



Article The Effects of Chromium and Vanadium on Biomarkers of Carbohydrate and Lipid Metabolism in Workers Exposed to Coal Fly Ash

Lulzim Zeneli¹, Majlinda Daci-Ajvazi², Ankica Sekovanić³, Jasna Jurasović³ and Demush Bajraktari^{4,*}

- ¹ Faculty of Education, University Fehmi Agani, 50000 Gjakova, Kosovo
- ² Faculty of Mathematics and Natural Sciences, University of Prishtina, 10000 Prishtina, Kosovo
- ³ Institute for Medical Research and Occupational Health, 10000 Zagreb, Croatia

⁴ Faculty of Pharmacy, UBT Higher Education Institution, 10000 Prishtina, Kosovo

* Correspondence: demush.bajraktari@ubt-uni.net

Abstract: Chromium (Cr) and vanadium (V) are micronutrients playing a role in carbohydrate and lipid metabolism but can be toxic at high concentrations, especially in specific forms. The study documents the effect of Cr and V concentrations on glucose and lipid metabolism in workers exposed to coal fly ash. We quantified selected metals (Cr, V) in the blood and serum of workers from a thermal power plant in Kosovo and compared them with the reference biological values. We determined fasting serum glucose and lipid profiles using a biochemical analyzer Synchron CX7 (Beckman Coulter). We quantified blood and serum Cr and V by inductively coupled plasma mass spectrometry. We also evaluated the association between carbohydrate and lipid metabolism biomarkers (glucose, cholesterol, and triglycerides) and co-exposure to coal fly ash. Power plant workers had significantly higher blood Cr and V levels (p < 0.0001) and significantly lower serum Cr and V levels (p < 0.0001) than the controls. We also found statistically significant (p < 0.0001) correlations between high blood Cr levels and low glucose/blood Cr ratios as well as between high serum Cr levels and low glucose/serum Cr ratios. Finally, in power plant workers, high blood V levels significantly correlated with low triglycerides/blood V and cholesterol/blood V ratios (p < 0.0001), while high serum V levels correlated with low cholesterol/serum V ratios (p = 0.005). Based on these findings, we concluded that the glucose/Cr, triglycerides/V and cholesterol/V ratios should be considered when evaluating carbohydrate and lipid metabolism disorders in occupationally-exposed workers.

Keywords: chromium; vanadium; carbohydrate; lipid; coal fly ash; correlation

1. Introduction

Human biomonitoring has its roots in the analysis of biological samples and can considerably help reduce uncertainty in health risk assessment [1–3]. Biological monitoring is defined as the repeated, controlled measurement of chemical or biochemical markers in biological samples or other accessible samples from subjects exposed to chemical, physical, or biological risk factors in the workplace and/or the general environment [4–6]. Cr and V are next to each other on the periodic table and share many characteristics [7,8]. Both of these elements, naturally present in our environment and predominantly excreted by the kidneys, are associated with normal human health and the pathogenesis of several diseases [9,10].

Cr and V metals do not occur naturally; instead, these elements are found in different valence states (Cr from +1 to +6 and V from -1 to +5) [11–13]. As transition metals, their chemistry is complex. They can participate in redox processes and produce free radicals [14], thus inducing lipid peroxidation both in vitro [15] and in vivo [16,17] and disturbing the antioxidative balance in the organism. Humans can be exposed to Cr and V through the air, but most of the exposure stems from food and water [11,12]. The hexavalent chromium



Citation: Zeneli, L.; Daci-Ajvazi, M.; Sekovanić, A.; Jurasović, J.; Bajraktari, D. The Effects of Chromium and Vanadium on Biomarkers of Carbohydrate and Lipid Metabolism in Workers Exposed to Coal Fly Ash. J. Xenobiot. 2022, 12, 307–316. https://doi.org/10.3390/jox 12040021

Academic Editor: Ana Catarina Sousa

Received: 6 September 2022 Accepted: 8 October 2022 Published: 19 October 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). form (Cr^{6+}) easily enters cells through facilitated uptake, which is more efficient than the simple diffusion undergone by the trivalent form (Cr³⁺). It travels to and accumulates in the erythrocytes, but the highest Cr concentrations are found in the kidneys and liver [18,19]. Vanadium (V^{5+}) readily enters cells via phosphate and sulfate ion channels; it also interferes with phosphate-containing enzymes [20], can activate several genes, and participates in the inflammatory response. The initial V concentrations are found in the kidneys, liver, and lungs, and in the long-term, it is stored in the bones and muscles [21]. The major route of elimination of absorbed Cr and V is urine, whereas unabsorbed Cr and V are excreted through the feces [22]. Vanadium has many systemic effects including gastrointestinal, respiratory, hematological, immunological, and cardiovascular effects, whereas trivalent and hexavalent Cr cause gastrointestinal, immunological, hematological (including anemia), reproductive, developmental, and other serious effects [12,23,24]. Vanadium compounds may prevent chemical carcinogenesis and act as potential anti-metastatic agents, and the estrogenic role of V and its possible therapeutic application in osteoporosis have also been described [25–27]. Supplementation with Cr decreases the plasma thiobarbituric acid reactive substance levels and minimizes the oxidative stress increase in patients with type 2 diabetes mellitus [28–31].

