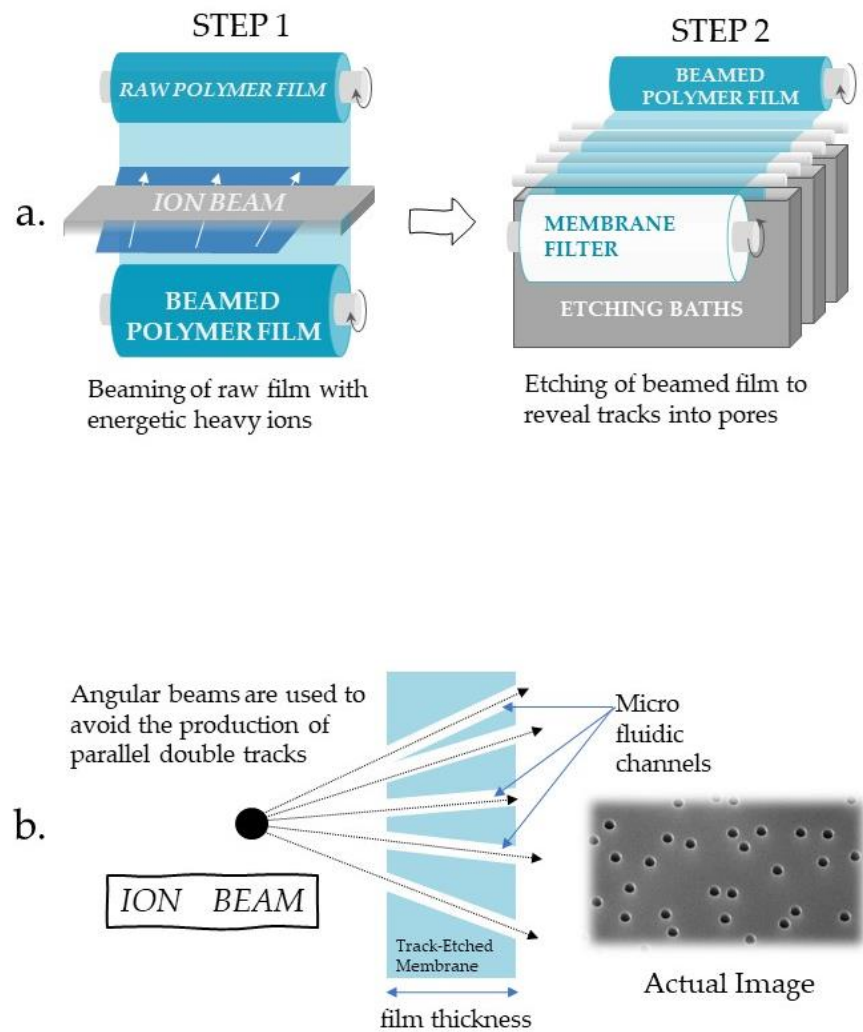


Figure S2. (a) Track-etched membranes were fabricated by irradiating a raw polymer film with energetic heavy ions. (b) The beamed film was then chemically etched to make uniform and precise pores. Pore density and pore orientation were determined during the beaming step, while the etching conditions defined the pore size.

