

Supplementary Materials: Identification of Immunoreactive Peptides of Toxins to Simultaneously Assess the Neutralization Potency of Antivenoms against Neurotoxicity and Cytotoxicity of *Naja atra* Venom

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Table S1. Identification of *N. atra* venom components within the HPLC chromatographic fractions using LC-MS/MS.

^a Peak	Protein identity	^b AC no.	Significant peptide	Ion, m/z	MS score	Toxin family	^c Toxicity score
1	Cobrotoxin	P60770	K.NGIEINCCTTDR.C R.GCGCPVK.N	726.7767 (2+) 432.6666 (2+)	88	3FTX	60.3
2	Cobrotoxin-b	P80958	K.VKPGVNLNCCTTDR.C K.TCSGETNCYK.K K.TCSGETNCYKK.W	545.2432 (3+) 610.2364 (2+) 674.2805 (2+)	170	3FTX	5.0
3	Phospholipase A ₂	P00598	R.SWWDFADYGCYCGR.G R.LAAIFAGAPYNNNNYNID LK.AR.CCQVHDNCYNEAEK.I K.TYSYECSQGTLTCK.G K.GGNNACAAAVCDCDR.L K.NMIQCTVPSR.S R.GGSGTPVDDLDR.C K.ISGCWPYFK.T	921.8533 (2+) 1178.5278 (2+) 609.5383 (2+) 849.3058 (2+) 805.7562 (2+) 603.2667 (2+) 594.7442 (2+) 579.2611(2+)	859	PLA ₂	N.D.
4	Cardiotoxin A5	P62375	K.CHNTQLPFIYK.T K.YVCCSTDKCN.- R.GCADNCPK.N K.FPLKFPVK.R K.YVCCSTDK.C	710.8403 (2+) 653.7474 (2+) 461.1946 (2+) 488.3006 (2+) 516.6811 (2+)	163	3FTX	
5	Cardiotoxin A1	P60304	K.MFMMSDLTIPVK.R K.LIPIASK.T R.GCIDVCPK.N K.YVCCNTDR.C K.RGCIDVCPK.N K.MFMMSDLTIPVKR.G	706.8523 (2+) 371.2426 (2+) 474.6818 (2+) 544.2131(2+) 552.7282 (2+) 523.5909 (3+)	308	3FTX	25.95
6	Cardiotoxin A3	P60301	K.LVPLFYK.T K.MFMVATPK.V K.MFMVATPK.V	440.2615 (2+) 462.7289 (2+) 478.7285 (2+)	274	3FTX	
7	Cardiotoxin A6	P80245	K.MFMVAAPK.V K.CNQLIPPFYK.A	447.7211 (2+) 640.3278 (2+)	284	3FTX	
8	Cysteine-rich secretory protein	Q7T1K6	R.WANTCSLNHSPDNLR.V R.AGCAVSYCPSSAWSYFYVCQ YCPSGNFQGK.T R.VSPTASNMLK.M K.LTNCDSSLK.Q K.EIVDLHNSLR.R K.SNCPASCFCR.N K.QSSCQDDWIK.S	595.5896 (3+) 1167.8121 (3+) 524.2760 (2+) 532.2622 (2+) 598.3191 (2+) 629.7216 (2+) 633.7682 (2+)	223	CRISP	N.D.
9	Zinc metalloproteinase-disintegrin-like kaouthiagin-like	D3TTC1	R.VAKDDCDLPELCTGQSAECP TDSLQR.N R.NDNAQLLTGIDFNGNTVGR. A K.FEVKPAASVTLK.S R.TAPAFQFSSCSIR.E K.DKFEVKPAASVTLK.S	989.0843 (3+) 1009.9933 (2+) 430.5788 (3+) 736.3360 (2+) 511.6191 (3+)	192	SVMP	N.D.

