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Salinity Tolerance and Ion Accumulation of Coastal and Inland Accessions of Clonal Climbing Plant Species *Calystegia sepium* in Comparison with a Coastal-Specific Clonal Species *Calystegia soldanella*

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Abstract: Plant species adapted to saline habitats represent an important resource in the assessment of salinity tolerance mechanisms. The aim of the present study was to analyze salinity tolerance and ion accumulation characteristics for various accessions of *Calystegia sepium* from different habitats in comparison to these of *Calystegia soldanella* in controlled conditions. Plants were introduced in culture using stem explants with leaf and were cultivated in controlled conditions under six different substrate salinities. Salinity tolerance of both *C. sepium* and *C. soldanella* plants was relatively high, but the tolerance of particular accessions did not depend on the substrate salinity level in their natural habitats. *C. sepium* accession from a mesophytic non-saline habitat was only slightly negatively affected by increasing substrate salinity. However, coastal accession of *C. sepium* and coastal-specific species *C. soldanella* had some similarities in ion accumulation characteristics, both accumulating a high concentration of soluble ions in aboveground parts and excluding them from underground parts. All *C. sepium* accessions from different habitats represented varied phenotypes, possibly associated with their genetic differences. *C. sepium* accessions from different habitats can be suggested as models for further studies aiming at dissecting possible genetic, epigenetic and physiological mechanisms of adaptation to heterogeneous environmental conditions.

Keywords: *Calystegia sepium*; *Calystegia soldanella*; clonal plants; ion accumulation; osmotic protection; salinity



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1. Introduction

Physiology of salinity tolerance is a fundamentally relevant aspect in plant biology with immense practical importance for nature conservation, agriculture as well as environmental biotechnology. Plant species adapted to saline habitats represent an important resource in this respect [1–4]. So far, a majority of salinity tolerance studies have focused on obligate halophyte species from hypersaline habitats, such as inland saline deserts or coastal salt marshes, but the mechanisms of salinity tolerance of facultative halophytic species occurring both in saline and non-saline habitats are a mostly unexplored topic. In contrast to obligate halophytes, it is expected to find a wider diversity of different physiological salinity tolerance-related mechanisms in plants from habitats along a border between saline and non-saline environments [5].

Calystegia sepium (L.) R. Br. (Convolvulaceae) is a clonal climbing plant species found in a variety of habitats. *C. sepium* is an umbrella species of the European-protected habitat EUH 6430: “Hydrophilous tall herb fringe communities of plains and the montane to alpine levels” [6]. However, the species exhibits high diversity in respect to a range of habitats where it can be found. Thus, at one extreme, *C. sepium* is a component of nitrophilous mesophytic vegetation of riverbanks and other similar habitats [7]; at the other, it is frequently located in relatively arid semi-ruderal habitats like road verges [8]. Moreover, *C. sepium* is often found on coastal habitats, being an important species of salt-adapted vegetation complexes

of the Southern Baltics [9]. These facts raise the question on existence of different ecotypes of *C. sepium* based on genetical variation in certain adaptive characteristics. Due to the predominantly vegetative mode of propagation of the species [10], it is highly possible that certain populations consist of a single locally well-adapted clone of *C. sepium*. The presence of *C. sepium* plants in habitats with different salinity levels gives the opportunity to consider the existence of differences in their salinity tolerance, but there is no experimental data available on the salinity tolerance of *C. sepium*. However, according to the recently established ecological indicators of Swedish vascular plants, the species is characterized as only moderately salinity tolerant (indicator value 2 of 5) [11].

The taxonomically closely related species, *Calystegia soldanella* (L.) R. Br., is a coastal-specific creeping clonal plant characteristic for sand dunes [12]. The species has pronounced drought [13] and, presumably, salinity tolerance [14,15], but the latter has been assessed experimentally only in a hydroponic cultivation system [16].