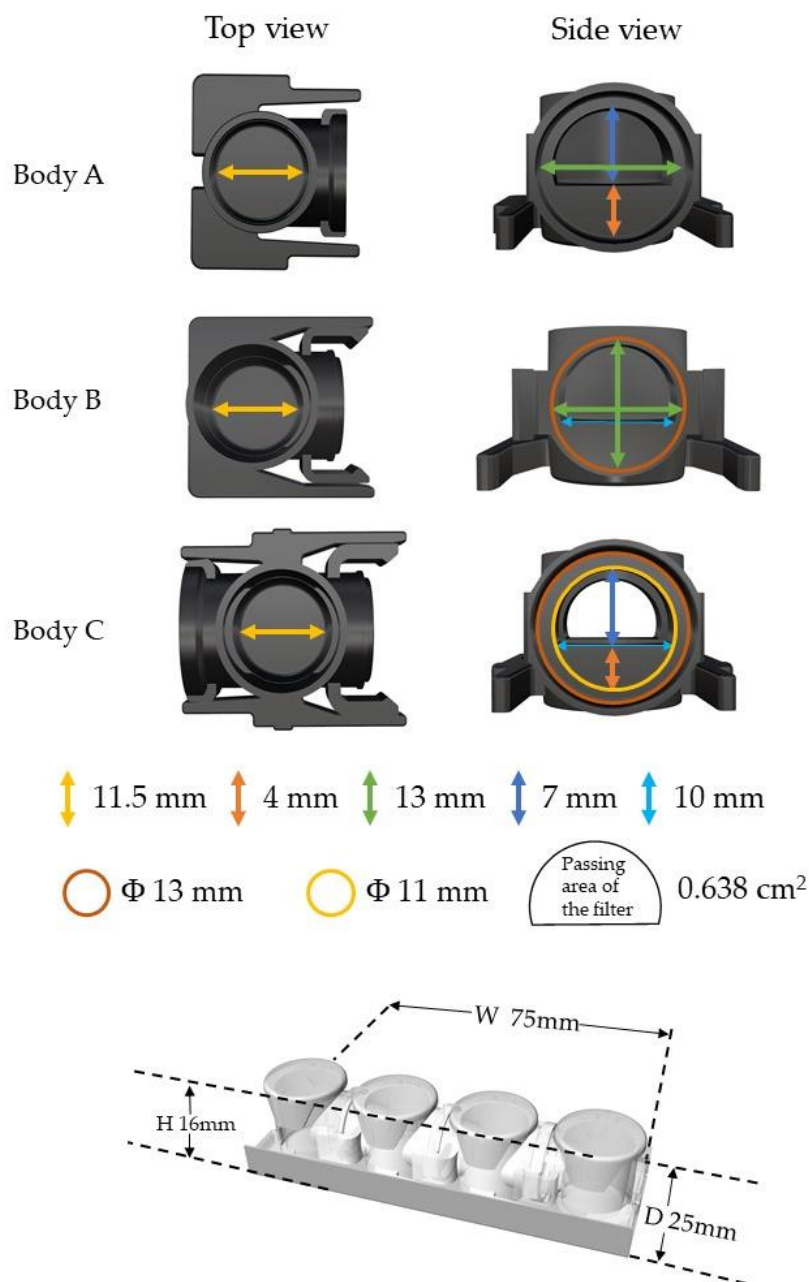


Figure S3. Size of each component. The size of each component is described in the figure. The top cover of the main body was made of polystyrene, the common sidewall cover was constructed using low-density polyethylene, and the O-ring was made of silicone. The O-ring was hydrophylized via corona discharge. All materials used for building the main body of the HTCP were sterilized with electron beams.

How to use

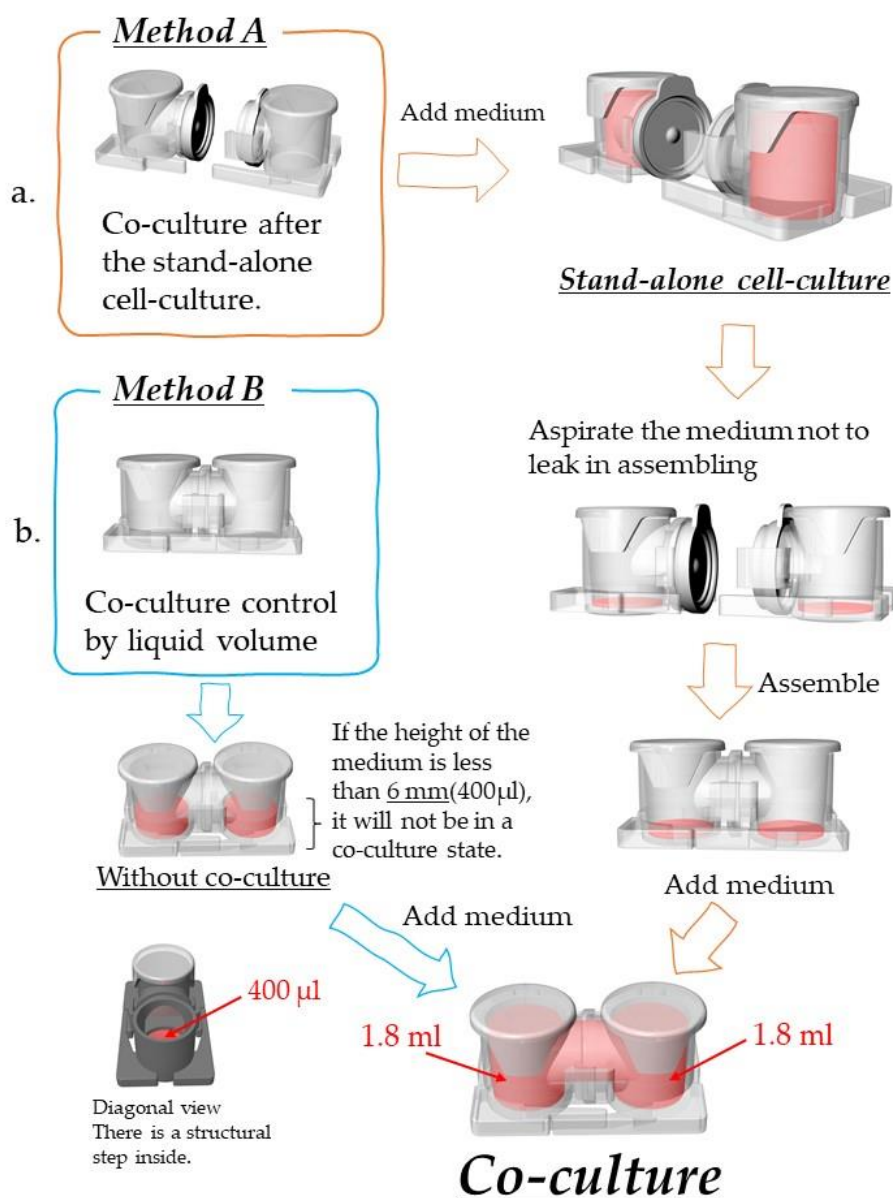
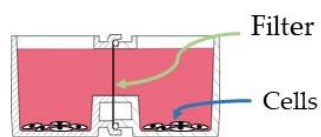
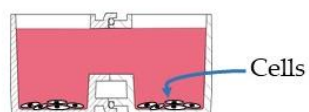