			K.ASCICIPGPCIMLK.K	810.3913 (2+)			
10	Zinc metalloproteinase-disintegrin-like atragin	D3TTC2	R.AAKDDCDLPELCTGQSAECP TDVFQR.N K.TSAAVVQDYSSR.T R.KIPCAAK.D R.GFCTCGFNK.C R.DSCFTLNQR.T R.ATLNLFGEWR.E R.TKPAYQFSSCSV.R.E K.LQHEAQCDSEECCEK.C K.DDCDLPELCTGQSAECPTDV FQR.N	995.0845 (3+) 642.3005 (2+) 394.2237 (2+) 545.7274 (2+) 570.7505 (2+) 603.8043 (2+) 765.8667 (2+) 641.5699 (3+) 905.0342 (3+)	281	SVMP	N.D.

^a The peak refers to the peak number highlighted in Figure S1. ^b AC is the abbreviation of accession number in Uniprot database. ^c Toxicity score was calculated according to Laustsen, A et al. [25] by the ratio of protein abundance (%) estimated from the reverse phase HPLC chromatography to its medium lethal dose (LD_{50}). The toxicity score of the crude venom, which abundance was defined as 100%, was 149.3. N.D. represented LD_{50} was undetectable at a cut-off value of 50 μ g per mouse. Abbreviations: SVMP indicates snake venom metalloproteinase. CRISP is cysteine-rich secretory protein. 3FTX means three finger toxins.

Table S2. List of synthetic peptides used for the immunoreactive peptide mapping study.

Index	Peptide sequence
CTXA3 ₁₋₁₅	LKCNKLVPLFYKTCP
CTXA3 ₅₋₁₉	KLVPLFYKTCPAGKN
CTXA3 ₁₁₋₂₅	YKTCPAGKNLCYKMF
CTXA3 ₁₅₋₂₉	PAGKNLCYKMFMVAT
CTXA3 ₂₁₋₃₅	CYKMFMVATPKVPVK
CTXA3 ₂₆₋₄₀	MVATPKVPVKRGCID
CTXA3 ₃₁₋₄₅	KVPVKRGCIDVCPKS
CTXA3 ₃₆₋₅₀	RGCIDVCPKSSLVVK
CTXA3 ₄₃₋₅₇	PKSSLLVKYVCCNTD
CTXA3 ₄₆₋₆₀	SLLVKYVCCNTDRCN
sNTX ₁₋₁₅	LECHNQQSSQTPTTT
sNTX ₄₋₁₈	HNQQSSQTPTTGCS
sNTX ₁₁₋₂₅	TPTTTGCSGETNCY
sNTX ₁₆₋₃₀	GCSGETNCYKKRWR
sNTX ₂₁₋₃₅	ETNCYKKWRDHRGY
sNTX ₂₆₋₄₀	KKWRDHRGYRTERG
sNTX ₃₁₋₄₅	DHRGYRTERGCGPS
sNTX ₃₆₋₅₀	RTERGCCPSPVKNGI
sNTX ₃₉₋₅₃	RGCGPSVKNGIEIN
sNTX ₄₅₋₅₉	SVKNGIEINCCTTDR
sNTX ₄₈₋₆₂	NGIEINCCTTDRCNN
^a TFF	TFFLTQGALLNDK
^b GIL	GILGFVFTLTVPSER

^aTFF is the abbreviation of TFFLTQGALLNDK which is a partial sequence of neuraminidase of H1N1 influenza virus. ^b GIL is the abbreviation of GILGFVFTLTVPSER which is a partial sequence of M1 protein of H5N1 influenza virus.

