



Article Decomposition Behavior of Biodegradable and Single-Use Tableware Items in the Warnow Estuary (Baltic Sea)

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Abstract: Due to the increasing demand for alternative and supposedly more sustainable single-use tableware compared to conventional fossil-based plastic items, there now exists a broad variety of innovative new bio-based and biodegradable items. The aim of this research is to conduct a monitoring of the biodegradability of selected tableware items: polylactic acid (PLA), crystallized polylactic acid (CPLA), wood, palm leaf, paper/cellulose and sugar cane bagasse, as well as a common fossil-based plastic polystyrene (PS) as a comparative material in a water column, to determine their possible use as a substitute for single-use plastic flatware. In situ experiments were performed over a one-year period to consider changes in water temperature, salinity gradients, pH values and dissolved oxygen (DO). The main findings show that tableware items composed of palm leaf, sugar cane bagasse and paper are less resistant to estuary conditions. However, wood, PLA, CPLA and PS items showed a certain resistance during the experimentation period. Furthermore, items placed in the upper incubators degraded relatively faster than those of the same material but placed in the lower incubators. The results indicate that temperature and exposure to natural light could be an important factor in terms of accelerating the degradation of the materials considered.

Keywords: biodegradability; bio-based tableware; conventional plastic; degradation rate; marine litter; Warnow River estuary

1. Introduction

Marine litter is recognized worldwide as a rising pollution problem affecting all the oceans and coastal areas of the world [1–3]. The increasing production and use of durable synthetic materials such as plastics has led to a gradual but significant accumulation of litter in the marine environment, making it ever more difficult to tackle [4,5].

Several studies have shown that marine litter exists in most oceans and seas, and their source and pathways have become an important focus of research [6]. This same work indicates that the identification of the origin of litter can help tackle the problem by implementing strategies to stop and avoid any pollutant reaching the water. Marine litter originates mainly from public littering, poor waste management practices, industrial activities, sewage-related debris and storm water discharge, all of which can be transported via rivers [7–9], until it reaches the seas and the oceans.

According to the Helsinki Commission (HELCOM), marine litter is a clearly visible problem along the Baltic Sea coastline. Around 70% of the marine litter in the Baltic Sea is plastic and represents a special concern due to its slow rate of degradation and its risk to the environment [10,11]. Therefore, a regional goal has been agreed within the area dealt by the Baltic Marine Environment Protection Commission on significantly reducing the amount of marine litter by 2025 and preventing harm from litter in the coastal and marine environment [12].



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). In fact, plastic pollution in particular is globally distributed across all oceans due to its properties of buoyancy and durability, and the sorption of toxicants to plastic while traveling through the environment [10,11].

In the Warnow estuary of Rostock, research carried out on existing plastics in the sediments showed that most the abundant items were plastic fragments (32.6%), followed by flat pieces of plastic (strapping bands, 20%), cups and cup lids (6.3%), corrugated plastic (5.9%), food containers incl. fast food containers (5.6%), etc. In fact, 26% of the litter is generated by residents and tourists [13].

In addition, according to the European Union, 80% to 85 % of marine litter is plastic and single-use plastic (representing 50% of the total) which results from a diverse range of commonly used fast-moving consumer products that are discarded after having been used once for the purpose for which they were provided. These are rarely recycled and are prone to becoming litter [14]. Furthermore, most plastics do not fully degrade, but split into tiny fragments [15]. For example, Rochman et al. [16] revealed that synthetic polymers in the ocean should be considered as hazardous waste. Some studies report little or no physical or chemical harm to marine biota [17], while others, which include the use of a thermodynamic approach and the simulation of physiological conditions in the gut, suggest that chemicals in plastics might be released to organisms after ingestion [18,19].

Despite the advantages of using plastics, their harmful effects on nature remain a problem to be solved. Hence, several research studies have been aimed at the development of alternative materials to single-use plastic items. Research and development are on-going to create harmonized standards for these items to be classified as biodegradable in water, which is needed before relevant products can be introduced to the market [20].