Figure S4. How to use. Methods A (Initial monoculture mode) and B (Controlling co-culture with medium volume). (a) In a stand-alone way, the cell types are cultured separately and then co-cultured by connecting the two containers (combined state) without damaging the cells. Bodies A and B are designed to allow the use of separated cell culturing. (b) In the connected mode, the two containers are joined from the beginning of cell culture. The height (volume) of the culture medium is used to control the amount of liquid that traverses the two containers, controlling the extent of co-culturing. Re-separation is possible. After reducing the amount of culture medium, one can separate bodies A and B by applying force to pull them apart to the left and right, and then cover the sides with a side cover and increase the amount of culture medium to create a monoculture. However, as this is a plastic product, recombination after separation could be problematic.

a. Filter separation



b. Without separation



c. Solid separation : 3D culture

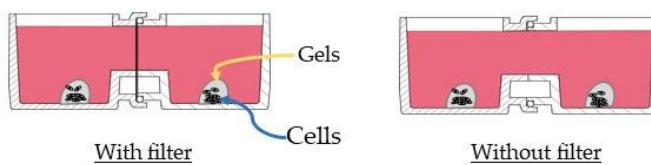


Figure S5. Schematic of various use methods: (a) filter separation, (b) without separation, and (c) solid separation with gels (three-dimensional culture).

