

Article

Joint Effects of Prenatal Folic Acid Supplement with Prenatal Multivitamin and Iron Supplement on Obesity in Preschoolers Born SGA: Sex Specific Difference

Qing Lu ¹, Esben Strodl ² , Yang Liang ¹, Li-Hua Huang ¹, Bing-Jie Hu ^{3,*} and Wei-Qing Chen ^{1,4,*} ¹ Department of Epidemiology, School of Public Health, Sun Yat-Sen University, Guangzhou 510080, China² School of Psychology and Counselling, Queensland University of Technology, Brisbane, QLD 4059, Australia³ School of Basic Medical Science, Guangzhou Medical University, Guangzhou 511436, China⁴ School of Health, Xinhua College of Guangzhou, Guangzhou 510080, China

* Correspondence: hubingjie@126.com (B.-J.H.); chenwq@mail.sysu.edu.cn (W.-Q.C.);

Tel.: +86-(20)-81343008 (B.-J.H.); +86-(20)-8733-2199 (W.-Q.C.)

Abstract: Prenatal maternal nutrient supplementation has been reported to be associated with offspring obesity, but the reports are inconsistent and have mainly ignored the differences between the total children population and children born small for gestational age (SGA). This study aimed to examine the joint effects of folic acid, iron, and multivitamin supplementation during pregnancy on the risk of obesity in preschoolers born SGA. A total of 8918 children aged 3–6.5 years born SGA were recruited from Longhua District in Shenzhen of China in 2021. Their mothers completed a structured questionnaire about the child's and parents' socio-demographic characteristics, maternal prepregnant obesity, and mothers' prenatal supplementation of folic acid, iron, and multivitamin. In addition, the children's current weight and height were measured by trained nurses. Logistic regression models were used to analyze the associations between prenatal supplementations and the current presence of childhood obesity. After controlling for potential confounders, the results of the logistic regression analysis showed that prenatal supplement of folic acid (OR = 0.72, 95% CI = 0.55–0.93) was associated with a lower likelihood of being an obese preschooler born SGA. In contrast, the ingestion of multivitamin or iron supplements during pregnancy did not seem to be related to the likelihood of childhood obesity in preschoolers born SGA. Moreover, cross-over analysis of prenatal folic acid and multivitamin obtained significant negative associations of prenatal folic acid supplement only (OR = 0.73, 95% CI = 0.55–0.97) and combination supplement of folic acid and multivitamin (OR = 0.67, 95% CI = 0.50–0.90) with obesity of preschoolers born SGA; while the cross-over analysis of prenatal folic acid and iron observed significant negative associations between obesity of preschoolers born SGA and a combination supplement of folic acid and iron (OR = 0.70, 95% CI = 0.52–0.96). Furthermore, the aforementioned significant associations were only found in girls and not in boys when the analyses were stratified by sex. Our findings suggest that the prenatal folic acid supplementation may decrease the risk of obesity in preschool girls born SGA, and that this effect may be modified by prenatal multivitamin or iron supplementation.

Keywords: obesity; prenatal supplement; folic acid; iron; multivitamin; preschoolers; SGA



Citation: Lu, Q.; Strodl, E.; Liang, Y.; Huang, L.-H.; Hu, B.-J.; Chen, W.-Q. Joint Effects of Prenatal Folic Acid Supplement with Prenatal Multivitamin and Iron Supplement on Obesity in Preschoolers Born SGA: Sex Specific Difference. *Nutrients* **2023**, *15*, 380. <https://doi.org/10.3390/nu15020380>

Received: 24 December 2022

Revised: 5 January 2023

Accepted: 10 January 2023

Published: 12 January 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Obesity is one of the most severe public health concerns in the world, ranking fourth among risk factors for mortality and accounting for 4.72 million deaths worldwide in 2017 [1]. This is concerning given that the global prevalence of obesity in children and adolescents increased from 0.7% in 1975 to 5.6% in 2016 for girls and from 0.9% to 7.8% for boys [2]. For Chinese preschoolers, overweight and obesity prevalence has been estimated as high as 8.4% and 4.2%, respectively [3]. While obesity is mainly a risk factor for the development of chronic diseases in adulthood [4], childhood obesity is a major public

health issue given that obese children are very likely to remain obese into adulthood [5]. Given this trajectory, reported lifetime healthcare cost and loss of productivity of an obese child or adolescent is high (approximately EUR 149,000) compared with a normal weight child or adolescent [6]. Early prevention and intervention of childhood obesity is, therefore, critical for both the individual and society. To guide the development of effective prevention programs it is important to identify significant risk and protective factors for childhood obesity.

It has been well documented that childhood obesity is caused by synergistical effects between genetic, prenatal, and postnatal factors. A body of evidence has revealed that specific genes correlate with childhood obesity. For example, a meta-analysis reported that a fat-mass and obesity-associated gene (FTO) increased the risk of obesity among children [7]. Similarly, a genome-wide association study in China linked single nucleotide polymorphisms near SEC16B, RBJ, CDKAL1, TFAP2B, MAP2K5, and FTO to childhood obesity [8]. In addition, there are several genes related to energy metabolism or appetite regulation, such as melanocortin 4 receptor gene leptin and the leptin receptor gene, that have been associated with childhood obesity [9,10]. With regard to prenatal factors, studies have shown that deficient maternal exercise, exposure to environmental tobacco smoke, unhealthy dietary pattern, prepregnancy maternal BMI, maternal weight gain during pregnancy, as well as poor birth outcomes, such as low birth weight (LBW) or small for gestational age (SGA) and preterm birth, are all associated with an increased likelihood of childhood obesity [11–15]. For postnatal factors, prior research indicates that childhood obesity is positively associated with early-life rapid weight gain of babies born SGA [16]. In addition, artificial feeding in early life—in the first six months—unbalanced nutrition, short sleep duration, and insufficient physical activity in childhood are also all related to childhood obesity [11–13].

SGA, defined as a birth weight less than the 10th percentile for that gestational age, is typically considered a fetal growth restriction (FGR) at birth [17]. Epidemiological studies have shown that SGA is associated with neurodevelopmental delay [18], obesity [19,20], and other cardiometabolic risk factors in childhood [21], and with chronic diseases in adulthood, such as cardiovascular diseases and type 2 diabetes [22]. Based on the Developmental Origins of Health and Disease Hypothesis (DOHaD) [19], an undesirable intrauterine environment increases the incidence of SGA and later high-risk metabolic patterns through possible mechanisms of placental dysfunction [17] and epigenetic modification (e.g., DNA methylation) [23,24].

Increasing evidence indicates that prenatal nutrients play a crucial role in fetal growth, offspring's development, and the likelihood of experiencing disease [11,25–27]. For example, several experimental studies have found that prenatal micronutrient supplements improved fetal growth [17,28–30] and reduced the incidence of SGA in fetuses diagnosed as FGR [31]. A randomized trial in Nepal [27] suggested that a prenatal micronutrients supplement might promote the metabolic status in young children. However, another study by Sauder et al. found that prenatal multivitamin use might slow the growth of an offspring during infancy [32]. Recently, a prospective study showed an inverse association between prenatal maternal iron supplementation and the infant's fat mass at birth, and 3 months and 6 months post-birth [33].

Folic acid is widely used to prevent neural tube defects [34]. Additionally, two population-based studies in China reported that maternal folic acid supplementation during pregnancy reduced SGA and low birth weight [35,36]. An animal trial found that prenatal folic acid or methyl donor supplements could prevent the offspring from developing obesity [37]. A study of 4449 school-age children reported that a relative higher folate concentration in maternal plasma during pregnancy decreased the risk of the offspring being overweight [38]. Conversely, a study by Yajnik et al. suggested higher circulating concentrations of maternal erythrocyte folate during pregnancy increased offspring adiposity [39].

