

Article

# Supplementation with *Saccharomyces boulardii* Increases the Maximal Oxygen Consumption and Maximal Aerobic Speed Attained by Rats Subjected to an Incremental-Speed Exercise

Anne Danieli Nascimento Soares <sup>1,2</sup>, Samuel Penna Wanner <sup>3,\*</sup> , Elissa Stefane Silva Morais <sup>1</sup>, Alexandre Sérvulo Ribeiro Hudson <sup>3</sup> , Flaviano Santos Martins <sup>4</sup> and Valbert Nascimento Cardoso <sup>1</sup>

<sup>1</sup> Department of Clinical and Toxicological Analysis, Faculty of Pharmacy, Universidade Federal de Minas Gerais, Belo Horizonte, MG, 31270-901, Brazil; annedanieli@gmail.com (A.D.N.S.); elissamorais\_hotmail.com (E.S.S.M.); valbertcardoso@yahoo.com.br (V.N.C.)

<sup>2</sup> Instituto Federal de Educação, Ciência e Tecnologia do Sudeste de Minas Gerais, Barbacena, MG, 36205-018, Brazil

<sup>3</sup> Exercise Physiology Laboratory, School of Physical Education, Physiotherapy and Occupational Therapy, Universidade Federal de Minas Gerais, Belo Horizonte, MG, 31270-901, Brazil; alexandre.servulo@yahoo.com.br

<sup>4</sup> Department of Microbiology, Institute of Biological Sciences, Universidade Federal de Minas Gerais, Belo Horizonte, MG, 31270-901, Brazil; flaviano@icb.ufmg.br

\* Correspondence: samuelwanner@eefito.ufmg.br; Tel.: +55-31-3409-2328

Received: 8 August 2019; Accepted: 12 September 2019; Published: 2 October 2019



**Abstract:** Benefits to the host metabolism resulting from *Saccharomyces boulardii* (Sb) supplementation have been described; however, no study has investigated the effects of this supplementation on aerobic metabolism and performance during physical exercise. Thus, in the present study, we addressed the effects of Sb supplementation on the rate of oxygen consumption ( $\text{VO}_2$ ), mechanical efficiency (external work divided by  $\text{VO}_2$ ), and aerobic performance of rats subjected to fatiguing, incremental-speed exercise. Twenty-six male Wistar rats were randomly divided into two groups: (1) non-supplemented, in which rats received 0.1 mL of a saline solution, and (2) Sb-supplemented, in which rats received 0.1 mL of a suspension containing  $8.0 \log_{10}$  colony-forming units. The rats received the treatments by gavage for 10 consecutive days; they were then subjected to fatiguing treadmill running. Sb supplementation did not change the  $\text{VO}_2$  values or mechanical efficiency during submaximal exercise intensities. In contrast, at fatigue,  $\text{VO}_{2\text{MAX}}$  was increased by 12.7% in supplemented rats compared with controls ( $p = 0.01$ ). Moreover, Sb improved aerobic performance, as evidenced by a 12.4% increase in maximal running speed attained by the supplemented rats ( $p < 0.05$ ). We conclude that Sb supplementation for 10 days increases  $\text{VO}_{2\text{MAX}}$  and aerobic performance in rats.

**Keywords:** aerobic capacity; metabolic rate; microbiota; physical performance; probiotics; yeast

## 1. Introduction

Probiotics are defined as live microorganisms that, when administered in adequate amounts, confer a health benefit to the host [1,2]. In this context, *Saccharomyces boulardii* (Sb) is a nonpathogenic yeast marketed worldwide due to its probiotic properties [3]. Sb regulates intestinal microbial homeostasis and, therefore, modulates both local and systemic immune responses [3].

Benefits to the host metabolism resulting from Sb supplementation have also been described. For instance, Sb supplementation was shown to attenuate the increase in serum concentrations of

triglycerides and total cholesterol induced by a cholesterol-enriched diet in hamsters [4] and to reduce the body weight, fat mass, hepatic steatosis, and inflammatory tone in obese/type 2 diabetic mice [5]. Moreover, Sb supplementation increases the fecal concentrations of short-chain fatty acids (SCFAs) in patients undergoing enteral nutrition [6], and these gut-derived SCFAs play important roles as substrates for glucose, cholesterol, and lipid metabolism [7]. However, whether these benefits promoted by Sb also result from changes in the aerobic metabolic rate remains to be investigated.

Physical exercise is a condition that places a burden on body metabolism due to the augmented energy demand of contracting muscles. The ability to perform prolonged exercise (i.e., endurance or aerobic capacity) is paramount in many sports and occupations. Recent literature has provided evidence that supplementation with a multi-strain probiotic for four weeks can improve human aerobic performance in the heat, as indicated by an increased running time to fatigue [8]. Of note, these findings did not confirm the findings of a previous study by Cox et al. [9], where no differences were reported in the treadmill performance of trained distance runners following supplementation; these two studies treated the individuals with different bacterial genera. As far as we know, no study has investigated the effects of supplementation with a yeast that acts as a probiotic (i.e., Sb) on aerobic performance.

The mechanisms underlying improved aerobic performance involve adaptations in multiple physiological systems, including skeletal muscles. For example, muscle adaptations are characterized by greater mitochondrial volume and increased expression of mitochondrial enzymes involved in  $\beta$ -oxidation, the tricarboxylic acid cycle, and the electron transport system [10]. The ability to increase the rate of chemical to mechanical energy conversion via aerobic metabolism, to spend less energy when exercising (i.e., improved running economy), and to depend less on the glycolytic pathway for energy conversion (i.e., a rightward shift in the lactate threshold) are some of the factors that are dependent on muscle functioning and determine the capacity to exercise for prolonged periods [11]. Previous studies showed that aerobically trained rats present a lower energy cost of running [12,13] and greater maximal rate of oxygen consumption ( $VO_{2MAX}$ ) [13]. These adaptations may underlie the performance-enhancing effects of probiotics, which modulate body metabolism as described earlier.