Accumulation of monovalent cations K^+ and Na^+ is an important constituent of salinity tolerance of plant species native to salt-affected habitats with consequences for both osmotic as well as ion balance [17–19]. There are extremely large differences in ion accumulation rates, patterns and specificity for various salt-adapted plant species [20–23]. Most intriguing, although it is usually justified that retention of K^+ in cytoplasm is a critical feature for salinity tolerance, several species native to saline habitats predominantly accumulate Na^+ and exclude K^+ at high salinity [24,25]. Previously, during a survey of 102 coastal plant species from sea-affected habitats, it was found that *C. sepium* had relatively high Na^+ uptake potential in leaves, with ion accumulation characteristics similar to these of the coastal-specific species *Trifolium fragiferum*, *Schoenoplectus tabernaemontani*, *Atriplex littoralis* etc. [26].

The aim of the present study was to analyze salinity tolerance and ion accumulation characteristics for various accessions of *C. sepium* from different habitats in comparison to these of *C. soldanella*. in controlled conditions. It was hypothesized that salt tolerance and ion accumulation characteristics of coastal accession of *C. sepium* will be comparable to these of *C. soldanella*, but putatively drought tolerant inland accession of *C. sepium* will be more salt tolerant than the accession from a mesophytic habitat.

2. Materials and Methods

2.1. Plant Material

As model plants, three accessions of *C. sepium* from different habitats and *C. soldanella* were used (Table 1). Mature rhizome fragments from various accessions of *C. sepium* were collected in three different habitats in June and July. *C. sepium* 1 (CSe1) was a component of a salt-affected coastal sandy beach perennial vegetation, located about 10 m from a water line, together with *Phragmites australis*. *C. sepium* 2 (CSe2) was from a mesophytic vegetation on the banks of a pond together with *P. australis* and other perennial vascular plant species. *C. sepium* 3 (CSe3) plants were located on a drought-exposed steep riverbank growing in sand-gravel soil. The fragments were used to establish a laboratory stock culture of *C. sepium*. Seeds of *C. soldanella* (CSo) were purchased from Plant World Seeds (Newton Abbot, Devon, UK).

In addition, five leaves from different shoots were collected from all accessions of *C. sepium* plants in their native habitats and were used for measurements, as described further.

2.2. Establishment of Stock Cultures

Rhizomes were washed in tap water to remove all soil particles, rinsed with deionized water and cut in 2–3 cm fragments with visible presence of vegetative meristems (nodes). After immersing for 30 s in ethanol (70%), rhizome fragments were washed with tap water for 2 min, followed by deionized water, and were blotted dry. Rhizome fragments (30 for each accession) were planted in plastic tissue culture containers (1 L, 10 fragments per container) containing heated (60 °C, 24 h) garden soil (Biolan, Eura, Finland) and closed with lids. Containers were placed in a growth cabinet (light/dark period of 16/8 h, photosynthetically active radiation with a photon flux density $100 \mu\text{mol m}^{-2} \text{s}^{-1}$, day/night temperature 15/20 °C). After the appearance of shoots, the lids were removed and the

containers were acclimated to greenhouse conditions. When shoots reached 10–15 cm length, plants were transferred to 5 L containers with garden soil (Biolan, Eura, Finland), 5 plants per container. Plants were cultivated in an experimental automated greenhouse (HortiMaX, Maasdijk, Netherlands) with supplemented light from Master SON-TPIA Green Power CG T 400 W (Philips, Amsterdam, Netherlands) and Powerstar HQI-BT 400 W/D PRO (Osram, Munich, Germany) lamps ($380 \mu\text{mol m}^{-2} \text{s}^{-1}$ at the plant level) for a 16 h photoperiod, with day/night temperature of 24/16 °C and relative air humidity of 60–70%. Support for plants was provided by bamboo poles. Necessary soil moisture was provided by deionized water. Stock cultures of *C. sepium* were used for the propagation of plant material for experiments when plagiotropic above-ground stems formed.

Table 1. Plant material used in the present study for the establishment of a laboratory culture of *Calystegia sepium* and *Calystegia soldanella*.