Several studies have shown that chromium and vanadium as transition metals have been directly implicated in the regulation of glucose and lipid metabolism, whereas increasing or decreasing levels of lipids and glucoses cause various health effects in the human body, which are called disorders. This study documents the effects of chronic exposure to anthropogenic pollution on the Cr and V levels and the relationship with lipid and carbohydrate metabolism biomarkers (glucose (Glu), cholesterol (Chol), and triglycerides (TG)) in the blood of power plant workers.

2. Materials and Methods

2.1. Study Population

The main characteristics of the study population have previously been reported [32]. Blood samples from 100 male workers participating in the study were available for metal analysis. The age of the subjects ranged from 30 to 65 years (mean 51 years), while the occupational exposure duration ranged from 4 to 43 years (mean 21 years). Among the 100 subjects, 70 were male workers employed at the thermal power plant (ThPP) in "Kosova" (Obiliq, Kosovo) who had been exposed to traces of heavy metals, and 30 were healthy inhabitants from the rural municipality of Dragash, Kosovo (environment without pollutants found at the power plant). The study protocol was carried out in accordance with the regulations in force of the University Fehmi Agani, Gjakova, Kosovo, and prior written consent was obtained from each participant in the research. Each subject completed a questionnaire regarding age, occupational exposure duration, and medical history. The candidates declared that they did not have any known acute or chronic diseases and in addition, each subject was informed about the purposes of the study. The exclusion criteria from the study were as follows: alcoholism, gender, food, and lifestyle of the study group.

2.2. Blood Sampling and Trace Elements Analysis

We sampled the venous blood of all subjects between 8:00 am and 9:00 am. Next, we placed 15 mL of blood in a K₂EDTA-containing tube (BD Vacutainer K₃E) to quantify the blood Cr (BCr) and blood V (BV) levels. We also placed the same amount of blood in an anticoagulant-free tube (BD Vacutainer Trace Element Serum, Heidelberg, Germany) to quantify the serum Cr (SCr) and serum V (SV) levels and into an anticoagulant-free tube (BD Vacutainer, SARSTEDT—Nümbrecht, Germany) for the biochemical parameter analysis. After allowing for 60 min of spontaneous blood clotting, we separated the serum from the blood cells via centrifugation at 3000 rpm/h for 10 min, decanted again, and stored in a metal-free polypropylene tube at -20 °C until SCr and SV quantification. We took special care to avoid any contamination with metals during the blood sampling, storage, and analyses. All chemicals used were of analytical grade for spectroscopy (Merck,

Darmstadt, Germany). We quantified trace elements using inductively coupled plasma mass spectrometry [33]. We used a biochemical analyzer Synchron CX7 (Beckman Coulter Company, Indianapolis, IN, USA), with reagents obtained from Beckman Instrumental, Inc. (Galway, Ireland) to determine the biochemical parameters such as fasting serum glucose and lipid profiles.

2.3. Statistical Methods

Because of the skewed distribution of most of the measured parameters, we presented the results within groups as the median and range, while we calculated the difference between groups using the *t*-test (t, p). We employed Pearson's correlation (r, p) to explore the relationships between each of the measured parameters. The hypothesis testing in the statistical analysis was based on a 0.05 significance level.

3. Results

The blood concentrations of metals and biological markers in the individuals studied in the present work are summarized in Table 1.

Table 1. Characteristics of the study groups based on the exposure duration of and concentrations of Cr and V in the blood and serum samples and biological markers.

Characteristic	ThPP Workers	Control Subjects	Т	р
Total Number of Individuals	70	30	-	-
Occupational Exposure Duration (years)	21 (4–43)	0	-	-
Age (years)	51 (31-64)	42 (30-65)	0.67	< 0.5
BMI	27.4 ± 0.17	26.9 ± 3.8	4.5	< 0.0001
BCr (µg/L)	1.0 ± 0.3	0.73 ± 0.12	7.3	< 0.0001
$BV (\mu g/L)$	0.7 ± 0.16	0.47 ± 3.6	6.6	< 0.0001
$SCr(\mu g/L)$	0.87 ± 0.1	1.07 ± 0.07	-9.3	< 0.0001
$SV(\mu g/L)$	1.38 ± 0.1	1.70 ± 0.07	-14	< 0.0001
Glu (mmol/L)	5.62 ± 1.5	5.88 ± 1.4	-1.0	0.283
TG (µg/L)	1.65 ± 0.4	1.84 ± 0.5	-3.2	< 0.002
Chol ($\mu g/L$)	5.63 ± 1.1	6.4 ± 1.0	-1.9	< 0.05

Workers exposed to coal fly ash had significantly higher BCr and BV levels (p < 0.0001) and significantly lower SCr and SV levels (p < 0.0001) than the control subjects. The BCr and BV concentrations were 1.4 and 1.5 times higher in the study group than in the control group, respectively. Moreover, the SCr and SV concentrations were both 1.2 times lower in the study group than in the control group. Table 2 presents the biological markers to trace elements ratios.

