

Article

## Molecular Cloning and Pharmacological Properties of an Acidic PLA<sub>2</sub> from *Bothrops pauloensis* Snake Venom

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**Abstract:** In this work, we describe the molecular cloning and pharmacological properties of an acidic phospholipase A<sub>2</sub> (PLA<sub>2</sub>) isolated from *Bothrops pauloensis* snake venom. This enzyme, denominated BpPLA<sub>2</sub>-TXI, was purified by four chromatographic steps and represents 2.4% of the total snake venom protein content. BpPLA<sub>2</sub>-TXI is a monomeric protein with a molecular mass of 13.6 kDa, as demonstrated by Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF) analysis and its theoretical isoelectric point was 4.98. BpPLA<sub>2</sub>-TXI was catalytically active and showed some pharmacological effects such as inhibition of platelet aggregation induced by collagen or ADP and also induced edema and myotoxicity. BpPLA<sub>2</sub>-TXI displayed low cytotoxicity on TG-180 (CCRF S 180 II) and Ovarian Carcinoma (OVCAR-3), whereas no cytotoxicity was found in regard to MEF (Mouse Embryonic Fibroblast) and Sarcoma 180 (TIB-66). The N-terminal sequence of forty-eight amino acid residues was determined by

Edman degradation. In addition, the complete primary structure of 122 amino acids was deduced by cDNA from the total RNA of the venom gland using specific primers, and it was significantly similar to other acidic D49 PLA<sub>2</sub>s. The phylogenetic analyses showed that BpPLA<sub>2</sub>-TXI forms a group with other acidic D49 PLA<sub>2</sub>s from the genus *Bothrops*, which are characterized by a catalytic activity associated with anti-platelet effects.

**Keywords:** *Bothrops pauloensis*; phospholipase A<sub>2</sub>; molecular cloning

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## 1. Introduction

*Bothrops pauloensis* is a venomous snake widely distributed throughout the Brazilian territory, except in the Amazonian region of Brazil. This species is particularly common in the Triangulo Mineiro region, in the southwest of Minas Gerais State [1].

Rodrigues *et al.* [2] described the repertoire of venom toxins from *B. pauloensis* by snake proteomics and venom gland transcriptomic surveys. The main toxins present in *B. pauloensis* snake venom include metalloproteinases, phospholipases A<sub>2</sub>, serine proteinases, L-amino acid oxidases, disintegrins, nucleotidases and hyaluronidases, among others [2]. Both approaches indicated metalloproteinases, vasoactive (bradykinin-potentiating) peptides and phospholipases A<sub>2</sub> as the major toxin classes.

Phospholipases A<sub>2</sub> (PLA<sub>2</sub>) (E.C. 3.1.1.4) represent a superfamily of lipolytic enzymes that catalyze the hydrolysis of the 2-acyl ester of the phospholipids, releasing free fatty acids and lysophosphatids [3,4]. PLA<sub>2</sub>s are classified into 15 groups that are further subdivided into several groups (sPLA<sub>2</sub>—secreted; cPLA<sub>2</sub>—cytosolic; iPLA<sub>2</sub>—Ca<sup>2+</sup> independent; LpPLA<sub>2</sub>—lipoprotein-associated), all of which display differences in amino acid sequence, disulfide bonds, tissue specificity and cellular functions [5,6].

PLA<sub>2</sub>s are present in snake venoms and are characterized by low molecular mass (13–18 kDa), histidine at the catalytic site, Ca<sup>2+</sup> bound at the active site, as well as six conserved disulfide bonds with one or two variable disulfide bonds [7]. These PLA<sub>2</sub>s can be divided into two groups, I and II, whereby the second is subdivided into two classes, and are present in snake venoms from the Viperidae family. These classes are D49 PLA<sub>2</sub>s, which display an Asp residue at position 49, with high catalytic activity upon artificial substrates; and Lys49 PLA<sub>2</sub>s, that presents a Lys residue at position 49, with low or no catalytic activity [8,9].

The different PLA<sub>2</sub>s isoforms that have already been isolated from *B. pauloensis* snake venom include: BnpTX-I and BpnTX-II (D-49 basic) [10], BnSP-6 and BnSP-7 (K-49 basic) [11] and Bp-PLA<sub>2</sub> (D-49 acidic) [12]. These toxins present some toxic and/or pharmacological effects characterized as neurotoxicity, myotoxicity, cytotoxicity, inhibition of platelet aggregation, anticoagulation, edema, convulsion and hypotension [3,8,9]. The present study describes the molecular cloning, as well as the enzymatic and pharmacological properties of an acidic phospholipase A<sub>2</sub> from *B. pauloensis* snake venom.

## 2. Results and Discussion

An acidic PLA<sub>2</sub> isolated from the venom of *B. pauloensis* was obtained by four chromatographic procedures in the present work. The first step consisted of an ion exchange chromatography on CM-Sepharose column and resulted in six major protein peaks (Figure 1A). The fraction named CM-1, with phospholipase A<sub>2</sub> activity (Table 1), was further fractionated on Sephacryl-S300 HR HiPrep 26/60 (filtration chromatography) and resulted in seven new protein fractions, denominated S1–S7 (Figure 1B). S4, which displayed high PLA<sub>2</sub> activity, was applied on a Hi-Trap Q FF ion-exchange column and resulted in two peaks, Q1 and Q2 (Figure 1C). The Q2 fraction was then applied on a reverse phase high performance liquid chromatography (HPLC) C2–C18  $\mu$ RPC 4.6/100 (GE HealthCare) and the acidic PLA<sub>2</sub>, named BpPLA<sub>2</sub>-TXI, was eluted with approximately 80% of the Solvent B (Figure 1D). BpPLA<sub>2</sub>-TXI was shown to be homogeneous by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) (Figure 1E) with an apparent molecular mass of 15,000 Da in the presence of the reducing agent. Analysis by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF) (Figure 2) of the intact protein revealed a purified protein with a molecular mass of 13,682 Da. The theoretical *pI* of BpPLA<sub>2</sub>-TXI was found to be 4.98, as calculated based on a deduced sequence of cDNA by using the program CLC Sequence Viewer 6 (CLC bio, Aarhus, Denmark; <http://www.clcbio.com/index.php?id=28>), confirming that BpPLA<sub>2</sub>-TXI is an acidic PLA<sub>2</sub>.