Species, Accession	Code	Habitat	Soil Electrical Conductivity (mS m^{-1})	Establishment of Stock Culture/Propagation	Characteristics	Locality (Coordinates)
<i>Calystegia sepium</i> 1	CSe1	Coastal, sandy beach with perennial vegetation	623 ± 105	Rhizome fragments/stem explants with leaf	White flowers	Mērsrags, Latvia (N 57°21'57" E 23°7'21")
<i>Calystegia sepium</i> 2	CSe2	Inland, mesophytic vegetation on banks of pond	86 ± 5	Rhizome fragments/stem explants with leaf	White flowers	Salaspils, Latvia (N 56°51'32" E 24°30'38")
<i>Calystegia sepium</i> 3	CSe3	Inland, dry grassland vegetation on a steep riverbank	83 ± 2	Rhizome fragments/stem explants with leaf	Light pink flowers	Ogre, Latvia (N 56°48'59" E 24°36'59")
<i>Calystegia soldanella</i>	CSo	Coastal, sand dunes	NA	Seeds/stem explants with leaf	NA	NA

For *C. soldanella*, seeds were treated with concentrated H_2SO_4 for 3 h to interrupt physical dormancy, as described previously [27]. Seeds were planted in plastic tissue culture containers (1 L, 5 seeds per container) containing heated (60 °C, 24 h) garden soil (Biolan, Eura, Finland) and closed with lids. Containers were placed in a growth cabinet (light/dark period of 16/8 h, photosynthetically active radiation with a photon flux density $100 \mu\text{mol m}^{-2} \text{s}^{-1}$, day/night temperature 15/20 °C). After the appearance of seedlings, the lids were removed, and the containers were acclimated to greenhouse conditions. Well-developed seedlings with 2–3 leaves were transferred to 5 L containers with garden soil (Biolan, Eura, Finland), 3 plants per container, and cultivated in greenhouse for 10 weeks. Necessary soil moisture was provided by deionized water.

2.3. Plant Propagation for Experiments and Cultivation

Preliminary experiments were performed to establish the most suitable method for vegetative propagation of *C. sepium* and *C. soldanella* from a stock culture. It was established that fragments of plagiotropic stem with a single leaf and fragments of creeping stem with a single leaf were the most appropriate material for propagation of *C. sepium* and *C. soldanella*, respectively. Stem explants (about 5–6 and 3–4 cm in length for *C. sepium* and *C. soldanella*, respectively) were inserted in quartz sand (Saulkalne S, Saulkalne, Latvia), moistened with deionized water in plant growing trays and placed in closed 48 L containers. Containers were put in a growth chamber (light/dark period of 16/8 h, photosynthetically active radiation with a photon flux density $100 \mu\text{mol m}^{-2} \text{s}^{-1}$, day/night temperature 15/20 °C). After 12 days, explants with well-developed roots and vigorous shoot growth were transplanted to 1.2 L plastic containers filled with 1 L garden soil (Biolan, Eura, Finland) and quartz sand (Saulkalne S, Saulkalne, Latvia) (2:1, v/v).

During all experiments, plants were kept in a greenhouse in the same conditions as indicated above and irrigated with deionized water every other day. Substrate water content was monitored with a HH2 moisture meter equipped with a WET-2 sensor (Delta-T Devices, Burwell, UK) and kept at 50 to 60%. Every other week, plants were fertilized

with Yara Tera Kristalon Red and Yara Tera Calcinit fertilizers (Yara International, Oslo, Norway). A stock solution was prepared for each fertilizer (100 g L^{-1}) and the working solution contained 25 mL of each per 10 L deionized water, used with a rate 100 mL per container. Individual containers were randomly redistributed weekly on a greenhouse bench. Support for *C. sepium* plants was provided by bamboo poles.

2.4. Plant Treatment and Harvesting

Ten days after the final transplanting, plants from three accessions of *C. sepium* and *C. soldanella* plants were randomly distributed in six groups (treatments), with five individuals per group (control without Na^+ addition, 0.5, 1, 2, 4, 6 $\text{g Na}^+ \text{ L}^{-1}$). Treatment with NaCl dissolved in deionized water was performed once a week, using doses not larger than 1 g Na^+ (2.54 g NaCl) per individual container until final treatment doses were reached. Plants were cultivated for an additional 7 weeks after reaching the full treatment.