In fact, several products have been developed as environmentally friendly that can substitute the use of non-degradable materials, particularly in the water. For example, biodegradable items are defined as any material likely to be degraded under appropriate conditions of temperature, humidity, oxygenation and pH, and under the effect of microorganisms, fungi or bacteria [21]. The outputs of biodegradability are the formation of water, CO_2 and/or methane and possibly by-products (residues, new biomass) that are not toxic to the environment [22]. Moreover, eco-friendly plastics, namely bio-based plastics, can be produced from renewable resources, and biodegradable plastics degrade in the environment to solve global environmental and waste management problems [23]. With society's growing concern for sustainable consumption, production and waste management, biodegradable packaging may appear as an environmentally friendly option and could rapidly become an attractive commercial feature [24]. In addition, the escalating problems caused by non-degradable plastics have led to the development of biodegradable plastics [25]. These polymers have the property of being biodegradable and are named biodegradable polymers (BDPs) or biodegradable plastics, which refers to polymeric materials that are capable of undergoing decomposition into carbon dioxide, methane, water, inorganic compounds or biomass in which the predominant mechanism is the enzymatic action of microorganisms [26].

Nowadays, the term 'biodegradable' is still under discussion between experts and decision makers. It is a term that should be distinguished from similar terms (biodegradation, degradability, etc.) and used to describe a material with some care. Biodegradability was first defined in terms of use in the soil; however, further studies have proved its use in water columns.

According to the United Nation's (UN) environment program, polymers, which biodegrade under favorable conditions on land, are much slower to break up in the ocean, and their widespread adoption is likely to contribute to marine litter and consequent undesirable consequences for marine ecosystems [27].

These experiences could lead to different hypotheses based on the origin of the materials, their behavior in the natural environment and to the affecting environmental changes. Regarding the PLA and CPLA as well as plastic items (in this case PS), their biodegradability in water columns does not seem to be achievable in one year. Other products (palm leaves, sugar canes, wood and paper) could be degradable after a shorter period, for the reason that this is linked to their original state and their behavior in nature.

The objective of the current research is to test the biodegradability of plastic tablewarereplacement items, PLA, CPLA, palm leaf, sugar cane (bleached/unbleached), wood, paper and PS, in a natural estuarine environment in Rostock city. To do this, the biodegradability of selected items was monitored via in situ experiments over a one-year period while monitoring the physicochemical parameters of the water (temperature, salinity, DO and pH) and the natural habitat of marine organisms on the incubators (barnacles, mussel, algae, etc.). The motivation behind this research is to understand the resistance of the selected items to the marine and natural conditions, and to reinforce international research into finding biodegradable alternatives to single-use plastic.

2. Study Site and Methods

2.1. Methods

The methodology of work began by setting the items to be tested. Used samples are already commercialized and used at some coastal events in Rostock (festivals, markets, street-food events, Hanse Sail, etc.). Once the items were provided by the municipality, a documentation with the description and the characteristics of each sample were noted.

To ensure a secure enrolment of the experiments in the water column, it was essential to develop a specific and rigid tool that ensures the natural circulation of the water and the contact with the product. Therefore, adapted incubators were designed and constructed within the workshop of the Leibniz Institute for Baltic Sea Research, described in Section 2.3. In addition, choosing the areas had to be very precise by respecting some criteria such as water depth, salinity, access to the station, water clarity and surrounding activities. Finally, incubators were prepared to be placed according to the research protocol.

Installed incubators were controlled periodically to clean them and to ensure an adequate running of the experiments. Additionally, the items' degradation rate (DR) was monitored in parallel with the water's physicochemical parameters. The work was complemented by data analyses to arrive at possible conclusions and to allow us to make appropriate recommendations.



The concept approach of this study is summarized in Figure 1.

Figure 1. The concept approach.

2.2. Study Site

The incubation of different samples took place in the Warnow River estuary crossing Rostock city, which is located in the German Federal state of Mecklenburg-Western Pomerania on the Baltic German coast. The Warnow River, which is 155 km long, is the second longest river in Mecklenburg-Western Pomerania. Through this channel salty ocean water reaches the estuary and spreads over the whole river. The salt gradient between the Baltic Sea and fresh water from the Warnow River drives an estuarine circulation with an inflow (upstream) of saline water near the surface, and an outflow (seaward) of near-surface water [28].