**Figure 1.** Sequential purification steps of BpPLA<sub>2</sub>-TXI. **(A)** *Bothrops pauloensis* venom (170 mg) on CM-Sepharose Fast Flow previously equilibrated with ammonium bicarbonate (AMBIC) buffer 0.05M pH 7.8. A gradient was then applied up to 0.5 M (AMBIC) buffer, pH 7.8. Fractions of 1 mL/tube were collected in a 6 mL/h flow rate at room temperature; **(B)** CM1 (60 mg) on Sephacryl-S300 previously equilibrated and eluted with AMBIC buffer (0.05 M; pH 7.8) at a flow rate of 12.0 mL/h and fractions of 2.0 mL/tube were collected; **(C)** S4 was rechromatographed on HiTrap Q FF column, equilibrated with AMBIC (0.05 M; pH 7.8). A gradient was then applied up to the column using AMBIC buffer (0.5 M, pH 7.8) at flow rate of 6.0 mL/h and fractions of 1 mL/tube were collected; **(D)** The active fraction (Q2) on reverse phase-high performance liquid chromatography (RP-HPLC) C2–C18 and PLA<sub>2</sub> was eluted using Solvent A (0.1% TFA, 4% acetonitrile) to 100% of the Solvent B (0.1% TFA, 80% acetonitrile) at the flow rate of 0.5 mL/min for 33 min and fractions of 0.5 mL/tube; **(E)** sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) at 12% (w/v). Line 1: molecular mass markers; Line 2: acidic PLA<sub>2</sub> (BpPLA<sub>2</sub>-TXI).

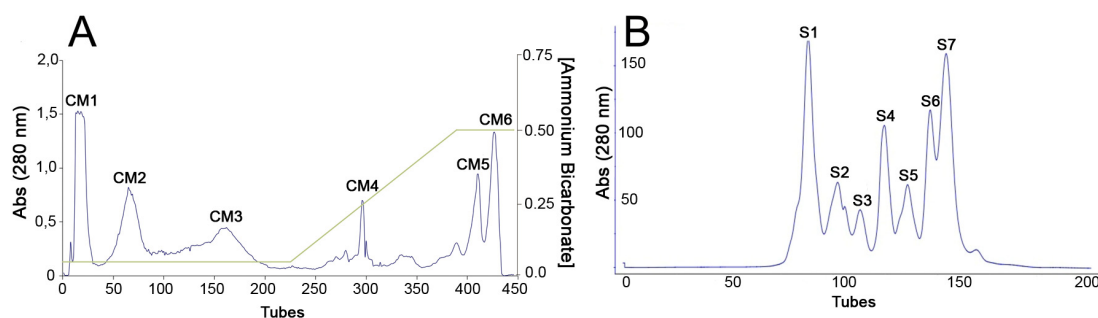


Figure 1. Cont.

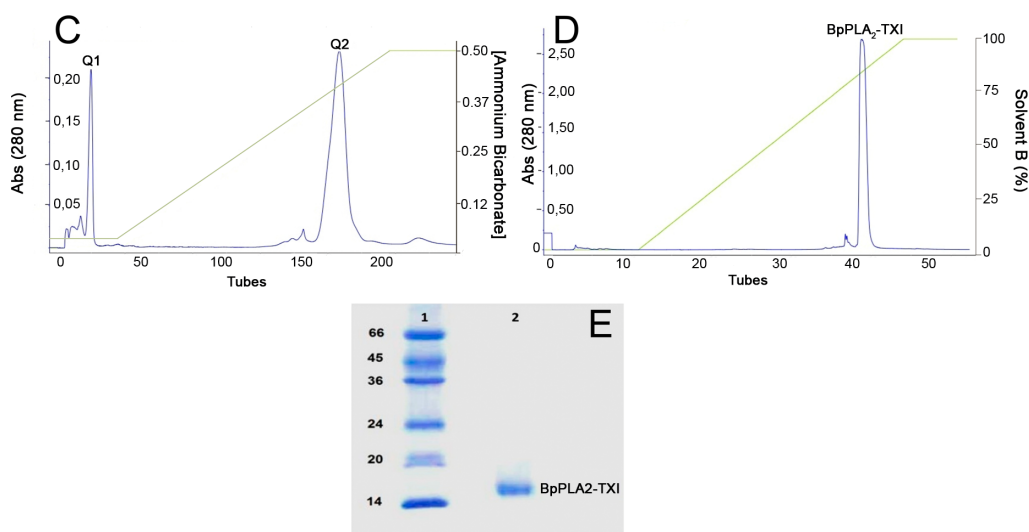


Table 1. Protein yield and recovery of enzymatic activity of the crude venom and fractions.

Fraction	Protein *		Phospholipase activity **		
	mg	% Rec	U/mg	U-total	Purification factor
Crude Venom	170	100	51	8670	1.0
CM1	60	35	87	5220	1.7
S4	20	11.7	113	2260	2.2
Q2	7.5	4.4	135	1012	2.6
BpPLA <sub>2</sub> -TXI	4.5	2.6	142	639	2.78

\* Protein concentration was determined by Bradford (1976); \*\* Phospholipase A<sub>2</sub> activity was determined by DE HAAS (1968).

**Figure 2.** Molecular mass determination of BpPLA<sub>2</sub>-TXI by Matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF) (Bruker Daltonics, Bremen, Germany).