At termination of the experiments, plants were individually separated in different parts (stems, leaves, rhizomes, roots). For CSe3, also flowers, and for CSo, leaf blades and leaf petioles were harvested separately. The number of individual stems and leaves, as well as the length of stems, were measured. All above-ground parts were rinsed with deionized water and blotted dry. Roots and rhizomes were carefully washed with running tap water to remove any substrate particles, rinsed with deionized water and blotted dry. All individual parts were weighed before and after drying in an oven at $60 \text{ }^\circ\text{C}$ for 72 h. Water content in plant parts was calculated in $\text{g H}_2\text{O}$ per g dry mass.

2.5. Measurements

Measurements were performed in three biological replicates (tissue samples from individual plants) per treatment. Tissue samples were cut to small pieces using scissors, thoroughly homogenized, and a sample of 0.2 g was randomly taken from the total amount of tissue material. Tissues were ground with mortar and pestle to a fine powder, and 10 mL of deionized water was added. The homogenate was stirred with a pestle for 1 min. After filtration through nylon mesh cloth, (No. 80) homogenate was used for measurement of ion concentration by LAQUAtwin compact meters B-722 (Na^+) and B-731 (K^+), and electrical conductivity by LAQUAtwin conductivity meter B-771 (Horiba, Kyoto, Japan) and measurement of osmotic value. For osmotic value analysis, 50 μL of extract were transferred in a 1.5 mL Eppendorf tube and placed in a freezing point osmometer, Osmomat 3000 Basic (Gonotec Meß- und Regeltechnik, Berlin, Germany), and operated according to the manufacturer's instructions. Using a standard curve for different concentrations of NaCl and KCl, the osmotic value caused by the total concentration of Na^+ and K^+ was calculated according to the actual Na^+ and K^+ concentration of each sample extract. For each sample, the difference between the total osmotic value and the osmotic value due to Na^+ and K^+ ions was calculated and designated as "non-ionic osmotic value", which showed the osmotic effect of other osmotically active ions (besides Na^+ , K^+ and Cl^-) or non-ionic compounds. At least three analytical replicates were performed for each sample and the average value was calculated.

2.6. Data Analysis

Results were analyzed by KaleidaGraph (v. 5.0, Synergy Software, Reading, PA, USA). Statistical significance of differences was evaluated by one-way ANOVA using post-hoc analysis with minimum significant difference. Principal component analysis was performed by a freely available web program, ClustVis (<http://biit.cs.ut.ee/clustvis/>) (accessed on 30 June 2022) [28]. For graphs of principal component analysis, prediction ellipses were such that, with probability 0.95, a new observation from the same group will fall inside the ellipse. Unit variance scaling was applied to rows; singular value decomposition with imputation was used to calculate principal components.

3. Results

Plants from three different *C. sepium* accessions showed significant differences in ion accumulation character in leaves in natural conditions (Table 2). Significant higher tissue water content in leaves of coastal accession CSe1 tended to equate to differences in ion content and electrolytical activity that were visible on dry mass basis.

Table 2. Water content, Na⁺ and K⁺ concentration and electrical conductivity (EC) in leaf blades of different accessions of *Calystegia sepium* in natural conditions, expressed either on dry mass or tissue water basis.

Parameter	<i>C. sepium</i> 1	<i>C. sepium</i> 2	<i>C. sepium</i> 3
H ₂ O (g g ⁻¹ DM)	7.5 ± 0.8 ^a	5.5 ± 0.1 ^b	4.4 ± 0.2 ^c
Na ⁺ (g kg ⁻¹ DM)	26.33 ± 1.97 ^a	0.95 ± 0.07 ^c	2.23 ± 0.07 ^b
K ⁺ (g kg ⁻¹ DM)	32.8 ± 2.9 ^b	26.6 ± 4.3 ^c	41.0 ± 1.0 ^a
Na ⁺ (mol L ⁻¹)	0.156 ± 0.011 ^a	0.008 ± 0.001 ^c	0.022 ± 0.001 ^b
EC (mS cm ⁻¹ kg ⁻¹ DM)	113 ± 6 ^a	57 ± 5 ^b	60 ± 1 ^b
K ⁺ (mol L ⁻¹)	0.115 ± 0.010 ^b	0.123 ± 0.018 ^b	0.237 ± 0.006 ^a
EC (mS cm ⁻¹ L ⁻¹)	15.4 ± 0.9 ^a	10.2 ± 0.7 ^b	13.5 ± 0.1 ^a
K:Na (mol)	0.8 ± 0.1 ^c	17.2 ± 3.6 ^a	10.8 ± 0.3 ^b

Data are means from three samples ± SE, at least five leaves each from different shoots. Different letters for a particular parameter indicate statistically significant differences between the accessions ($p < 0.05$). DM, dry mass.