Thus, the present study investigated the effects of Sb supplementation on the rate of oxygen consumption ( $VO_2$ ) of rats kept under resting conditions or subjected to fatiguing, incremental-speed treadmill running. We also evaluated the running economy, mechanical efficiency, and maximal speed attained by rats subjected to exercise. We expected greater aerobic performance in rats treated with Sb, which would be associated with greater  $VO_{2MAX}$ , running economy, and mechanical efficiency.

## 2. Materials and Methods

### 2.1. Microorganisms

Viable *Saccharomyces boulardii* cells were used after isolation on Sabouraud dextrose agar (Difco) from a lyophilized commercial preparation (Floratil, Merck S.A., Kenilworth, NJ, USA). The yeast was grown overnight at 37 °C with constant shaking (180 rpm) in YPD (yeast extract 1%, peptone 1%, and dextrose 2%) broth. The culture was then concentrated to obtain  $10^9$  ( $9.0 \log_{10}$ ) colony-forming units (CFU)·mL<sup>-1</sup>; for treatment, rats received 0.1 mL of this suspension by gavage, which means that each animal was treated with  $10^8$  CFU (or approximately  $3 \times 10^8$  CFU·kg<sup>-1</sup>·day<sup>-1</sup>).

### 2.2. Animals

Twenty-six male Wistar rats (11 weeks old) weighing 250 to 350 g were provided by the Animal Care Center at the Faculty of Pharmacy of the Universidade Federal de Minas Gerais. The rats were housed in individual cages under controlled light (05:00–19:00 h) and temperature ( $24.0 \pm 2.0$  °C) conditions with water and commercial chow (Nuvilab CR-1) provided ad libitum. The experiments were approved by the local Ethics Commission on Animal Use (protocol number: 34/2014) and complied with the policies determined by the National Council for the Control of Animal Experimentation (CONCEA/Brazil).

### 2.3. Experimental Design

The rats were randomly allocated into two groups, with 13 rats in each group: (1) non-supplemented (NS), in which rats received 0.1 mL of a saline solution, and (2) supplemented (Sb), in which rats received a suspension containing Sb. The rats received the treatments by gavage and had food intake and body mass recorded once per day for 10 consecutive days. Since our objective was to test the effectiveness of a preventive intervention aimed at protecting an organism during subsequent exposure to stressful stimuli, a 10-day supplementation period was chosen. In this sense, we demonstrated that mice supplemented with Sb for 10 days had decreased disease and death caused by an experimental model of typhoid fever [14].

During 5 consecutive days of the supplementation period (from the 4th until the 9th day), all of the rats were familiarized with running on a treadmill. On the 10th day of the supplementation, each rat was subjected to an incremental-speed exercise. All experimental trials were performed during the light phase of the day. Following the trial, each rat was euthanized with a lethal dose of anesthetic (a cocktail containing 240 mg·kg<sup>-1</sup> of ketamine and 31.5 mg·kg<sup>-1</sup> of xylazine) administered via intraperitoneal injection.

### 2.4. Familiarization with the Running Exercise on a Treadmill

The familiarization protocol consisted of running on a treadmill designed for small rodents (Panlab, Harvard Apparatus, Cornellà, Spain) over 5 consecutive days. Rats were encouraged to run by light electrical stimulation (0.2 mA) provided by a grid located at the rear end of the treadmill belt. Each familiarization session initially consisted of 5 min during which the rat could move freely on the treadmill belt; the treadmill was then turned on, and the speed was gradually increased up to 15 m·min<sup>-1</sup>, at which point the speed was held constant for 5 min [15]. At the last familiarization session, the rats could run with minimal exposure to electrical stimulation. The incline was always set at 5% during all familiarization sessions and experimental trials.

### 2.5. Experimental Trials

On the day of the experimental trials, the rats were weighed, administered the assigned treatment by gavage, and then transferred from their home cages to the motor-driven treadmill. The chamber that contained the treadmill belt was sealed. VO<sub>2</sub> was measured by open-circuit indirect calorimetry. For this measurement, the motor-driven treadmill chamber was coupled to a gas analyzer (Panlab, Harvard Apparatus), and the air flow inside the chamber was maintained at 2.0 L·min<sup>-1</sup>. The gas sensors were calibrated with primary gas standards containing known concentrations of O<sub>2</sub> and CO<sub>2</sub>. The air leaving the chamber was automatically sampled and passed through the gas analyzer to determine the O<sub>2</sub> and CO<sub>2</sub> content of the air. The VO<sub>2</sub> was measured every second, and 1-min average values were calculated with the help of Metabolism 2.2v software.

The rats were allowed to rest for 90 min and were then subjected to an incremental-speed exercise. The exercise started at a speed of 10 m·min<sup>-1</sup> with increments of 1 m·min<sup>-1</sup> every 3 min until the animals were fatigued [16]. Fatigue was determined as the moment when the rat could not maintain the pace with the treadmill, subjecting itself to the light electrical stimuli for 10 s [15]. VO<sub>2</sub> was measured continuously during resting and exercise sessions. The ambient temperature was controlled at 24 °C. It is worth mentioning that physical performance measured during this exercise (i.e., workload) has been shown to be positively correlated with the VO<sub>2MAX</sub> of untrained rats [12].

Three different indices were used to determine aerobic performance: Time to fatigue, the maximal speed attained, and the external work performed by the rats. The maximal aerobic speed (S<sub>MAX</sub>) attained during the incremental exercises was calculated according to the equation described by Kunstetter et al. [16]:  $S_{MAX} = S1 + (S2 \times t/180 \text{ seconds})$ , where S1 is the speed reached in the last completed stage, S2 is the increment in the treadmill speed at each stage, and t is the time spent (in seconds) in the incomplete stage. The external work was calculated in Joules as  $bm \times g \times s \times \sin\theta \times t$ ,

where  $bm$  is the animal's body mass (kg),  $g$  is the acceleration of gravity ( $9.8 \text{ m}\cdot\text{s}^{-2}$ ),  $s$  is the treadmill speed ( $\text{m}\cdot\text{min}^{-1}$ ),  $\theta$  is the angle of treadmill inclination, and  $t$  is the time spent in each stage [12,17]. Values for workload were calculated for each stage of incremental exercise, including the incomplete stage, and were then summed; the value obtained after the summation corresponded to the total external work.