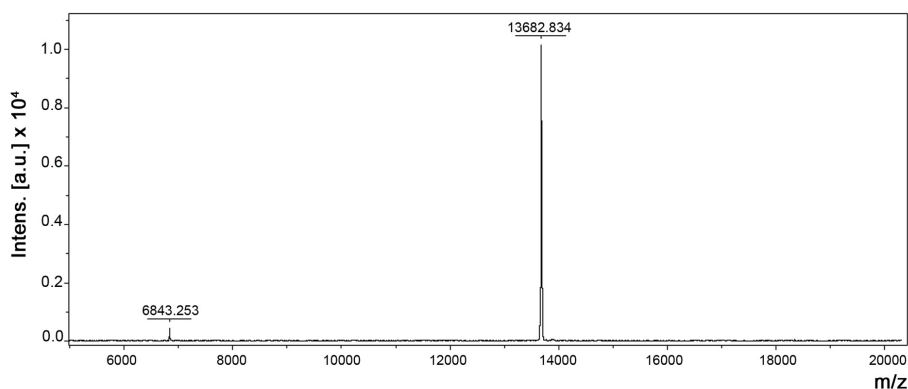
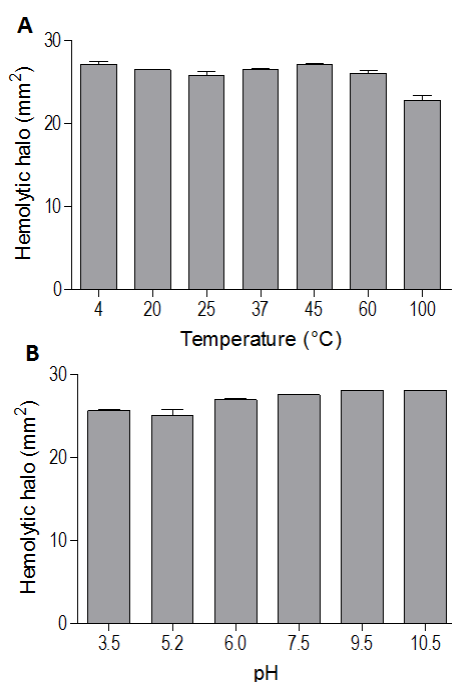


Table 1 shows total protein yield and recovery of PLA<sub>2</sub> activity. BpPLA<sub>2</sub>-TXI exhibited a specific PLA<sub>2</sub> activity of 142 U/mg, three-fold higher than the crude venom (51 U/mg). BpPLA<sub>2</sub>-TXI, an acidic PLA<sub>2</sub> isoenzyme isolated from *B. pauloensis* venom, presented a considerable recovery of the specific phospholipase activity (Table 1). These results are in agreement with previous studies

involving PLA<sub>2</sub> from snake venoms, in which the catalytic activity of D49 acidic PLA<sub>2</sub>s are higher than that related to D49 basic PLA<sub>2</sub>s [8,9,11,12].

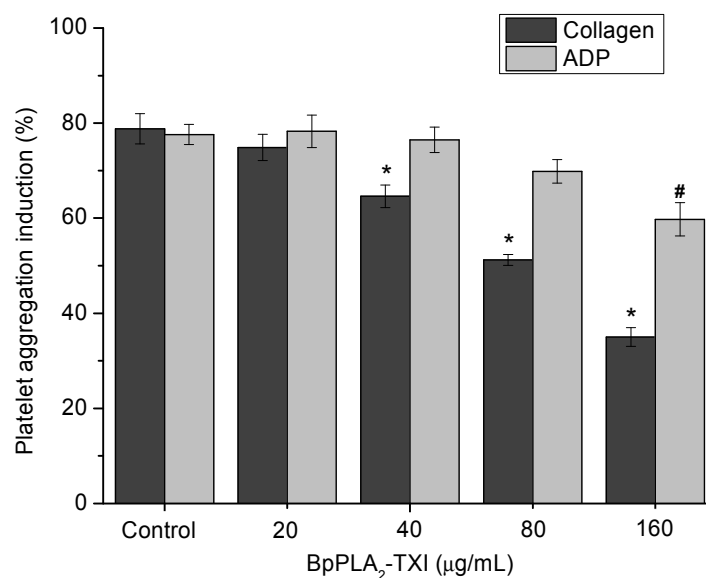
The indirect hemolytic activity was performed to assess the stability of BpPLA<sub>2</sub>-TXI. The protein was stable when submitted to different pH values (3.5, 5.2, 6.0, 7.5, 9.5, 10.5) (Figure 3A) or temperatures (4, 20, 25, 37, 45, 60 and 100 °C) (Figure 3B). However, PLA<sub>2</sub> activity was reduced when BpPLA<sub>2</sub>-TXI was subjected to 100 °C for 30 min, suggesting that denaturation causes loss of protein stability, thereby decreasing its activity. The majority of PLA<sub>2</sub>s from snake venoms are highly stable due to the presence of intra-chain disulfide bonds present in their structures [8,13].

**Figure 3.** Effects of different pH values and temperatures on phospholipase activity induced by Bp-PLA<sub>2</sub>-TxI from *Bothrops pauloensis* snake venom. The indirect hemolysis assay was determined by measuring the hemolytic halo in mm<sup>2</sup> after 24 h of incubation of 5 µg of BpPLA<sub>2</sub>-TXI previously incubated with different buffers (A) and temperatures (B). For all the experiments, data are expressed as means ± SEM (*n* = 3).