When cultivated in controlled conditions, the model species showed different biomass accumulation potential (Figure 1) and biomass partitioning characteristics even without added salt (Figure 2). In non-saline conditions, CSe1 plants had the highest total biomass, and more than 80% was allocated to stems. CSe2 plants also had high contribution of stems, but no rhizomes were present. In contrast, *C. soldanella* plants had the lowest biomass and accumulated most of the matter in leaves and rhizomes. Only CSe3 plants developed flowers in the current conditions.

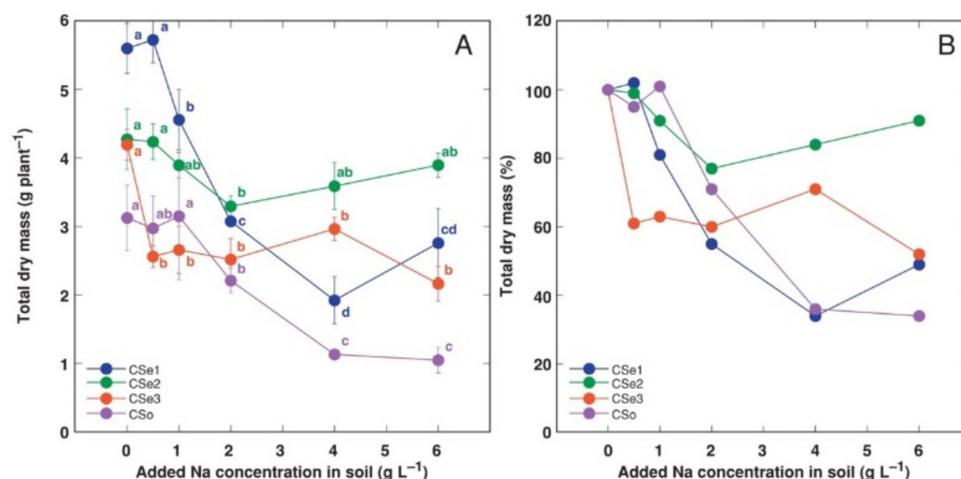


Figure 1. Effect of increasing soil salinity on total dry mass (A) and total dry mass relative to control plants (B) of *Calystegia sepium* accessions CSe1, CSe2, CSe3 and *Calystegia soldanella* (CSo). Na⁺ was added in a form of NaCl. Data are means from five replicates for each concentration ± SE. Different letters of the respective color indicate statistically significant differences ($p < 0.05$) between treatments for the respective model plant.