From both sides, the coastal area of the Warnow estuary can be divided according to the different activities such as ship building industry, waste industry, the Rostock port, tourism and recreation, beaches, green areas, reed belts, military area and others. This variety of the sections affects the Warnow directly or indirectly, a reason to do more research and studies about this small and semi-closed area to preserve it and avoid all types of pollution.

The Figure 2 shows the exact location of each chosen station as well as the incubators emplacement.



Figure 2. Location of selected stations S1, S2 to S3 in the Warnow River estuary at Rostock city in addition to the hanging incubators in the water column.

The land-side coastal zone area is the most heavily built-on and used urban environment in the world. From both sides, the coastal area of the estuary can be divided according to different industrial activities such as ship building, waste treatment, Rostock port, tourism and recreation, beaches, green areas, reed belts, military area and others. This variety of activities affects the Warnow River directly and indirectly, hence a reason for undertaking more research and studies with regard to this small and semi-enclosed area in order to preserve it and to avoid all types of pollution.

Three different stations were chosen for the location of the incubators and were named in terms of the area in which they were located. They are distributed, respectively from North to South along the Warnow River estuary, as follows: Hohe Düne, IGA Park and Gehlsdorf.

S1. Hohe Düne: This station is at the Marine Science Center which is a seal research center which is part of the Institute for Biosciences at the University of Rostock. It is located at the Marina of Hohe Düne right on the Warnow's east breakwater. The incubators are hung on plastic floating buoys which surround the seal and penguin cages. The geographical coordinates are $54^{\circ}11'02.4''$ N, $12^{\circ}05'43.4''$ E. The characteristics of this station are the water depth (around 5 m), the water clarity (transparency of the water is high all year round), mechanical effect due to the transport flow involving small boats and yachts (which could be a source of invasive species) and different weather conditions. During the verification of the incubators, the presence of several species, such as small shrimps, medusa, etc., was evident. This station is known by scientific research activities related to different animals such as seals and penguins, it is also an open spot for visitors.

S2. IGA Park: This station is located at the IGA Park as indicated by its name. This park is located at Schmarl Dorf, known for different activities, such as open-air concerts and musical events, food exposures and even a place of attraction for swimmers and water activities performed throughout the year, etc. On the coastal part of the park, which is exposed to the Warnow River estuary, there is a platform next to the Maritime Museum which extends into the estuary for approximately 165 m. The incubators are hung at the extremity of this platform with the geographic coordinates: 54°08′27.4″ N, 12°05′21.2″ E. It is characterized by its exposure to the different currents of the Warnow River and to the sun; parameters that are important and play a role in the degradation of the materials to be tested. The water depth is around 4 m, and water clarity is not high and depends on the season. This station is also characterized by an important growth of mussels. Access is supported by the park administration.

S3. Gehlsdorf: This station is located at the rowing club opposite Rostock city. The boat dock used belongs to the rowing club, and the incubators were hung to the outer end. The geographic coordinates are $54^{\circ}05'47.8''$ N, $12^{\circ}07'54.1''$ E. The water depth is approximately 3 m, and the water transparency is very low. This area is known for an intense activity in terms of rowers training throughout the year, and recreational boats being active during the summer season.

The choice of the three stations was based on different relevant criteria that may influence the rapidity of the degradation or the general progress of the experiments. In addition, the accessibility to the station and the security of the incubators during the experimentation period represents a crucial factor that future projects should consider.

2.3. Incubators Preparation

Incubators were designed and constructed within the IOW Institute [29]. They are made of a circular hard plastic cage (black part) and a 2 mm mesh size plastic net which was used to build the walls of each incubator. The mesh size allows good water circulation, which is considered an important mechanical parameter. The steps to get a finalized incubator are presented in the following Figure 3.



Figure 3. Incubator preparation for experiments.

Tableware items were cut into small pieces in order to fit into the incubators and were weighed and measured. For each item, a sample was placed in an upper incubator (close to the surface, more exposed to sunlight and to pelagic organisms) and in a lower one (closer to the bottom, less exposed to sunlight and to demersal organisms). In total and at each station, 12 incubators were placed in the water in 3 groups of 4 incubators and were attached to ropes. They were hung from the top and fixed with chains to the bottom as shown in Figure 4.