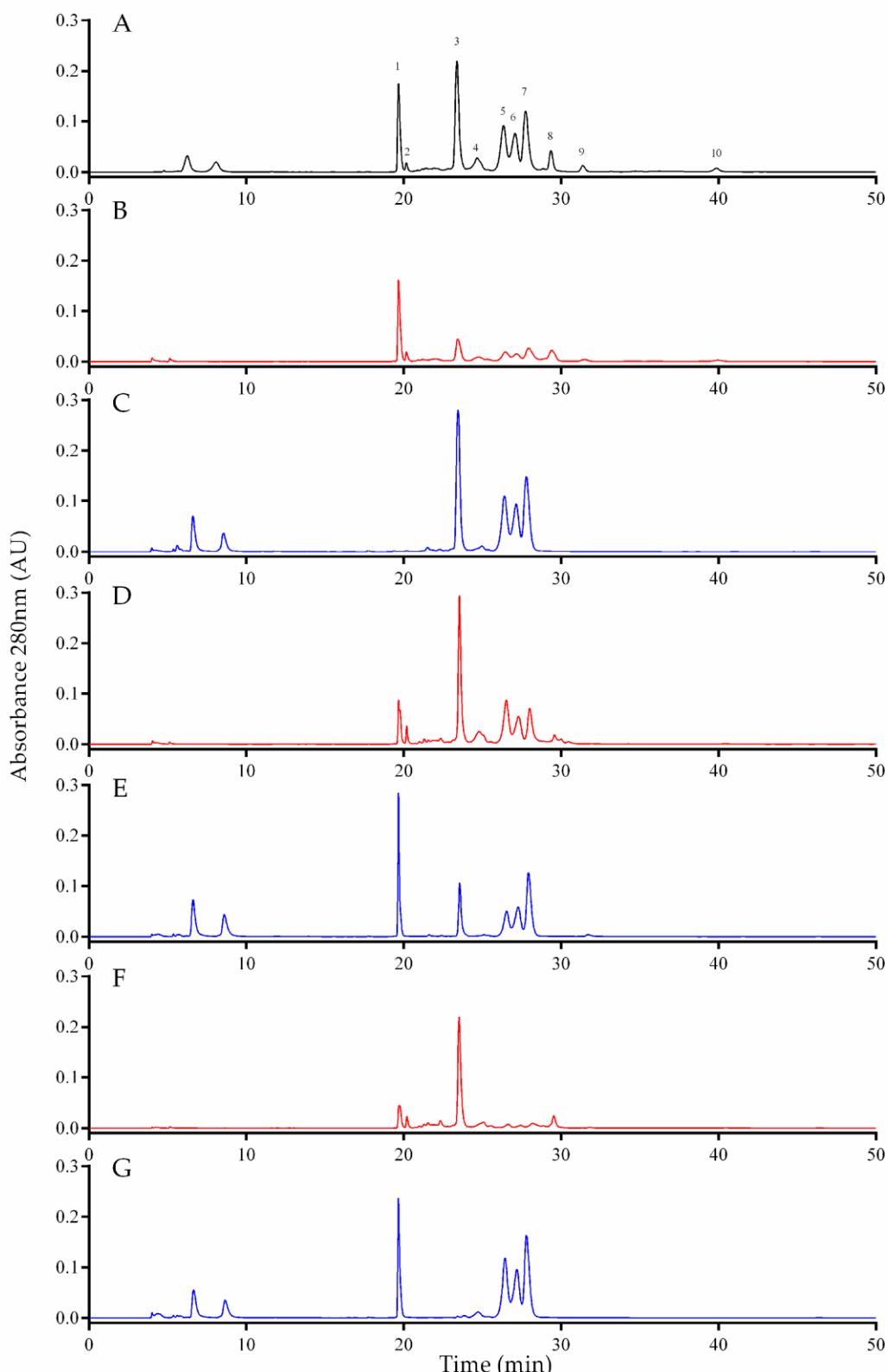


Figure S1. Evaluation of antivenom efficacy in capturing *N. atra* venom components. (A) HPLC chromatogram of 500 µg of *N. atra* crude venom. The component within the chromatographic peaks (no.1-10) were identified using LC MS/MS (Table S1). The elution (Elu) and flow through (FT) fractions of (B,C) BAV-, (D,E) SAV-Naja-, and (F,G) NPAV-immobilized affinity columns were collected and analyzed by reverse phase HPLC to determine the retained percentage of venom components.