The gross oxygen cost of running, which is inversely associated with the running economy, was calculated by dividing the  $\text{VO}_2$  ( $\text{mL}\cdot\text{kg}^{-1}\cdot\text{min}^{-1}$ ) by the running speed ( $\text{m}\cdot\text{min}^{-1}$ ), and the resulting value was expressed in  $\text{mL}\cdot\text{kg}^{-1}\cdot\text{m}^{-1}$  [12,18]. Although the oxygen cost of running is commonly determined during constant-speed exercises, we calculated this parameter in the last minute of the 3-min stages during the incremental exercise, since Wisloff et al. [13] reported that  $\text{VO}_2$  leveled off in running rats ~3 min after the external work was changed. Gross mechanical efficiency was calculated by dividing the caloric equivalent of external work by the caloric equivalent of  $\text{VO}_2$  [19]; the value obtained was then multiplied by 100. According to Brooks et al. [19], the gross efficiency of rats running on a treadmill ranges from 1.3% to 4.5%.

## 2.6. Statistical Analysis

The variables studied were tested for normality using the Shapiro–Wilk test. All variables were normally distributed; therefore, the data are expressed as the means  $\pm$  SEM. The data presented in Table 1 (i.e., nutritional parameters, physical performance indices,  $\text{VO}_{2\text{MAX}}$ , oxygen cost of running, and mechanical efficiency) were compared between the two groups using unpaired Student's  $t$ -tests. The  $\text{VO}_2$  curves were compared between the two experimental groups and across time points using two-way analysis of variance (ANOVA) with repeated measures for only the time factor. Tukey's test was used as the post hoc analysis. The significance level was set at  $\alpha < 0.05$ . The effect sizes (Cohen's  $d$  for two independent means) were calculated for the data presented in Table 1. Effect size allowed the assessment of the magnitude of differences between experimental trials and was calculated by subtracting the mean value of one group from the mean value of the group it was being compared to. The result was then divided by a combined standard deviation for the data. The effect size values were classified as trivial ( $ES < 0.2$ ), small ( $ES = 0.2\text{--}0.6$ ), moderate ( $ES = 0.6\text{--}1.2$ ), or large ( $ES \geq 1.2$ ) [20].

**Table 1.** The nutritional parameters during the 10 days before the experimental trials, the physical performance indices, running economy, and the mechanical efficiency values of rats from the non-supplemented (NS) and the *Saccharomyces boulardii*-supplemented (Sb) groups during the incremental exercise.

| Parameter  | NS               | Sb               | $p$ -Value | Cohen's $d$ |
|--|------------------|------------------|------------|-------------|
| Nutritional parameters   |                  |                  |            |             |
| Body mass gain (g)   | 33.6 $\pm$ 4.8   | 34.9 $\pm$ 3.7   | 0.827      | 0.084       |
| Chow intake ( $\text{g}\cdot\text{day}^{-1}$ )   | 28.3 $\pm$ 1.9   | 26.4 $\pm$ 0.5   | 0.357      | 0.381       |
| Physical performance indices   |                  |                  |            |             |
| Time to fatigue (min)  | 36.7 $\pm$ 1.8   | 44.6 $\pm$ 3.4   | 0.047      | 0.816       |
| Maximum speed attained ( $\text{m}\cdot\text{min}^{-1}$ )                                  | 21.2 $\pm$ 0.6   | 23.9 $\pm$ 1.1   | 0.047      | 0.845       |
| Total external work (J)  | 162.9 $\pm$ 10.6 | 217.6 $\pm$ 22.1 | 0.036      | 0.874       |
| $\text{VO}_{2\text{MAX}}$ ( $\text{mL}\text{O}_2\cdot\text{kg}^{-1}\cdot\text{min}^{-1}$ ) | 61.09 $\pm$ 1.83 | 68.82 $\pm$ 2.08 | 0.010      | 1.095       |
| Gross oxygen cost of running   |                  |                  |            |             |
| First stage ( $\text{mL}\text{O}_2\cdot\text{kg}^{-1}\cdot\text{m}^{-1}$ )                 | 3.48 $\pm$ 0.15  | 3.64 $\pm$ 0.10  | 0.392      | 0.349       |
| Last stage ( $\text{mL}\text{O}_2\cdot\text{kg}^{-1}\cdot\text{m}^{-1}$ )                  | 2.91 $\pm$ 0.05  | 2.91 $\pm$ 0.08  | 0.965      | 0.000       |
| Gross mechanical efficiency  |                  |                  |            |             |
| First stage (%)  | 1.24 $\pm$ 0.07  | 1.16 $\pm$ 0.03  | 0.315      | 0.414       |
| Last stage (%)   | 1.40 $\pm$ 0.03  | 1.40 $\pm$ 0.04  | 0.908      | 0.000       |

Data are expressed as the means  $\pm$  SEM.

### 3. Results

#### 3.1. Body Mass Gain and Chow Intake

The initial body mass of the rats was similar between groups (NS:  $296.7 \pm 4.2$  g vs. Sb:  $297.6 \pm 5.4$  g;  $p = 0.887$ ). The control, non-supplemented rats had an average chow intake of 28.3 g per day and gained an average of 3.4 g per day. The group supplemented with Sb did not show a change in chow intake or body mass gain during the 10 days of treatment relative to the group treated with saline (Table 1); indeed, the intergroup differences were classified as trivial or small. These data indicate that the animals tolerated the supplementation well and that the experimental findings can be attributed exclusively to the actions of Sb.