The power plant workers had significantly lower Glu/BCr, Chol/BCr, TG/BCr, Glu/BV, Chol/BV, and TG/BV ratios ($p \le 0.0001$) and significantly higher Glu/SCr and Glu/SV ratios ($p \le 0.005$) than the control subjects. The Pearson's correlation analysis (Table 3) showed that high BCr levels were significantly (p < 0.001) associated with low Glu/BCr ratios (r = -0.713). We observed a similar relationship with the SCr levels and the Glu/SCr (r = -0.678), Chol/SCr (r = -0.955), and TG/SCr (r = -0.358) ratios. Meanwhile, we found no significant correlation between the BCr levels and the Chol/BCr or TG/BCr ratios.

Table 2. A comparison of the biological markers glucose, cholesterol, and triglycerides to the trace elements ratios of the study group (ThPP workers, n = 70) and the control group (n = 30).

	ThPP Workers	Control Subjects	Т	р
Glu/BCr	5.62	8.05	-6.8	< 0.0001
Chol/BCr	5.63	8.76	-7.9	< 0.0001

	ThPP Workers	Control Subjects	Т	р
TG/BCr	1.56	2.52	-5.7	< 0.0001
Glu/SCr	6.45	5.49	3.4	< 0.005
Chol/SCr	6.47	5.98	1.64	0.115
TG/SCr	1.79	1.71	1.13	0.275
Glu/BV	8.08	13.36	-9.1	< 0.0001
Chol/BV	8.04	13.61	-9.3	< 0.0001
TG/BV	2.22	3.91	-5.6	< 0.0001
Glu/SV	4.19	3.45	-3.8	0.0002
Chol/SV	4.08	3.76	-1.65	0.105
TG/SV	1.13	1.02	-1.06	0.226

Table 2. Cont.

Table 3. The Pearson's correlation coefficients and significance levels (r, p) for the relationships between the Cr and V concentrations and biological markers (glucose, cholesterol and triglycerides) to trace elements ratios in whole blood and serum.

	B ¹ Cr (μg/L)	S ² Cr (µg/L)	BV (μg/L)	SV (μg/L)
Glu/BCr	-0.713 ^a			
Chol/BCr				
TG/BCr				
Glu/SCr		-0.678 ^a		
Chol/SCr		-0.955 ^a		
TG/SCr		-0.358 ^c		
Glu/BV			-0.895 ^a	
Chol/BV			-0.825 ^a	
TG/BV			-0.672^{b}	
Glu/SV				$-0.288 ^{\text{d}}$
Chol/SV				-0.203 ^d
TG/SV				

^a $p \le 0.00001$; ^b $p \le 0.0001$; ^c $p \le 0.001$; ^d $p \le 0.05$; ¹ B—blood; ² S—serum.

Finally, in ThPP workers, high BV concentrations were significantly correlated (p < 0.05) with the low Glu/BV (r = -0.895), Chol/BV (r = -0.825), and TG/BV (r = -0.672) ratios. Similarly, high SV concentrations (Figure 1) were associated with the low Glu/SV (r = -0.288) and Chol/SV (r = -0.203) ratios. In contrast, we found no significant correlation between the SCr levels and TG/SV ratios.



Figure 1. The correlation between the SCr levels and the Glu/SCr ratio in ThPP workers (r = -0.67, p < 0.0001).

4. Discussion

Cr and V are micronutrients that play a significant role in health maintenance [34], but excessive concentrations can lead to the development of diseases [35,36]. Vanadium pentoxide (V_2O_5), as a component of particles derived from the combustion of different types of fuel, and Cr⁶⁺, generated in the process of burning coal at high temperature in thermal power plants as the dominant form in fly ash, are a source of occupational exposure in humans [37-41]. In the present study, these forms of the two elements were analyzed, and we assessed the effect of V and Cr co-exposure on the carbohydrate and lipid metabolism biomarkers. Biomarkers are associated with an alteration in the cellular or biochemical components, processes, structures, or functions. Moreover, biological indicators can be any quantifiable substance, structure, or process tissues or fluids that reflect the health or the incidence or biological behavior of a disease [42]. For this purpose, we calculated the ratios of Glu, Chol, and TG to the Cr and V blood concentrations. Trace elements, especially Cr and V, play a significant role in the metabolism of lipids and carbohydrates [43,44]. Table 1 shows the relevant parameters of the investigation group. The investigation group had been significantly more exposed to nonessential elements than the control group, as indicated by the higher BCr and BV concentrations (p < 0.0001) and significantly lower SCr and SV concentration (p < 0.0001).

The higher BCr levels of the investigation group could be due to the propensity of erythrocytes to bind and uptake Cr(VI) [45]. Cr(VI) enters erythrocytes through a sulfate ion channel. Inside the cell, it is rapidly reduced into the reactive intermediates Cr(V) and Cr(IV) and binds to the beta chain of human hemoglobin [45–47] and other ligands such as proteins and glutathione. Furthermore, the lower SCr levels of the investigation group can be explained by the fact that ascorbate and glutathione can reduce inhaled Cr(VI) to Cr(III) in the epithelial lining fluid of the lungs [48,49]. The higher BV levels in the study group compared with the control group (Table 1) may result from vanadium emissions through coal fly ash. For most organisms including mammals, V is not known to have any essential biological function. However, many biomolecules contain vanadium.