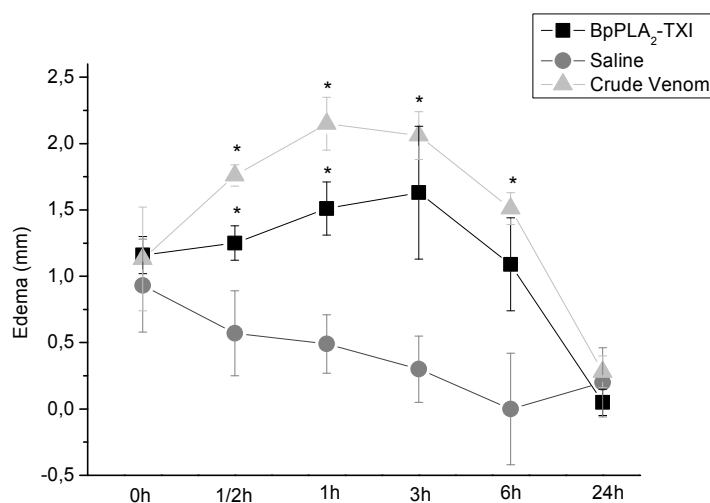
BpPLA<sub>2</sub>-TXI inhibited platelet aggregation induced by collagen and ADP in a dose-dependent manner (Figure 4). Acidic PLA<sub>2</sub>s Asp-49 from several bothropic snake venoms are able to inhibit platelet aggregation induced by physiological agonists such as collagen and ADP [8,14]. The anti-platelet effect induced by PLA<sub>2</sub>s might be due to direct interference of the catalytic site in this activity or because of the C-terminal region of these proteins [8]. Some studies demonstrated that, when PLA<sub>2</sub>s are modified by bromophenacyl bromide (p-BPB), the His48 residue present in the active site is alkylated, resulting in a drastic loss or reduction of the enzymatic activity and/or pharmacological effects of PLA<sub>2</sub>s [8,15–17]. BpPLA<sub>2</sub>-TXI was efficient in inhibiting the platelet aggregation, although further experiments should be conducted to increase the knowledge of the mechanism of action of BpPLA<sub>2</sub>-TX on platelet aggregation.

**Figure 4.** Platelet aggregation inhibition induced by BpPLA<sub>2</sub>-TXI. Different concentrations (20, 40, 80 and 160 µg/mL) of BpPLA<sub>2</sub>-TXI were preincubated at 37 °C for 10 min under stirring with washed platelets ((3–4) × 10<sup>5</sup> cells/µL) and then platelet aggregation was initiated with ADP (20 µM) or collagen (10 µg/mL). Results are presented as percent of platelet aggregation (mean ± S.D., *n* = 3). Statistically significant results compared to collagen (\*) or ADP (#) (*p* < 0.05) are shown.



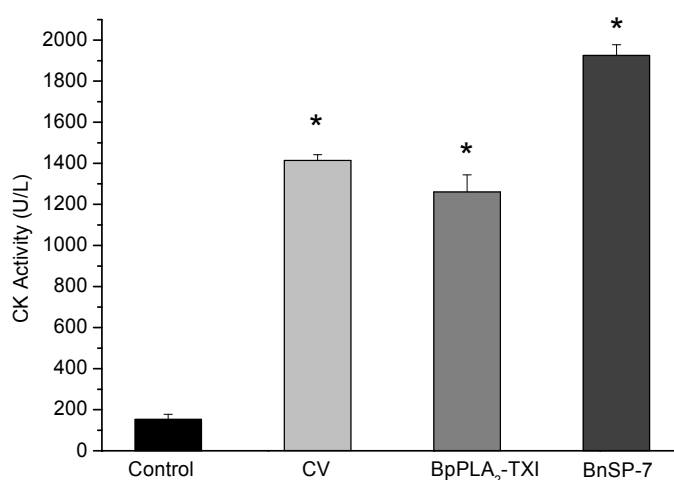
BpPLA<sub>2</sub>-TXI also induced mouse paw edema (Figure 5), which was more extensive 3 h after administration of the toxin. Some D49 acidic phospholipase A<sub>2</sub> forms have been characterized and have also shown similar effects [11,12,14]. Several factors may influence edema induction by these toxins that catalyze the breakdown of the phospholipids and release arachidonic acid, a precursor of eicosanoids, which can mediate the inflammatory response and induce edema [18].

**Figure 5.** Edema-inducing activities of BpPLA<sub>2</sub>-TXI or crude venom. Paw edema in Swiss mice after injection of BpPLA<sub>2</sub>-TXI or crude venom (10 µg/50 µL Saline). Results are reported as mean ± S.D. (*n* = 3). (\*) Statistically significant results compared to saline (*p* < 0.05) are shown.



BpPLA<sub>2</sub>-TXI induced significant increases in the plasma CK (creatine kinase) levels 3h after administration of the toxin in the gastrocnemius muscle. However, this effect was reduced when compared with CK levels induced by BnSP-7, a basic K49 PLA<sub>2</sub> (Figure 6). Some D49 PLA<sub>2</sub>s do not present myotoxicity [12,19,20], but some studies have reported that these toxins may injure muscle fibers to a lesser degree [11,21].

**Figure 6.** Myotoxic activity of BpPLA<sub>2</sub>-TXI in mice. Animals were injected i.m. with crude venom. Acidic BpPLA<sub>2</sub>-TXI or basic BnSP-7 (20 µg/50 µL Saline) and plasma CK activity was measured 3h after injection. Control mice were injected with only saline. Results are reported as mean ± S.D. (*n* = 3). (\*) Statistically significant results compared to saline (*p* < 0.05) are shown.



The mechanism by which these toxins induced the myotoxicity is not precisely defined. Some works suggest an independent mechanism from the catalytic site, provoking a disturbance of the sarcolemma through the insertion of the toxin in the C-terminal region inside the plasmatic membrane, leading to influx of calcium and consequent muscular necrosis [22]. However, other authors who have accomplished modification in the PLA<sub>2</sub> with *p*-BPB have observed a loss or decrease of myotoxic activity [21], suggesting that myotoxicity of the acidic D49 PLA<sub>2</sub> is directly related to the catalytic site.

