

Supplementary Materials: A Gold Growth-Based Plasmonic ELISA for The Sensitive Detection of Fumonisin B₁ in Maize

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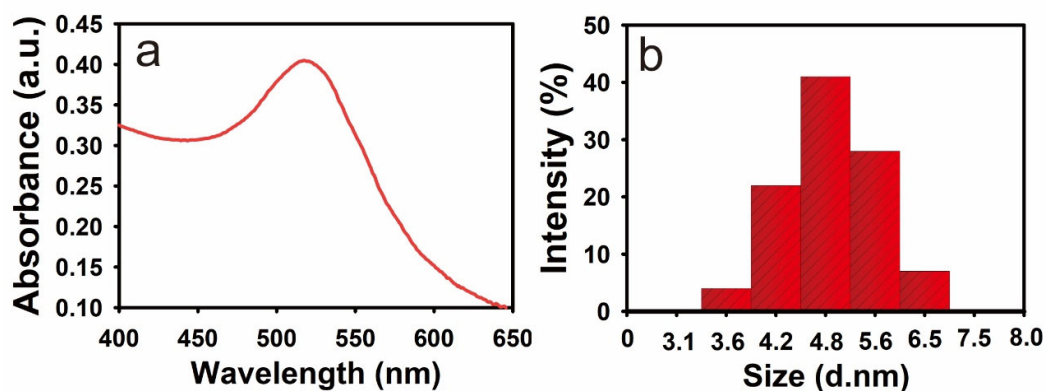


Figure S1. Characterization of the synthesised 5 nm AuNPs with Uv-vis spectrum (a) and hydrodynamic diameter (b).

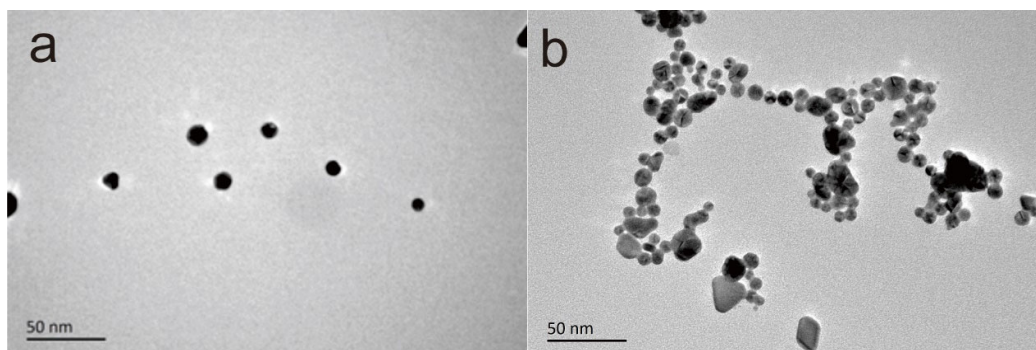


Figure S2. The morphology of 5 nm AuNPs before (a) and after grown (b).

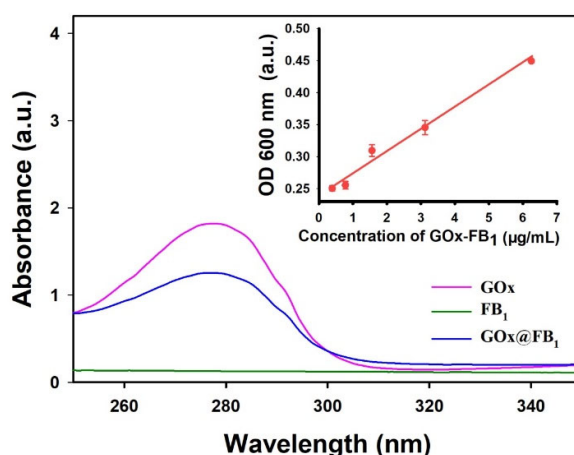


Figure S3. The UV-vis spectrum of glucose oxidase (GOx), fumonisin B₁ (FB₁) and GOx@FB₁, indicating that GOx has been successfully immobilized onto the FB₁.