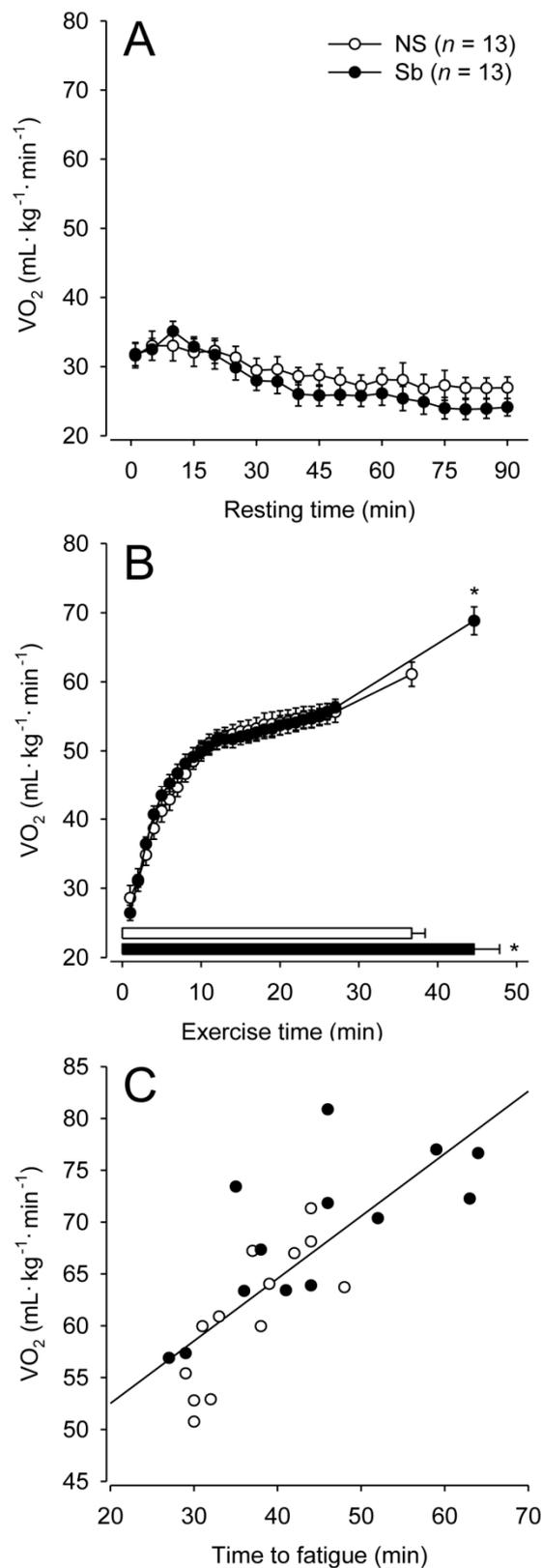
#### 3.2. Resting Experiments

A significant main effect of time was observed for  $VO_2$  data in resting rats ( $F = 9.859$ ;  $p < 0.001$ ). When the rats were placed on the treadmill, they displayed an average  $VO_2$  that was above  $30 \text{ mL}O_2 \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$  irrespective of their experimental group. Then, the  $VO_2$  gradually decreased until values of  $26.95 \pm 1.55$  and  $24.13 \pm 1.26 \text{ mL}O_2 \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$  for the NS and Sb groups, respectively, were reached after the 90-min rest period (Figure 1A). There was no difference in  $VO_2$  between the two groups throughout the resting period ( $F = 0.926$ ;  $p = 0.346$ ) and no interaction between time and groups ( $F = 0.754$ ;  $p = 0.949$ ).

#### 3.3. Incremental-Speed Exercise Sessions

The duration of fatiguing and incremental exercise differed between the two groups; the Sb group ran approximately 8 min longer than the NS group (Table 1). Supplementation with Sb also increased the  $S_{MAX}$  attained and the external work performed by the rats by 12.4% and 33.6%, respectively. The improvements caused by Sb in the three indices of aerobic performance were classified as moderate.

A significant interaction between time and group was observed for  $VO_2$  data in rats subjected to exercise ( $F = 3.725$ ;  $p < 0.001$ ). Treadmill running induced an immediate and marked increase in  $VO_2$  in both groups. The  $VO_2$  value was not different between the two groups up to the 27th minute of exercise when all rats from both groups were still running. However, the  $VO_2$  value attained at fatigue, which corresponded to the  $VO_{2MAX}$ , was 12.7% higher in the Sb group than in the NS group ( $68.82 \pm 2.08 \text{ mL}O_2 \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$  vs.  $61.09 \pm 1.83 \text{ mL}O_2 \cdot \text{kg}^{-1} \cdot \text{min}^{-1}$ ;  $p = 0.010$ ; Figure 1B); this difference in  $VO_{2MAX}$  was also classified as moderate. In agreement with the latter findings, significant and positive correlations were observed between  $VO_{2MAX}$  and the three aerobic performance indices evaluated: Time to fatigue ( $r = 0.778$ ;  $p < 0.001$ ; Figure 1C),  $S_{MAX}$  ( $r = 0.778$ ;  $p < 0.001$ ), and total external work ( $r = 0.752$ ;  $p < 0.001$ ).



**Figure 1.** The rate of oxygen consumption in rats during the 90-min rest (Figure 1A) or in rats subjected to an incremental-speed exercise (Figure 1B). NS, non-supplemented group (○); Sb, *Saccharomyces boulardii*-supplemented group (●). Data are expressed as the means ± SEM. The horizontal bars at the bottom of panel B represent the time to fatigue. \* denotes a significant difference compared with the NS group. Figure 1C shows the significant and positive correlation between the maximal rate of oxygen consumption (VO<sub>2</sub>MAX) and time to fatigue. Data are expressed as individual values.

