



Case Report

The First Case of Haemophagocytic Lymphohistiocytosis Triggered by the Booster Dose of Anti-SARS-CoV-2 Vaccine in a Patient with β -Thalassemia

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Abstract: Background: Haemophagocytic lymphohistiocytosis (HLH) is a rare and potentially life-threatening systemic hyperinflammatory disease, which can have several aetiologies. Clinical case: a 48-year-old woman affected by a transfusion-dependent β -thalassemia was hospitalized in our haematology unit presenting with intermittent fever, hepatosplenomegaly and pancytopenia, which developed a few days after the booster dose of anti-SARS-CoV-2 mRNA vaccine. The investigations performed during hospitalization led to a diagnosis of HLH and steroid therapy where IV dexamethasone was initiated and provided benefits. Conclusions: the severity of HLH mandates early treatment, but the management of patients with post-vaccine HLH is still challenging and requires further study. No cases of HLH in patients with thalassemia were previously described.

Keywords: thalassemia; haemophagocytic lymphohistiocytosis; SARS-CoV-2; vaccine



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1. Introduction

Haemophagocytic lymphohistiocytosis (HLH) is a rare systemic hyperinflammatory disease characterized by the excessive but ineffective stimulation of the immune system, which leads to an accumulation of macrophages and lymphocytes in target organs. The resulting cytokine storm is responsible for severe, life-threatening clinical manifestations; therefore, early diagnosis and treatment are essential. We report a case of a patient with thalassemia who developed pancytopenia, hepatosplenomegaly and fever a few days after receiving anti-SARS-CoV-2 vaccination and was later diagnosed with HLH.

2. Clinical Case

A 48-year-old woman affected by transfusion-dependent β -thalassemia (genotype IVS-I-110 [G > A]/Cod39 [C > T], pre-transfusion haemoglobin levels of about 9 g/dL, ferritin levels kept around 700 ng/mL with deferasirox) was hospitalized in our haematology unit. She presented with intermittent fever and a maximum temperature of 39 °C, preceded by chills, which started six days after receiving anti-SARS-CoV-2 vaccine (booster dose with Comirnaty) and were accompanied by worsening of the anaemia to haemoglobin levels of 6.8 g/dL (9.3 g/dL at the previous test 7 days before vaccination). At the time of hospitalization, the fever had lasted for over a month and had not been responsive to multiple empiric antibiotic treatments. From the onset of fever to hospitalization, the patient

had undergone thirteen blood transfusions, about twice her periodic need. Moreover, before admission, a chest radiography was negative for any pathological findings, while abdomen ultrasonography showed an enlargement of the spleen, compared to a previous abdominal ultrasonography performed 6 months earlier (longitudinal diameter increased from 13 to 20 cm). A physical examination showed no clinical evidence suggestive of infection sites and there were no significant history or additional symptoms in the two weeks prior to the vaccination. The patient had not taken any cytotoxic drugs, had no nutritional deficiencies and no family or personal history of autoimmune diseases or other immunological disorders. Table 1 summarizes the most relevant laboratory examinations upon admission.

Table 1. Main laboratory tests upon admission.

Test	Results	Normal Values
Hb	10.1 g/dL	12–16 g/dL
MCV	81 fL	78–100 fL
WBC	$2.8 \times 10^3/\mu\text{L}$	$4\text{--}9 \times 10^3/\mu\text{L}$
Neutrophils, lymphocytes	$1.12 \times 10^3/\mu\text{L}$, $1.09 \times 10^3/\mu\text{L}$	$2\text{--}8 \times 10^3/\mu\text{L}$, $1\text{--}5 \times 10^3/\mu\text{L}$
PLT	$107 \times 10^3/\mu\text{L}$	$150\text{--}450 \times 10^3/\mu\text{L}$
Reticulocytes	$13.4 \times 10^3/\mu\text{L}$, 0.46%	$25\text{--}75 \times 10^3/\mu\text{L}$, 1–2%
Direct and indirect Coombs test	Positive	Negative
Bilirubin total/indirect	2.3/1.6 mg/dL	<1.2/<0.7 mg/dL
AST/ALT	35/24 U/L	<35 U/L
LDH	287 U/L	125–220 U/L
Haptoglobin	<8 mg/dL	>25 mg/dL
Hs-CRP	1.3 mg/dL	<0.5 mg/dL
Ferritin	1590 ng/mL	15–150 ng/mL
Triglycerides	146 mg/dL	<150 mg/dL
AP, INR	82%, 1.14	80–120%
PTT	56"	22–34"
Fibrinogen	276 mg/dL	150–450 mg/dL
D-dimer	327 ng/mL	<250 ng/mL