According to occupational exposure studies, human experimental studies, and laboratory animal studies, the respiratory tract is one of the primary targets of V toxicity after exposure by inhalation. Experimental studies indicate that, inside erythrocytes, the vanadium (V^{5+}) is mainly bound to hemoglobin [50–52], but probably also to other intracellular bioligands [53,54]. The vanadium biomedical importance has been proven by numerous studies and is mainly based on its interaction with proteins, along with enzymatic systems and cellar components, which can affect the synthesis of lipids and lipoproteins or interact with catabolism [55].

In contrast, the lower SV levels observed in the study group (Table 1) compared with the control group may result from an interconversion between vanadium species (mostly V^{4+}/V^{5+} and to a lesser degree V^{+3}) inside the biomolecules [56–58].

We also evaluated the classification (ThPP workers vs. control group) performance of the Glu, Chol, TR, V, and Cr concentrations and ratios and performed a discriminant analysis (Table 1). The Glu/BCr, Chol/BCr, TG/BCr, Glu/BV, Chol/BV, and TG/BV ratios had smaller *p*-values ($p \le 0.0001$) than Glu, Chol, TR, V, or Cr concentrations, indicating that the biological markers to trace elements ratio had superior discriminating power over the Glu, Chol, TR, V, or Cr concentrations alone. The fact that the study group had significantly lower Glu/BCr, Chol/BCr, TG/BCr, Glu/BV, Chol/BV, and TG/BV ratios than the control group could indicate a higher risk of hyperglycemia and lipid metabolism disorders.

Recent studies have demonstrated that Cr metabolism is affected by several factors including lifestyle (stress, diet, activities) and diabetes [36,59,60]. Additionally, chromium is well-known to play substantial roles in the lipid and metabolism of carbohydrates and affects the etiology of diabetes, obesity, and cardiovascular diseases [61].

Our results show that a highly significant correlation exists between the BCr levels and Glu/BCr ratios (r = -0.713) as well as between the SCr levels and the Glu/SCr (r = -0.678), Chol/SCr (r = -0.955), and TG/SCr (r = -0.358) ratios. A correlation such as

that between BCr and Glu/BCr (Figure 2) and SCr and Glu/SCr (Figure 1) could result from predispositions to diabetes, as the ability to convert inorganic chromium into a useable organic form (organic chromium) that activates insulin is reduced in people predisposed to diabetes [62–64]. Moreover, the study group displayed lower SCr levels than the control group (Table 1), which supports this hypothesis. In addition, the correlation between the SCr levels and the Chol/SCr and TG/SCr ratios may result from Cr metabolism and thus be a predictor of cardiovascular risk.



Figure 2. The correlation between the BCr levels and the Glu/BCr ratio in the ThPP workers (r = -0.70, p < 0.0001).

In the investigation group, the BV concentration was significantly correlated (p < 0.05) with the Glu/BV (r = -0.895), Chol/BV (r = -0.825), and TG/BV (r = -0.672) ratios, while the SV concentration (Figure 1) was correlated with the Glu/SV (r = -0.288) and Chol/SV (r = -0.203) ratios. The role of V in carbohydrate hemostasis can explain the correlation between BV and Glu/BV and SV and Glu/SV. Recent studies have shown that administering V to diabetic animals or humans decreased hepatic glucose production [65–67]. Additionally, the direct biochemical control of glucose homeostasis during the V treatments is associated with the enhancement of glycolysis and glucose oxidation [68,69]. In contrast, the correlations between the BV and SV concentrations and the Chol/BV and Chol/SV (Figures 3 and 4) and TG/BV ratios could be explained by the role of V in the process of reducing the total and free cholesterol levels, or through the inhibition of the cholesterol biosynthesis steps [70].

Glu/Cr and Tr/V and Chol/V ratios would be more valuable than the biochemical parameters (Glu, TR, Chol) levels alone for predicting the early stage of disorders of carbohydrate and lipid metabolism risk. Future studies should explore the underlying mechanisms of this relationship, compare genders (male and female), assess the impacts of alcohol consumption, and monitor the long-term impacts of exposure.



Figure 3. The correlation between the BV levels and the Chol/BV ratio (r = -0.78, *p* < 0.0001) in the ThPP workers.



Figure 4. The correlation between the SV levels and the Chol/SV ratio (r = -0.31, p = 0.0048) in the ThPP workers.

5. Conclusions

Our results indicate that the Glu/Cr, TG/V, and Chol/V ratios should be considered when evaluating carbohydrate and lipid metabolism disorders in occupationally-exposed workers. Furthermore, this study demonstrated that coal fly ash significantly altered the Cr and V blood levels. Cr affects the biochemical parameters, leading to various diseases. Nevertheless, the biochemistry of chromium and vanadium requires more exploration. The Glu/Cr, TG/V, and Chol/V ratios should be considered in the evaluation of disorders of carbohydrate and lipid metabolism expressly in the environments that are affected by several factors including environmental pollutions, stress, diet, etc.