Regarding the label attributed to each incubator, and as an example, at station S1, paper samples were divided into two fragments, one is in the upper incubator (U), hence named M.1.1.U, and the other in the lower one (D) and named M.1.3.D.



Figure 4. Example of preparation and installation of the attached incubators to ropes.

2.4. Incubators Installation

The incubators were installed at a precise spot at each station in order to ensure easy access for parameter monitoring and incubator opening. The ropes were attached at each station into different boarders: at S1 they were attached to floating buoys, at S2 to a wooden barrier and at S3 to a rope fixed at the end of the dock. The experiments started at the same period (end of April for S2 and S3 and beginning of May for S1). The idea is to have the same weather conditions simultaneously for the different experiment locations.

At IGA Park, the incubators were hung against a wall (Figure 3), but in the two other stations they were suspended freely in the water column.

2.5. Used Tableware Items

Provided tableware items are from a range of different sizes, shapes, colors and densities. They were made of PLA, CPLA, palm leaf, sugar cane bagasse (bleached/unbleached), wood, paper/cellulose and single-used fossil-based conventional plastics (PS). Some examples of used tableware items are presented in Figure 5.



Cardboard coating from starch

Wood Pressed / without any coating

PLA and CPLA Bio-plastic synthesized from renewable raw materials e.g. lactic acid from corn starch



Palm leaves Pressed / without any coating

Sugar cane un-/bleached pressed fibers natural binder

Figure 5. Tableware used for experiments.



PLA or polylactic acid is a biodegradable polyester which is very versatile and is derived from 100% renewable resources [30]. It is produced from corn, starch and sugars [31,32]. It has a density value between 1200–1450 kg/m³, and it is used for packaging food products given its high permeability to water [33]. According to [34], soil burial tests show that the degradation of PLA is slow, and that it takes a long time for degradation to start. For instance, no degradation was observed on the material sheets after 6 weeks in soil.

CPLA or crystallized poly lactic acid is obtained through sugars and corn starches. When producing PLA, lime is added. The material then becomes crystallized and resistant to heat. CPLA, when used for tableware, is considered to be a bioplastic. In a natural environment (soil), it starts to degrade in 5 to 6 months, with complete decomposition after 12 months. If composted with food garbage, it begins to decompose after 2 weeks [35].

According to the technical file relating to the product used:

- The plates and bowls were made from discarded and pressed palm leaf from the "Adaka" nut palm. Depending on the source of the palm leaf used, its density is between 0.7–1.55 g/cm³ [36].
- The sugar cane bagasse products used in these experiments are obtained from extracting sugar from the cane's marrow leaf "bio-mass" (fiber), which is usually burned. The density of sugar cane is 1.20 g/m³ [37].

Paper is composed of lignocellulose. The density of paper is 700 kg/m³ according to [38,39]. According to the technical file of the product used, cardboard cups are made from pure cellulose. In order to make the cups resistant to moisture, the cardboard is coated on the inside with a starch film.

Wood packaging is mainly used for the transport of goods and is mainly made of cellulose [24]. The wooden tableware used was made or punched from birch wood, having a density of 623 kg/m³ [40], using a pressing process. There is no further coating or surface treatment.

Polystyrene (PS) is a synthetic hydrophobic polymer with a high molecular weight and a density of 1060 kg/m³ [41]. It is recyclable but not biodegradable, and at room temperature, it exists in a solid state. PS, being a transparent hard plastic, is commonly used as disposable tableware [42].

The values of densities give a better understanding of biodegradable items' behavior in water. Some tend to sink while others float. This affects their fragmentation and the behavior of living organisms (which could use the products as habitats or nutrition elements) throughout the experimental period. PLA and sugar cane tend to sink after a certain time, while wood, paper and PS items tend to float. In the case of palm leaf materials, differences in density margins mean that they can either float or sink.