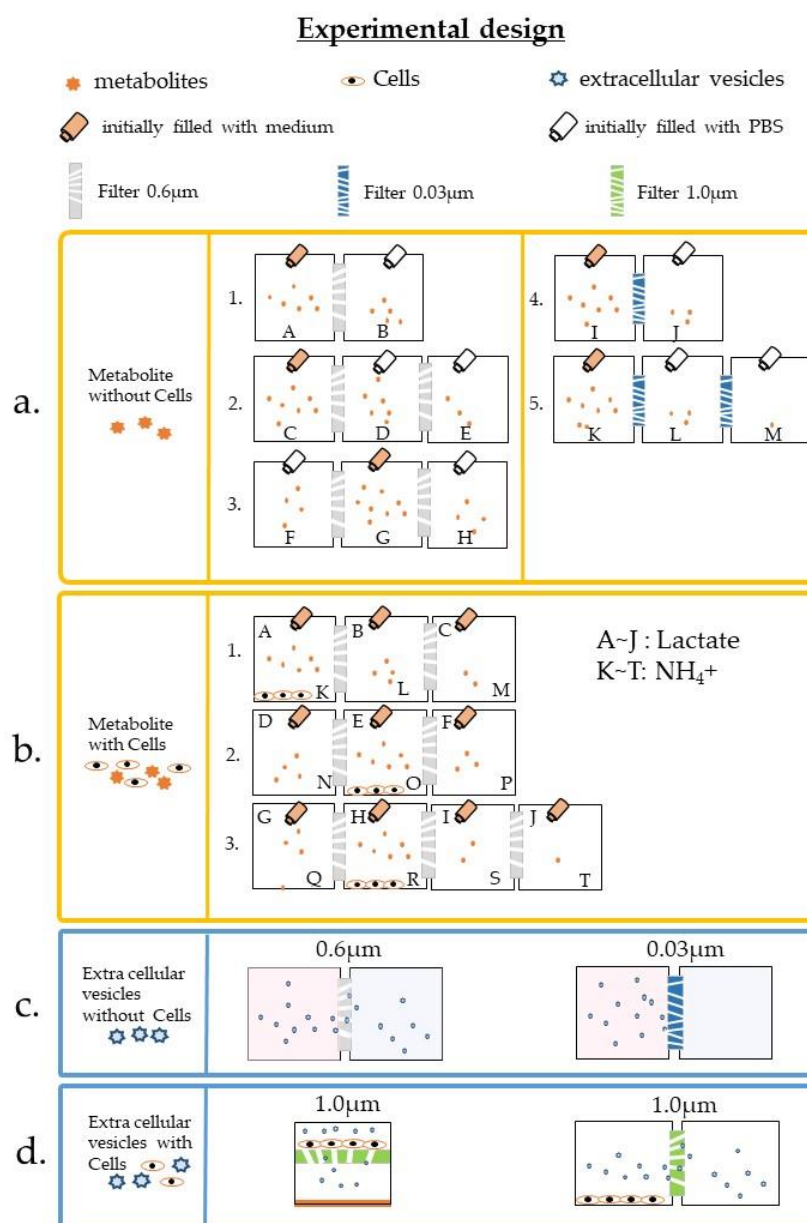


Figure S6. Summary of experimental design. (a) Differences in mass transfer due to the number of containers, position of the medium, and pore size of the filter were measured. All the measurements were performed under cell-free conditions. (b) Differences in metabolite concentrations due to differences in the positions of the containers in which the cells were placed were measured. (c) The filter passability of exosomes was measured. (d) Comparison of the passability of exosomes between VTCP and HTCP.

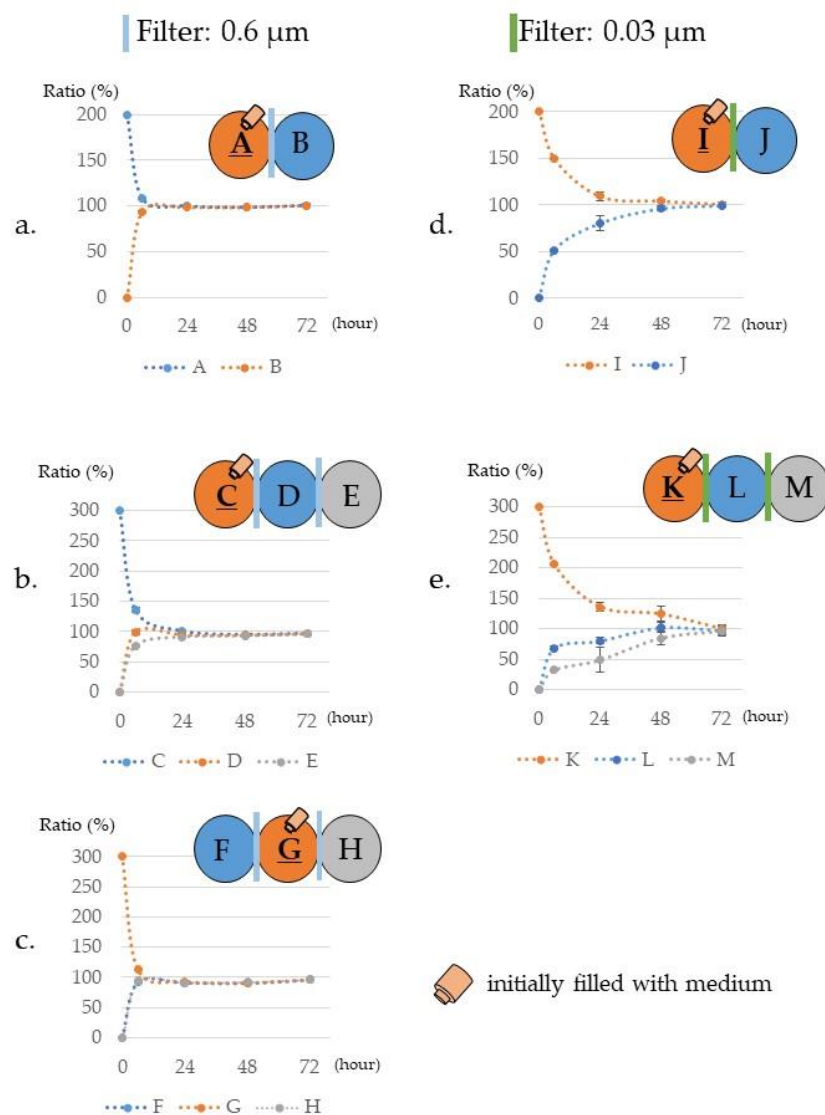


Figure S7. The results of NH_4^+ transfer without cells. Orange-colored wells with clip art of bottle were filled with medium including glucose. The remaining wells were filled with phosphate-buffered saline (PBS), which did not contain glucose. When all wells were mixed, the final concentration was converted to 100 %. The vertical axis shows the converted concentration in %. Glucose concentration in the medium was measured after 6–72 h. The vertical axis is the concentration (%), and the horizontal axis is the time (h). Filter pore size is 0.6 μm or 0.03 μm .