It has been recognized that children born SGA have a higher tendency to develop obesity; however, not all individuals born SGA develop obesity. Moreover, prenatal

supplements of folic acid, multivitamin, and iron may be associated with the likelihood of offspring obesity, but the results are inconsistent. Furthermore, most studies have mainly focused on the total population of children, and not just on children with SGA. As such, an unanswered question remains as to whether prenatal supplementation of folic acid, multivitamin, and iron affects the development of obesity in offspring born SGA. Therefore, this study aimed to investigate the joint effects of prenatal folic acid supplementation with prenatal multivitamin and iron supplementation on obesity in Chinese preschoolers born SGA.

2. Materials and Methods

2.1. Study Population

Participants of this research were recruited from all of the kindergartens in the Longhua District of Shenzhen, China, with a total of 69,639 child–mother dyads recruited. The study was approved by the Ethics Committee of School of Public Health, Sun Yat-sen University in Guangzhou, China. Written informed consent was obtained from the mothers of all of the preschoolers. The study was carried out in accordance with the Declaration of Helsinki.

This paper only included 8919 preschoolers born singleton and classified as SGA based upon being under the 10th percentile of Chinese birth weight for gestational age [40]. Due to missing data, 143 child–mother pairs were excluded for lacking information of weight or height, 214 pairs were excluded for missing paternal age at childbirth, 445 for missing maternal age at childbirth, 12 for missing paternal education level, and 88 for missing maternal prepregnant BMI. Finally, a total of 8016 child–mother pairs were included in the final analysis.

2.2. Data Collection

The enrolled mothers were asked to complete a self-report structured questionnaire collecting the socio-demographic characteristics of the child and the parents (such as age, gender, maternal prepregnant BMI, parents' marital status, parents' education level, family income, and single child or not), prenatal maternal supplementations of multivitamin, folic acid, and iron, and birth-related variables (mode of delivery, gestational age, and birth weight).

2.3. Measurement of Prenatal Maternal Nutrient Supplementations

Based upon previous research [41–44], we collected the data on folic acid, multivitamin, and iron supplementations during pregnancy using the following questions: (1) Did the mother take folic acid during pregnancy? (2) Did the mother take a multivitamin during pregnancy? (3) Did the mother take iron during pregnancy? The answers for each question were: 1 for “yes”, and 0 for “no”.

2.4. Measurement and Definition of Obesity

Standardized measurements of the weight and height of the child were taken by trained nurses from Longhua Maternity & Child Healthcare Hospital. A portable electronic weight scale (fractional value = 0.01 kg) was used to measure the child's weight by placing the scale on level ground and asking the preschoolers to stand in the center of the scale bareheaded, barefooted, and wearing close-fitting light clothes. After the value stabilized, the nurses read and recorded the measurement accurate to 0.1 kg. Height was measured by column human altimeter (fractional value = 0.1 cm). The column human altimeter was placed vertically against the wall on horizontal ground. Preschoolers were asked to stand on the pedal, with heels close and feet separated by a 60-degree angle, chest lifted, abdomen pulled in, and eyes straight ahead. Nurses slid the slider to the apex of the measured child's skull, and read the measurements with their line of sight the same height as the slide board.

Body mass index (BMI) was calculated by dividing weight in kilograms by height in meters squared. We employed BMI reference values based upon the normative data obtained from two national representative cross-sectional surveys in China: The National

Growth Survey of Children under 7 years in the Nine Cities of China in 2005 and The Physical Fitness and Health Surveillance of Chinese School Students in 2005. The researchers used the LMS method to smooth the curve of BMI, calculated values of percentile, and generated the screening cut-offs of obesity in Chinese children by integrating with adult cut-offs for obesity at 18 years in China [45]. Based on the aforementioned investigation in China, this research defined obesity as a BMI equal to or greater than the reference values for sex and age [45].

2.5. Potential Confounding Variables

In the light of previously findings [46,47], potential confounding variables included in this study were the child's sex, child's age, prepregnant maternal BMI, mode of delivery, parents' marital status, parents' age at the childbirth, parents' education level, single child or not, and family income.

2.6. Statistical Analyses

Means and standard deviations (SD) were used to describe the continuous variables, while frequencies and percentages were used to describe the categorical variables. Categorical covariates and numeric covariates were compared using a chi-square test and *t*-test, respectively.

A series of binary logistic regression analyses were conducted to examine the association of maternal supplementations of multivitamin, folic acid, and iron during pregnancy with obesity in SGA children after adjusting for the potential confounding variables. Their multiplicative interaction effects on obesity were tested by establishing multiplicative terms in logistic regression models with the interaction of odds ratio (IOR) used to indicate the strength of multiplicative interaction effects. Moreover, crossover analyses were employed to assess the additive interaction effects and modification effects among the combination of the three nutrients. Adjusted odds ratios (AOR), the relative excess risk due to interaction (RERI), and the attributable proportion due to interaction (AP) were used to indicate the strength of these effects.

Furthermore, after stratification by sex, the aforementioned analyses were repeated to evaluate sex-specific associations of prenatal supplementations with obesity in SGA preschoolers.

All statistical analyses were performed in RStudio versions 4.1.2 (Poist, BOSTON, MA, USA) and two-tailed *p*-values < 0.05 were deemed statistically significant.

3. Results

3.1. Characteristics of Participants

Table 1 presents the demographic characteristics and pregnancy condition of the participants. The mean age was 4.85 years (SD = 0.84) old for the children, 30.55 years (SD = 4.83) old for their fathers and 28.44 years (SD = 4.27) old for their mothers, respectively. The mean birthweight was 2.72 kg (SD = 0.41) and the mean gestational age at childbirth was 39.65 weeks (SD = 1.90). There were only 204 mothers (2.54%) with acknowledged gestational hypertension history. Over 60% of the preschoolers were male. The mean value was 15.33 kg/m² (SD = 2.18) for children's BMI, 18.07 kg (SD = 3.64) for children's weight, 108.34 cm (SD = 7.69) for children's height, and 20.31 kg/m² (SD = 2.81) for their mothers before pregnancy, respectively. There were only 119 children (1.48%) born to obese mothers. Most mothers (99.09%) and fathers (98.52%) were married. The proportion of mothers and fathers with high school education and above was 83.16% and 85.32%, respectively. More than half of participants had a sibling(s) (63.91%) and a high household income (82.92%).

Table 1. Comparison of demographic characteristics between obese and non-obese children born SGA.