Hb: haemoglobin; MCV: mean corpuscular volume; WBC: white blood cells; PLT: platelets; AST: aspartate aminotransferase; ALT: alanine aminotransferase; LDH: lactate dehydrogenase; Hs-CRP: high-sensitivity C-reactive protein; AP: prothrombin activity; INR: international normalized ratio; PTT: partial thromboplastin time.

The chest and abdomen computer tomography (CT) scan with contrast showed the already known paravertebral foci of extramedullary erythropoiesis and hepatosplenomegaly with homogeneous density. A positron emission tomography (PET)-CT showed liver and spleen of increased size with heterogeneous hypercaptation of the radiotracer (^{18}F -FDG), mainly affecting the spleen, in absence of other significant findings. Four sets of blood cultures tested negative, and the echocardiogram showed no endocardial vegetations. QuantiFERON-TB Gold urine and stool cultures also tested negative. Several serological tests (via immunofluorescence or enzyme immunoassays) and molecular biology tests (via polymerase chain reaction) were performed on peripheral blood and bone marrow, and the following infectious agents were ruled out as active pathogens: Cytomegalovirus, Epstein–Barr virus, parvovirus B19, hepatitis B virus, hepatitis C virus, *Leishmania* spp., *Rickettsia* spp., *Aspergillus* spp., *Bartonella* spp., *Borrelia burgdorferi*, *Brucella* spp., *Toxoplasma gondii*, *Plasmodium* spp., human immunodeficiency virus 1–2. SARS-CoV-2 RNA was not detected on nasopharyngeal swabs.

During her hospital stay, haemoglobin level, platelet and neutrophil counts dropped, respectively, to a minimum of 7.7 g/dL, $88 \times 10^3/\mu\text{L}$ and $0.94 \times 10^3/\mu\text{L}$. The bone marrow examination showed a rich histiocytic infiltrate with interstitial distribution, haemophago-

cytosis and T-prevalent lymphoid infiltrate with interstitial distribution. Moreover, perforin and CD107a expression were tested, showing a normal intracytoplasmic perforin expression but a defective NK cells degranulation. With an H-score [1] of 203 points, defining an 88–93% probability of haemophagocytic syndrome, the patient fulfilled 6 of 8 diagnostic criteria according to the HLH-2004 protocol [2]: fever $\geq 38.5^{\circ}\text{C}$, splenomegaly, peripheral blood trilinear cytopenia, haemophagocytosis in bone marrow, low NK cells activity and ferritin $> 500\text{ ng/mL}$, evaluated as a timing of fast and repeated increase after vaccination (727 ng/mL before vaccine; 927 ng/mL, 1590 ng/mL, 2247 ng/mL at 2, 3, 4 weeks later, respectively) in the presence of normal values of liver iron concentration assessed by magnetic resonance imaging (LIC-R2 3.3 mg/g dry tissue). Hypertriglyceridemia and hypofibrinogenemia were absent, and soluble CD25 was not tested. No genetic tests were performed to rule out hereditary HLH since family history was negative.

Therefore, as the clinical features were chronic by this stage, and the patient's condition was stable, a steroid therapy with IV dexamethasone 10 mg sid was initiated. After a few days, the complete blood count recovered to normal, and the patient remained afebrile (Figure 1). The patient was treated with dexamethasone for a total of 6 weeks, during which a gradual tapering was performed. No additional supportive care was needed.