Author Contributions: Conceptualization, L.Z. and M.D.-A.; Methodology, L.Z., M.D.-A. and D.B.; Software, L.Z.; Validation, L.Z., A.S. and J.J.; Formal analysis, L.Z., A.S. and J.J.; Investigation, L.Z., A.S. and J.J.; Resources, L.Z. and M.D.-A.; Data curation, L.Z.; Writing—original draft preparation, L.Z. and D.B.; Writing—review and editing, L.Z. and D.B.; Visualization, D.B.; Supervision, L.Z.; Project administration, L.Z.; Funding acquisition, L.Z. and M.D.-A. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki. The ethical review for this study was based on the standard procedures supported by University Regulation 05//1896 for Ethical Code dated 9 December 2020, and Regulation 05/1092 for Research—Scientific activity dated 30 June 2021.

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declares no potential conflict of interest with respect to the research, authorship, and/or publication of this article.

References

- 1. Gurusankar, R.; Yenugadhati, N.; Krishnan, K.; Hays, S.; Haines, D.; Zidek, A.; Kuchta, S.; Kinniburgh, D.; Gabos, S.; Mattison, D.; et al. The role of human biological monitoring in health risk assessment. *Int. J. Risk Assess. Manag.* **2017**, *20*, 136–197. [CrossRef]
- 2. Sexton, K.; Needham, L.; Pirkle, J. Human Biomonitoring of Environmental Chemicals. Am. Sci. 2004, 92, 38–45. [CrossRef]
- 3. Zeneli, L.; Daci, N.; Pacarizi, H.; Daci-Ajvazi, M. Impact of Environmental Pollution on Human Health of the Population Which Lives Nearby Kosovo Thermopower Plants. *Indoor Built Environ.* **2011**, *20*, 479–482. [CrossRef]
- 4. Manno, M.; Viau, C.; Cocker, J.; Colosio, C.; Lowry, L.; Mutti, A.; Nordberg, M.; Wang, S. Biomonitoring for occupational health risk assessment (BOHRA). *Toxicol. Lett.* **2010**, *192*, 3–16. [CrossRef] [PubMed]
- 5. Zeneli, L.; Sekovanić, A.; Ajvazi, M.; Kurti, L.; Daci, N. Alterations in antioxidant defense system of workers chronically exposed to arsenic, cadmium and mercury from coal flying ash. *Environ. Geochem. Health* **2016**, *38*, 65–72. [CrossRef] [PubMed]
- 6. Ladeira, C.; Viegas, S. Human Biomonitoring—An overview on biomarkers and their application in Occupational and Environmental Health. *Biomonitoring* **2016**, *3*, 15–24. [CrossRef]
- Filler, G.; Kobrzynski, M.; Sidhu, H.K.; Belostotsky, V.; Huang, S.-H.S.; McIntyre, C.; Yang, L. A cross-sectional study measuring vanadium and chromium levels in paediatric patients with CKD. *BMJ Open* 2017, 7, e014821. [CrossRef] [PubMed]
- 8. Lagerkvist, B.; Nordberg, G.F.; Vouk, V.V. Handbook on the Toxicology of Metals; Elsevier: Amsterdam, The Netherlands, 1986.
- 9. Hosokawa, S.; Yamaguchi, O.; Yoshida, O. Vanadium transfer during haemodialysis. *Int. Urol. Nephrol.* **1991**, 23, 407–409. [CrossRef] [PubMed]
- 10. Ochi, A.; Ishimura, E.; Tsujimoto, Y.; Kakiya, R.; Tabata, T.; Mori, K.; Shoji, T.; Yasuda, H.; Nishizawa, Y.; Inaba, M. Trace elements in the hair of hemodialysis patients. *Biol. Trace Elem. Res.* **2011**, *143*, 825–834. [CrossRef]
- 11. Services USDoHaH. Toxicological Profile for Chromium. Available online: https://www.atsdr.cdc.gov/ToxProfiles/tp7.pdf (accessed on 10 July 2020).
- Services USDoHaH. Toxicological Profile for Vanadium. Available online: http://www.atsdr.cdc.gov/ToxProfiles/tp58.pdf (accessed on 10 July 2020).
- 13. Barceloux, D.G.; Barceloux, D. Vanadium. J. Toxicol. Clin. Toxicol. 1999, 37, 265–278. [CrossRef] [PubMed]
- 14. Byczkowski, J.Z.; Kulkarni, A.P. Oxidative stress and pro-oxidantbiological effects of vanadium. In *Vanadium in the Environment*. *Part 1: Chemistry and Biochemistry*; Nriagu, J.O., Ed.; John Wiley & Sons: Hoboken, NJ, USA, 1998; pp. 235–263.
- 15. Donaldson, J.; LaBella, F. Prooxidant properties of vanadate in vitro on catecholamines and on lipid peroxidation by mouse and rat tissues. *J. Toxicol. Environ. Health* **1983**, *12*, 119–126. [CrossRef] [PubMed]
- Soares, S.S.; Martins, H.; Duarte, R.O.; Moura, J.J.G.; Coucelo, J.; Gutiérrez-Merino, C.; Aureliano, M. Vanadium distribution, lipid peroxidation and oxidative stress markers upon decavanadate in vivo administration. *J. Inorg. Biochem.* 2007, 101, 80–88. [CrossRef] [PubMed]
- 17. Huang, Y.L.; Chen, C.Y.; Sheu, J.Y.; Chuang, I.C.; Pan, J.H.; Lin, T.H. Lipid peroxidation in workers exposed to hexavalent chromium. *J. Toxicol. Environ. Health* **1999**, *56*, 235–247. [CrossRef]
- 18. O'Flaherty, E.J. A Physiologically Based Model of Chromium Kinetics in the Rat. *Toxicol. Appl. Pharmacol.* **1996**, *138*, 54–64. [CrossRef] [PubMed]
- 19. Wu, F.Y.; Wu, W.Y.; Kuo, H.W.; Liu, C.-S.; Wang, R.-Y.; Lai, J.-S. Effect of genotoxic exposure to chromium among electroplating workers in Taiwan. *Sci. Total Environ.* **2001**, 279, 21–28. [CrossRef]
- 20. Rehder, D. Biological and medicinal aspects of vanadium. Inorg. Chem. Commun. 2003, 6, 604-617. [CrossRef]
- Crebelli, R.; Leopardi, P. Long-term risks of metal contaminants in drinking water: A critical appraisal of guideline values for arsenic and vanadium. Ann. Ist Super Sanita 2012, 48, 354–361. [CrossRef] [PubMed]