We then calculated the oxygen cost of running (which represents the running economy) and mechanical efficiency in an attempt to better understand our data. Regardless of the timing of the calculation (in the first stage or last completed stage of the exercise), no significant intergroup differences were observed (Table 1; trivial and small-magnitude differences). In addition, we investigated whether these parameters were associated with  $VO_{2MAX}$ . No significant correlations were observed between  $VO_{2MAX}$  and running economy ( $r = 0.099$ ;  $p = 0.631$ ) or mechanical efficiency ( $r = -0.051$ ;  $p = 0.804$ ) measured in the last completed stage.

#### 4. Discussion

In the present study, supplementation with Sb did not alter the  $VO_2$  of rats kept undisturbed on the treadmill belt. In addition, during submaximal intensities of the incremental-speed running exercise, the probiotic did not change the  $VO_2$  values, running economy, or mechanical efficiency. In contrast, at fatigue,  $VO_{2MAX}$  was increased in supplemented rats compared with controls, despite the absence of changes in running economy or mechanical efficiency. Interestingly, Sb supplementation improved aerobic performance, as evidenced by the increases in time to fatigue, maximal speed attained, and external work performed by the supplemented rats during the incremental exercise.

The lack of changes in the resting  $VO_2$  of the supplemented rats suggests that the beneficial metabolic effects mediated by Sb are not due to changes in energy expenditure via oxidative pathways. It is relevant to state that the rats used in the present experiments were young, fed a conventional chow diet, and likely did not exhibit any metabolic dysfunction. This is an important difference from previous studies in which beneficial effects of Sb on metabolism were observed; these studies used hamsters subjected to a cholesterol-enriched diet [4] or obese and diabetic mice [5]. Thus, further investigation into the effects of Sb on the aerobic metabolism of unhealthy animals or human participants is warranted.

The running time to fatigue and  $VO_{2MAX}$  were, respectively, 21.6% and 12.7% higher in supplemented rats compared to non-supplemented rats. Because no other study has investigated whether Sb supplementation influences physical performance, the following discussion was based on the effects of different probiotics administered at doses similar to that used in the present study. Our findings are in agreement with those reported by Chen et al. [21]; these authors observed that the endurance swimming time in mice was increased following six weeks of oral administration of *Lactobacillus plantarum* TWK10 compared with control mice treated with a vehicle. Similarly, in humans, Shing et al. [8] reported an extended time to fatigue in trained runners during a fixed-intensity exercise performed in a hot environment following 4 weeks of supplementation with a multi-strain probiotic (*Lactobacillus*, *Bifidobacterium*, and *Streptococcus* strains). Nevertheless, an improvement in physical performance is not a universal finding after supplementation with probiotics. Cox et al. [9] did not report any differences in the treadmill performance of trained distance runners following 4 weeks of supplementation with *Lactobacillus fermentum* VRI-003 relative to the placebo treatment. The differences between our findings and those of previously published studies may be explained by methodological differences. Factors, such as probiotic type (bacteria or yeast species), supplementation time and duration, exercise type, species (human, rat, or mouse), and sample characteristics, may influence the experimental outcomes [22].

Shing et al. [8] suggested the maintenance of gastrointestinal structural integrity, endotoxin translocation, and immune modulation as possible causes for the probiotic-mediated improvement of performance in the heat. Indeed, these factors are suggested to modulate performance when animals are subjected to high thermoregulatory strain, particularly during prolonged exercise under uncompensable heat stress. However, because our experiments were conducted in a temperate environment (24 °C) and the exercise consisted of an incremental-speed running exercise, these factors likely do not explain the ergogenic effects mediated by Sb. Under the present conditions, the ability of the cardiovascular system to specifically increase coronary and skeletal muscle blood flow is more of a determinant for physical performance than thermoregulatory responses [16].

A plausible hypothesis to explain the increase in  $VO_{2MAX}$  caused by Sb is a greater ability to transform energy using lipid substrates. Sb supplementation increases the fecal concentrations of SCFAs [6], which have several modulatory actions on metabolism. For example, mice fed a high-fat diet supplemented with sodium butyrate, an SCFA, present increased levels of phosphorylated adenosine monophosphate-activated protein kinase (AMPK) in oxidative muscles [23]. The greater activation of AMPK stimulates peroxisome proliferator-activated receptor-gamma coactivator-1 $\alpha$  (PGC-1 $\alpha$ ) [23], which, in turn, promotes the gene expression of proteins involved in lipid oxidation and mitochondrial respiration. The latter responses represent a physiological mechanism that may underlie the increased aerobic performance in Sb-treated rats, although we recognize that rodents fed a standard diet will oxidize carbohydrates to generate energy at high exercise intensities [13], including the intensities close to the maximal aerobic speed.

Although Sb increased  $VO_{2MAX}$  in our rats, the running economy and mechanical efficiency remained unaltered. Notably, aerobically trained rats [12] or rats with higher intrinsic exercise capacity [24] perform better and spend less energy while running on a treadmill than their respective controls. Improvements in running economy and mechanical efficiency following aerobic training are explained by biomechanical factors, including improved technique and the transfer of elastic energy during stretch–shortening cycles [25], as well as by physiological adaptations in skeletal muscle, including an increase in mitochondrial content, which results in increased respiratory capacity [26,27]. Thus, Sb supplementation does not seem to affect the biomechanical and physiological aspects mentioned above, and most likely, the Sb-mediated increase in  $VO_{2MAX}$  does not result from activation of the AMPK/PGC-1 $\alpha$  pathway.

The significant and positive correlation between  $VO_{2MAX}$  and time to fatigue does not allow us to identify which one is the cause and which one is the consequence. The rationale presented in the previous paragraphs supports the idea that increased muscle function, characterized by greater  $VO_{2MAX}$ , leads to improved performance. In contrast, one can argue that increased performance may have resulted from changes in non-metabolic factors (e.g., motivation), forcing rats to reach greater rates of oxygen consumption. This rationale (discussed in the next paragraph) comes with the assumption that non-supplemented rats did not attain their  $VO_{2MAX}$  during the incremental exercise. Thus, the greater performance caused by supplementation with probiotics may also have resulted from changes in neural function.