Characteristics	Total	Obesity, N (%)		t/χ^2	p
		Yes	No		
Total	8016	702 (8.76)	7314 (91.24)	-	-
Age [(Mean $\pm$ SD) (years)]	4.85 $\pm$ 0.84	4.89 $\pm$ 0.83	4.85 $\pm$ 0.84	-1.200	0.230
Sex				15.152	<0.001
Male	4863	474 (67.52)	4389 (60.01)		
Female	3153	228 (32.48)	2925 (39.99)		
Current weight of child [(Mean $\pm$ SD) (kg)]	18.07 $\pm$ 3.64	24.32 $\pm$ 5.12	17.47 $\pm$ 2.81	-56.287	<0.001
Current height of child [(Mean $\pm$ SD) (cm)]	108.34 $\pm$ 7.69	108.51 $\pm$ 9.27	108.33 $\pm$ 7.53	-0.614	0.539
Current BMI of child [(Mean $\pm$ SD) (kg/m ²)]	15.33 $\pm$ 2.18	20.47 $\pm$ 2.29	14.84 $\pm$ 1.38	-96.150	<0.001
Birthweight [(Mean $\pm$ SD) (kg)]	2.72 $\pm$ 0.41	2.71 $\pm$ 0.50	2.72 $\pm$ 0.40	0.469	0.639
Gestational age at birth [(Mean $\pm$ SD) (weeks)]	39.65 $\pm$ 1.90	39.58 $\pm$ 2.38	39.65 $\pm$ 1.85	0.962	0.336
Maternal age [(Mean $\pm$ SD) (years)]	28.44 $\pm$ 4.27	28.64 $\pm$ 4.41	28.42 $\pm$ 4.25	-1.310	0.190
Paternal age [(Mean $\pm$ SD) (years)]	30.55 $\pm$ 4.83	30.79 $\pm$ 4.92	30.52 $\pm$ 4.82	-1.418	0.156
Maternal prepregnancy obesity				7.856	0.005
No	7897	683 (97.29)	7214 (98.63)		
Yes	119	19 (2.71)	100 (1.37)		
Maternal prepregnancy BMI [(Mean $\pm$ SD) (kg/m ²)]	20.31 $\pm$ 2.81	20.87 $\pm$ 3.22	20.26 $\pm$ 2.76	-5.486	<0.001
Gestational hypertension				2.651	0.266
No	7743	683 (97.43)	7060 (96.74)		
Yes	204	12 (1.71)	192 (2.63)		
Uncertain	52	6 (0.86)	46 (0.63)		
Mode of delivery				0.373	0.542
Vaginal delivery	5848	1324 (61.07)	3539 (60.52)		
Cesarean delivery	2168	844 (38.93)	2309 (39.48)		
Maternal marital state				2.251	0.134
Married	7943	692 (98.58)	7251 (99.14)		
Others *	73	10 (1.42)	63 (0.86)		
Paternal marital state				0.567	0.451
Married	7935	693 (98.72)	7242 (99.02)		
Others *	81	9 (1.28)	72 (0.98)		
Maternal education level				3.345	0.188
Junior high school or lower	1350	135 (19.23)	1215 (16.61)		
High school	1628	143 (20.37)	1485 (20.30)		
College or higher	5038	424 (60.40)	4614 (63.08)		
Paternal education level				10.542	0.005
Junior high school or lower	1177	131 (18.66)	1046 (14.30)		
High school	1687	150 (21.37)	1537 (21.01)		
College or higher	5152	421 (59.97)	4731 (64.68)		
Household income [(Chinese Yuan)]				10.785	0.005
0–9999	1369	151 (21.51)	1218 (16.65)		
10,000–29,999	4613	386 (54.99)	4227 (57.79)		
$\geq 30,000$	2034	165 (23.50)	1869 (25.55)		
Single child				37.421	<0.001
Yes	2893	179 (25.50)	2714 (37.11)		
No	5123	523 (74.50)	4600 (62.89)		

* Including divorced, remarried, spouse loss, unmarried.

The prevalence of obesity among the preschoolers was 8.76%. Significant differences of the following characteristics were observed between obese and normal weight preschoolers, including the child's sex, maternal prepregnant obesity/BMI, paternal education level, monthly household income, and being a single child.

3.2. Associations between Prenatal Supplementation of the Micronutrients and Obesity in Preschoolers Born SGA

After controlling for the potential confounding variables, the results of the logistic regression analysis showed that prenatal folic acid supplement was significantly and nega-

tively associated with obesity in preschoolers born SGA (AOR = 0.72, 95% CI = 0.55~0.93). No significant associations were found between prenatal maternal iron and multivitamin supplementation and childhood obesity (Table 2).

Table 2. The association of prenatal maternal supplementation of the three micronutrients with obesity in preschoolers born SGA.

Nutrients Supplementation	Total, N = 8016	Obesity, N (%)	AOR (95% CI) ^a
Folic acid			
No	567	71 (12.52)	1.00
Yes	7449	631 (8.47)	0.72 (0.55, 0.93) *
Multivitamin			
No	4873	454 (9.32)	1.00
Yes	3143	248 (7.89)	0.89 (0.75, 1.05)
Iron			
No	4914	453 (9.22)	1.00
Yes	3102	249 (8.03)	0.91 (0.77, 1.07)

^a: adjusted for child's sex, child's age, mode of delivery, parents' age at the childbirth, maternal prepregnancy BMI, marital status, parents' education level, family income, and single child or not in models. * $p < 0.05$.

3.3. Combination Effects of Maternal Micronutrients Supplementation during Pregnancy on Obesity in Preschoolers Born SGA

Table 3 presents the combination effects of maternal micronutrients supplementation during pregnancy on obesity in preschoolers born SGA. The results of the crossover analyses indicated that a maternal supplement of a combination of folic acid and multivitamin (AOR = 0.67, 95% CI = 0.50~0.90), as well as maternal supplement of a combination of folic acid and iron (AOR = 0.70, 95% CI = 0.52~0.96) significantly decreased the risk of obesity in preschoolers born SGA; while only maternal folic acid supplementation during pregnancy decreased the risk of obesity with significance (AOR = 0.73, 95% CI = 0.55~0.97) in the crossover analysis on a combination of folic acid and multivitamin, and marginal significance (AOR = 0.77, 95% CI = 0.57~1.04) in the crossover analysis on a combination of folic acid and iron, respectively. There was no significant multiplicative and additive interaction between prenatal maternal supplementation of the three nutrients on obesity in preschool children born SGA.

Table 3. The combination effect of maternal supplementation of nutrients on obesity in preschoolers born SGA.

Nutrients Supplementation		AOR (95% CI) ^a	IOR (95% CI) ^a	RERI (95% CI) ^a	AP (95% CI) ^a
Folic acid	Multivitamin		1.07 (0.40, 2.84)	0.08 (−0.76, 0.92)	0.12 (−1.14, 1.39)
	No	1.00			
	No	0.86 (0.33, 2.25)			
	Yes	0.73 (0.55, 0.97) *			
Folic acid	Iron		0.69 (0.35, 1.39)	−0.38 (−1.28, 0.52)	−0.54 (−1.78, 0.69)
	No	1.00			
	No	1.31 (0.67, 2.58)			
	Yes	0.77 (0.57, 1.04)			
		0.67 (0.50, 0.90) **			
		0.70 (0.52, 0.96) *			

^a: adjusted for child's sex, child's age, mode of delivery, parents' age at the childbirth, maternal prepregnancy BMI, marital status, parents' education level, family income, and single child or not in models. * $p < 0.05$, ** $p < 0.01$.

3.4. Combination Effects of Maternal Micronutrients Supplementation during Pregnancy on Obesity in Preschoolers Born SGA

Table 4 presents the results of the stratification analysis by sex. After controlling for the potential confounding variables, the results of the logistic analysis showed that prenatal maternal folic acid supplement was significantly and negatively associated with obesity in preschoolers born SGA (AOR = 0.57, 95% CI = 0.36~0.90) in girls, but not in boys.

Table 4. The association of prenatal maternal supplementation of the three micronutrients with obesity in preschoolers born SGA in stratified analysis by sex.

Male				Female			
Nutrients Supplementation	Total, N = 4863	Obesity, N (%)	AOR (95% CI) ^a	Nutrients Supplementation	Total, N = 3153	Obesity, N (%)	AOR (95% CI) ^a
Folic acid				Folic acid			
No	361	46 (12.74)	1.00	No	206	25 (12.14)	1.00
Yes	4502	428 (9.51)	0.78 (0.56, 1.09)	Yes	2947	203 (6.89)	0.57 (0.36, 0.90) *
Multivitamin				Multivitamin			
No	2999	319 (10.64)	1.00	No	1874	135 (7.20)	1.00
Yes	1864	155 (8.32)	0.83 (0.67, 1.02)	Yes	1279	93 (7.27)	1.01 (0.76, 1.34)
Iron				Iron			
No	3010	307 (10.20)	1.00	No	1904	146 (7.67)	1.00
Yes	1853	167 (9.01)	0.94 (0.77, 1.15)	Yes	1249	82 (6.57)	0.85 (0.64, 1.13)

^a: adjusted for child's age, mode of delivery, parents' age at the childbirth, maternal prepregnancy BMI, marital status, parents' education level, family income, and single child or not in models. * $p < 0.05$.