The cytotoxicity of the BpPLA<sub>2</sub>-TXI on some tumor cells was evaluated on MEF (Mouse Embryonic Fibroblast), Sarcoma 180 (TIB-66), TG-180 (CCRF S 180 II) and Ovarian Carcinoma (OVCAR-3). BpPLA<sub>2</sub>-TXI presented low cytotoxicity against TG-180 (CCRF S 180 II) and Ovarian Carcinoma (OVCAR-3) and did not show such activity against MEF (Mouse Embryonic Fibroblast) and Sarcoma 180 (TIB-66) (data not shown). Several authors have reported that some basic snake venom PLA<sub>2</sub>s produce high cytotoxic activity against tumor cells [23,24]. Meanwhile, other studies have revealed that the acidic PLA<sub>2</sub> present low or no cytotoxicity [8,19].

The sequence of the first 48 amino acid residues from BpPLA<sub>2</sub>-TXI was determined by Edman degradation (Figure 7) and the complete sequence was deduced further by cDNA. This sequence was recorded in the GenBank under Accession No. JK998826.

**Figure 7.** Complete sequence of the cDNA coding BpPLA<sub>2</sub>-TXI. The N-terminal sequence of the mature protein obtained by the Edman method is underlined. The initial region corresponds to the signal peptide. The symbol (\*) represents the stop codon.

>BpPLA <sub>2</sub> -TXI	
1	ATG AGG ACT CTC TGG ATA ATG GCC GTG TTG CTG GTG GGC GTC GAG 45
1	<u>M R T L W I M A V L L V G V E</u> 15
46	GGG AAC CTG GTG CAA TTT GAG ACG TTG ATC ATG AAA ATT GCG GGG 90
16	<u>G N L V Q F E T L I M K I A G</u> 30
91	AGA AGT GGT GTT TGG TAC TAC GGC TCT TAC GGA TGC TAC TGC GGC 135
31	<u>R S G V W Y Y G S Y G C Y C G</u> 45
136	TCG GGA GGC CAA GGC CGG CCA CAG GAC GCC AGC GAC CGC TGC TGC 180
46	<u>S G G Q G R P Q D A S D R C C</u> 60
181	TTT GTG CAC GAC TGC TGT TAT GGA AAA GTG ACC GAC TGC GAC CCC 225
61	<u>F V H D C C Y G K V T D C D P</u> 75
226	AAA GCG GAC GTC TAC ACC TAC AGC GAG GAG AAC GGG GTT GTC GTC 270
76	K A D V Y T Y S E E N G V V V 90
271	TGC GGA GGG GAC GAC CCG TGC AAG AAG CAG ATT TGT GAG TGC GAC 315
91	C G G D D P C K K Q I C E C D 105
316	AGG GTT GCG GCA ACC TGC TTC CGA GAC AAT AAG GAC ACA TAC GAA 360
106	R V A A T C F R D N K D T Y D 120
361	AAC AAA TAT TGG TTT TCC CCG GCC AAA AAT TGC CAG GAG GAA TCA 405
121	N K Y W V F P A K N C Q E E S 135
406	GAG CCA TGC TAA 417
136	E P C *

The nucleotide sequence of the BpPLA<sub>2</sub>-TXI showed 417 bp, codifying a mature protein of 122 amino acid residues (Figure 7). These results are in agreement with other works concerning acidic PLA<sub>2</sub>s, which presents 120–135 amino acid residues and 400–500 bp [9,19,20,25]. Analysis of the PLA<sub>2</sub> primary sequence reveals that this enzyme possesses not only an Asp residue in the 49th position (D49) but also many other conserved residues (24YGCYCGWGG32; H48) involved at the Ca<sup>2+</sup> binding and catalytic sites.

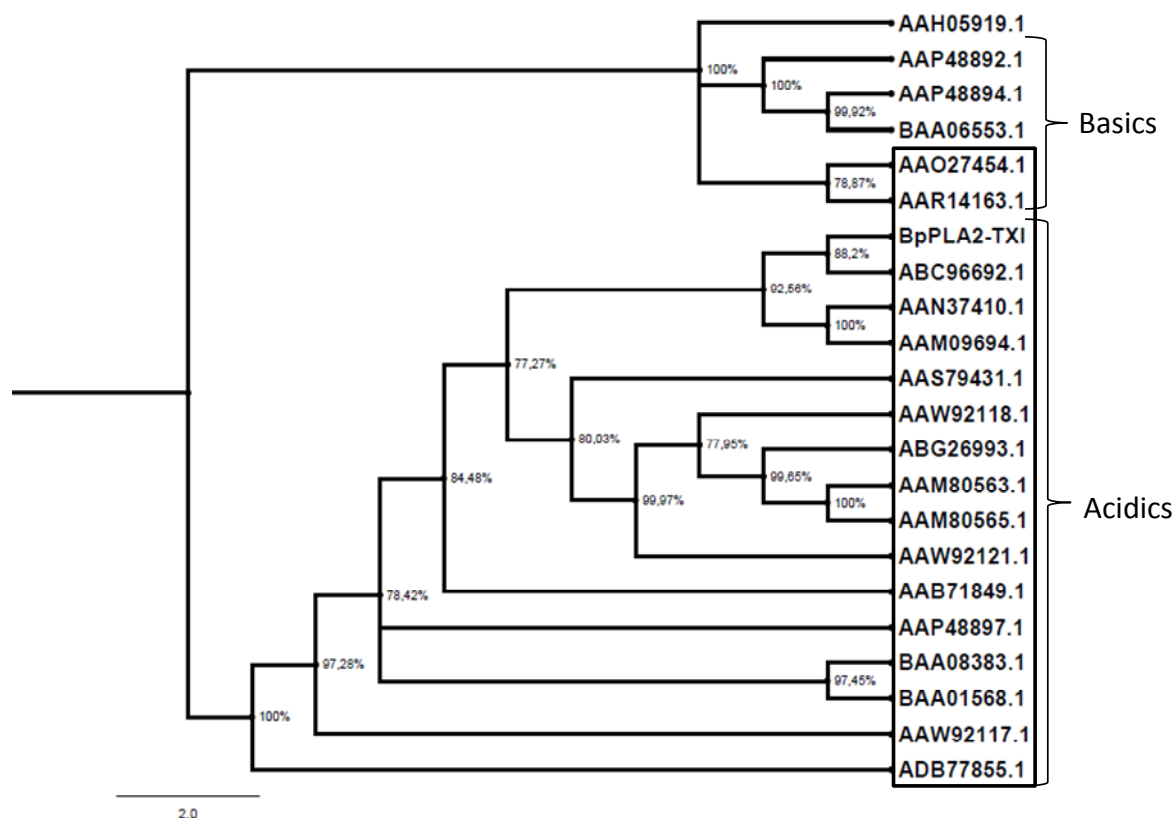
BpPLA<sub>2</sub>-TXI was aligned with other PLA<sub>2</sub> sequences from *B. pauloensis* (results not shown). High identity was visualized (about 77%) with Bp-PLA<sub>2</sub> [11], as expected, since both are acidic D49 PLA<sub>2</sub>s. Multiple alignments of the BpPLA<sub>2</sub>-TXI sequence with other acidic D49 PLA<sub>2</sub>s are shown in Figure 8. The best alignment scores pointed by ClustalW were with PLA<sub>2</sub> from *Bothrops erythromelas* and *Sistrurus catenatus*, representing 99% and 89%, respectively. BpPLA<sub>2</sub>-TXI possesses many conserved domains that are common to D49 PLA<sub>2</sub>s, including 14 cystein residues which form seven disulfide bonds, and presents other residues involved with the calcium binding and catalytic sites as well. According to the results published by Rodrigues *et al.* [11] and of the present study, we suggest that Bp-PLA<sub>2</sub> and BpPLA<sub>2</sub>-TXI may be isoforms present in *B. pauloensis* snake venom.





