

Article

Gene Influence in the Effectiveness of Plant Sterols Treatment in Children: Pilot Interventional Study

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Abstract: Cardiovascular disease is linked to high serum low density lipoprotein (LDL)-cholesterol levels. Cardiovascular risk may be indirectly influenced by genetic load. Serum LDL-cholesterol levels may be reduced by the consumption of food enriched with plant sterols (PS). The aim was to test a plant sterol treatment on cholesterol levels according to different genetic polymorphisms. A pilot interventional trial was performed in 26 children (n = 16 girls, n = 10 boys). Seven hundred milliliters/day of commercial skimmed milk with added plant sterols delivering 2.2 g plant sterols were ingested for three weeks. Blood draws were performed at the baseline and end of the study. Significant modifications of non-high density lipoprotein (HDL)-cholesterol (p = 0.010; p = 0.013) and LDL-cholesterol (p = 0.004; p = 0.013) levels appeared in the genes *LIPC C-514T* and *PPAR-a L162V* carriers. No statistically significant differences were observed for other genes. *LIPC C-514T* and *PPAR-alpha L162V* carriers could benefit from a plant sterol supplement to ameliorate hypercholesterolemia.

Keywords: children; cholesterol; genetics; low-density lipoprotein cholesterol; sterol

1. Introduction

Cardiovascular diseases (CVD) have gained relevance through the past years due to their importance in public health. They are the main cause of death around the globe, according to the World Health Organization [1].

Cholesterol levels are known determinants of cardiovascular health [2]. Atherosclerotic lesions start appearing in childhood, making cholesterol levels especially important on pediatric age [3].

Familiar hypercholesterolemia is the most prevalent genetic disorder among children and teenagers [4]. These children are a risk group for the development of CVD. Familial hypercholesterolemia results in alterations in carotid intima-media thickness, vessel walls and flow-mediated dilatation [2]. When strategies for lowering low-density lipoprotein cholesterol (LDL-cholesterol) levels are applied to these children, there is a deceleration in the progress of the lesions [5].

Since the modification of lifestyle factors is generally useful to control the lipidic profile, public health strategies base their recommendations on changes in dietary habits and physical activity [6]. Those dietary changes include limiting saturated fat and cholesterol intake [2]. Also, plant sterols (PS) seem to be beneficial for regulating hypercholesterolemia [7].



When a person ingests a daily dose of 1.5–2.4 g of PS, blood cholesterol may be lowered by 7–10.5% within 2–3 weeks, according to EFSA (European Food Safety Authority) [8]. The reduction of serum cholesterol could be sustained for 85 weeks, as mid- and long-term studies show [9]. It is important to note that the success of those changes depend on the genetic heritage [6] and its interactions with nutrients [10,11].

Apolipoprotein A5 gene, in its naturally occurring variants, confers risk for CVD and is associated with high triglyceride levels. Congenital heart diseases and CVD risk are increased when carrying *MTHFR* gene polymorphisms [12], more specifically when carrying the *C677T* polymorphism [13]. Another gene relevant to cholesterol metabolism is the hepatic triglyceride lipase gene (*LIPC*). LIPC not only works as a hydrolase but also as a ligand factor for lipoprotein uptake [14]. *Lipoprotein (a)* is a well-known predictor for stroke, coronary heart disease and other atherosclerotic diseases [15]. This is due to its connection with low-density lipoprotein cholesterol (LDL-cholesterol) [16]. Additionally, *APOE* directly influences the catabolism of lipoproteins [17].

Previous research has been carried out on the effect of variations in genes such as *ABCG5/G8*, *NPC1L1*, *APOA4*, *SR-BI*, *HMG-CoA*, *CETP*, *APOE* and *CYP7A1* in relation to the cholesterol-lowering response to plant sterols/stanols [18–22]. In addition, several reviews have addressed the impact of genes on the cholesterol-lowering efficacy of plant sterols/stanols [23].

The aim of this trial was to determine whether the intake of 2.2 g of PS in milk would influence cholesterol levels in children, according to different genetic polymorphisms.

2. Materials and Methods

2.1. Study Design

This pilot study was designed as an interventional study. Participants were recruited at Hospital El Escorial in Madrid. Parents' consent and children's assent were obtained before the commencement of the trial.

Naturcol milk with PS (Corporación Alimentaria Peñasanta, S.A., Spain) was supplied to the participants throughout the study. Naturcol milk nutritional composition, per 250 mL, was: 36 kcal, 0.5 g fat, 0.2 g saturated fat, 4.7 g carbohydrates of which 4.7 g were sugars, 3.2 g protein, 0.13 g salt, and 120 mg calcium.