- 22. Ryan, L.K. Vanadium Pentoxide Effects on Lungs. In *Encyclopedia of Metalloproteins*; Kretsinger, R.H., Uversky, V.N., Permyakov, E.A., Eds.; Springer: Berlin/Heidelberg, Germany, 2013; pp. 2293–2297.
- Ray, R.R. Adverse hematological effects of hexavalent chromium: An overview. *Interdiscip. Toxicol.* 2016, 9, 55–65. [CrossRef] [PubMed]
- Jacobs, L.; Buczynska, A.; Walgraeve, C.; Delcloo, A.; Potgieter-Vermaak, S.; Van Grieken, R.; Demeestere, K.; Dewulf, J.; Van Langenhove, H.; De Backer, H.; et al. Acute changes in pulse pressure in relation to constituents of particulate air pollution in elderly persons. *Environ. Res.* 2012, 117, 60–67. [CrossRef]
- Facchini, D.M.; Yuen, V.G.; Battell, M.L.; McNeill, J.; Grynpas, M. The effects of vanadium treatment on bone in diabetic and non-diabetic rats. *Bone* 2006, *38*, 368–377. [CrossRef] [PubMed]
- Beam, H.A.; Parsons, J.R.; Lin, S.S. The effects of blood glucose control upon fracture healing in the BB Wistar rat with diabetes mellitus. J. Orthop. Res. 2002, 20, 1210–1216. [CrossRef]
- 27. Rehder, D. Perspectives for vanadium in health issues. Future Med. Chem. 2016, 8, 325–338. [CrossRef] [PubMed]
- 28. Lai, M.H. Antioxidant effects and insulin resistance improvement of chromium combined with vitamin C and e supplementation for type 2 diabetes mellitus. *J. Clin. Biochem. Nutr.* **2008**, *43*, 191–198. [CrossRef]
- 29. Pieper, G.M. Peroxidative stress in diabetic blood vessels. *Diabetes* **1995**, *44*, 884–889. [CrossRef] [PubMed]
- Duman, B.S. Thiols, malonaldehyde and total antioxidant status in the Turkish patients with type 2 diabetes mellitus. *Tohoku J. Exp. Med.* 2003, 201, 147–155. [CrossRef] [PubMed]
- 31. Oberley, L.W. Free radicals and diabetes. Free Radic. Biol. Med. 1988, 5, 113–124. [CrossRef]
- 32. Zeneli, L.; Sekovanić, A.; Daci, N. Chronic exposure to aluminum, nickel, thallium and uranium and their relationship with essential elements in human whole blood and blood serum. *J. Environ. Sci. Health A Tox. Hazard. Subst. Environ. Eng.* **2015**, *50*, 540–546.
- Živković, T.; Tariba, B.; Pizent, A. Multielement analysis of human seminal plasma by octopole reaction cell ICP-MS. J. Anal. At. Spectrom. 2014, 9, 2114–2126. [CrossRef]
- Di Bona, K.R.; Love, S.; Rhodes, N.R.; McAdory, D.A.; Sinha, S.H.; Kern, N.; Kent, J.; Strickland, J.; Wilson, A.; Beaird, J.; et al. Chromium is not an essential trace element for mammals: Effects of a low-chromium diet. *J. Biol. Inorg. Chem.* 2011, 16, 381–390. [CrossRef] [PubMed]
- 35. Vincent, J.B. Chromium: Celebrating 50 years as an essential element? Dalton Trans. 2010, 39, 3787–3794. [CrossRef] [PubMed]
- 36. Anderson, R.A. Chromium as an essential nutrient for humans. Regul. Toxicol. Pharmacol. 1997, 26, 35–41. [CrossRef] [PubMed]
- 37. Sørensen, M.; Schins, R.P.F.; Hertel, O.; Loft, S. Transition metals in personal samples of PM2.5 and oxidative stress in human volunteers. *Cancer Epidemiol. Biomark. Prev.* 2005, *14*, 1340–1343. [CrossRef] [PubMed]