Evidence indicates that the ingestion of probiotics is associated with changes in the concentrations of serotonin and dopamine metabolites in specific brain areas [28] and promotes anxiolytic-like effects in rats and beneficial psychological effects in healthy human volunteers [29]. In fact, there is evidence pointing to a potential impact of the enteric microbiota on brain function [30]; more specifically, many of the gut microbiota or potential probiotics-mediated effects on brain function are dependent on vagal activation [31], although vagus-independent mechanisms have already been reported [32]. Therefore, it is possible that the ergogenic effects mediated by Sb may be dependent on the modulation of serotonergic and dopaminergic neurotransmission, both of which have been associated with aerobic performance [33]. As previously identified, Sb administration changes gut microbiota composition [5,34], which may affect the activity of vagal afferent fibers in the gut, thereby influencing monoaminergic brain systems [35]. This hypothesis is based on similar observations regarding physical performance and  $VO_{2MAX}$  of rats treated with central dopamine [36]. If this hypothesis is correct, increased motivation to exercise would allow the rats to achieve faster speeds and therefore, a higher  $VO_{2MAX}$ .

Another hypothesis that could explain the increased performance in Sb-treated rats is the modulation of immune function, as evidenced by the observations that Sb supplementation reduces the concentrations of proinflammatory cytokines, including interleukin-8 (IL-8), IL-6, IL-1 $\beta$ , tumor necrosis factor alpha (TNF- $\alpha$ ), and interferon gamma (IFN- $\gamma$ ) [37–40]. Recently, a neuroinflammatory model was proposed by Vargas and Marino [41] to explain fatigue during an acute physical exercise session. In this model, augmented concentrations of circulating IL-6 and other inflammatory mediators would

contribute to increased perceived fatigue during exercise, thus leading to less muscle recruitment and consequently, reduced aerobic performance.

Of note, all the hypotheses raised to explain the augmented performance and  $VO_{2MAX}$  in supplemented rats are merely speculative because we did not analyze the concentrations of intramuscular substrates, cytokines, and neurotransmitters. Future investigations should identify the mechanisms underlying the benefits mediated by Sb and evaluate whether the present findings can be translated to human physiology. At the present moment, our findings cannot be used to endorse Sb supplementation as an efficient strategy for improving aerobic performance of recreational or professional athletes. In addition, it is not clear whether the Sb-induced increases in  $VO_{2MAX}$  and  $S_{MAX}$  are indeed beneficial outcomes, as these increases were not accompanied by adaptations that are commonly caused by aerobic training, such as greater running economy and mechanical efficiency. For example, a dopamine/noradrenaline reuptake inhibitor enhances aerobic performance but also core body temperatures during exercise in a hot environment, suggesting that this drug overrides the inhibitory signals arising from the central nervous system that cause exercise to stop when close to critical temperature values [42]. This may also be the case for Sb supplementation, which may allow the rats to exercise beyond safe limits.

## 5. Conclusions

We conclude that Sb supplementation does not affect resting aerobic metabolism but does increase  $VO_{2MAX}$  and aerobic performance in rats. In addition, the present data rule out the suggestion that increased running economy and mechanical efficiency could explain the enhanced performance of supplemented rats.

**Author Contributions:** The individual author contributions are listed as follows: Conceptualization, A.D.N.S., S.P.W., F.S.M. and V.N.C.; methodology, A.D.N.S., S.P.W., E.S.S.M., A.S.R.H. and F.S.M.; formal analysis, A.D.N.S., S.P.W., A.S.R.H. and V.N.C.; investigation, A.D.N.S., E.S.S.M. and A.S.R.H.; data curation, A.D.N.S., S.P.W. and A.S.R.H.; writing—original draft preparation, A.D.N.S., S.P.W. and A.S.R.H.; writing—review and editing, A.D.N.S., S.P.W., E.S.S.M., A.S.R.H., F.S.M. and V.N.C. and A.S.R.H.; supervision, S.P.W. and V.N.C.; project administration, V.N.C.; funding acquisition, S.P.W. and V.N.C.

**Funding:** This study was funded by the following agencies: Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG, Brazil; grant number APQ-01983-18), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq, Brazil), and Pró-Reitoria de Pesquisa da Universidade Federal de Minas Gerais. In addition, this study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior—Brasil (CAPES)—Finance Code 001.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Food and Agriculture Organization (FAO). *Guidelines for the Evaluation of Probiotics in Food*; Report of a Joint FAO/WHO Working Group on Drafting Guidelines for the Evaluation of Probiotics in Food; FAO: London, ON, Canada, 2002.
2. Hill, C.; Guarner, F.; Reid, G.; Gibson, G.R.; Merenstein, D.J.; Pot, B.; Morelli, L.; Canani, R.B.; Flint, H.J.; Salminen, S.; et al. Expert consensus document: The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat. Rev. Gastroenterol. Hepatol.* **2014**, *11*, 506–514. [[CrossRef](#)] [[PubMed](#)]
3. McFarland, L.V. Systematic review and meta-analysis of *Saccharomyces boulardii* in adult patients. *World J. Gastroenterol.* **2010**, *16*, 2202–2222. [[CrossRef](#)]
4. Girard, P.; Pansart, Y.; Verleye, M. Anti-hypercholesterolemic effect of *Saccharomyces boulardii* in the hamster. *Pharmacology* **2014**, *94*, 239–244. [[CrossRef](#)] [[PubMed](#)]
5. Everard, A.; Matamoros, S.; Geurts, L.; Delzenne, N.M.; Cani, P.D. *Saccharomyces boulardii* administration changes gut microbiota and reduces hepatic steatosis, low-grade inflammation, and fat mass in obese and type 2 diabetic db/db mice. *mBio* **2014**, *5*, e01011–e01014. [[CrossRef](#)] [[PubMed](#)]