Two glasses, of 350 mL capacity each, were ingested by each participant on a daily basis for 3 weeks. The participants had a daily consumption of 2.24 g of PS, contained in 700 mL of regular skim milk. To be eligible for analysis, compliance had to be met and was defined as a minimum consumption of 80% according to empty milk packages returned. Blood tests were performed before and after the intervention (see Figure 1).

A total of 58 participants, 22 boys and 36 girls with a mean age of 8.82 ± 2.28 y were enrolled. Total cholesterol was used as a primary biomarker, based on the clinical practice guidelines by the Spanish Pediatrics Association (AEP) [24], to recruit participants.

2.2. Inclusion and Exclusion Criteria

Inclusion criteria: 5 to 12 years-old children, with a total cholesterol (TC) >170 mg/dL and/or LDL-cholesterol >110 mg/dL.

Exclusion criteria: TC <170 mg/dL and/or LDL-cholesterol <110 mg/dL, pubertal development at the beginning of the trial (Tanner Stage II), intolerance to lactose, allergic to proteins originating from cow's milk, galactosemia, celiac disease, any known chromosomopathies and growth hormone therapy for short stature.



Figure 1. Flow diagram of study design.

2.3. Clinical Analyses

Blood samples were taken at Hospital El Escorial in San Lorenzo de El Escorial (Madrid, Spain) by trained personnel. Participants were asked to fast 12 h prior to the extraction.

Blood was collected in gel serum tubes with an s-Monovette in aspiration [25–27]. Blood samples were refrigerated at 5 ± 3 °C after extraction and during shipment to the laboratory. Samples were centrifuged at $1200 \times g$ for 10 min at 20 ± 5 °C. The stability of the samples was 1 week at 5 ± 3 °C.

Serum total cholesterol (TC) was determined after enzymatic hydrolysis and oxidation, indicator quinoneimine is formed from hydrogen peroxide and 4-aminoantipyrine in the presence of phenol and peroxidase. Analytical ultracentrifugation with density gradient flotation was used to isolate LDL-cholesterol and HDL-cholesterol subfractions, simultaneously. Non–HDL-C was calculated by the following equation: (non-HDL-cholesterol = TC – HDL-cholesterol).

2.4. Genetics

The response level to the hypercholesterolemia plant sterol treatment according to different genetic polymorphisms was studied by means of genomic analysis. The different genetic polymorphisms studied were PPAR α L162V, LIPC C-514T, APOE APOE2/3/4, APOE 2,3,4 APOA5 C56G Ser19Trp, APOA5 1131T>C, MTHFR C677T, Prothrombin G20210A, F5 Arg506Gln, and LPA I4300M. Out of 58 participants, 26 were recruited for the genetic test: 10 boys and 16 girls with a mean age of 8.7 ± 2.06 years.

Genomic DNA was extracted from saliva samples, for genotyping the single nucleotide polymorphism (SNP) using the Biobank Axiom1 96-Array from Affymetrix. Genotype calling was

performed as stated in Affymetrix's best practice guidelines, including analysis with SNPolisher, assuming a quality control rate of >0.97 [28,29].

To extract saliva-derived DNA samples under high molecular weight and high-quality conditions, each sample was run on an agarose gel. Then, the optical density of the extracted DNA (OD260/280 ratio) was analyzed at 260 nm and 280 nm wavelengths to ensure maximum purity of the extracted DNA (ratio >1.7). Duplicate analyses were performed to obtain precise and reliable results.

2.5. Study Variables

Anthropometric measurements, including weight (kg), height (m), Body Mass Index (BMI) (kg/m²), total body fat (%), visceral fat (%) and lean body mass (kg), were taken by trained staff who used precise measuring techniques. An electrical bioimpedance InBody Model 270, tetrapolar multi-frequency (20 and 100 kHz), was used to measure weight, BMI and body composition. Manufacturer's recommendations and a standard protocol were followed to undertake the analysis.

Anthropometric measurements, along with other study variables such as age, sex, health habits, sleep quality, clinical and pharmacological history, use of tobacco and alcohol, food consumption frequency, intestinal transit and physical activity, were recorded in an ad hoc questionnaire designed for this study. The analytical markers of interest obtained from the blood analysis were total cholesterol, high-density lipoprotein cholesterol (HDL-cholesterol), LDL-cholesterol, and non-HDL-cholesterol. Confounding factors were also considered with an affinity table after ingestion (>95%), monitoring of the non-modification of baseline habits during the trial, and a record of food consumption frequencies to control the ingestion of foods that may influence the metabolism of cholesterol upwards or downwards.