- IARC. Cobalt in Hard Metals and Cobalt Sulfate, Gallium Arsenide, Indium Phosphide and Vanadium Pentoxide; International Agency for Research on Cancer (IARC): Lyon, France, 2006.
- 39. Mukherjee, A.B. Chromium in the environment of Finland. Sci. Total Environ. 1998, 217, 9–19. [CrossRef]
- Ellis, A.S.; Johnson, T.M.; Bullen, T.D. Chromium isotopes and the fate of hexavalent chromium in the environment. *Science* 2002, 295, 2060–2062. [CrossRef] [PubMed]
- 41. Rowbotham, A.L.; Levy, L.S.; Shuker, L.K. Chromium in the environment: An evaluation of exposure of the UK general population and possible adverse health effects. *J. Toxicol. Environ. Health B Crit. Rev.* **2000**, *3*, 145–178. [PubMed]
- 42. Strimbu, K.; Tavel, J.A. What are biomarkers? Curr. Opin. HIV AIDS 2010, 5, 463–466. [CrossRef]
- 43. Davis, C.M.; Vincent, J.B. Chromium in carbohydrate and lipid metabolism. JBIC J. Biol. Inorga. Chem. 1997, 2, 675–679. [CrossRef]
- 44. Sidorova, Y.S.; Skalnaya, M.G.; Tinkov, A.A.; Mazo, V.K. The effect of vanadium compounds on carbohydrate and lipid metabolism disorders. *Probl. Endokrinol.* **2019**, *65*, 184–190. [CrossRef] [PubMed]
- 45. Kerger, B.D.; Paustenbach, D.J.; Corbett, G.E.; Finley, B.L. Absorption and elimination of trivalent and hexavalent chromium in humans following ingestion of a bolus dose in drinking water. *Toxicol. Appl. Pharmacol.* **1996**, *141*, 145–158. [CrossRef]
- 46. Wetterhahn, K.E.; Hamilton, J.W.; Aiyar, J.; Borges, K.M.; Floyd, R. Mechanisms of Chromium(VI) carcinogenesis—Reactive intermediates and effect on gene-expression. *Biol. Trace Elem. Res.* **1989**, *21*, 405–411. [CrossRef] [PubMed]
- Reynolds, M.; Zhitkovich, A. Cellular vitamin C increases chromate toxicity via a death program requiring mismatch repair but not p53. *Carcinogenesis* 2007, 28, 1613–1620. [CrossRef] [PubMed]
- Petrilli, F.L.; Rossi, G.A.; Camoirano, A.; Romano, M.; Serra, D.; Bennicelli, C.; De Flora, A.; De Flora, S. Metabolic reduction of chromium by alveolar macrophages and its relationships to cigarette smoke. *J. Clin. Investig.* 1986, 77, 1917–1924. [CrossRef] [PubMed]
- Suzuki, Y.; Fukuda, K. Reduction of hexavalent chromium by ascorbic acid and glutathione with special reference to the rat lung. *Arch. Toxicol.* 1990, 64, 169–176. [CrossRef] [PubMed]
- 50. Sanna, D.; Serra, M.; Micera, G.; Garribba, E. Interaction of antidiabetic vanadium compounds with hemoglobin and red blood cells and their distribution between plasma and erythrocytes. *Inorg. Chem.* **2014**, *53*, 1449–1464. [CrossRef] [PubMed]
- De Cremer, K.; Van Hulle, M.; Chéry, C.; Cornelis, R.; Strijckmans, K.; Dams, R.; Lameire, N.; Vanholder, R. Fractionation of vanadium complexes in serum, packed cells and tissues of Wistar rats by means of gel filtration and anion-exchange chromatography. J. Biol. Inorg. Chem. 2002, 7, 884–890. [CrossRef]
- Delgado, T.C.; Tomaz, A.I.; Correia, I.; Pessoa, J.C.; Jones, J.G.; Geraldes, C.F.; Castro, M.M.C. Uptake and metabolic effects of insulin mimetic oxovanadium compounds in human erythrocytes. J. Inorg. Biochem. 2005, 99, 2328–2339. [CrossRef]