The PLA<sub>2</sub> gene is involved in an accelerated evolution by substitutions in protein-encoding regions and appears to be universal for all the PLA<sub>2</sub>-encoding genes in snake venom glands of the crotaline snakes [26–29]. As shown in Figure 9, BpPLA<sub>2</sub>-TXI, together with other acidic PLA<sub>2</sub>s, formed a separate group from basic PLA<sub>2</sub>s, suggesting that the replacement of the D49 by the K49 residue is a derived characteristic, although additional studies need to be accomplished in order to elucidate this aspect. Tsai *et al.* [30], when isolating and characterizing several PLA<sub>2</sub>s from *Crotalus viridis viridis* snake venoms from different regions, showed that these acidic PLA<sub>2</sub>s were capable of inhibiting platelet aggregation and presented a glutamate (E) residue at Position 6. BpPLA<sub>2</sub>-TXI also possesses the E6 residue and is able to inhibit platelet aggregation.

**Figure 9.** Evolutionary PLA<sub>2</sub> relationships. The evolutionary history was inferred using the neighbor-joining method. Phylogenetic analyses were conducted by the program MrBayes. The PLA<sub>2</sub>s from snake venoms (deposited in GenBank) were grouped into two branches (acidic and basic); those shown inside the box are D49 PLA<sub>2</sub> and those outside the box are K49. Human Synovial PLA<sub>2</sub> (NCBI: AAH05919.1) sequence was included as outside group.



### 3. Experimental Section

#### 3.1. Material and Animals

*B. pauloensis* dried crude venom was purchased from Serpentarium Bioagents (Batatais, SP, Brazil) and maintained at -20 °C. This serpentarium is registered with the Brazilian Institute of the Environment and Renewable Natural Resources (n 471301). Male Swiss mice (18–25 g) were obtained from and maintained in the Center for Animal Experimentation and Housing (CEBEA) at the Federal

University of Uberlândia (UFU), Uberlândia, MG, Brazil. Animal experimentation procedures were approved by the UFU Ethics Committee for Animal Utilization (protocol number 046/09). CK-Nac Kit was purchased from Analisa (Belo Horizonte, Brazil). Adenosine diphosphate (ADP) and Collagen were acquired from Helena Laboratories (Beaumont, TX, USA). CM-Sepharose resin was purchased from Amersham Biotec (São Paulo, Brazil). All other reagents used were of analytical grade.

### 3.2. Isolation of Acidic Phospholipase A<sub>2</sub>

The acidic PLA<sub>2</sub> was isolated by a combination of an ion exchange chromatography on CM-Sepharose fast flow [31], a gel filtration Sephacryl S-300 [32] and an ion exchange Capto-Q column (Amersham Biotec, São Paulo, Brazil), according to methods previously standardized by our research group. Initially, a sample containing 170 mg of desiccated *B. pauloensis* venom was dispersed into 2.0 mL of 0.05 M ammonium bicarbonate (AMBIC, Sigma, St. Louis, MO, USA) buffer, pH 7.8, cleared by centrifugation at 10,000 × *g* for 10 min at 4 °C and submitted to CM-Sepharose Fast Flow column (2.0 × 20 cm) which was previously equilibrated and initially eluted with the same buffer. A linear gradient was then applied up to 0.05 M Ambic buffer, pH 7.8 and fractions of 1 mL/tube were collected at a flow rate of 6.6 mL/h.

The fraction with PLA<sub>2</sub> activity (CM1) was collected and gel filtrated on a Sephacryl S-300 HR HiPrep 26/60 (GE HealthCare, Uppsala, Sweden), equilibrated and eluted with 0.05 M Ambic buffer, pH 7.8. Fractions of 2.0 mL/tube were collected at a flow rate of 12.0 mL/h. The fraction with PLA<sub>2</sub> activity (S4) was further submitted to an anionic exchange HiTrap Q FF column (GE HealthCare, Uppsala, Sweden) equilibrated with 0.05 M AMBIC buffer, pH 7.8. A linear gradient was then applied up to 0.5 M AMBIC buffer (AMBIC, Sigma, St. Louis, MO, USA), pH 7.8 and fractions of 1.0 mL/tube were collected at a flow rate of 6.0 mL/h. The active fraction (Q2) was rechromatographed by reverse-phase chromatography on a C2–C18 μRPC 4.6/100 (GE HealthCare, Uppsala, Sweden) column, previously equilibrated with solvent A (0.1% trifluoroacetic acid and 4% acetonitrile). The elution of the protein was conducted using a linear gradient of solvent B (0.1% trifluoroacetic acid and 80% acetonitrile) from 0% to 100%, at a flow rate of 0.5 mL/tube for 33 min. All steps of the purification procedure were carried out at room temperature (25 °C). The acidic PLA<sub>2</sub> derived from RP-HPLC, named BpPLA<sub>2</sub>-TXI, was lyophilized and used for biochemical characterization and pharmacological studies.