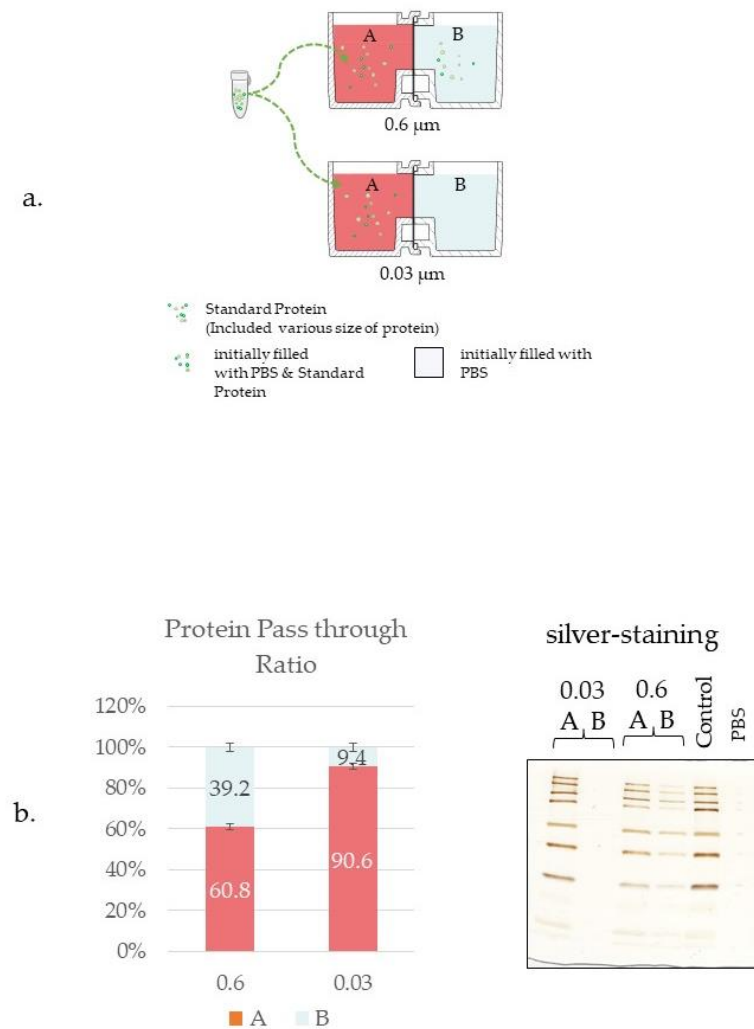


Figure S8. Migration of proteins through filters (A) Equal amounts of standard proteins were added to one side of the horizontal type co-culture system (HTCP) to test the ability of proteins to pass through filters of different sizes. (B) Proteins passing through the 0.6 μm and 0.03 μm filters were measured with silver staining.

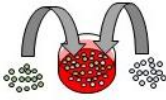

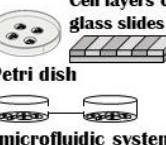
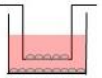
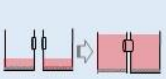

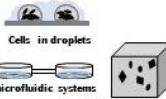
How to co-culture		Direction	Comparison of co-culture		
			Separating Agent	Method and name of interaction	Feature
Contact	Both Direction	None	Mix cells 	Direct impact can be observed. Since cells are mixed, it becomes difficult to distinguish cells. Whether it is an action by secretion or an action by contact is unknown. Notice that action by humoral factors is also added	
	One Way		Add supernatant to the other 	Standard method of adding cell supernatant. We can examine the effect at the time of attachment, but it is impossible to observe sustained effects.	
Non-Contact	Both Direction		Structure specific type separation 	There is no guarantee that cells will not be mixed, but co-culture is carried out by utilizing structural features.	
			Filter	Vertical Type Co-Culture Plate 	Although interaction by secretion can be continuously seen, both living cells can not be observed simultaneously with a time lapse microscope.
				Horizontal Type Co-Culture Plate 	After incubating each living cell with different conditions and drugs, it is possible to observe sustained interaction with a time lapse microscope together. A filter can also be used between them.
		Micro-Flow Type  microfluidic systems	Co-cultivate using microchannel. Some have filters or some make microstructures similar to filters.		
		Gels	Solid separate type  microfluidic systems Cells encased in gel	Co-culture is performed utilizing the characteristics of solid substances such as gel.	

Figure S9. Various co-culture methods have been introduced in this study. Co-culture methods can be divided into two categories based on whether the cells are in direct contact or not. Next, they are divided by based on the nature of the material exchange i. e. bidi-rectional or unidirectional. Finally, different types of separation materials exist. Some use filters, while others use microfluidic channels, and some use no separating material. Modified from Shimasaki et al. [1] with permission.