Table 5 presents the combination effects of maternal micronutrients supplementation during pregnancy on obesity in preschoolers born SGA stratified by sex. The results of the crossover analyses indicate that combining the supplements of folic acid and multivitamin (AOR = 0.55, 95% CI = 0.33~0.92), as well as combining the supplements of folic acid and iron (AOR = 0.51, 95% CI = 0.30~0.86) significantly decreased the risk of obesity in girls, while only maternal folic acid supplementation during pregnancy decreased the risk of obesity with significance in the crossover analysis on a combination of folic acid and multivitamin, as well as in the crossover analysis on a combination of folic acid and iron in girls, and their ORs were 0.50 (95% CI = 0.31~0.82) and 0.57 (95% CI = 0.35~0.94), respectively. There was no significant multiplicative and additive interaction between prenatal maternal supplement of three nutrients on obesity in both male and female preschool children born SGA.

Table 5. The combination effect of maternal supplementation of nutrients on obesity in preschoolers born SGA in stratified analysis by sex.

Sex	Nutrients Supplementation		AOR (95% CI) ^a	IOR (95% CI) ^a	RERI (95% CI) ^a	AP (95% CI) ^a
Male	Folic acid	Multivitamin				
	No	No	1.00			
	No	Yes	1.22 (0.40, 3.73)			
	Yes	No	0.85 (0.60, 1.20)			
	Yes	Yes	0.71 (0.49, 1.03)			
	Folic acid	Iron				
	No	No	1.00	0.56 (0.24, 1.28)	−0.73 (−2.08, 0.63)	−0.90 (−2.48, 0.69)
	No	Yes	1.66 (0.74, 3.71)			
Female	Yes	No	0.87 (0.60, 1.27)			
	Yes	Yes	0.81 (0.55, 1.20)			
	Folic acid	Multivitamin				
	No	No	1.00	3.55 (0.44, 28.50)	0.74 (0.08, 1.40)	1.34 (−0.07, 2.74)
	No	Yes	0.31 (0.04, 2.46)			
	Yes	No	0.50 (0.31, 0.82) **			
	Yes	Yes	0.55 (0.33, 0.92) *			
	Folic acid	Iron				
	No	No	1.00	1.16 (0.31, 4.33)	0.17 (−0.83, 1.17)	0.34 (−1.69, 2.37)
	No	Yes	0.76 (0.21, 2.76)			
	Yes	No	0.57 (0.35, 0.94) *			
	Yes	Yes	0.51 (0.30, 0.86) *			

^a: adjusted for child's age, mode of delivery, parents' age at the childbirth, maternal prepregnancy BMI, marital status, parents' education level, family income, and single child or not in models. * $p < 0.05$, ** $p < 0.01$.

4. Discussion

To the best of our knowledge, this is the first study to examine the joint effects of prenatal maternal supplements of multivitamin, folic acid, and iron on childhood obesity in a large sample of Chinese preschoolers born SGA. Our results show that maternal folic acid supplementation in pregnancy is significantly and negatively associated with obesity in preschool girls born SGA, but not in preschool boys born SGA. Moreover, this association may be modified by a prenatal supplement of iron or multivitamin.

4.1. Associations of Prenatal Maternal Supplementation of Folic Acid with Preschool Obesity in SGA

There have been several previous studies examining the associations between maternal folate concentration and children obesity, but they have failed to yield consistent findings [48]. For example, a study involving 4449 school-age children reported that an increase in one standard deviation score (SDS) on maternal serum concentrations of folate was associated with a decreased BMI in the offspring (-0.04 SDS, 95% CI = -0.08 ~ -0.01) [38]. Similarly, a cohort study with 1517 mother–child dyads in Boston, USA further showed that pregnant mothers with the lowest quantile of plasma folate concentration (6.64~20.36 nmol/L) were significantly more likely to have offspring who were overweight or obese (OR = 1.45, 95% CI = 1.13~1.87) [49]. In contrast, a study in Pune, India found that maternal erythrocyte folate concentrations at 28 weeks of pregnancy was positively related to the offspring's fat mass at 6 years old [39]. Interestingly, our study found that a prenatal maternal folic acid supplement reduced the risk of obesity in preschool girls born SGA. Inconsistent findings of these studies might arise from differences of prenatal maternal folic acid measurement, and the selection of adjusted confounders in these studies.

Our findings raise the question of what the biomedical mechanism may be by which prenatal supplements of folic acid affect childhood obesity? Folic acid is pivotal for cellular growth, nucleic acid synthesis, and as a classic one-carbon metabolite [50]. It is also important in the synthesis of S-adenosylmethionine, which is the main methyl donor and affects DNA methylation reactions, gene expression, and chronic disease development [51–54]. Prior studies have demonstrated the influence of prenatal maternal folic acid intake on offspring's DNA methylation of IGF2, leptin, and retinoid X receptor- α gene, which have all been shown to be related to growth, energy metabolism, and appetite regulation that affected energy balance and obesity [52,55]. For example, hypomethylation of leptin is recognized as decreasing the risk of obesity through increasing leptin expression and restraining appetite [52]. However, a rat model has revealed that maternal intake of folic acid, at more than the recommended dose, may be associated with a lower expression of leptin receptor and proopiomelanocortin, failing to suppress food intake [56]. Moreover, a prenatal supplement of folic acid appears to maintain normal placenta structure and functions through anti-inflammatory effects [57,58], and ultimately works in programming chronic diseases [59]. Unfortunately, we are far from confirming the precise biomedical mechanisms that explain the associations identified in our study, so further studies are needed to answer questions about causal mechanisms as well as dose-dependent responses.

4.2. Modification Effects of Prenatal Supplement of Iron or Multivitamin on Associations between Prenatal Maternal Supplementation of Folic Acid and Obesity

A systematic analysis indicated that multi-micronutrient (MMN) containing iron and folic acid could reduce more preterm birth, SGA, and low birth weight (LBW) more than supplementation of iron or folic acid alone did in pregnant mothers [42]. This finding has been supported by an intervention study by Yijun Kang et al., who also reported similar results [60]. Additionally, a double-blind randomized controlled trial in China found that the incidence of preterm birth was 4.2% for prenatal supplementation of iron and folic acid and 4.6% prenatal supplementation of folic acid, respectively [61]. In line with these previous findings, our study found that maternal supplementation of both folic acid and iron in pregnancy reduced the risk of obesity among preschoolers born SGA in girls with statistical significance, and in boys without significance, compared

with maternal supplementation of folic acid during pregnancy only. The mechanisms for prenatal supplement of iron strengthening the effect on prenatal supplementation of folic acid reducing the risk of childhood obesity may be related to iron supplementation affecting the transcription of folic acid transporters [62] and regulating the metabolism of folate-activated one-carbon units [63].

There is no report on the combination effects of prenatal supplement of folic acid and multivitamin on childhood obesity. Our study found that maternal supplementation of both folic acid and iron in pregnancy significantly reduced the risk of obesity among preschoolers born SGA in girls, but its extent was less than maternal supplementation of folic acid only. Regarding the mechanisms for prenatal multivitamin supplement modifying the prenatal supplementation of folic acid reducing the risk of childhood obesity, vitamin B₂ could promote the metabolism of folate in body [64], while vitamin B₆ and vitamin B₁₂ could modulate one-carbon metabolism to affect DNA methylation [65,66]. All of these evidences implicated that B-vitamins contained in a multivitamin might modify prenatal folic acid supplement, decreasing the risk of offspring's obesity.