2.6. Statistical Analysis

Statistical analysis of the data collected was run using the SPSS[®] (v. 21) statistical software (IBM Corp., Armonk, NY, USA). Descriptive analysis of the sociodemographic variables, anthropometric measurements, and the decrease percentage from baseline to final measures of lipid values after the ingestion of Naturcol were included. The Shapiro–Wilk test was used to determine lipid values normality. Student's T-test, for paired samples, or the Wilcoxon rank sum test according to compliance with the assumption of normality of the dependent variables was applied to analyze the efficacy of the consumption of Naturcol. The effect size and the proportion of the mean differences were estimated regarding the standard deviation of the baseline, or milk with PS. A value of p < 0.05 was considered a significant difference. The statistical power was set at 90%. Distributions calculation of the usual genetics was not taken into account, since genetic studies need larger samples to be carried out.

2.7. Ethical Standards

This study followed the ethical principles stated in the Helsinki Declaration. All procedures involving human subjects/patients were approved by the Bioethics Committee of Hospital Universitario Puerta de Hierro Majadahonda, Majadahonda, Madrid, Spain. Written informed consent was obtained from all subjects.

3. Results

Twenty-six subjects (16 girls, 10 boys) completed the trial. The remaining 32 subjects did not meet the inclusion criteria (n = 21), declined to participate (n = 2) or were not available when needed (n = 9). They had an average age of 8.7 ± 2.06 years and weighed 33.08 ± 13.00 kg (BMI 18.79 ± 4.20 kg/m²) (Table 1). Baseline TC was 236.6 mg/dL, LDL-cholesterol was 157.3 mg/dL and HDL-cholesterol 58.2 mg/dL. Demographic variables did not differ significantly between subjects.

	Total (<i>n</i> = 26) Mean	SD ^a	Males (<i>n</i> = 10) Mean	SD	Females (<i>n</i> = 16) Mean	SD	
Age (years)	8.7	2.06	8.5	2.44	8.8	1.89	
Weight (kg)	33.1	13.00	30.1	10.96	35.3	14.39	
Height (m)	1.3	0.12	1.3	10.13	1.3	0.12	
BMI b (Kg/m ²)	18.8	4.20	17.2	3.18	20.0	4.59	
Body fat (%)	25.5	9.66	20.8	6.99	29.0	10.16	
Visceral fat (kg)	3.8	4.04	2.4	2.83	5.0	4.54	
Muscle (kg)	11.5	4.46	11.1	4.1	11.9	4.86	
	Change from Baseline (%)						
Total cholesterol	-12.1	-9.71	-12.5	-7.52	-12.2	-12.62	
LDL ^c -cholesterol	-16.2	-12.78	-17.3	-11.95	-3.2	-16.42	
HDL ^d -cholesterol	-2.1	-14.44	3.3	-13.19	-15.9	-15.75	
Non-HDL-cholesterol	-15.9	-11.66	-17.3	-10.90	-15.6	-14.47	

Table 1. Descriptive statistics of the anthropometric measurements and lipid profile.

^a SD, standard deviation;
 ^b BMI, body mass index;
 ^c LDL-cholesterol, low-density lipoprotein cholesterol;
 ^d HDL-cholesterol, high-density lipoprotein cholesterol.

In Table 2, the descriptive statistics of genes and haplotypes can be found.

Gene	Haplotype	Frequency (n)	Percentage (%)
ADOA5C56CSar10Trn (rc3135506)	CG	5	19.2
AFOA5 C50G 5er1911p (155155500)	GG	21	80.8
	CC	8	30.8
MTHFR C677T (rs1801133)	CT	15	57.7
	TT	3	11.5
	CC	6	23.1
LIPC C-514T (rs1800588)	CT	13	50
	TT	7	26.9
I DA IA200 M (m 2708220)	TT	25	96.2
LPA 14500101 (7557 96220)	TC	1	3.8
DDAR alpha I 162V (rc1800206)	CC	22	84.6
11AR-upnu L102V (151800200)	CG	4	15.4
ADOAE 1121T > C (match2700)	TT	24	92.3
APOA5 11511 > C (rs662799)	TC	2	7.7
ADOE Hanlotung $ADOE2/3/4$ (rc/20358)	TT	22	84.6
AT OL Huplotype AT OL2/3/4 (1942)538)	TC	4	15.4
ADOF Hanlatime ADOF2 3 A (re7.12)	TC	5	19.2
л ос пирюкуре лг ос2,3,4 (15/412)	CC	21	80.8

Table 2. Descriptive statistics of genes and haplotypes.