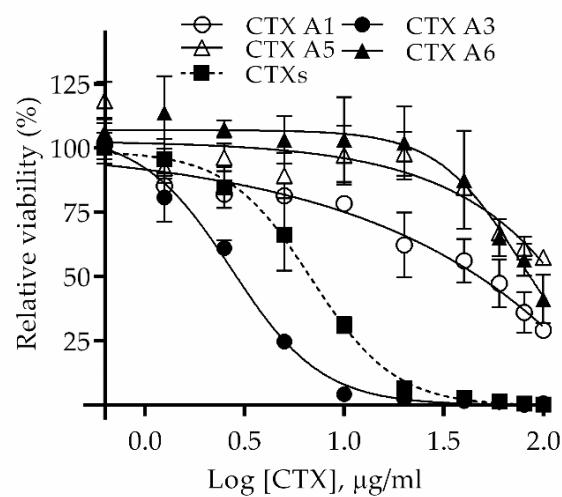


Figure S2. Analyzing the cytotoxicity of CTX analogs of *N. atra* venom using the cell-based assay.

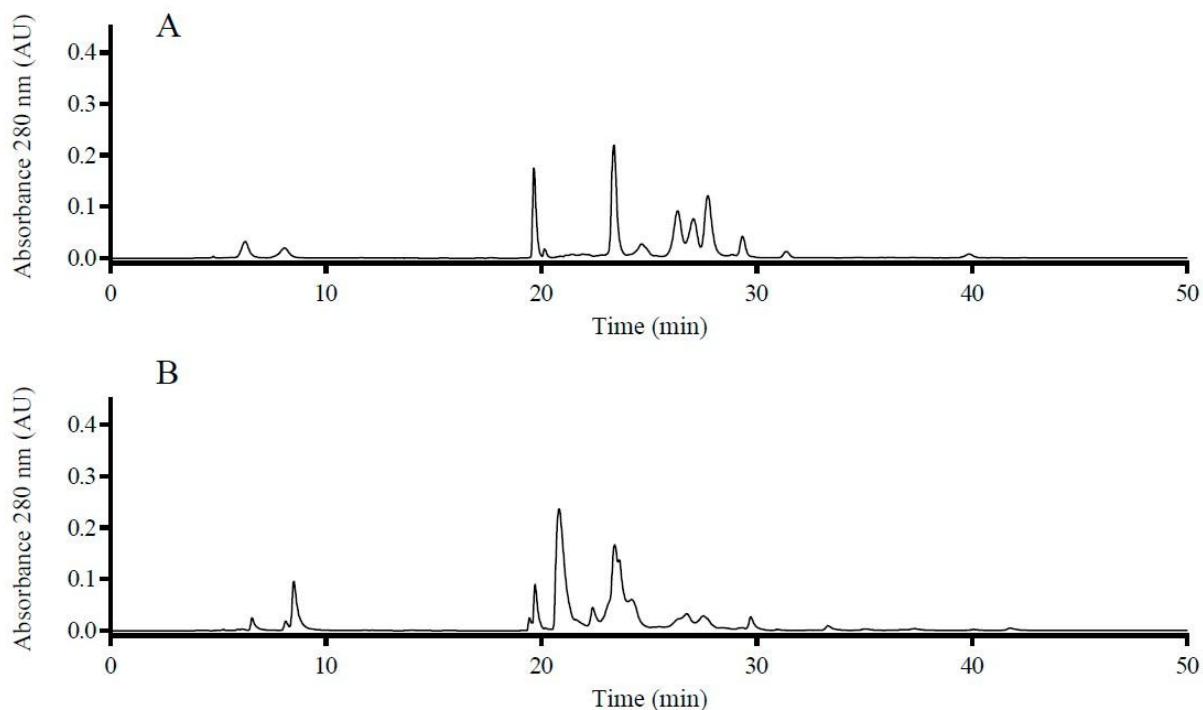


Figure S3. Reverse phase HPLC chromatograms of 300 μ g of (A) *N. atra* and (B) *N. kaouthia* venom used in this study.