Of course, further in-depth animal experiments and epidemiological research are needed to clarify the mechanisms for prenatal supplement of iron or multivitamin modifying prenatal maternal supplementation of folic acid reducing the risk of obesity in offspring.

4.3. Sex Specific Differences of Associations between Prenatal Supplement of Folic Acid, Iron, and Multivitamin and Offspring's Obesity

There have been several studies investigating the sex-specific effects of prenatal folic acid supplement on offspring's health outcomes. For example, a Spanish multicenter longitudinal study reported that low (<400 µg/day) folic acid supplements use during pregnancy was associated with poorer attentional function in girls and poorer working memory in boys, and high (≥1000 µg/day) folic acid supplement was associated with a better working memory only in girls [67]. Moreover, the effects of prenatal folic acid supplementation on body weight and insulin resistance were sex-dependent in a rat experiment, which showed that maternal folic acid supplementation could increase body weight and insulin sensitivity in male offspring, but decrease body weight and increase insulin sensitivity in female offspring [56]. Similarly, our study found that maternal folic acid supplementation in pregnancy was significantly and negatively associated with obesity in preschooler girls born SGA, but not in preschooler boys born SGA.

The sex-specific effects of prenatal folic acid supplement on offspring's health outcomes might be explained by the following reasons. First, the aforementioned rat study showed that genes related to appetite regulation, such as proopiomelanocortin, neuropeptide Y, leptin receptor, and agouti-related protein, were expressed differently in males and females [56]. Second, several researchers have suggested that different sex-specific methylation patterns might explain the sex-specific effects. For example, a trial in mice showed that a high fat diet could lead to global DNA hypomethylation, but only in females [68]. Similarly, one of our previous studies found that placenta areas mediated the positive association between DNA methylation of FGFR2 in placenta and full-term low birth weight only in girls, but not in boys [69]. Moreover, Sinclair's sheep models showed that restricting folate in pregnancy caused 53% of changed loci of DNA methylation exclusive for males, while only 12% specific to females [70]. Third, adipose or body composition diverged between boys and girls as early as the fetus period, which might be another biological basis for the sex differences identified [71]. Further studies are needed to decipher the sex-specific responses to the maternal prenatal supplementations of nutrients and the mechanisms lying behind these sex differences.

4.4. Limitation

Some limitations should be considered when interpreting the results. First, our findings are based upon a retrospective observation study, so caution is required when interpreting a causal relationship. Second, all variables in this study were collected by a

self-administered questionnaire, which might be affected by memory bias and so influence the validity and reliability of our measures of prenatal nutrient supplementation. Third, unfortunately we did not collect any detailed information about the dose, frequency, and period of maternal nutrients supplement, limiting our ability to fully assess the association between prenatal nutrients supplement and offspring' obesity. Fourth, we did not collect the information on the specific vitamins that comprised the multivitamin used by the mothers during pregnancy, so we were unable to identify the associations between specific combinations of vitamin supplementation and offspring' obesity. Fifth, we were unable to assess the level of relevant nutrients (e.g., folic acid, vitamins, iron, etc.) ingested from diet during pregnancy, and so we could not control for their influence on our findings [52]. Sixth, although BMI measurement is a standard method of categorizing obese and non-obese children, it is imprecise in measuring the degree and location of body fat [72]. As such, other measures, such as bioelectric impedance and skin fold thickness, should be included in future studies. Notably, different definitions of SGA and obesity may lead to the different classification of some participants, which would undoubtedly affect the results. The differences in definitions should be considered when comparing our results with other studies. Seventh, we have excluded 902 pairs (10.11%) for missing information, which might have led to selection bias. Eighth, obesity is a complex disorder caused by multiple factors, and we acknowledged other potential confounders, such as paternal obesity, maternal diet, and genetic factors, which were not available for this study sample, and may disturb the true association between prenatal supplement of nutrients and preschool obesity. Lastly, data collection was limited to the Longhua District of Shenzhen, and so, consequently, it is possible there may be some limits to the generalizability of our findings to other regions due to cultural differences in diet (e.g., folic acid fortification cereal in North America).

5. Conclusions

In summary, prenatal supplementation of folic acid was found to be associated with a decreased risk of obesity in preschool girls born SGA. This effect was modified by prenatal supplementations of multivitamin and iron. These findings support public health programs that encourage appropriate prenatal maternal folic acid supplementation to reduce obesity levels in SGA girls.

Author Contributions: Conceptualization, W.-Q.C. and Q.L.; methodology, W.-Q.C. and Q.L.; formal analysis, Q.L.; investigation, Q.L. and Y.L.; resources, W.-Q.C.; data curation, B.-J.H.; writing—original draft preparation, Q.L.; writing—review and editing, W.-Q.C. and E.S.; visualization, Q.L.; supervision, W.-Q.C. and L.-H.H.; project administration, W.-Q.C. and B.-J.H. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Science and Technology Planning Project of Guangdong Province, grant number 2019A1515011915. The funding source was non-profit scientific research management and academic institutions, it had no role in the design of this study, and did not have any role during its execution, analyses, interpretation of the data, or decision to submit results.

Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki, and approved by the Institutional Review Board of the School of Public Health, Sun Yat-sen University, Guangzhou, China (protocol code: No.2015-016).

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study. Parents/guardians provided written assent for children.

Data Availability Statement: The datasets generated and/or analyzed during the current study are not publicly available due to privacy protection of the participants, but are available from the corresponding author on reasonable request.

Acknowledgments: The authors would like to thank all participants in the studies and the clinicians of the Longhua Maternity & Child Healthcare Hospital involved in recruiting participants and collecting data.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