No statistically significant differences (p > 0.05) were seen between genotypes of *APOEA5 MTHFR C677T*, *PPAR_ALPHA L162V*, and *APOE APOE2*, *3*, *4* genes and the lower percentage of lipid parameters (TC, HDL-cholesterol, and non-HDL-cholesterol) (Table 3).

No statistically significant differences (p > 0.05) were seen in genotypes *LIPC C-514T* and HDL-cholesterol and *PPAR-alpha L162V* and TC and HDL-cholesterol. *LIPC C-514T* showed statistically significant changes for the levels of TC (p = 0.0414), LDL-cholesterol (p = 0.004) and non-HDL-cholesterol (p = 0.010); *PPAR-alpha L162V* showed a significant difference for the levels of LDL-cholesterol and non-HDL-cholesterol (p = 0.013; p = 0.013), respectively (Table 3).

Since genes *Prothrombin G20210A* and *F5 Arg506Gln* only had one haplotype, an analytical study could not be applied. Even though there are visible differences in HDL-cholesterol and LDL-cholesterol, Table 3 shows a lack of statistical significance for many of the analysed genes. These could be due to the uneven frequency of the haplotypes in this sample.

6 of 10

Genes		Change from Baseline (%)			
		HDL-c ^a	Total	LDL-c ^b	No-HDL-c ^c
APOA5 C56G	CG	-0.30	-10.07	-15.98	-13.78
Ser19Trp	GG	-0.96	-12.8	-16.53	-16.75
	CC	9.76	-10.60	-17.04	-17.71
	CT	-6.33	-14.60	-18.21	-17.46
MTHFR C677T	TT	-3.40	-5.25	-6.39	-1.93
	CC	3.10	-5.33 *	-4.17 *	-5.58 *
LDCC514T	CT	-0.03	-11.45 *	-15.97 *	-15.67 *
LIPC C-5141	TT	-5.12	-19.82	-26.00	-24.86
1.04.1420014	TT	-0.29	-12.90	-17.49	-17.39
LPA 145001VI	TC	-13.73	2.92	9.18	10.00
DDAR almha 1162V	CC	-1.45	-13.74	-18.44 *	-18.23 *
111111- <i>uipnu</i> E102V	CG	3.70	-4.33	-1.62 *	-2.36 *
ADOAE 1121T > C	TT	-0.38	-11.84	-16.01	-15.75
APOA5 11511 > C	TC	-5.99	-17.67	-21.08	-21.77
ADOF Hanlotime ADOF2/3/1	TT	0.93	-11.58	-15.72	-15.97
AI OL Hupiolype AFOL2/3/4	TC	-13.70	-16.22	-21.58	-18.23
APOF Hanlotume APOF2 3 Λ	TC	-12.32	-16.29	-21.18	-18.71
AT OL Huplotype AFOL2,3,4	CC	1.36	-11.34	-15.51	-15.76

Table 3. Mean changes (in percent) of serum lipids after the plant sterol intervention.

^a HDL-c, high-density lipoprotein cholesterol; ^b LDL-c, low-density lipoprotein cholesterol; ^c no-HDL-c, non-high-density lipoprotein cholesterol. * Results with an asterisk indicate a p-value < 0.05.

All of the participants analyzed complied with the treatment under the guidelines given and none of them incurred adverse events.

4. Discussion

Plant sterols are known to compete with cholesterol for incorporation into micelles in the intestine and hence in the presence of plant sterols less cholesterol can be transported in micelles leading to a lesser absorption through the gut wall [30] without changing HDL-cholesterol levels. The efficacy of phytosterols to reduce significantly the serum LDL-cholesterol when incorporated into foods and supplements is well-documented [31]. An analysis of PS added to different food matrices was carried out by Clifton et al. (2004) [32]. They reached the conclusion that milk is the matrix with better results in reducing LDL-cholesterol. However, Demonty et al. (2009) [9] showed that there is no difference in the cholesterol-lowering efficacy between different food formats.

The use of PS for the treatment of CVD risk biomarkers has been covered extensively by recent scientific literature [33–36].

APOA5T-1131C is a risk factor for stroke disease sufferers, independent of the subgroup. More precisely, the C allele variant of *APOA5T-1131C* is a risk factor for ischemic stroke and heart disease [37,38], and it increases triglyceride levels in serum. This polymorphism is strongly associated with coronary heart disease risk [39]. However, the 56G allele is only recognized as a risk factor for large-vessel-associated stroke, according to Maasz et al. (2008) [35]. Non-carriers of the minor allele benefit from lower very low-density lipoprotein cholesterol (VLDL-cholesterol) concentrations ($0.6 \pm 0.22 \text{ mmol/L}$) than the carriers of the minor allele, who present higher concentrations ($0.7 \pm 0.32 \text{ mmol/L}$) (p = 0.01) [39].