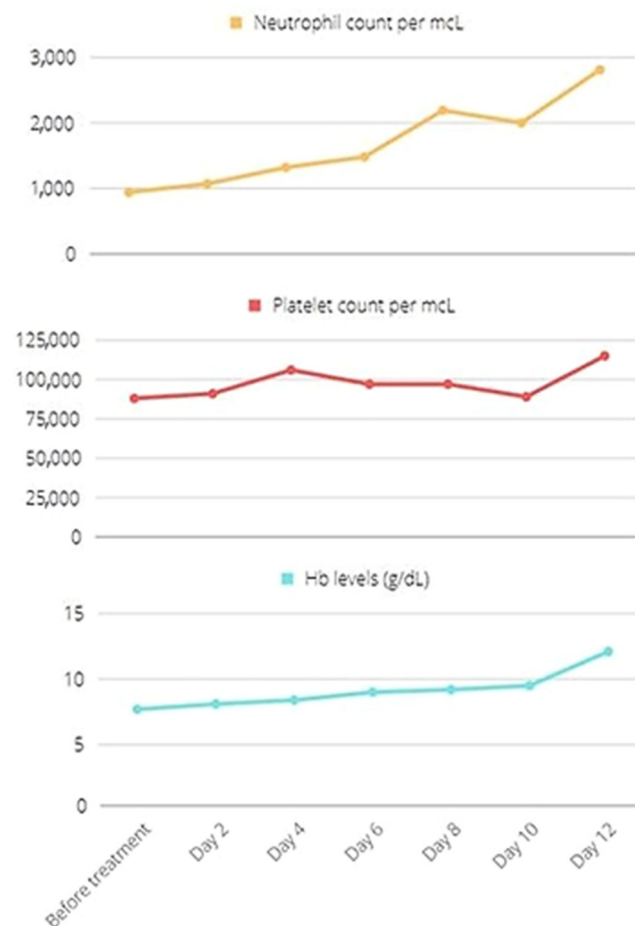


Figure 1. Resolution of pancytopenia with dexamethasone. Hb: haemoglobin.

3. Discussion

Given the close temporal correlation between the administration of the booster dose of anti-SARS-CoV-2 vaccine and the onset of fever and the worsening of anaemia with subsequent neutropenia and thrombocytopenia, the clinical presentation compatible with HLH was likely triggered by vaccination. Since the patient was admitted to our unit about a month after the onset of symptoms, useful laboratory data that could have been collected in the acute phase were lacking. The timing of the fast and repeated increase in serum

ferritin levels associated with normal values of LIC, after vaccination, may be related to the inflammatory state of HLH rather than to iron overloading.

Fortunately, the disease did not have a rapid and ominous course as in most cases. This made it possible to exclude other potential causes of secondary HLH and to attempt a treatment with glucocorticoids only, avoiding classic therapeutic schemes that involve the combination of dexamethasone and etoposide for eight weeks of induction [3,4].

To our knowledge, there are no data concerning the management of HLH in patients with thalassemia, and this is the first case report of a patient with HLH occurring after anti-SARS-CoV-2 booster dose. Other authors have recently described similar cases associated with the first doses of m-RNA or virus-vectored vaccines: some of them were successfully treated with corticosteroid therapy alone, obtaining a rapid remission of symptoms and improvement in blood counts [5–8]. Some recovered with support therapy alone [9], while others required etoposide or stronger combined therapies with steroids, IVIG and IL-1 receptor antagonists [8,10–12]. To date, unfortunately, there are no management guidelines for patients who develop HLH after anti-SARS-CoV-2 vaccination.

4. Conclusions

The immune stimulation caused by vaccines can sometimes lead to systemic hyperinflammatory diseases. The relationship between the immune response to anti-SARS-CoV-2 vaccines and HLH requires further study, but HLH should be considered in patients who present with long-lasting fever after vaccination in the absence of any other potential causes, even after the administration of first doses without adverse reactions.

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Conflicts of Interest: Aurelio Maggio has been or is a member of advisory boards for Novartis, Celgene Corp (Bristol Meyers Squibb), Vertex and Bluebird Bio. The remaining authors have no conflicts of interest to disclose.

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