- 53. Cakir, Y.; Yildiz, D. Efflux of glutathione and glutathione complexes from human erythrocytes in response to vanadate. *Blood Cells Mol. Dis.* **2013**, *50*, 1–7. [CrossRef]
- 54. Sanna, D.; Micera, G.; Garribba, E. On the transport of vanadium in blood serum. Inorg. Chem. 2009, 48, 5747–5757. [CrossRef]
- 55. Zhang, Y.; Zhang, Q.; Feng, C.; Ren, X.; Li, H.; He, K.; Wang, F.; Zhou, D.; Lan, Y. Influence of vanadium on serum lipid and lipoprotein profiles: A population-based study among vanadium exposed workers. *Lipids Health Dis.* **2014**, *13*, 39. [CrossRef]
- 56. Yang, X.G.; Yuan, L.; Wang, K.; Crans, D. The permeability and cytotoxicity of insulin-mimetic vanadium compounds. *Pharm. Res.* **2004**, *21*, 1026–1033. [CrossRef]
- Cohen, M.D.; Sisco, M.; Prophete, C.; Chen, L.C.; Zelikoff, J.T.; Ghio, A.J.; Stonehuerner, J.D.; Smee, J.; Holder, A.; Crans, D. Pulmonary immunotoxic potentials of metals are governed by select physicochemical properties: Vanadium agents. *J. Immunotoxicol.* 2007, *4*, 49–60. [CrossRef] [PubMed]
- Treviño, S.; Díaz, A.; Sánchez-Lara, E.; Sanchez-Gaytan, B.L.; Perez-Aguilar, J.M.; González-Vergara, E. Vanadium in Biological Action: Chemical, Pharmacological Aspects, and Metabolic Implications in Diabetes Mellitus. *Biol. Trace Elem. Res.* 2019, 188, 68–98. [CrossRef] [PubMed]
- 59. Wang, Z.Q.; Cefalu, W.T. Current concepts about chromium supplementation in type 2 diabetes and insulin resistance. *Curr. Diab. Rep.* **2010**, *10*, 145–151. [CrossRef] [PubMed]
- 60. Balk, E.M.; Tatsioni, A.; Lichtenstein, A.H.; Lau, J.; Pittas, A.G. Effect of chromium supplementation on glucose metabolism and lipids: A systematic review of randomized controlled trials. *Diabetes Care* **2007**, *30*, 2154–2163. [CrossRef] [PubMed]
- 61. Ngala, R.A.; Awe, M.A.; Nsiah, P. Chromium is well known to play substantial roles in lipid and metabolism if carbohydrates and it affects etiology of diabetes, obesity and cardiovascular diseases. *PLoS ONE* **2018**, *13*, e0197977. [CrossRef]
- 62. Anderson, R.A.; Cheng, N.; Bryden, N.A.; Polansky, M.M.; Cheng, N.; Chi, J.; Feng, J. Elevated intakes of supplemental chromium improve glucose and insulin variables in individuals with type 2 diabetes. *Diabetes* **1997**, *46*, 1786–1791. [CrossRef]
- 63. Lai, M.H.; Chen, Y.Y.; Cheng, H.H. Chromium yeast supplementation improves fasting plasma glucose and LDL-cholesterol in streptozotocin-induced diabetic rats. *Int. J. Vitam. Nutr. Res.* 2006, *76*, 391–397. [CrossRef]
- Yang, J.; Xu, Y.; Qian, K.; Zhang, W.; Wu, D.; Wang, C. Effects of chromium-enriched Bacillus subtilis KT260179 supplementation on growth performance, caecal microbiology, tissue chromium level, insulin receptor expression and plasma biochemical profile of mice under heat stress. *Br. J. Nutr.* 2016, 115, 774–781. [CrossRef]
- Cong, X.-Q.; Piao, M.-H.; Li, Y.; Xie, L.; Liu, Y. Bis(maltolato)oxovanadium(IV) (BMOV) Attenuates Apoptosis in High Glucose-Treated Cardiac Cells and Diabetic Rat Hearts by Regulating the Unfolded Protein Responses (UPRs). *Biol. Trace Element Res.* 2016, 173, 390–398. [CrossRef]
- Bâlici, Ş.; Wankeu-Nya, M.; Rusu, D.; Nicula, G.Z.; Rusu, M.; Florea, A.; Matei, H. Ultrastructural analysis of in vivo hypoglycemiant effect of two polyoxometalates in rats with streptozotocin-induced diabetes. *Microsc. Microanal.* 2015, 21, 1236–1248. [CrossRef]
- 67. Blondel, O.; Simon, J.; Chevalier, B.; Portha, B. Impaired insulin action but normal insulin receptor activity in diabetic rat liver: Effect of vanadate. *Am. J. Phys.* **1990**, 258, 459–467. [CrossRef] [PubMed]
- 68. Korbecki, J.; Baranowska-Bosiacka, I.; Gutowska, I.; Chlubek, D. Biochemical and medical importance of vanadium compounds. *Acta Biochim. Pol.* **2012**, 59, 195–200. [CrossRef] [PubMed]
- 69. Xie, M.; Chen, D.; Zhang, F.; Willsky, G.R.; Crans, D.C.; Ding, W. Effects of vanadium (III, IV, V)-chlorodipicolinate on glycolysis and antioxidant status in the liver of STZ-induced diabetic rats. *J. Inorg. Biochem.* **2014**, *136*, 47–56. [CrossRef]
- Francik, R.; Kryczyk-Kozioł, J.; Francik, S.; Gryboś, R.; Krośniak, M. Bis(4,4'-dimethyl-2,2'-bipyridine)oxidovanadium(IV) sulfate dehydrate: Potential candidate for controlling lipid metabolism? *BioMed Res. Int.* 2017, 2017, 6950516. [CrossRef] [PubMed]