Carriers of the ϵ 4 allele of the *ApoE*4 have a higher risk of suffering coronary heart disease. On the contrary, *ApoE*2 polymorphisms are not significantly related to coronary risk [40].

There seems to be an inverse relationship between LIPC and HDL-cholesterol [41], even though its influence has not been clearly described. Since the metabolism of glycerophospholipids is influenced by the hepatic lipase gene, it is believed that it may also alter HDL-cholesterol plasma concentrations [42]. As suggested by Posadas-Sanchez et al. [43], the TT genotype of *LIPC C-154T* polymorphism is related

to higher levels of triglycerides/HDL-cholesterol index (p = 0.046) and triglycerides (p = 0.0002), under a dominant model. Since the metabolism of glycerophospholipids is influenced by the hepatic lipase gene, it is believed that it may also alter HDL-cholesterol plasma concentrations [42]. As suggested by Posadas-Sanchez et al. [43], the TT genotype of *LIPC C-154T* polymorphism is linked with increased levels of triglycerides/HDL-cholesterol index (p = 0.046) and triglycerides (p = 0.0002), under a dominant model. Wang et al. [44], on the other hand, associated the same genotype to the presence of LDL-cholesterol (p = 0.003). The *LIPCC-514T* polymorphism was involved with hypertriglyceridemia (OR = 1.36, p = 0.006) and coronary artery calcification (OR = 1.44, p = 0.015), under a dominant model. In addition, obese boys and non-obese girls carrying the T allele presented higher levels of LDL-cholesterol and TC and LDL-cholesterol, respectively (all p < 0.05).

Finally, carriers of the CT allele of the *C677T* polymorphism present a higher risk of congenital heart disease compared to the wild CC genotype (OR = 2.249, 95% CI 1.305–3.877, p = 0.003) [41]. Besides, the risk of congenital heart disease is significantly associated with the homozygous mutant genotype TT (OR = 3.121, 95% CI 1.612–6.043, p = 0.001) [39].

Studies with children show controversial results with *LIPC C-514T*. Riestra et al. (2009) [42] described a relationship between *LIPC* and HDL-cholesterol levels modulated by fat intake in children. Agirbasli et al. (2013) [45] on the contrary, could not find an association between *LIPC* and the lipidic profile in a Turkish cohort.

In Chinese children, a link between the TT haplotype of *LIPC C-514T* was related with an increase in triglycerides, TC, LDL-cholesterol and HDL-cholesterol levels. It should be noticed that this effect depended on the presence of obesity and male gender [44].

SNP within the *CYP7A1* and *ApoE* genes, as well as possibly genes including *ABC G5* and *G8*, have been found to be predictors of the LDL-cholesterol response to plant sterol/stanol treatment. However, this study found that other gene variants than those previously described seem to affect the response of plant sterols on serum lipids. It is worth noting that nutraceuticals can go beyond the genetic influence, they may influence lipid levels beyond the skills in managing sterols [46].

Limitations

This was a pilot-type study with a rather small number of children participating and a short duration of just eight weeks intervention. Other genotypes reported before to have an impact where not studied.

5. Conclusions

The results of this study show that only *LIPC C-514T* and *PPAR-alpha L162V* show a statistically significant effect on the lipidic profile. This could be due to the small size of our sample and the uneven distribution of the genetic haplotypes reviewed in this clinical trial.

Future research with bigger samples is needed to understand the interplay between genomics and cardiovascular health.

The reader should be aware that results cannot be extrapolated to the Spanish population, due to the small sample size. A smaller, humble pilot study was carried out since a bigger sample size and more sophisticated research would incur higher costs.