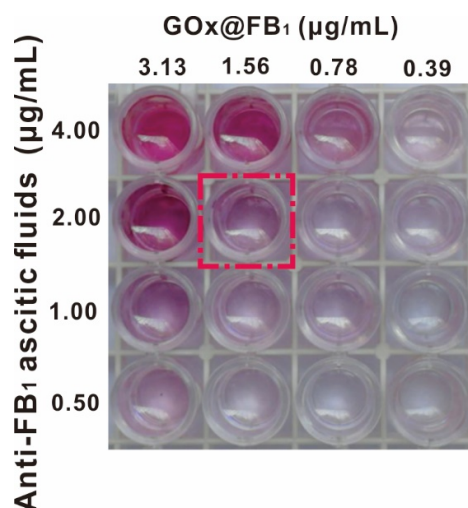


Figure S4. The chessboard titration experiment and the photo taken is in accordance with the figures displayed in Table S1.

1. The Verification of GOx Conjugated FB₁

The insert graph is the characterization of the H₂O₂ produced by the reaction between glucose oxidase (GOx) and glucose after the enzyme-linked immunosorbent assay using a H₂O₂ quantitative assay kit (water-compatible) method, details were as follow: 96-well microplate which was coated with protein G (20 μg/mL, diluted with 0.01 M PBS, pH 8.6) at 4 °C overnight, then the plate was washed with PBST (0.01 M PBS, with 0.05% tween-20, pH 7.4) thrice, anti-FB₁ mAbs (2.00 μg/mL, 100 μL per well) was incubated at 37 °C, 1 h. With the same procedure, then plate was blocked with 1 mg/mL BSA solution for 2 h at 37 °C. Different concentrations of GOx-FB₁ was added with 2-fold diluted (0, 0.39, 0.78, 1.56, 3.13, 6.25 μg/mL) at 37 °C, 1 h. After three times washing with PBST, 1 M of glucose solution (100 μL per well) was added and incubated at 37 °C for another 1 h, then 10 μL of each reaction solution was moved to the work solution (100 μL) from the H₂O₂ quantitative assay kit (water-compatible). After incubating 2 h at 37 °C, the absorbance value at 600 nm was recorded by a microplate reader (DNM-9602, PERLONG, Beijing, China).

2. The Conventional HRP Based ELISA

To compare the sensitivity with the proposed pELISA, we conducted the horse radish peroxidase (HRP) conventional ELISA based on the same molecular ratio of HRP to FB₁ compared to GOx to FB₁. In brief, the conventional ELISA was performed as follow: The 96-well microplate was incubated with 100 μ L per well of protein G (20 μ g/mL, diluted with 0.01 M PBS, pH 8.6) at 4 $^{\circ}$ C overnight, the unbounded protein was washed with PBST (0.01 M PBS, with 0.05% tween-20, pH 7.4) thrice, then 300 μ L of blocking buffer (1 mg/mL BSA solution) was added to block the redundant sites for a period of 1 h at 37 $^{\circ}$ C, after that, the plate was washed three times with washing buffer, and 100 μ L of anti-FB₁ ascitic fluids (2.03 μ g/mL) were added into each well and incubated at 37 $^{\circ}$ C for 1 h, subsequently, the plate was washed three times with washing buffer, and 50 μ L of FB₁-HRP and 50 μ L of sample solution was added to each well, followed by incubating at 37 $^{\circ}$ C for 60 min. The same washing procedure was performed to remove the redundant element, then 100 μ L TMB solution (50 μ L solution A and 50 μ L solution B mixed immediately) was added into each well, then after 15 min of incubation at 37 $^{\circ}$ C, 50 μ L stop buffer (sulfuric acid, 2 mg/mL) was added to stop the reaction, then the absorbance of the plate was recorded by a microplate reader (DNM-9602, PERLONG, Beijing, China).