2.6. Parameter Monitoring Tools

In order to obtain the required data regarding the measurements of DO, temperature, pH and salinity, specific tools were used involving cables and measuring device from the Hach HQD digital multimeter which combines reliability, flexibility and ease of use. The devices used for measurement in the water were an HQ40D Portable pH, Conductivity/TDS & Dissolved Oxygen Meter, Premium Field Kit for Environmental Monitoring with a 5 m Cable and a CTD48M gear. The follow up of these parameters was done according to a schedule. Initially measurements were done with a frequency of two to three times per month, while at the end of the experimental period, it was done about one to two times per month, depending on the availability of the equipment and access to the stations.

3. Results

3.1. Degradation Rate Monitoring

The incubators were removed periodically from the water column to clean them and to identify the status of the materials from Hohe Düne after 50 days, 202 days and 339 days after the first day of installation, from IGA Park after 65 days, 165 days and 399 days, and from Gehlsdorf after 64 days, 148 days and 356 days. At the end of the experimental period, the walls of the incubators were removed, opened at the IOW laboratory, and the results were documented. Samples were weighed and left in labelled bags. The following Tables 1–3 show the difference between the initial and the final weight of the samples after the experimentation period. The degradation rate was calculated using the following formula:

Degradation Rate :
$$(DR) = (Wo - Wt / Wo) \times 100$$
 (1)

where W₀ and W_t are the weights of the initial and degraded items, respectively.

Incubators and Content	Initial Weights of Samples	Final Total Weight	Degradation Rate (DR) (%)	Final Status of the Item
HD.1.1.U Paper	Plate (1.6 g), small cup (0.3 g)	1.1 g (-0.8 g)	42.1%	Soft to the touch
HD.1.2.U Plastics	Cup (1 g), box (2.1 g)	4.3 g (+1.2 g)	0%	Not degraded
HD.1.3.D Paper	Plate (1.8 g), small cup (0.4 g)	1.5 g (-0.7 g)	31.8%	Soft to the touch
HD.1.4.D Plastics	Cup (1 g), box (1.8 g)	3.5 g (+0.7 g)	0%	Not degraded
HD.2.1.U Wood	Spoon (2.5 g), fork (2 g)	4.8 g (+0.3 g)	0%	Soft to the touch
HD.2.2.U Palm leaf	Plate (3.3 g)	0.6 g (-2.7 g)	81.8%	Small fibers
HD.2.3.D Wood	Spoon (1.6 g), fork (2 g)	5.8 g (+2.2 g)	0%	Soft
HD.2.4.D Palm leaf	Plate (2.9 g)	0 g (-2.9 g)	100%	Totally degraded
HD.3.1.U PLA/CPLA	PLA (lid 0.9 g, cup 1.2 g)/ CPLA (lid 1.9 g, spoon 2.4 g)	6.6 g (+0.2 g)	0%	Not degraded
HD.3.2.U Sugar cane	Bleached (1.1 g)/ unbleached (1.2 g)	0 g (-2.3 g)	100%	Totally degraded
HD.3.3.D PLA/CPLA	PLA (lid 1 g, cup 1.1 g)/ CPLA (lid 1.7 g, spoon 2 g)	6 g (+0.2 g)	0%	Not degraded
HD.3.4.D Sugar cane	Bleached (1 g)/ unbleached (1.3 g)	0 g (-2.3 g)	100%	Totally degraded

Table 1. Results of the degradation of selected items in Station S1, Hohe Düne (HD).

Table 2. Results of the degradation of selected items in Station S2, IGA Park (I).