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References

- 1. WHO. Cardiovascular Diseases (CVDs); WHO: Geneva, Switzerland, 2017.
- 2. Luirink, I.K.; Hutten, B.A.; Wiegman, A. Optimizing Treatment of Familial Hypercholesterolemia in Children and Adolescents. *Curr. Cardiol. Rep.* **2015**, *17*, 78. [CrossRef] [PubMed]
- 3. McMahan, C.A.; Gidding, S.S.; Malcom, G.T.; Tracy, R.E.; Strong, J.P.; McGill, H.C. Pathobiological Determinants of Atherosclerosis in Youth Risk Scores Are Associated with Early and Advanced Atherosclerosis. *Pediatrics* **2006**, *118*, 1447–1455. [CrossRef] [PubMed]
- 4. Araujo, M.B.; Botto, P.M.; Mazza, C.S. Uso de ezetimibe en el tratamiento de la hipercolesterolemia. *An. Pediatr.* **2012**, 77, 37–42. [CrossRef] [PubMed]
- 5. Blumenfeld, J.A. *Plant Sterol-Enriched Milk in Paediatric Children with Hypercholesterolemia*; Double Blinded, randomized controlled clinical trial; Universidad Complutense de Madrid: Madrid, Spain, 2017.
- 6. Abdullah, M.M.H.; Jones, P.J.H.; Eck, P.K. Nutrigenetics of cholesterol metabolism: Observational and dietary intervention studies in the postgenomic era. *Nutr. Rev.* **2015**, *73*, 523–543. [CrossRef]
- Gylling, H.; Plat, J.; Turley, S.; Ginsberg, H.N.; Ellegard, L.; Jessuo, W.; Jones, P.J.; Lutjohann, D.; Maerz, W.; Masana, L.; et al. Plant sterols and plant stanols in the management of dyslipidaemia and prevention of cardiovascular disease. *Atherosclerosis* 2014, 232, 346–360. [CrossRef]
- 8. EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA). Scientific Opinion on the substantiation of health claims related to plant sterols and plant stanols and maintenance of normal blood cholesterol concentrations, and maintenance of normal prostate size and normal urination pursuant to Article 13(1) of Regu. *EFSA J.* **2010**, *8*, 1–22.
- 9. Karlezi, R.A.A.; Pariente, N.M.; López, P.M. Control de las hiperlipemias en la práctica clínica. *Rev. Esp. Cardiol. Supl.* **2006**, *6*, 24G–35G.
- Demonty, I.; Ras, R.T.; van der Knaap, H.C.; Duchateau, G.S.; Meijer, L.; Zock, P.L.; Geleijnse, J.M.; Trautwein, E.A. Continuous dose-response relationship of the LDL-cholesterol-lowering effect of phytosterol intake. J. Nutr. 2009, 139, 271–284. [CrossRef]
- Ras, R.T.; Geleijnse, J.M.; Trautwein, E.A. LDL-cholesterol-lowering effect of plant sterols and stanols across different dose ranges: A meta-analysis of randomised controlled studies. *Br. J. Nutr.* 2014, 112, 214–219. [CrossRef]
- Chen, K.H.; Chen, L.L.; Li, W.G.; Fang, Y.; Huang, G.Y. Maternal MTHFR C677T polymorphism and congenital heart defect risk in the Chinese Han population: A meta-analysis. *Genet. Mol. Res.* 2013, 12, 6212–6219. [CrossRef]
- Zhang, M.J.; Li, J.C.; Yin, Y.W.; Li, B.H.; Liu, Y.; Liao, S.Q.; Gao, C.Y.; Zhang, L.L. Association of MTHFR C677T polymorphism and risk of cerebrovascular disease in Chinese population: An updated meta-analysis. *J. Neurol.* 2014, 261, 925–935. [CrossRef] [PubMed]
- 14. Kumar, A.; Kumar, P.; Prasad, M.; Sagar, R.; Yadav, A.K.; Pandit, A.K.; Jali, V.P.; Pathak, A. Association of C677T polymorphism in the methylenetetrahydrofolate reductase gene (MTHFR gene) with ischemic stroke: A meta-analysis. *Neurol. Res.* **2015**, *37*, 568–577. [CrossRef] [PubMed]
- 15. Edmondson, A.C.; Braund, P.S.; Stylianou, I.M.; Khera, A.V.; Nelson, C.P.; Wolfe, M.L.; DerOhannessian, S.L.; Keating, B.J.; Qu, L.; He, J.; et al. Dense genotyping of candidate gene loci identifies variants associated with high-density lipoprotein cholesterol. *Circ. Cardiovasc. Genet.* **2011**, *4*, 145–155. [CrossRef] [PubMed]
- Nordestgaard, B.G.; Chapman, M.J.; Ray, K.; Boré, J.; Andreotti, F.; Watts, G.F.; Ginsberg, H.; Amarenco, P. Lipoprotein(a) as a cardiovascular risk factor: Current status. *Eur. Heart J.* 2010, *31*, 2844–2853. [CrossRef] [PubMed]
- 17. San Mauro-Martin, I.; Sanz Rojo, S.; Garicano-Vilar, E.; Collado-Yurrita, L. Enfoque genómico en la enfermedad cardiovascular. *Nutr. Hosp.* **2016**, *33*, 148–155.
- 18. Plat, J.; Bragt, M.C.; Mensink, R.P. Common sequence variations in ABCG8 are related to plant sterol metabolism in healthy volunteers. *J. Lipid Res.* **2005**, *46*, 68–75. [CrossRef]