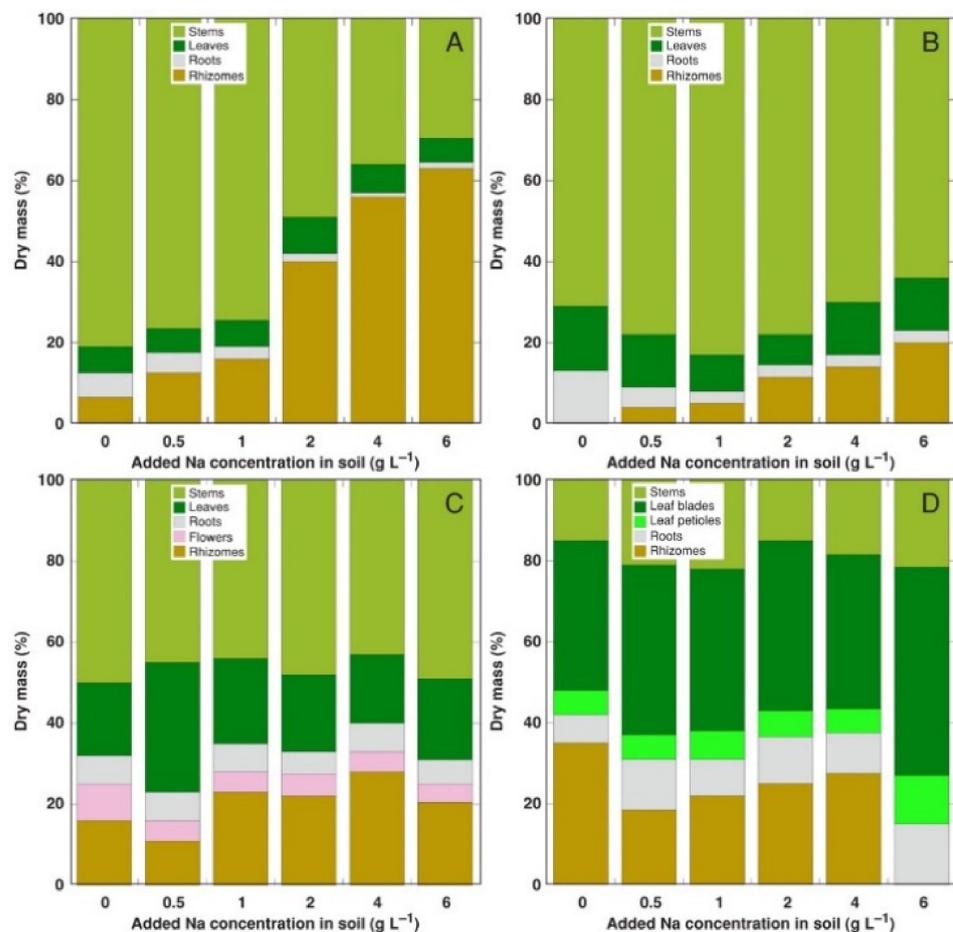


Figure 2. Relative changes in biomass partitioning in *Calystegia sepium* plants from accessions CSe1 (A), CSe2 (B), CSe3 (C) and *Calystegia soldanella* (CSo) (D) plants due to increasing substrate salinity. Na⁺ was added in a form of NaCl.

In addition, the growth of different model plants was differentially affected by increasing soil salinity. Total biomass of CSe1 plants showed the most dramatic decrease as a result of increasing soil salinity both in absolute (Figure 1A) and relative (Figure 1B) terms. *C. soldanella* (CSo) plants also were highly negatively affected by increasing soil NaCl concentration. In contrast, CSe2 plants showed significantly lower total biomass, at only 2 g Na⁺ L⁻¹. However, the CSe3 plants were sensitive even at 0.5 g Na⁺ L⁻¹, but the effect was not concentration-dependent.

The most pronounced change in biomass partitioning due to increasing soil salinity was the Na⁺ concentration-dependent relative increase of rhizome biomass in CSe1, accompanied by a decrease in both leaf and root biomass (Figure 2A). A similar effect was evident also for CSe2, but to a lesser extent (Figure 2B). Individual variability in biomass partitioning was rather high for CSe3 plants (Figure 2C); therefore, no pronounced effect of increasing salinity was evident. For CSo plants, sudden disappearance of rhizomes occurred at the highest salinity, but no other changes were evident (Figure 2D).

Other morphological parameters also showed plant genotype-specific changes due to increasing soil salinity (Figure 3). The number of leaves was the most sensitive indicator, with significant decrease by salinity for all model plants (Figure 3C). Coastal accession *C. sepium* had the highest number of stems (Figure 3A) and total stem length (Figure 3B) for control plants, and these parameters were dramatically decreased with increasing soil salinity.