Incubators and Content	Initial Weights of Samples	Final Total Weight	Degradation Rate (%)	Final Status of the Item
I.1.1.U Plastics/CPLA	Cup (0.6 g), box (1.5 g), knife (1.6 g)	4 g (+0.3 g)	0%	Not degraded
I.1.2.U Sugar cane	Bleached (2.8 g)/ unbleached (1.3 g)	0 g (-4.1 g)	100%	Totally degraded
I.1.3.D Plastics/CPLA	Cup (0.6 g), box (1.5 g), knife (1.8 g)	4.5 g (+0.6 g)	0%	Not degraded
I.1.4.D Sugar cane	Bleached (2.8 g)/ unbleached (1.3 g)	1.4 g (-2.7 g)	65.8%	Soft to the touch
I.2.1.U Wood	Tableware (1.3 g)/(2.2 g)	7.1 g (+3.6 g)	0%	Soft to the touch
I.2.2.U Palm leaf	Rectangular plate (1.5 g)	0 g (-1.5 g)	100%	Totally degraded
I.2.3.D Wood	Tableware (1.8 g)/(1.7 g)	6.8 g (+3.3 g)	0%	Soft to the touch
I.2.4.D Palm leaf	Plate (1.6 g)	1.1 g (−0.5 g)	31.2%	Small fibers
I.3.1.U Paper	Plate (1.7 g), cup (2 g)	0 g (-3.7 g)	100%	Totally degraded
I.3.2.U PLA/CPLA	PLA (cup 2.5 g)/ CPLA (knife 2.3 g, lid 1.7 g)	18.2 g (+11.7 g)	0%	Not degraded
I.3.3.D Paper	Plate (1.7 g), cup (1.9 g)	0 g (-3.6 g)	100%	Soft to the touch
I.3.4.D PLA/CPLA	PLA (cup 2.5 g)/ CPLA (knife 2.3 g, lid 1.7 g)	9.1 g (+2.6 g)	0%	Not degraded

Incubators and Content	Initial Weights of Samples	Final Total Weight	Degradation Rate (%)	Final Status of the Item
G.1.1.U Sugar cane	Bleached (1.6 g)/unbleached (1.9 g)	0 g (-3.5 g)	100%	Totally degraded
G.1.2.U PLA/CPLA	PLA (cup 1 g, box 1.8 g)/CPLA (lid 1.5 g, spoon 3.1 g, knife 2.5 g)	11.6 g (+1++6.7 g)	0%	Not degraded
G.1.3.D Sugar cane	Bleached (1.6 g), unbleached (1.8 g)	0 g (-3.4 g)	100%	Totally degraded
G.1.4.D PLA/CPLA	PLA (cup 1 g, box 1.8 g)/CPLA (lid 1.6 g, spoon 2.7 g, knife 1.9 g)	10.6 g (+1.6 g)	0%	Not degraded
G.2.1.U Palm leaf	Plate (4 g)	0 g (-4 g)	100%	Totally degraded
G.2.2.U Wood	Spoon (2.3 g), stick (0.7 g)	6.7 g (+3.7 g)	0%	Soft to the touch
G.2.3.D Palm leaf	Plate (4.2 g)	1.3 g (-2.9 g)	69%	Small fibers
G.2.4.D Wood	Spoon (1.5 g), stick (0.8 g)	4.6 g (+2.3 g)	0%	Soft to the touch
G.3.1.U Paper	Plate (1.6 g), cup (1.7 g)	0 g (-3.3 g)	100%	Soft to the touch
G.3.2.U Plastics	Cup (0.8 g), box (1.6 g)	3.3 g (+0.9 g)	0%	Not degraded
G.3.3.D Paper	Plate (1.7 g), cup (1.7 g)	1.3 g (-2.1 g)	61.7%	Soft to the touch
G.3.4.D Plastics	Cup (0.9 g), box (1.7 g)	3.2 g (+0.6 g)	0%	Not degraded

Table 3. Results of the degradation of selected items in Station S3, Gehlsdorf (G).

At S1, the results show that the DR of sugar cane and palm leaf placed in the upper and lower incubators was high. Paper was relatively resistant to the water conditions (42.1% and 31.8%), but its morphology changed considerably, and it tended to degrade. However, PS, PLA/CPLA and wood in both upper and lower incubators were not degraded (DR: 0%). In addition, it was noted that barnacles were growing on its surface, which explain the gain in weight (+0.2 to +2.2). In fact, as larvae, barnacles have a size between 0.35 and 1 mm [43], they can easily get inside the incubators which have a 2 mm mesh size. Barnacles were then able to fix themselves to materials.

The found results are also presented by examples of the final status of the items at S1 in Figure 6.



Figure 6. Examples of degraded items from the upper incubators at S1.