References

1. Stanaway, J.D.; Afshin, A.; Gakidou, E.; Lim, S.S.; Abate, D.; Abate, K.H. Global, regional, and national comparative risk assessment of 84 behavioural, environmental and occupational, and metabolic risks or clusters of risks for 195 countries and territories, 1990–2017: A systematic analysis for the Global Burden of Disease Study 2017. *Lancet* **2018**, *392*, 1923–1994. [\[CrossRef\]](#)
2. Bentham, J.; Di Cesare, M.; Bilano, V.; Bixby, H.; Zhou, B.; Stevens, G.A.; Riley, L.M.; Taddei, C.; Hajifathalian, K.; Lu, Y.; et al. Worldwide trends in body-mass index, underweight, overweight, and obesity from 1975 to 2016: A pooled analysis of 2416 population-based measurement studies in 128.9 million children, adolescents, and adults. *Lancet* **2017**, *390*, 2627–2642. [\[CrossRef\]](#)
3. Capital Institute of Pediatrics; The Coordinating Study Group of Nine Cities on the Physical. A national epidemiological survey on obesity of children under seven years of age in nine cities of China in 2016. *Zhonghua Er Ke Za Zhi* **2018**, *56*, 745–752. [\[CrossRef\]](#)
4. Weihrauch-Blüher, S.; Schwarz, P.; Klusmann, J.H. Childhood obesity: Increased risk for cardiometabolic disease and cancer in adulthood. *Metabolism* **2019**, *92*, 147–152. [\[CrossRef\]](#)
5. Simmonds, M.; Llewellyn, A.; Owen, C.G.; Woolacott, N. Predicting adult obesity from childhood obesity: A systematic review and meta-analysis. *Obes. Rev.* **2016**, *17*, 95–107. [\[CrossRef\]](#) [\[PubMed\]](#)
6. Hamilton, D.; Dee, A.; Perry, I.J. The lifetime costs of overweight and obesity in childhood and adolescence: A systematic review. *Obes. Rev.* **2018**, *19*, 452–463. [\[CrossRef\]](#) [\[PubMed\]](#)
7. Quan, L.L.; Wang, H.; Tian, Y.; Mu, X.; Zhang, Y.; Tao, K. Association of fat-mass and obesity-associated gene FTO rs9939609 polymorphism with the risk of obesity among children and adolescents: A meta-analysis. *Eur. Rev. Med. Pharmacol. Sci.* **2015**, *19*, 614–623.
8. Wang, H.J.; Hinney, A.; Song, J.Y.; Scherag, A.; Meng, X.R.; Grallert, H.; Illig, T.; Hebebrand, J.; Wang, Y.; Ma, J. Association of common variants identified by recent genome-wide association studies with obesity in Chinese children: A case-control study. *BMC Med. Genet.* **2016**, *17*, 7. [\[CrossRef\]](#)
9. Raskilene, A.; Smalinskiene, A.; Kriaucioniene, V.; Lesauskaite, V.; Petkeviciene, J. Associations of MC4R, LEP, and LEPR Polymorphisms with Obesity-Related Parameters in Childhood and Adulthood. *Genes* **2021**, *12*, 949. [\[CrossRef\]](#)
10. Mărginean, C.O.; Mărginean, C.; Meliț, L.E. New Insights Regarding Genetic Aspects of Childhood Obesity: A Minireview. *Front. Pediatr.* **2018**, *6*, 271. [\[CrossRef\]](#)
11. Larqué, E.; Labayen, I.; Flodmark, C.E.; Lissau, I.; Czernin, S.; Moreno, L.A.; Pietrobelli, A.; Widhalm, K. From conception to infancy—Early risk factors for childhood obesity. *Nat. Rev. Endocrinol.* **2019**, *15*, 456–478. [\[CrossRef\]](#) [\[PubMed\]](#)
12. Weng, S.F.; Redsell, S.A.; Swift, J.A.; Yang, M.; Glazebrook, C.P. Systematic review and meta-analyses of risk factors for childhood overweight identifiable during infancy. *Arch. Dis. Child.* **2012**, *97*, 1019–1026. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Gurnani, M.; Birken, C.; Hamilton, J. Childhood Obesity: Causes, Consequences, and Management. *Pediatr. Clin. N. Am.* **2015**, *62*, 821–840. [\[CrossRef\]](#) [\[PubMed\]](#)
14. Biosca, M.; Rodríguez, G.; Ventura, P.; Samper, M.P.; Labayen, I.; Collado, M.P.; Valle, S.; Bueno, O.; Santabábara, J.; Moreno, L.A. Central adiposity in children born small and large for gestational age. *Nutr. Hosp.* **2011**, *26*, 971–976. [\[CrossRef\]](#)
15. Yuan, Z.P.; Yang, M.; Liang, L.; Fu, J.F.; Xiong, F.; Liu, G.L.; Gong, C.X.; Luo, F.H.; Chen, S.K.; Zhang, D.D.; et al. Possible role of birth weight on general and central obesity in Chinese children and adolescents: A cross-sectional study. *Ann. Epidemiol.* **2015**, *25*, 748–752. [\[CrossRef\]](#)
16. Goedegebuure, W.J.; Van der Steen, M.; Smeets, C.C.J.; Kerkhof, G.F.; Hokken-Koelega, A.C.S. SGA-born adults with postnatal catch-up have a persistently unfavourable metabolic health profile and increased adiposity at age 32 years. *Eur. J. Endocrinol.* **2022**, *187*, 15–26. [\[CrossRef\]](#) [\[PubMed\]](#)
17. Sharma, D.; Shastri, S.; Farahbakhsh, N.; Sharma, P. Intrauterine growth restriction—Part 1. *J. Matern Fetal Neonatal Med.* **2016**, *29*, 3977–3987. [\[CrossRef\]](#)
18. Sacchi, C.; Marino, C.; Nosarti, C.; Vieno, A.; Visentin, S.; Simonelli, A. Association of Intrauterine Growth Restriction and Small for Gestational Age Status With Childhood Cognitive Outcomes: A Systematic Review and Meta-analysis. *JAMA Pediatr.* **2020**, *174*, 772–781. [\[CrossRef\]](#)
19. Kemp, M.W.; Kallapur, S.G.; Jobe, A.H.; Newnham, J.P. Obesity and the developmental origins of health and disease. *J. Paediatr. Child. Health* **2012**, *48*, 86–90. [\[CrossRef\]](#)
20. Nam, H.K.; Lee, K.H. Small for gestational age and obesity: Epidemiology and general risks. *Ann. Pediatr. Endocrinol. Metab.* **2018**, *23*, 9–13. [\[CrossRef\]](#)
21. Maguolo, A.; Olivieri, F.; Zusi, C.; Miraglia Del Giudice, E.; Morandi, A.; Maffei, C. The risk of metabolic derangements is higher in children and adolescents with overweight or obesity born small for gestational age. *Nutr. Metab. Cardiovasc. Dis.* **2021**, *31*, 1903–1910. [\[CrossRef\]](#)
22. Colella, M.; Frérot, A.; Novais, A.R.B.; Baud, O. Neonatal and Long-Term Consequences of Fetal Growth Restriction. *Curr. Pediatr. Rev.* **2018**, *14*, 212–218. [\[CrossRef\]](#) [\[PubMed\]](#)
23. Banister, C.E.; Koestler, D.C.; Maccani, M.A.; Padbury, J.F.; Houseman, E.A.; Marsit, C.J. Infant growth restriction is associated with distinct patterns of DNA methylation in human placentas. *Epigenetics* **2011**, *6*, 920–927. [\[CrossRef\]](#) [\[PubMed\]](#)