Differences in Co-culture Vessels by Vertical and Horizontal Binding Directions.

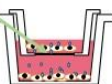
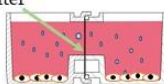
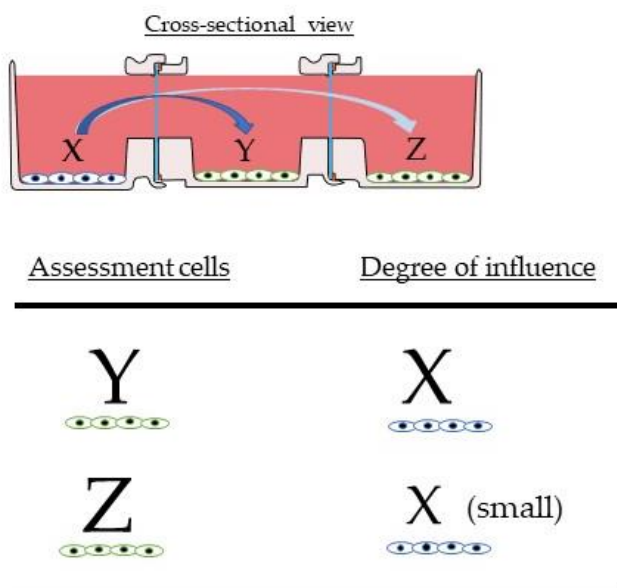
Type of vessels Connection type	VTCP Vertical connection	HTCP Horizontal connection
Image	Filter 	Filter 
The morphology of the cells in the two chambers can be easily visualized by microscope	X morphology of the cells in upper chamber can not be visualized.	O Time-lapse video can be taken of the growing cell cultures in both chamber.
Both cell cultures grow on same type of plastic surface.	X	O
A variety of different filter pore sizes can be used; changing the maximum size of molecules shared between the two culture vessels.	△	O
Growth of cells do not prevent the exchange of liquid materials such as EVs by blocking the pore of filter	△	O
Multi connection	X	O

Figure S10. List of differences in features between VTCP and HTCP modified from Shimasaki et al. [1] with permission.

a. Experimental design 1



b. Experimental design 2

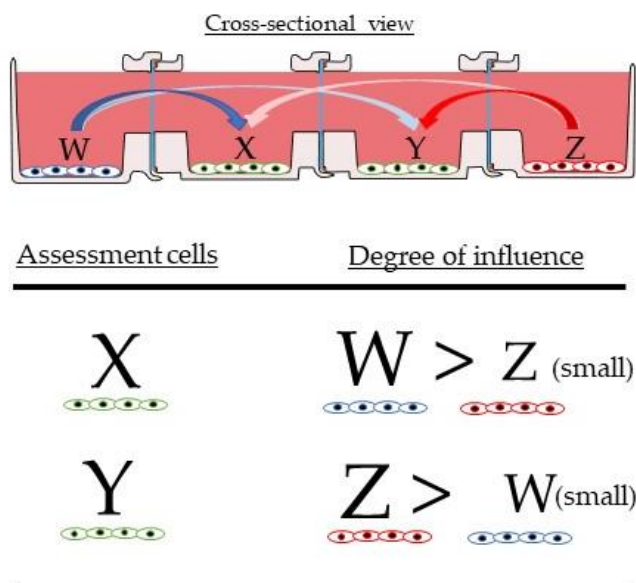


Figure S11. (a) Experimental design 1: A method to check for changes in the cells due to different degrees of influence. Different distances from the cells produce different concentrations, and changes due to different degrees of influence can be observed. (b) Experimental design 2: How to check the change in the two types of cells due to different degrees of influence. For example, suppose X and Y are the same cells, and W and Z are different cells from X and Y. We can confirm that X and Y show different changes owing to the different influences of W and Z.

Filter material/ pore size/ density/ area

Material	Filter Size(Φ) (area)	Pore Size(Φ)	Pore density (Number)/ cm^2	Number of effective pores	Total area of effective pore area	Ratio of open pore area to filter area
Polycarbonate	13mm (1.327 cm^2)	0.03 μm	6.00 $\times 10^8$	4.00 $\times 10^8$	2.71 $\times 10^{-3}$ cm^2	0.424%
Polycarbonate	13mm (1.327 cm^2)	0.6 μm	4.00 $\times 10^8$	2.55 $\times 10^8$	7.22 $\times 10^{-1}$ cm^2	11.311%

Table S1. The density and number of pores in each filter were described. The area of the filter pores as a percentage of the effective filter area in the passage was calculated and expressed as the percentage of open pores.