The parameters of conventional ELISA were optimized by a checkerboard titration method. The optimized concentrations of anti-FB₁ ascitic fluids and FB₁-HRP were 5 μ g/mL and 2.03 μ g/mL, respectively. Under the optimized, the IC₅₀ value of the conventional ELISA was 25 ng/mL, which was about 13-fold higher than that of the proposed plasmonic-ELISA method (Figure S2).

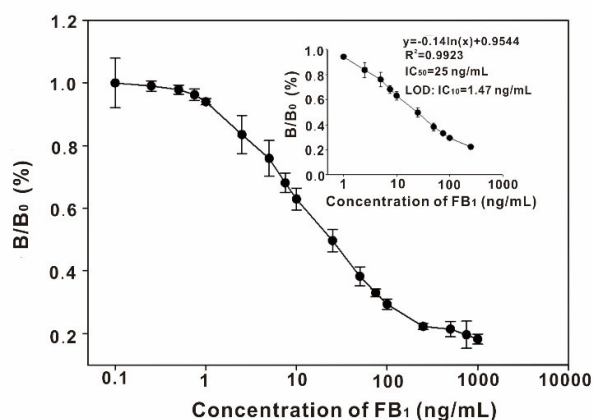


Figure S5. The calibration curve of conventional ELISA. Vertical bars indicate the standard deviation ($n = 3$).

Table S1. The selection for the working conditions of GOx@FB₁ and anti-FB₁ ascitic fluids based plasmonic enzyme-linked immunoassay (pELISA) using the checkerboard method.

Ascitic Fluids Concentration (μ g/mL)	GOx@FB ₁ (μ g/mL)			
	3.13	1.56	0.78	0.39
4.00	0.29	0.19	0.12	0.08
2.00	0.21	0.15*	0.09	0.08
1.00	0.14	0.11	0.09	0.07
0.50	0.09	0.08	0.06	0.05

* The data in red, whose corresponding parameters of anti-FB₁ ascitic fluids and GOx@FB₁ were the final selected ones.

Table S2. Comparison of this work with some established immunoassays for FB₁ detection

Method	Material Used	LOD	LOD by Naked Eyes	References
ELISA	Magnetic beads	0.24 ng/mL	/	[1]
Fluorescence-based ICA	Fluorescent microspheres	0.12 ng/mL	/	[2]
Fluorescence polarization immunoassay	Fluorescein	157.4 µg/kg	/	[3]
Electrochemical	Single-walled carbon nanotubes/chitosan	2 pg/mL	/	[4]
Microarray	Mimotope, fluorescein	11.1 ng/mL	/	[5]
Chemiluminescence	Gold-coated magnetic nanoparticles	0.027 ng/mL	/	[6]
Electrochemiluminescence	Ru(bpy) ₃ ²⁺ -doped silica nanoparticles	0.35 µg/kg	/	[7]
Immunochromatographic strip	Gold nanoparticles	25 ng/mL	/	[8]
Paper sensor	Gold nanoparticle	0.53–1.05 µg/kg	5–25 µg/kg	[9]
Plasmonic ELISA	Gold nanoparticles	0.31 ng/mL	1.25 ng/mL	This study

Table S3. Comparison of ELISA with pELISA analysis of fumonisin B₁ in 16 artificially contaminated maize samples.

Sample	Contaminated	Detected	
	Concentration (µg/kg)	ELISA (µg/kg)	pELISA (µg/kg)
1	5	5.14	6.19
2	10	11.49	9.59
3	15	13.85	16.19
4	15	17.28	15.48
5	15	20.55	17.78
6	20	26.19	23.54
7	20	30.32	34.94
8	25	32.66	29.10
9	25	44.30	36.90
10	32	48.42	61.27
11	35	48.50	54.50
12	50	57.01	37.46
13	60	89.08	76.45
14	100	115.19	131.56
15	150	126.89	115.70

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