6. Schneider, S.M.; Girard-Pipau, F.; Filippi, J.; Hebuterne, X.; Moyses, D.; Hinojosa, G.C.; Pompei, A.; Rampal, P. Effects of *Saccharomyces boulardii* on fecal short-chain fatty acids and microflora in patients on long-term total enteral nutrition. *World J. Gastroenterol.* **2005**, *11*, 6165–6169. [[CrossRef](#)] [[PubMed](#)]
7. Den Besten, G.; Van Eunen, K.; Groen, A.K.; Venema, K.; Reijngoud, D.J.; Bakker, B.M. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *J. Lipid Res.* **2013**, *54*, 2325–2340. [[CrossRef](#)]
8. Shing, C.M.; Peake, J.M.; Lim, C.L.; Briskey, D.; Walsh, N.P.; Fortes, M.B.; Vitetta, L. Effects of probiotics supplementation on gastrointestinal permeability, inflammation and exercise performance in the heat. *Eur. J. Appl. Physiol.* **2014**, *114*, 93–103. [[CrossRef](#)]
9. Cox, A.J.; Pyne, D.B.; Saunders, P.U.; Fricker, P.A. Oral administration of the probiotic *Lactobacillus fermentum* VRI-003 and mucosal immunity in endurance athletes. *Br. J. Sports Med.* **2010**, *44*, 222–226. [[CrossRef](#)]
10. Lundby, C.; Jacobs, R.A. Adaptations of skeletal muscle mitochondria to exercise training. *Exp. Physiol.* **2016**, *101*, 17–22. [[CrossRef](#)]
11. Joyner, M.J.; Coyle, E.F. Endurance exercise performance: The physiology of champions. *J. Physiol.* **2008**, *586*, 35–44. [[CrossRef](#)]
12. Teixeira-Coelho, F.; Fonseca, C.G.; Barbosa, N.H.S.; Vaz, F.F.; Cordeiro, L.M.S.; Coimbra, C.C.; Pires, W.; Soares, D.D.; Wanner, S.P. Effects of manipulating the duration and intensity of aerobic training sessions on the physical performance of rats. *PLoS ONE* **2017**, *12*, e0183763. [[CrossRef](#)]
13. Wisloff, U.; Helgerud, J.; Kemi, O.J.; Ellingsen, O. Intensity-controlled treadmill running in rats:  $\text{VO}_{2\text{max}}$  and cardiac hypertrophy. *Am. J. Physiol. Heart Circ. Physiol.* **2001**, *280*, H1301–H1310. [[CrossRef](#)]
14. Martins, F.S.; Vieira, A.T.; Elian, S.D.; Arantes, R.M.; Tiago, F.C.; Sousa, L.P.; Araújo, H.R.; Pimenta, P.F.; Bonjardim, C.A.; Nicoli, J.R.; et al. Inhibition of tissue inflammation and bacterial translocation as one of the protective mechanisms of *Saccharomyces boulardii* against *Salmonella* infection in mice. *Microbes Infect.* **2013**, *15*, 270–279. [[CrossRef](#)]
15. Wanner, S.P.; Leite, L.H.; Guimarães, J.B.; Coimbra, C.C. Increased brain L-arginine availability facilitates cutaneous heat loss induced by running exercise. *Clin. Exp. Pharmacol. Physiol.* **2015**, *42*, 609–616. [[CrossRef](#)]
16. Kunstetter, A.C.; Wanner, S.P.; Madeira, L.G.; Wilke, C.F.; Rodrigues, L.O.; Lima, N.R. Association between the increase in brain temperature and physical performance at different exercise intensities and protocols in a temperate environment. *Braz. J. Med. Biol. Res.* **2014**, *47*, 679–688. [[CrossRef](#)]
17. Kunstetter, A.C.; Barbosa, N.H.S.; Moraes, M.M.; Pinto, V.A.; Soares, D.D.; Pires, W.; Wanner, S.P. Pre-exercise exposure to the treadmill setup changes the cardiovascular and thermoregulatory responses induced by subsequent treadmill running in rats. *Temperature* **2017**, *5*, 109–122. [[CrossRef](#)]
18. Helgerud, J. Maximal oxygen uptake, anaerobic threshold and running performance in women and men with similar performances levels in marathons. *Eur. J. Appl. Physiol.* **1994**, *68*, 155–161. [[CrossRef](#)]
19. Brooks, G.A.; Donovan, C.M.; White, T.P. Estimation of anaerobic energy production and efficiency in rats during exercise. *J. Appl. Physiol. Respir. Environ. Exerc. Physiol.* **1984**, *56*, 520–525. [[CrossRef](#)]
20. Hopkins, W.G.; Marshall, S.W.; Batterham, A.M.; Hanin, J. Progressive statistics for studies in sports medicine and exercise science. *Med. Sci. Sports Exerc.* **2009**, *41*, 3–13. [[CrossRef](#)]
21. Chen, Y.M.; Wei, L.; Chiu, Y.S.; Hsu, Y.J.; Tsai, T.Y.; Wang, M.F.; Huang, C.C. *Lactobacillus plantarum* TWK10 supplementation improves exercise performance and increases muscle mass in mice. *Nutrients* **2016**, *8*, 205. [[CrossRef](#)]
22. Pyne, D.B.; West, N.P.; Cox, A.J.; Cripps, A.W. Probiotics supplementation for athletes—Clinical and physiological effects. *Eur. J. Sport Sci.* **2015**, *15*, 63–72. [[CrossRef](#)]
23. Gao, Z.; Yin, J.; Zhang, J.; Ward, R.E.; Martin, R.J.; Lefevre, M.; Cefalu, W.T.; Ye, J. Butyrate improves insulin sensitivity and increases energy expenditure in mice. *Diabetes* **2009**, *58*, 1509–1517. [[CrossRef](#)]
24. Rabelo, P.C.R.; Cordeiro, L.M.S.; Aquino, N.S.S.; Fonseca, B.B.B.; Coimbra, C.C.; Wanner, S.P.; Szawka, R.E.; Soares, D.D. Rats with higher intrinsic exercise capacities exhibit greater preoptic dopamine levels and greater mechanical and thermoregulatory efficiencies while running. *J. Appl. Physiol.* **2019**, *126*, 393–402. [[CrossRef](#)]
25. Saunders, P.U.; Pyne, D.B.; Telford, R.D.; Hawley, J.A. Factors affecting running economy in trained distance runners. *Sports Med.* **2004**, *34*, 465–485. [[CrossRef](#)]
26. Baar, K. Involvement of PPAR $\gamma$  co-activator-1, nuclear respiratory factors 1 and 2, and PPAR $\alpha$  in the adaptive response to endurance exercise. *Proc. Nutr. Soc.* **2004**, *63*, 269–273. [[CrossRef](#)]