Results at S2 showed that the DR of paper, palm leaf and sugar canes items of the upper incubators was high (100%) and were relatively important in most lower incubators. Palm leaf degraded to small fibers which explains the registered weight. However, PS, wood and PLA/CPLA were resistant to the water conditions and degradation did not take place during the experimentation period. In fact, the process of degradation seemed to start



at that period of time. Figure 7 shows examples of items and differences noted between first time opening the incubators and the end of experiments.

Figure 7. Examples of degraded items from the upper incubators at S2.

At S3, sugar cane, palm leaf and paper items in the upper incubators totally degraded (DR: 100%). However, palm leaf items placed in the lower incubator degraded to a form of small fibers resulting in a DR of 65%. In addition, paper items in the lower incubators lost weight and changed their morphology. However, they did not degrade totally. No change was registered regarding PLA/CPLA, wood and PS items in terms of DR. On the contrary, their color changed, showing also a growth of barnacles, algae and mussels on top. As for S1 and S2, examples of difference between the items is presented in Figure 8.



Figure 8. Examples of degraded items from the upper incubators at S3.

3.2. Parameters and DR Monitoring

In this research it was not possible to follow the degradation rate of items periodically through weighing. In fact, most items were very wet, fragile and soft to the touch. The weighing operation could have lost some parts of the material and distorted the results. However, and in order to assess the influence of the monitored parameters on the DR, an estimation of the degradation is noted, based on the photographs taken during the field visits. The results of the parameter monitoring and the DR of the materials used are presented in Figures 9–11:



Figure 9. Parameter variation and DR in Upper and Lower incubators at S1.



Figure 10. Parameter variation and DR in Upper and Lower incubators at S2.

The characteristic parameters of natural water have a certain fluctuation caused by seasonal changes. Among them, water temperature shows the most obvious fluctuation between 4.7 °C and 24.4 °C (in the upper layer) and 5.2 °C and 22.7 °C (in the lower layer). The pH does not change significantly. The same is true in terms of salinity, with average values of 2.8‰ and 12.8‰ (in upper layer) and 6.7‰ and 13.2‰ (in the lower layer), respectively. The DO was not stable in either the upper or lower incubators.



Figure 11. Parameter variation and DR in Upper and Lower incubators at S3.

4. Discussion

Degradation is assessed by measuring the decay of relevant physical properties, such as changes in molecular weight mass and molecular mass distribution, tensile properties, mass loss, morphological changes, etc. [44–46]. In this research, both mass loss and morphological changes were considered. It is important then to characterize the degradation rate of several bio-based items because these materials can be applied in several economic activities in coastal areas, where the possible end-of-life is actually the sea. Therefore, suitable test methodologies are required.

The current study has been carried out during one year of incubation of the items in the Warnow estuary. The study confirms the results highlighted by Law. K.L. et al., that prolonged exposure to environmental factors such as heat, light or microbial action causes polymers to be degraded into smaller pieces [47].

After completion of the experiment, the main findings indicate that palm leaf, sugar cane bagasse and paper/cellulose almost degraded in most incubator locations, but not with the same rapidity of degradation. This depended on the station location and the position in the water column (in the upper or lower level). In fact, according to Jambeck et al., the exposure to UV radiation and oxygen causes photodegradation. These processes occur to a depth of 50–100 μ m and result in molecular weight reduction [48]. In addition, the morphology of some partially degraded materials changed, either slightly or considerably.

4.1. Final Morphology of Tableware Items

The morphology represents an important criterion for predicting the possible future of the tested tableware items. This changes differently, depending on the material and its characteristics. For example, most of the partial degraded palm leaf samples were transformed into fibers or into pieces which were very soft to the touch. Likewise, the nondegraded parts of sugar cane were very thin, and decomposed into pieces, with a brown color. In addition, incubators containing wood items were partly filled with barnacles and mussels using the samples as habitats. The morphology of wood items slightly changed, their color became darker and a bit softer to the touch. These changes indicated that the degradation process could have been started. At some stations, paper samples were covered by a thin layer of biofilm and some barnacles. The material was very soft and easily crushed, resulting in a complete loss of mechanical properties.

Finally, no changes were noted in terms of the shape of PLA with some change of color, which was the case for [49]. Same for the common plastic PS and CPLA. An important growth of mussels and barnacles on top of the samples and inside the incubators was noted.