24. Sharma, D.; Farahbakhsh, N.; Shastri, S.; Sharma, P. Intrauterine growth restriction—Part 2. *J. Matern Fetal Neonatal. Med.* **2016**, *29*, 4037–4048. [\[CrossRef\]](#)
25. Christian, P.; Stewart, C.P. Maternal micronutrient deficiency, fetal development, and the risk of chronic disease. *J. Nutr.* **2010**, *140*, 437–445. [\[CrossRef\]](#)
26. Priliani, L.; Oktavianthi, S.; Prado, E.L.; Malik, S.G.; Shankar, A.H. Maternal biomarker patterns for metabolism and inflammation in pregnancy are influenced by multiple micronutrient supplementation and associated with child biomarker patterns and nutritional status at 9–12 years of age. *PLoS ONE* **2020**, *15*, e0216848. [\[CrossRef\]](#)
27. Stewart, C.P.; Christian, P.; Schulze, K.J.; Leclercq, S.C.; West, K.P., Jr.; Khatry, S.K. Antenatal micronutrient supplementation reduces metabolic syndrome in 6- to 8-year-old children in rural Nepal. *J. Nutr.* **2009**, *139*, 1575–1581. [\[CrossRef\]](#)
28. Georgieff, M.K. Iron deficiency in pregnancy. *Am. J. Obstet. Gynecol.* **2020**, *223*, 516–524. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Roberfroid, D.; Huybregts, L.; Lanou, H.; Habicht, J.P.; Henry, M.C.; Meda, N.; Kolsteren, P. Prenatal micronutrient supplements cumulatively increase fetal growth. *J. Nutr.* **2012**, *142*, 548–554. [\[CrossRef\]](#)
30. Papadopoulou, E.; Stratakis, N.; Roumeliotaki, T.; Sarri, K.; Merlo, D.F.; Kogevinas, M.; Chatzi, L. The effect of high doses of folic acid and iron supplementation in early-to-mid pregnancy on prematurity and fetal growth retardation: The mother-child cohort study in Crete, Greece (Rhea study). *Eur. J. Nutr.* **2013**, *52*, 327–336. [\[CrossRef\]](#)
31. Wang, C.; Gao, R.; Huang, L.; Hu, P.; Zhu, L.; Chen, W.Q. Effect of prenatal nutritional intervention on foetal growth restriction: A real-world study in Shenzhen, China. *J. Matern Fetal Neonatal Med.* **2022**, *35*, 2435–2444. [\[CrossRef\]](#) [\[PubMed\]](#)
32. Sauder, K.A.; Starling, A.P.; Shapiro, A.L.; Kaar, J.L.; Ringham, B.M.; Glueck, D.H.; Dabelea, D. Exploring the association between maternal prenatal multivitamin use and early infant growth: The Healthy Start Study. *Pediatr. Obes.* **2016**, *11*, 434–441. [\[CrossRef\]](#)
33. Herath, M.P.; Ahuja, K.D.K.; Beckett, J.M.; Jayasinghe, S.; Byrne, N.M.; Hills, A.P. Determinants of Infant Adiposity across the First 6 Months of Life: Evidence from the Baby-bod study. *J. Clin. Med.* **2021**, *10*, 1770. [\[CrossRef\]](#) [\[PubMed\]](#)
34. Balashova, O.A.; Visina, O.; Borodinsky, L.N. Folate action in nervous system development and disease. *Dev. Neurobiol.* **2018**, *78*, 391–402. [\[CrossRef\]](#)
35. Li, N.; Li, Z.; Ye, R.; Liu, J.; Ren, A. Impact of Periconceptional Folic Acid Supplementation on Low Birth Weight and Small-for-Gestational-Age Infants in China: A Large Prospective Cohort Study. *J. Pediatr.* **2017**, *187*, 105–110. [\[CrossRef\]](#) [\[PubMed\]](#)
36. Li, S.; Liu, D.; Zhang, R.; Lei, F.; Liu, X.; Cheng, Y.; Li, C.; Xiao, M.; Guo, L.; Li, M.; et al. The association of maternal dietary folate intake and folic acid supplementation with small-for-gestational-age births: A cross-sectional study in Northwest China. *Br. J. Nutr.* **2019**, *122*, 459–467. [\[CrossRef\]](#)
37. Waterland, R.A.; Travisano, M.; Tahiliani, K.G.; Rached, M.T.; Mirza, S. Methyl donor supplementation prevents transgenerational amplification of obesity. *Int. J. Obes.* **2008**, *32*, 1373–1379. [\[CrossRef\]](#)
38. Monasso, G.S.; Santos, S.; Geurtsen, M.L.; Heil, S.G.; Felix, J.F.; Jaddoe, V.W.V. Associations of Early Pregnancy and Neonatal Circulating Folate, Vitamin B-12, and Homocysteine Concentrations with Cardiometabolic Risk Factors in Children at 10 y of Age. *J. Nutr.* **2021**, *151*, 1628–1636. [\[CrossRef\]](#)
39. Yajnik, C.S.; Deshpande, S.S.; Jackson, A.A.; Refsum, H.; Rao, S.; Fisher, D.J.; Bhat, D.S.; Naik, S.S.; Coyaji, K.J.; Joglekar, C.V.; et al. Vitamin B12 and folate concentrations during pregnancy and insulin resistance in the offspring: The Pune Maternal Nutrition Study. *Diabetologia* **2008**, *51*, 29–38. [\[CrossRef\]](#)
40. Capital Institute of Pediatrics; The Coordinating Study Group of Nine Cities on the Physical. Growth standard curves of birth weight, length and head circumference of Chinese newborns of different gestation. *Zhonghua Er Ke Za Zhi* **2020**, *58*, 738–746. [\[CrossRef\]](#)
41. Bian, H.; Tang, Y.; Zhou, Y.; Li, H.; Liu, J. Demographic variations and temporal trends in prenatal use of multiple micronutrient supplements in Beijing, 2013–2017. *Public Health Nutr.* **2021**, *24*, 826–833. [\[CrossRef\]](#) [\[PubMed\]](#)
42. Keats, E.C.; Haider, B.A.; Tam, E.; Bhutta, Z.A. Multiple-micronutrient supplementation for women during pregnancy. *Cochrane Database Syst. Rev.* **2019**, *3*, Cd004905. [\[CrossRef\]](#) [\[PubMed\]](#)
43. Shand, A.W.; Walls, M.; Chatterjee, R.; Nassar, N.; Khambalia, A.Z. Dietary vitamin, mineral and herbal supplement use: A cross-sectional survey of before and during pregnancy use in Sydney, Australia. *Aust. New Zealand J. Obstet. Gynaecol.* **2016**, *56*, 154–161. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Koivuniemi, E.; Hart, K.; Mazanowska, N.; Ruggeri, S.; Egan, B.; Censi, L.; Roccaldo, R.; Mattila, L.; Buonocore, P.; Löyttyniemi, E.; et al. Food Supplement Use Differs from the Recommendations in Pregnant Women: A Multinational Survey. *Nutrients* **2022**, *14*, 2909. [\[CrossRef\]](#)
45. Li, H.; Ji, C.Y.; Zong, X.N.; Zhang, Y.Q. Body mass index growth curves for Chinese children and adolescents aged 0 to 18 years. *Zhonghua Er Ke Za Zhi* **2009**, *47*, 493–498. [\[CrossRef\]](#)
46. Kunaratnam, K.; Halaki, M.; Wen, L.M.; Baur, L.A.; Flood, V.M. Tracking Preschoolers' Lifestyle Behaviors and Testing Maternal Sociodemographics and BMI in Predicting Child Obesity Risk. *J. Nutr.* **2020**, *150*, 3068–3074. [\[CrossRef\]](#)
47. Morgen, C.S.; Ängquist, L.; Baker, J.L.; Andersen, A.M.N.; Michaelsen, K.F.; Sørensen, T.I.A. Prenatal risk factors influencing childhood BMI and overweight independent of birth weight and infancy BMI: A path analysis within the Danish National Birth Cohort. *Int. J. Obes.* **2018**, *42*, 594–602. [\[CrossRef\]](#)
48. Xie, R.H.; Liu, Y.J.; Retnakaran, R.; MacFarlane, A.J.; Hamilton, J.; Smith, G.; Walker, M.C.; Wen, S.W. Maternal folate status and obesity/insulin resistance in the offspring: A systematic review. *Int. J. Obes.* **2016**, *40*, 1–9. [\[CrossRef\]](#)