27. Holloszy, J.O. Regulation by exercise of skeletal muscle content of mitochondria and GLUT4. *J. Physiol. Pharmacol.* **2008**, *59*, 5–18.
28. Desbonnet, L.; Garrett, L.; Clarke, G.; Bienenstock, J.; Dinan, T.G. The probiotic *Bifidobacteria infantis*: An assessment of potential antidepressant properties in the rat. *J. Psychiatr. Res.* **2008**, *43*, 164–174. [[CrossRef](#)]
29. Messaoudi, M.; Lalonde, R.; Violle, N.; Javelot, H.; Desor, D.; Nejd, A.; Bisson, J.F.; Rougeot, C.; Pichelin, M.; Cazaubiel, M.; et al. Assessment of psychotropic-like properties of a probiotic formulation (*Lactobacillus helveticus* R0052 and *Bifidobacterium longum* R0175) in rats and human subjects. *Br. J. Nutr.* **2011**, *105*, 755–764. [[CrossRef](#)]
30. Khanna, S.; Tosh, P.K. A clinician’s primer on the role of the microbiome in human health and disease. *Mayo Clin. Proc.* **2014**, *89*, 107–114. [[CrossRef](#)]
31. Bercik, P.; Collins, S.M.; Verdu, E.F. Microbes and the gut-brain axis. *Neurogastroenterol. Motil.* **2012**, *24*, 405–413. [[CrossRef](#)]
32. Dinan, T.G.; Stilling, R.M.; Stanton, C.; Cryan, J.F. Collective unconscious: How gut microbes shape human behavior. *J. Psychiatr. Res.* **2015**, *63*, 1–9. [[CrossRef](#)]
33. Cordeiro, L.M.S.; Rabelo, P.C.R.; Moraes, M.M.; Teixeira-Coelho, F.; Coimbra, C.C.; Wanner, S.P.; Soares, D.D. Physical exercise-induced fatigue: The role of serotonergic and dopaminergic systems. *Braz. J. Med. Biol. Res.* **2017**, *50*, e6432. [[CrossRef](#)]
34. Girard-Pipau, F.; Pompei, A.; Schneider, S.; Nano, J.L.; Hebuterne, X.; Boquet, P.; Rampal, P. Intestinal microflora, short chain and cellular fatty acids, influence of a probiotic *Saccharomyces boulardii*. *Microb. Ecol. Health Dis.* **2002**, *14*, 221–228. [[CrossRef](#)]
35. Breit, S.; Kupferberg, A.; Rogler, G.; Hasler, G. Vagus nerve as modulator of the brain–gut axis in psychiatric and inflammatory disorders. *Front. Psychiatry* **2018**, *9*, 44. [[CrossRef](#)]
36. Balthazar, C.H.; Leite, L.H.; Rodrigues, A.G.; Coimbra, C.C. Performance-enhancing and thermoregulatory effects of intracerebroventricular dopamine in running rats. *Pharmacol. Biochem. Behav.* **2009**, *93*, 465–469. [[CrossRef](#)]
37. Dalmaso, G.; Cottrez, F.; Imbert, V.; Lagadec, P.; Peyron, J.F.; Rampal, P.; Czerucka, D.; Groux, H.; Foussat, A.; Brun, V. *Saccharomyces boulardii* inhibits inflammatory bowel disease by trapping T cells in mesenteric lymph nodes. *Gastroenterology* **2006**, *131*, 1812–1825. [[CrossRef](#)]
38. Dalmaso, G.; Loubat, A.; Dahan, S.; Calle, G.; Rampal, P.; Czerucka, D. *Saccharomyces boulardii* prevents TNF- $\alpha$ -induced apoptosis in EHEC-infected T84 cells. *Res. Microbiol.* **2006**, *157*, 456–465. [[CrossRef](#)]
39. Sougioultzis, S.; Simeonidis, S.; Bhaskar, K.R.; Chen, X.; Anton, P.M.; Keates, S.; Pothoulakis, C.; Kelly, C.P. *Saccharomyces boulardii* produces a soluble anti-inflammatory factor that inhibits NF- $\kappa$ B-mediated IL-8 gene expression. *Biochem. Biophys. Res. Commun.* **2006**, *343*, 69–76. [[CrossRef](#)]
40. Mumy, K.L.; Chen, X.; Kelly, C.P.; McCormick, B.A. *Saccharomyces boulardii* interferes with Shigella pathogenesis by postinvasion signaling events. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2008**, *294*, 599–609. [[CrossRef](#)]
41. Vargas, N.T.; Marino, F. A neuroinflammatory model for acute fatigue during exercise. *Sports Med.* **2014**, *44*, 1479–1487. [[CrossRef](#)]
42. Hasegawa, H.; Piacentini, M.F.; Sarre, S.; Michotte, Y.; Ishiwata, T.; Meeusen, R. Influence of brain catecholamines on the development of fatigue in exercising rats in the heat. *J. Physiol.* **2008**, *586*, 141–149. [[CrossRef](#)]