- Rudkowska, I.; AbuMweis, S.S.; Nicolle, C.; Jones, P.J. Association between non-responsiveness to plant sterol intervention and polymorphisms in cholesterol metabolism genes: A case-control study. *Appl. Physiol. Nutr. Metab.* 2008, 33, 728–734. [CrossRef]
- Zhao, H.L.; Houweling, A.H.; Vanstone, C.A.; Jew, S.; Trautwein, E.A.; Duchateau, G.S.; Jones, P.J. Genetic variation in ABC G5/G8 and NPC1L1 impact cholesterol response to plant sterols in hypercholesterolemic men. *Lipids* 2008, 43, 1155–1164. [CrossRef]
- Gylling, H.; Hallikainen, M.; Raitakari, O.T.; Laakso, M.; Vartiainen, E.; Salo, P.; Korpelainen, V.; Sundvall, J.; Miettinen, T.A. Long-term consumption of plant stanol and sterol esters, vascular function and genetic regulation. *Br. J. Nutr.* 2009, *101*, 1688–1695. [CrossRef]
- 22. MacKay, D.S.; Eck, P.K.; Gebauer, S.K.; Baer, D.J.; Jones, P.J. CYP7A1-rs3808607 and APOE isoform associate with LDL cholesterol lowering after plant sterol consumption in a randomized clinical trial. *Am. J. Clin. Nutr.* **2015**, *102*, 951–957. [CrossRef]
- 23. Rideout, C. Getting personal: Considering variable interindividual responsiveness to dietary lipid-lowering therapies. *Curr. Opin. Lipidol.* **2011**, *22*, 37–42. [CrossRef] [PubMed]
- Moráis López, A.; Lama More, R.A.; Dalmau Serra, J. Hipercolesterolemia. Abordaje terapéutico. *An. Pediatr.* 2009, 70, 488–496. [CrossRef] [PubMed]
- 25. Lippi, G.; Avanzini, P.; Musa, R.; Sandei, F.; Aloe, R.; Cervellin, G. Evaluation of sample hemolysis in blood collected by S-monovette[®] using vacuum or aspiration mode. *Biochem. Med.* **2013**, *23*, 64–69. [CrossRef]
- 26. Bowen, R.A.R.; Remaley, A.T. Interferences from blood collection tube components on clinical chemistry assays. *Biochem. Med.* 2014, 24, 31–44. [CrossRef]
- Sarstedt Blood Collection with the S-Monovette[®]. Available online: https://www.sarstedt.com/fileadmin/ user_upload/99_Gebrauchsanweisungen/Englisch_US_Code/644_c_PosterA3_AnleitungVenoeseBE_ SafetyKanuele_GB_US_0314.pdf (accessed on 8 March 2018).
- 28. Weale, M.E. Quality control for genome-wide association studies. Methods Mol. Biol. 2010, 628, 341–372.
- 29. Ziegler, A. Genome-wide association studies: Quality control and population-based measures. *Genet. Epidemiol.* **2009**, *33*, S45–S50. [CrossRef]
- 30. Ostlund, R.E. Phytosterols and cholesterol metabolism. Curr. Opin. Lipidol. 2004, 15, 37-41. [CrossRef]
- Gylling, H.; Simonen, P. Phytosterols, phytostanols, and lipoprotein metabolism. *Nutrients* 2015, 7, 7965–7977. [CrossRef]
- Clifton, P.M.; Noakes, M.; Sullivan, D.; Erichsen, N.; Ross, D.; Annison, G.; Fassoulakis, A.; Cehun, M.; Nestel, P. Cholesterol-lowering effects of plant sterol esters differ in milk, yoghurt, bread and cereal. *Eur. J. Clin. Nutr.* 2004, *58*, 503–509. [CrossRef]
- Katan, M.B.; Grundy, S.M.; Jones, P.; Law, M.; Miettinen, T.; Paoletti, R. Efficacy and Safety of Plant Stanols and Sterols in the Management of Blood Cholesterol Levels. *Mayo Clin. Proc.* 2003, 78, 965–978. [CrossRef]
- 34. Abumweis, S.S.; Barake, R.; Jones, P.J.H. Plant sterols/stanols as cholesterol lowering agents: A meta-analysis of randomized controlled trials. *Food Nutr. Res.* **2008**, 52. [CrossRef] [PubMed]
- Maász, A.; Kisfali, P.; Szolnoki, Z.; Hadarits, F.; Melegh, B. Apolipoprotein A5 gene C56G variant confers risk for the development of large-vessel associated ischemic stroke. *J. Neurol.* 2008, 255, 649–654. [CrossRef] [PubMed]
- San Mauro Martín, I.; Sanz Rojo, S.; Garicano Vilar, E.; Collado Yurrita, L.; Blumenfeld Olivares, J.A. Modulation of plasma triglycerides concentration by sterol-based treatment in children carrying different genes. *Ann. Pediatr. Cardiol.* 2019, 12, 83–89. [CrossRef] [PubMed]
- 37. Zhou, J.; Xu, L.; Huang, R.S.; Huang, Y.I.; Le, Y.; Jiang, D.; Yang, X.I.; Xu, W.; Huang, X.; Dong, C.; et al. Apolipoprotein A5 gene variants and the risk of coronary heart disease: A case-control study and meta-analysis. *Mol. Med. Rep.* **2013**, *8*, 1175–1182. [CrossRef] [PubMed]
- Pi, Y.; Zhang, L.; Yang, Q.; Li, B.; Guo, L.; Fang, C.; Gao, C.; Wang, J.; Xiang, J.; Li, J. Apolipoprotein A5 gene promoter region-1131T/C polymorphism is associated with risk of ischemic stroke and elevated triglyceride levels: A meta-analysis. *Cerebrovasc. Dis.* 2012, *33*, 558–565. [CrossRef]
- Sánchez-Moreno, C.; Ordovás, J.M.; Smith, C.E.; Baraza, J.C.; Lee, Y.C.; Garaulet, M. {APOA}5 Gene Variation Interacts with Dietary Fat Intake to Modulate Obesity and Circulating Triglycerides in a Mediterranean Population. *J. Nutr.* 2011, 141, 380–385. [CrossRef] [PubMed]