4.2. Influence of Temperature on DR

The degradation of the placed items in all stations started in June, when the temperature increased considerably. Before June, the DR was, in most cases, 0% for all tested materials, and reached higher percentages by the end of the summer period. The experimental findings showed the important role of temperature with regard to accelerating the degradation of palm leaf, sugar cane and paper, particularly in the upper incubators, where the temperature was relatively higher than in the bottom due to its greater exposure to the sun. However, the results indicate that the temperature parameter had no effects on the degradation of PLA, CPLA and PS tableware.

For instance, the results confirm the finding of Huang. D et al. [49] that for example PLA, which exhibits excellent degradation performance in controlled industrial composting, shows no degradation in the ocean. The main reason behind is that the temperature in seawater is considered relatively low and the specific microbial species and number are less, resulting in obvious inhibition of the degradation process. This is also the case of CPLA and plastics items tested in the context of this research.

The results of the research carried out by Bagheri. A.R et al., on different materials including PLA, aimed to test their biodegradability in sea water and freshwater in a thermostatic chamber at 25 °C and under fluorescent light (16 h light and 8 h dark) for one year, showed that PLA did not show any significant degradability under the testing conditions [50].

The influence of weathering on different materials represents a complex issue. The degradation of the material depends on some different parameters, such as the temperature, mechanical factors, sunlight and the sample depth, which makes the analyses of the influence a complex issue [51,52].

This research of the degradation of tested items in the water column in the Warnow estuary indicates the potential that certain environmental parameters have, such as seawater temperature, sample depth and sunlight, to accelerate or to slow the degradation. The potential of the rest of parameters is still not known and needs a deeper analysis.

5. Conclusions and Recommendations

The Warnow River estuary is an area of Rostock city where several important events and activities take place during the year, which leads to a generation of high emissions of single-use tableware. This type of pollution differs from one area to another in the Warnow estuary. Therefore, three different stations were selected based on different placements, parameters and type of activities.

In this regard, a number of lessons learned could be transferred to future studies, such as ensuring the accessibility to the incubators, as well as the safety of the incubators while selecting the areas of experimentation.

The field work requires a regular check-up of the installed equipment in order to ensure the continuity of the experiments without disturbing the degradation process.

The main findings of this research indicate that tableware items composed of palm leaf, sugar cane and paper are less resistant to the seawater physicochemical parameters than wood, PLA, CPLA and plastic (PS) items, which showed a certain resistance during the experimentation period. Furthermore, items placed in the upper incubators degraded relatively faster in most cases than those of the same material and placed in the lower incubators. Furthermore, the results showed that the temperature and exposure to sunlight could be the main factors that accelerate the degradation of the materials. The degradation could also be related to the presence of water current and a greater amount of pelagic organisms. However, in this study the potential of these parameters on the DR has not been proved. Therefore, further studies will be needed to complete and fully check the test methodology used, in terms of the determination of both physical and biological degradation.

Based on the findings, it is recommended to consider sugar cane, palm leaf and paper items since they mainly degraded in the water condition of the Warnow estuary. However, it is desirable to continue testing the rest of items for a longer period to ensure an accurate average in terms of DR based on weather condition changes. It is also recommended that more tableware items be tested, particularly those recently put on the market. This study has some other limitations that need to be addressed. For example, the fact that the selected items have not been tested in other areas or countries with different weather conditions, in particular a higher temperature.

Experimentally speaking, its recommended that there is a need to test the degradability of materials through placing an incubator at the bottom of the water column. In fact, according to Schernewski G. et al. [13], the growth of bacteria and algae on the littered item in the water column increases in density and causes its deposition at the bottom.

In addition, mechanical destruction experimentations within the laboratory and under controlled conditions could be relevant to test the effect of water movements and currents on the DR of the items.

Moreover, the importance of specific factors and conditions such as temperature, UV light, pH and salinity in accelerating the degradation of items in the water column are still largely unknown. Therefore, it is highly recommended to develop further research to test the relation of the degradation parameters, to ensure a better understanding of the degradation process, and consequently, to support innovation in terms of the optimization of biodegradable item production.

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