(A) $y = 6E-05X^2 + 0.0027X + 0.0043$	$R^2 = 0.9999$
(B) $y = 5E-05X^2 + 0.0004X - 0.0014$	$R^2 = 0.9999$
(C) $y = 4E-05X^2 + 0.0001X - 0.0038$	$R^2 = 0.9987$
(D) $y = 4E-05X^2 + 0.0002X + 0.0062$	$R^2 = 0.9992$
(E) $y = 5E-05X^2 + 0.0005X + 0.0154$	$R^2 = 1$
(F) $y = 4E-05X^2 + 0.0004X + 0.0065$	$R^2 = 1$
(G) $y = -2E-06X^3 + 0.0003X^2 - 0.0065X + 0.0368$	$R^2 = 1$
(H) $y = -1E-06X^3 + 0.0002X^2 - 0.0046X + 0.04$	$R^2 = 1$
(I) $y = -2E-06X^3 + 0.0002X^2 - 0.0056X + 0.0325$	$R^2 = 1$
(J) $y = -2E-06X^3 + 0.0002X^2 - 0.007X + 0.0333$	$R^2 = 1$
(K) $y = -2E-06X^3 + 0.0002X^2 + 0.0003X + 0.139$	$R^2 = 1$
(L) $y = -3E-06X^3 + 0.0003X^2 - 0.004X + 0.1603$	$R^2 = 1$
(M) $y = -1E-06X^3 + 0.0001X^2 - 0.0002X + 0.1397$	$R^2 = 1$
(N) $y = -2E-06X^3 + 0.0002X^2 - 0.0029X + 0.1461$	$R^2 = 1$
(O) $y = -9E-07X^3 + 0.0001X^2 + 0.0013X + 0.1251$	$R^2 = 1$
(P) $y = -2E-06X^3 + 0.0002X^2 - 0.0029X + 0.149$	$R^2 = 1$
(Q) $y = -2E-06X^3 + 0.0003X^2 - 0.0045X + 0.1574$	$R^2 = 1$
(R) $y = -2E-06X^3 + 0.0003X^2 - 0.0041X + 0.1582$	$R^2 = 1$
(S) $y = -3E-06X^3 + 0.0003X^2 - 0.0053X + 0.1476$	$R^2 = 1$
(T) $y = -2E-06X^3 + 0.0003X^2 - 0.0046X + 0.1455$	$R^2 = 1$

Y(g/L) X (Hour)

Table S2. The list of correlation approximation equations and correlation coefficients is shown in Figure 6.