- 40. Zhang, M.; Gu, W.; Qiao, S.; Zhu, E.; Zhao, Q.; Lv, S. Apolipoprotein E Gene Polymorphism and Risk for Coronary Heart Disease in the Chinese Population: A Meta-Analysis of 61 Studies Including 6634 Cases and 6393 Controls. *PLoS ONE* **2014**, *9*, e95463. [CrossRef]
- 41. Demirkan, A.; van Duijn, C.M.; Ugocsai, P.; Isaacs, A.; Pramstaller, P.P.; Liebisch, G.; Wilson, J.F.; Johansson, A.; Rudan, I.; Aulchenko, Y.S.; et al. Genome-wide association study identifies novel loci associated with circulating phospho- and sphingolipid concentrations. *PLoS Genet.* **2012**, *8*, e1002490. [CrossRef]
- 42. Riestra, P.; López-Simón, L.; Ortega, H.; Gorgojo, L.; Martin-Moreno, J.M.; Schoppen, S.; de Oya, M.; Garcés, C. Fat Intake Influences the Effect of the Hepatic Lipase C-514T Polymorphism on HDL-Cholesterol Levels in Children. *Exp. Biol. Med.* **2009**, 234, 744–749. [CrossRef]
- Posadas-Sánchez, R.; Ocampo-Arcos, W.A.; López-Uribe, Á.R.; Posadas-Romero, C.; Villarreal-Molina, T.; León, E.Á.; Pérez-Hernández, N.; Rodríguez-Pérez, J.M.; Cardoso-Saldaña, G.; Medina-Urrutia, A.; et al. Hepatic lipase (LIPC) C-514T gene polymorphism is associated with cardiometabolic parameters and cardiovascular risk factors but not with fatty liver in Mexican population. *Exp. Mol. Pathol.* 2015, *98*, 93–98. [CrossRef]
- 44. Wang, H.; Zhang, D.; Ling, J.; Lu, W.; Zhang, S.; Zhu, Y.; Lai, M. Gender specific effect of LIPC C-514T polymorphism on obesity and relationship with plasma lipid levels in Chinese children. *J. Cell. Mol. Med.* **2015**, *19*, 2296–2306. [CrossRef] [PubMed]
- 45. Agirbasli, M.; Eren, F.; Agirbasli, D.; White, M.J.; Williams, S.M. Multi-locus candidate gene analyses of lipid levels in a pediatric Turkish cohort: Lessons learned on LPL, CETP, LIPC, ABCA1, and SHBG. *OMICS* **2013**, 17, 636–645. [CrossRef] [PubMed]
- Scicchitano, P.; Cameli, M.; Maiello, M.; Modesti, P.A.; Muiesan, M.L.; Novo, S.; Palmiero, P.; Saba, P.S.; Pedrinelli, R.; Ciccone, M.M.; et al. Nutraceuticals and dyslipidaemia: Beyond the common therapeutics. *J. Func. Foods* 2014, *6*, 11–32. [CrossRef]



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