49. Wang, G.; Hu, F.B.; Mistry, K.B.; Zhang, C.; Ren, F.; Huo, Y.; Paige, D.; Bartell, T.; Hong, X.; Caruso, D.; et al. Association Between Maternal Prepregnancy Body Mass Index and Plasma Folate Concentrations With Child Metabolic Health. *JAMA Pediatr.* **2016**, *170*, e160845. [\[CrossRef\]](#)
50. Greenberg, J.A.; Bell, S.J.; Guan, Y.; Yu, Y.H. Folic Acid supplementation and pregnancy: More than just neural tube defect prevention. *Rev. Obstet. Gynecol.* **2011**, *4*, 52–59.
51. Qian, Y.Y.; Huang, X.L.; Liang, H.; Zhang, Z.F.; Xu, J.H.; Chen, J.P.; Yuan, W.; He, L.; Wang, L.; Miao, M.H.; et al. Effects of maternal folic acid supplementation on gene methylation and being small for gestational age. *J. Hum. Nutr. Diet* **2016**, *29*, 643–651. [\[CrossRef\]](#) [\[PubMed\]](#)
52. Pauwels, S.; Ghosh, M.; Duca, R.C.; Bekaert, B.; Freson, K.; Huybrechts, I.; Langie, S.A.S.; Koppen, G.; Devlieger, R.; Godderis, L. Maternal intake of methyl-group donors affects DNA methylation of metabolic genes in infants. *Clin. Epigenetics* **2017**, *9*, 16. [\[CrossRef\]](#) [\[PubMed\]](#)
53. McGee, M.; Bainbridge, S.; Fontaine-Bisson, B. A crucial role for maternal dietary methyl donor intake in epigenetic programming and fetal growth outcomes. *Nutr. Rev.* **2018**, *76*, 469–478. [\[CrossRef\]](#) [\[PubMed\]](#)
54. Salbaum, J.M.; Kappen, C. Genetic and epigenomic footprints of folate. *Prog. Mol. Biol. Transl. Sci.* **2012**, *108*, 129–158. [\[CrossRef\]](#)
55. Steegers-Theunissen, R.P.; Obermann-Borst, S.A.; Kremer, D.; Lindemans, J.; Siebel, C.; Steegers, E.A.; Slagboom, P.E.; Heijmans, B.T. Periconceptional maternal folic acid use of 400 microg per day is related to increased methylation of the IGF2 gene in the very young child. *PLoS ONE* **2009**, *4*, e7845. [\[CrossRef\]](#)
56. Huot, P.S.; Ly, A.; Szeto, I.M.; Reza-López, S.A.; Cho, D.; Kim, Y.I.; Anderson, G.H. Maternal and postweaning folic acid supplementation interact to influence body weight, insulin resistance, and food intake regulatory gene expression in rat offspring in a sex-specific manner. *Appl. Physiol. Nutr. Metab.* **2016**, *41*, 411–420. [\[CrossRef\]](#)
57. Pickell, L.; Li, D.; Brown, K.; Mikael, L.G.; Wang, X.L.; Wu, Q.; Luo, L.; Jerome-Majewska, L.; Rozen, R. Methylenetetrahydrofolate reductase deficiency and low dietary folate increase embryonic delay and placental abnormalities in mice. *Birth Defects Res. A Clin. Mol. Teratol.* **2009**, *85*, 531–541. [\[CrossRef\]](#)
58. Zhao, M.; Chen, Y.H.; Dong, X.T.; Zhou, J.; Chen, X.; Wang, H.; Wu, S.X.; Xia, M.Z.; Zhang, C.; Xu, D.X. Folic acid protects against lipopolysaccharide-induced preterm delivery and intrauterine growth restriction through its anti-inflammatory effect in mice. *PLoS ONE* **2013**, *8*, e82713. [\[CrossRef\]](#)
59. Hofstee, P.; McKeating, D.R.; Perkins, A.V.; Cuffe, J.S. Placental adaptations to micronutrient dysregulation in the programming of chronic disease. *Clin. Exp. Pharmacol. Physiol.* **2018**, *45*, 871–884. [\[CrossRef\]](#)
60. Kang, Y.; Dang, S.; Zeng, L.; Wang, D.; Li, Q.; Wang, J.; Ouzhu, L.; Yan, H. Multi-micronutrient supplementation during pregnancy for prevention of maternal anaemia and adverse birth outcomes in a high-altitude area: A prospective cohort study in rural Tibet of China. *Br. J. Nutr.* **2017**, *118*, 431–440. [\[CrossRef\]](#)
61. Li, Z.; Mei, Z.; Zhang, L.; Li, H.; Zhang, Y.; Li, N.; Ye, R.; Ren, A.; Liu, J.M.; Serdula, M.K. Effects of Prenatal Micronutrient Supplementation on Spontaneous Preterm Birth: A Double-Blind Randomized Controlled Trial in China. *Am. J. Epidemiol.* **2017**, *186*, 318–325. [\[CrossRef\]](#) [\[PubMed\]](#)
62. Radziejewska, A.; Suliburska, J.; Kołodziejski, P.; Chmurzynska, A. Simultaneous supplementation with iron and folic acid can affect Slc11a2 and Slc46a1 transcription and metabolite concentrations in rats. *Br. J. Nutr.* **2020**, *123*, 264–272. [\[CrossRef\]](#) [\[PubMed\]](#)
63. Oppenheim, E.W.; Adelman, C.; Liu, X.; Stover, P.J. Heavy chain ferritin enhances serine hydroxymethyltransferase expression and de novo thymidine biosynthesis. *J. Biol. Chem.* **2001**, *276*, 19855–19861. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Powers, H.J. Riboflavin (vitamin B-2) and health. *Am. J. Clin. Nutr.* **2003**, *77*, 1352–1360. [\[CrossRef\]](#) [\[PubMed\]](#)
65. Franco, C.N.; Seabrook, L.J.; Nguyen, S.T.; Leonard, J.T.; Albrecht, L.V. Simplifying the B Complex: How Vitamins B6 and B9 Modulate One Carbon Metabolism in Cancer and Beyond. *Metabolites* **2022**, *12*, 961. [\[CrossRef\]](#) [\[PubMed\]](#)
66. Mahajan, A.; Sapehia, D.; Thakur, S.; Mohanraj, P.S.; Bagga, R.; Kaur, J. Effect of imbalance in folate and vitamin B12 in maternal/parental diet on global methylation and regulatory miRNAs. *Sci. Rep.* **2019**, *9*, 17602. [\[CrossRef\]](#) [\[PubMed\]](#)
67. Compañ-Gabucio, L.M.; Torres-Collado, L.; Garcia-de la Hera, M.; Fernández-Somoano, A.; Tardón, A.; Julvez, J.; Sunyer, J.; Rebagliato, M.; Murcia, M.; Ibarluzea, J.; et al. Association between the Use of Folic Acid Supplements during Pregnancy and Children's Cognitive Function at 7-9 Years of Age in the INMA Cohort Study. *Int. J. Environ. Res. Public Health* **2022**, *19*, 12123. [\[CrossRef\]](#) [\[PubMed\]](#)
68. Gallou-Kabani, C.; Gabory, A.; Tost, J.; Karimi, M.; Mayeur, S.; Lesage, J.; Boudadi, E.; Gross, M.S.; Taurelle, J.; Vigé, A.; et al. Sex- and diet-specific changes of imprinted gene expression and DNA methylation in mouse placenta under a high-fat diet. *PLoS ONE* **2010**, *5*, e14398. [\[CrossRef\]](#) [\[PubMed\]](#)
69. Tian, F.Y.; Wang, X.M.; Xie, C.; Zhao, B.; Niu, Z.; Fan, L.; Hivert, M.F.; Chen, W.Q. Placental surface area mediates the association between FGFR2 methylation in placenta and full-term low birth weight in girls. *Clin. Epigenetics* **2018**, *10*, 39. [\[CrossRef\]](#) [\[PubMed\]](#)
70. Sinclair, K.D.; Allegrucci, C.; Singh, R.; Gardner, D.S.; Sebastian, S.; Bispham, J.; Thurston, A.; Huntley, J.F.; Rees, W.D.; Maloney, C.A.; et al. DNA methylation, insulin resistance, and blood pressure in offspring determined by maternal periconceptional B vitamin and methionine status. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 19351–19356. [\[CrossRef\]](#) [\[PubMed\]](#)

-
71. Wells, J.C. Sexual dimorphism of body composition. *Best Pract. Res. Clin. Endocrinol. Metab.* **2007**, *21*, 415–430. [[CrossRef](#)] [[PubMed](#)]
 72. Adab, P.; Pallan, M.; Whincup, P.H. Is BMI the best measure of obesity? *BMJ* **2018**, *360*, k1274. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.