### 3.3. Determination of Mr

Polyacrylamide gel electrophoresis (PAGE), in the presence of sodium dodecyl sulfate (SDS-PAGE), was performed according to Laemmli [33]. Samples were heated at 100 °C for 5 min and run under reducing (10% β-mercaptoethanol) and non-reducing conditions. The gel was stained with Coomassie Brilliant Blue R-250. The *Mr* was estimated by interpolation from a linear logarithmic plot of relative molecular mass *versus* migration distance. The molecular mass markers used were: bovine serum albumin (66 kDa), ovalbumin (45 kDa), glyceraldehyde-3-phosphate dehydrogenase (36 kDa), carbonic anhydrase (29 kDa), trypsinogen (24 kDa), trypsin inhibitor (20 kDa) and α-lactalbumin (14.2 kDa) (Amersham Biosciences, São Paulo, Brazil).

### 3.4. MALDI TOF Mass Spectrometry

BpPLA<sub>2</sub>-TXI was also submitted to mass spectrometric analysis, using an AutoFlex III MALDI-TOF mass spectrometer (Bruker Daltonics, Bremen, Germany) controlled by the software FlexControl 3.0 (Bruker Daltonics, Bremen, Germany). The sample was mixed with sinapinic acid matrix solution (1:1, v/v) directly onto a target plate (Bruker Daltonics, Bremen, Germany) and dried at room temperature. The mean mass of the protein was obtained in linear mode with external calibration, using Protein Calibration Standard (Bruker Daltonics, Bremen, Germany). The software Flex Analysis 3.0 (Bruker Daltonics, Bremen, Germany) was used for analysis of mass spectrometric data.

### 3.5. Phospholipase A<sub>2</sub> Activity

Phospholipase activity of BpPLA<sub>2</sub>-TXI was determined upon egg-yolk emulsion according to [34] and by the indirect hemolysis method, using washed mice erythrocytes and hen's egg-yolk emulsion as substrate as described by [35].

BpPLA<sub>2</sub>-TXI stability at various pH intervals (50 mM ammonium formate pH 3.5; 50 mM sodium acetate pH 5.2; pH 6.0; 50 mM Tris HCl pH 7.5, 9.5 and 50 mM sodium borate pH 10.5) and temperatures (4 °C, 20 °C, 25 °C, 37 °C, 45 °C, 60 °C and 100 °C) was carried out as described by [35]. For this assay, 5 µg of BpPLA<sub>2</sub>-TXI were first incubated for 30 min with 50 µL of different buffer solutions at various pH intervals and temperatures, and the enzymatic assays were subsequently performed as described above [35].

### 3.6. Platelet Aggregation

The platelet aggregation was determined by using the method previously described by Fuly *et al.* [21], with modifications. Platelet-rich plasma (PRP) was prepared from mouse blood collected in the presence of citrate (0.31% w/v) and centrifuged at 360 × g for 15 min at room temperature. Washed platelets (WP) were prepared by centrifugation of PRP at 1800 × g for 12 min at room temperature. Platelets were resuspended in a Tyrode-BSA buffer and the final pH was adjusted to 7.5. Platelets were counted and their density adjusted to 3–4 × 10<sup>5</sup> cells/µL. The platelet aggregation was measured turbidimetrically by AggRAM aggregometer (Helena Laboratories, Beaumont, TX, USA). WP suspended in Tyrode-BSA buffer (225 µL) were previously incubated at 37 °C for 2 min under stirring with 1 mM CaCl<sub>2</sub> (final concentration). Aggregation was triggered with agonists after preincubation of platelets with different concentrations of PLA<sub>2</sub> (20, 40, 80 and 160 µg/mL) at 37 °C for 30 min. Agonists ADP (20 µM) and collagen (10 µg/mL) (Helena Laboratories, Beaumont, TX, USA) were used as positive controls of reaction and their concentrations were determined according to the manufacturer's instructions.

### 3.7. Edema-Inducing Activity

Crude venom (10 µg/50 µL saline), BpPLA<sub>2</sub>-TXI (10 µg/50 µL saline) or saline were injected into the subplantar region of male Swiss mice (18–22 g, n = 5). After 0.5, 1, 2, 3, 6 and 24 h, paw edema was measured with the aid of a low-pressure spring caliper (Mitutoyo, Tokyo, Japan). Zero time values were then subtracted and the differences reported as mean ± S.D.

### 3.8. Myotoxic Activity

Four groups of male Swiss mice (18–25 g,  $n = 4$ ) were injected i.m. into the right *gastrocnemius* muscle with saline, crude venom (20  $\mu\text{g}/50 \mu\text{L}$  Saline), BpPLA<sub>2</sub>-TXI (20  $\mu\text{g}/50 \mu\text{L}$  Saline) or BnSP-7 (20  $\mu\text{g}/50 \mu\text{L}$ ), a Lys49 PLA<sub>2</sub> previously isolated from *B. pauloensis* snake venom according to Soares *et al.* [18]. The control was 0.9% NaCl. After 3 h, the animals were anesthetized (ketamine<sup>®</sup> 10% (0.05 mL/kg) + xylazine<sup>®</sup> 2% (0.025 mL/kg)) and the blood was collected by cardiac puncture in heparin-coated tubes and centrifuged ( $2500 \times g$  for 10 min at 4 °C) for separation of the serum. The amount of creatine kinase (CK) was then determined with 50  $\mu\text{L}$  of serum, which was incubated for 3 min at 37 °C with 1.0 mL of the reagent according to the CK-Nac protocol (Gold Analisa Diagnóstica LTDA, Belo Horizonte, Brazil). Activity was expressed in U/L, with one unit resulting from the phosphorylation of 1  $\mu\text{mole}$  of creatine/min at 25 °C.