Condition	Figure No.	Objects	Location	Time															
				0 h	6 h	24 h	48 h	72 h	6 h	24 h	48 h	72 h							
				mean	mean	mean	mean	mean	S.E.	S.E.	S.E.	S.E.							
Medium	Figure 4.	Glucose 0.6µm	A	200	119.54766	99.363057	97.160883	97.121212		1.6	1.0	0.3							
		Glucose 0.6µm	B	0	84.006462	94.267516	95.741325	96.969697		1.3990717	0.7297095	0.2731941							
		Glucose 0.6µm	C	300	169.67419	110	96.897375	93.693694		3.7089345	1.6898057	1.8018698							
		Glucose 0.6µm	D	0	93.233083	93.658537	93.556086	91.441441		4.186289	2.195122	1.6535091							
		Glucose 0.6µm	E	0	46.115288	82.682927	90.930788	89.864865		0.4340979	1.2673542	0.7159905							
		Glucose 0.6µm	F	0	84.210526	96.097561	97.852029	95.720721		0	1.1177014	1.0936935							
		Glucose 0.6µm	G	300	139.84962	101.70732	98.806683	95.720721		1.5037594	1.2673542	1.4319809							
		Glucose 0.6µm	H	0	83.9599	96.341463	98.329336	95.495495		0.4340979	1.5231702	1.8018698							
				mean	mean	mean	mean	mean		S.E.	S.E.	S.E.							
		Glucose 0.03µm	I	200	192.73021	166.40127	129.49527	115.60606		1.008885	0.988529	2.8522305							
		Glucose 0.03µm	J	0	11.470113	37.738854	65.457413	82.121212		0.2798143	0.8274128	4.8564051							
		Glucose 0.03µm	K	300	294.48622	247.07317	189.02148	156.53153		2.4169551	1.5231702	6.9047619							
		Glucose 0.03µm	L	0	18.045113	55.853659	80.668258	89.414414		2.2556391	10.094733	5.8314042							
		Glucose 0.03µm	M	0	4.3902439	27.684964	43.918919			0	2.6382083	5.3739047							
with Cell	Figure 5.	Lactate 0.6µm	A		0.0233333	0.1	0.2666667	0.4933333		0.008165	0.0141421	0.0285774							
		Lactate 0.6µm	B		0.0033333	0.0366667	0.14	0.3		0.0040825	0.008165	0.0254951							
		Lactate 0.6µm	C		0	0.0166667	0.09	0.1933333		0	0.0040825	0.0254951							
		Lactate 0.6µm	D		0.01	0.03	0.11	0.2266667		0	0	0.0070711							
		Lactate 0.6µm	E		0.02	0.0566667	0.1566667	0.3166667		0	0.0040825	0.0108012							
		Lactate 0.6µm	F		0.01	0.0366667	0.11	0.2266667		0	0.0040825	0.0122474							
		Lactate 0.6µm	G		0.0066667	0.0066667	0.13	0.2566667		0.0040825	0.008165	0.0141421							
		Lactate 0.6µm	H		0.02	0.0366667	0.1733333	0.34		0.0070711	0.0177951	0.0285774							
		Lactate 0.6µm	I		0.0066667	0.0066667	0.11	0.21		0.0040825	0.0108012	0.0212132							
		Lactate 0.6µm	J		0	-0.016667	0.07	0.1466667		0	0.0040825	0.0141421							
				mean	mean	mean	mean	mean		S.E.	S.E.	S.E.							
		NH4+ 0.6µm	K		0.1466667	0.2266667	0.3933333	0.5133333		0.0040825	0.0177951	0.0470815							
		NH4+ 0.6µm	L		0.1466667	0.2	0.3566667	0.4		0.0040825	0.0141421	0.0204124							
		NH4+ 0.6µm	M		0.1433333	0.1966667	0.3066667	0.3666667		0.0040825	0.0147196	0.0285774							
		NH4+ 0.6µm	N		0.1366667	0.1833333	0.32	0.3833333		0.008165	0.0147196	0.0070711							
		NH4+ 0.6µm	O		0.1366667	0.2066667	0.3366667	0.4366667		0.0108012	0.0227303	0.0040825							
		NH4+ 0.6µm	P		0.14	0.19	0.33	0.3866667		0	0.0070711	0.0187083							
		NH4+ 0.6µm	Q		0.14	0.18	0.3333333	0.42		0.0470815	0.0141421	0.0147196							
		NH4+ 0.6µm	R		0.1433333	0.1933333	0.3633333	0.4666667		0.0749487	0.0070711	0.0318852							
		NH4+ 0.6µm	S		0.1266667	0.1666667	0.3233333	0.3833333		0.0441588	0.0122474	0.0108012							
		NH4+ 0.6µm	T		0.1266667	0.1533333	0.2766667	0.3333333		0.0681909	0.0040825	0.0326599							
Medium	Figure 7.			mean	mean	mean	mean	mean		S.E.	S.E.	S.E.							
		EV in VTCP 1.0µm					7%	16%	19%			0%							
		EV in HTCP 1.0µm					23%	34%	41%			3%							
Medium	Supplemental Figure A7			0 h	6 h	24 h	48 h	72 h	6 h	24 h	48 h	72 h							
				mean	mean	mean	mean	mean	S.E.	S.E.	S.E.	S.E.							
		NH4+ 0.6µm	A	200	108.49673	100	98.484848	100.72402		1.1320594	1.0188534	0							
		NH4+ 0.6µm	B	0	93.464052	98.235294	98.484848	100.8092		1.1320594	1.0188534	1.5151515							
		NH4+ 0.6µm	C	300	135.45455	100.76923	94.83871	97.086777		4.1659779	2.6646936	0							
		NH4+ 0.6µm	D	0	99.090909	94.615385	93.548387	95.971074		4.1659779	2.3076923	1.174521							
		NH4+ 0.6µm	E	0	76.363636	90.769231	92.903226	95.929752		2.7272727	1.3323468	0							
		NH4+ 0.6µm	F	0	92.727273	91.538462	90.967742	96.714876		0	1.3323468	1.9354839							
		NH4+ 0.6µm	G	300	112.72727	93.076923	90.967742	96.301653		1.5745916	2.6646936	1.9354839							
		NH4+ 0.6µm	H	0	93.636364	91.538462	92.258065	96.528926		1.5745916	1.3323468	2.9565004							
				mean	mean	mean	mean	mean		S.E.	S.E.	S.E.							
		NH4+ 0.03µm	I	200	149.6732	109.41176	104.54545	100.91567		1.1320594	4.6689729	1.5151515							
		NH4+ 0.03µm	J	0	51.633987	80.588235	96.464646	99.63799		1.1320594	7.9574996	2.3144322							
		NH4+ 0.03µm	K	300	205.45455	135.38462	123.87097	99.380165		1.5745916	6.6617339	12.087093							
		NH4+ 0.03µm	L	0	68.181818	80	101.93548	97.004132		2.7272727	4.8038446	8.9396171							
		NH4+ 0.03µm	M	0	31.818182	48.461538	84.516129	95.929752		1.5745916	20.117995	10.659814							

Table S3. Raw data of the results An Excel file is available online from the MDPI.