### 3.9. Cell Culture

MEF (Mouse Embryonic Fibroblast), Sarcoma 180 (TIB-66), TG-180 (CCRF S 180 II) and human ovarian carcinoma (OVCAR-3) cancer cell lines were maintained on RPMI 1640 medium supplemented with 2 mM L-glutamine, 1.5 g/L sodium bicarbonate, 4.5 g/L glucose, 10 mM HEPES, 1.0 mM sodium pyruvate, 10% fetal bovine serum and 10  $\mu\text{g}/\text{mL}$  gentamicin. All cell culture reagents were purchased from Gibco (São Paulo, Brazil). All cell lines were maintained at 37 °C in 5% CO<sub>2</sub> and 95% air containing more than 95% humidity.

### 3.10. Cytotoxicity Activity

Tumor cells were cultivated in appropriate flasks and continuously maintained at exponential growth. The tumor cells then were transferred to tubes and washed three times with RPMI medium at  $500 \times g$  for 10 min at room temperature. Cells were suspended in 5 mL of RPMI complete medium and dispersed in 96-well plates at a density of  $1 \times 10^5$  cells/well. After 24 h, the media were removed and fresh media, with or without different concentrations of BpPLA<sub>2</sub>-TXI (200–0.01 mg/mL), were added into the wells and incubated for 24 h. Cytotoxic activity of PLA<sub>2</sub> was assayed with 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) staining and the MTT reduction was quantified at 540 nm. The cytotoxicity rate was calculated as follows: Cytotoxicity (%) =  $(1 - \text{absorbance of the treated wells}) / (\text{absorbance of the control wells}) \times 100\%$ .

### 3.11. Statistical Analysis

The results were presented as mean  $\pm$  standard deviation (S.D.). Statistical significance of results was evaluated using the Student's *t*-test. A value of  $p > 0.05$  was considered significant.

### 3.12. Determination of Complete cDNA Sequence

The venom gland from *B. pauloensis* adult snake was dissected 3 days after venom extraction, when transcription was most stimulated [36]. The pair of venom glands was homogenized under liquid nitrogen and the total RNA was extracted using the Trizol (Gibco) method. tRNA was dissolved in

sterile milli-Q water (Billerica, MA, USA) and submitted to reverse-transcription polymerase chain reaction (RT-PCR). Two microliters of this sample was used to amplify the specific PLA<sub>2</sub> cDNA, by using the specific primer PLA<sub>2</sub>-forward (5'-GAT CAT GAA AAT TGC GGG GA-3'), which was designed according to the N-terminal sequence, as determined for BpPLA<sub>2</sub>-TXI, and the universal M13 reverse primer (5'-CAG GAA ACA GCT ATG AC-3' GE Healthcare, Uppsala, Sweden) by PCR. After the amplification of the obtained cDNA, the PCR products were analyzed in regard to the amplified size by means of gel electrophoresis on 1% agarose. The gel was stained with ethidium bromide (0.5 mg/mL) and revealed under UV light. The PCR bands were purified by using the Concert Rapid PCR Purification System (Gibco BRL, São Paulo, Brazil) kit, according to the manufacturer's specifications. The amplified fragment was cloned into a p-GEM-T Easy Vector System (Promega<sup>®</sup>, São Paulo, Brazil). Bacteria were selected on a medium containing ampicillin and visualized after adding isopropylthio-β-galactoside (IPTG) and X-Gal in the culture medium. After selection, the colonies of recombinant bacteria were assayed by PCR and gel electrophoresis for cloning confirmation. The PCR products were purified and submitted to sequencing, using the kit DYEnamic ET Terminator Cycle Sequencing Kit (GE Healthcare Uppsala, Sweden) in a MEGA-BACE 1000 automated DNA sequencer (GE Healthcare, Uppsala, Sweden). The Base Caller Cimarron 3.12 software (Amersham Biosci., Sunnyvale, CA, USA) was used in order to analyze the electropherograms and generate the sequences. The cDNA sequences corresponding to acidic PLA<sub>2</sub>s were identified and the full-length sequence was obtained.

### 3.13. Phylogenetic Analysis

The predicted sequences of the acidic PLA<sub>2</sub> (BpPLA<sub>2</sub>-TXI) and other snake venom PLA<sub>2</sub>s were aligned by ClustalW program (UCD, Dublin, Ireland; <http://www.ebi.ac.uk/Tools/clustalw2/index.html>) while employing Human Synovial PLA<sub>2</sub> as the out-group. Datasets were analyzed using Bayesian inference implemented on MrBayes, version 3.2.1 [37] using Iset rate  $s = \text{invgamma}$  with prset aamodelpr = mixed, which enables the program to optimize between nine different amino acid substitution matrices. The analysis was performed by running a minimum of  $1.5 \times 10^7$  generations in four chains, and saving every 100th tree. The log-likelihood score of each saved tree was plotted against the number of generations to establish the point at which the log likelihood scores reached their asymptote, and the posterior probabilities for clades established by constructing a majority-rule consensus tree for all trees generated after completion of the burn-in phase.

## 4. Conclusions

BpPLA<sub>2</sub>-TXI isolated from *B. pauloensis* snake venom is an acidic D49 PLA<sub>2</sub> that shows high catalytic activity, platelet aggregation inhibition, edematogenic and myotoxic activities. This enzyme shares a high degree of sequence identity and a great similarity with other acidic PLA<sub>2</sub>s, especially with D49 and E6, thus forming a group distinct from the basic PLA<sub>2</sub>s.

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### Conflicts of Interest

The authors declare no conflict of interest.

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