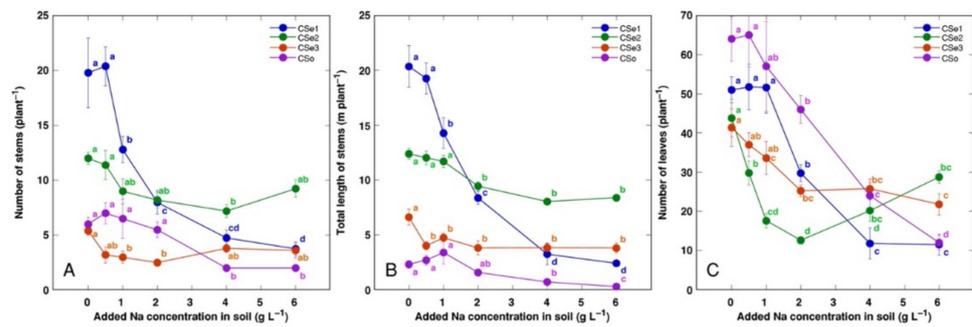


Figure 3. Effect of increasing soil salinity on the number of stems (A), total stem length (B), and number of leaves (C) of *Calystegia sepium* accessions CSe1, CSe2, CSe3 and *Calystegia soldanella* (CSo). Na^+ was added in a form of NaCl. Data are means from five replicates for each concentration \pm SE. Different letters of the respective color indicate statistically significant differences ($p < 0.05$) between treatments for the respective model plant.

Water content in leaves and stems changed relatively little with increasing soil salinity (data not shown); however, the roots showed salinity-related physiological differences between the model plants (Figure 4A). Root water content significantly increased by salinity in CSe1 and decreased in CSe2. In addition, root water content increased in CSe3 at low salinity and tended to decrease in CSo. Rhizome water content decreased both in CSe1 and CSe2, with a tendency to increase in CSo (Figure 4B).

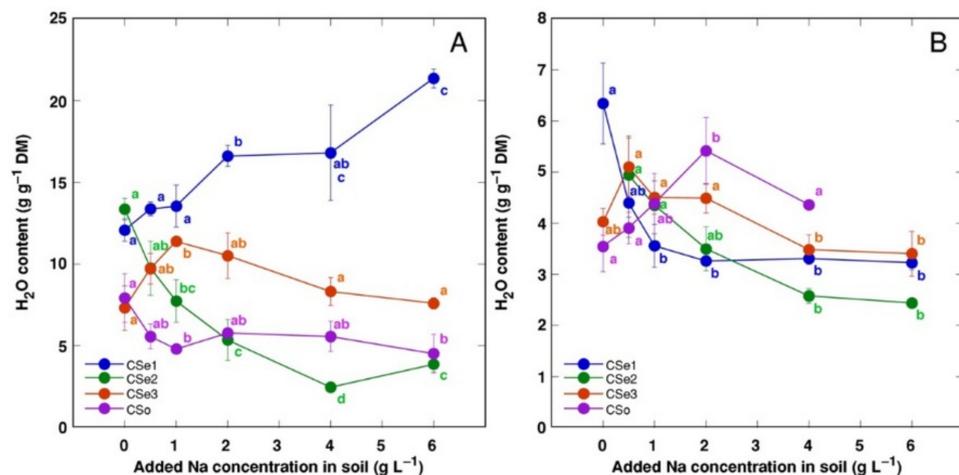


Figure 4. Effect of increasing soil salinity on water content in roots (A) and rhizomes (B) of *Calystegia sepium* accessions CSe1, CSe2, CSe3 and *Calystegia soldanella* (CSo). Na^+ was added in a form of NaCl. Data are means from five replicates for each concentration \pm SE. Different letters of the respective color indicate statistically significant differences ($p < 0.05$) between treatments for the respective model plant.

Changes in ion accumulation under salinity were evaluated in separate plant parts. Na^+ was efficiently excluded from rhizomes of all model plants, leaves of CSe3 and roots of CSe1 (Figure 5). For CSe1, Na^+ preferentially accumulated in leaves, with lower concentration in stems (Figure 5A). For CSe2 and CSo, similar concentration of Na^+ accumulated in roots, leaves and stems (Figure 5B,D). For CSe3, accumulation of Na^+ dominated in roots at low and medium soil Na^+ concentration, with stem Na^+ concentration reaching high levels at high treatment rate (Figure 5C). In general, CSe1 and CSe2 had higher Na^+ accumulation potential in comparison to that in CSe3 and CSo.

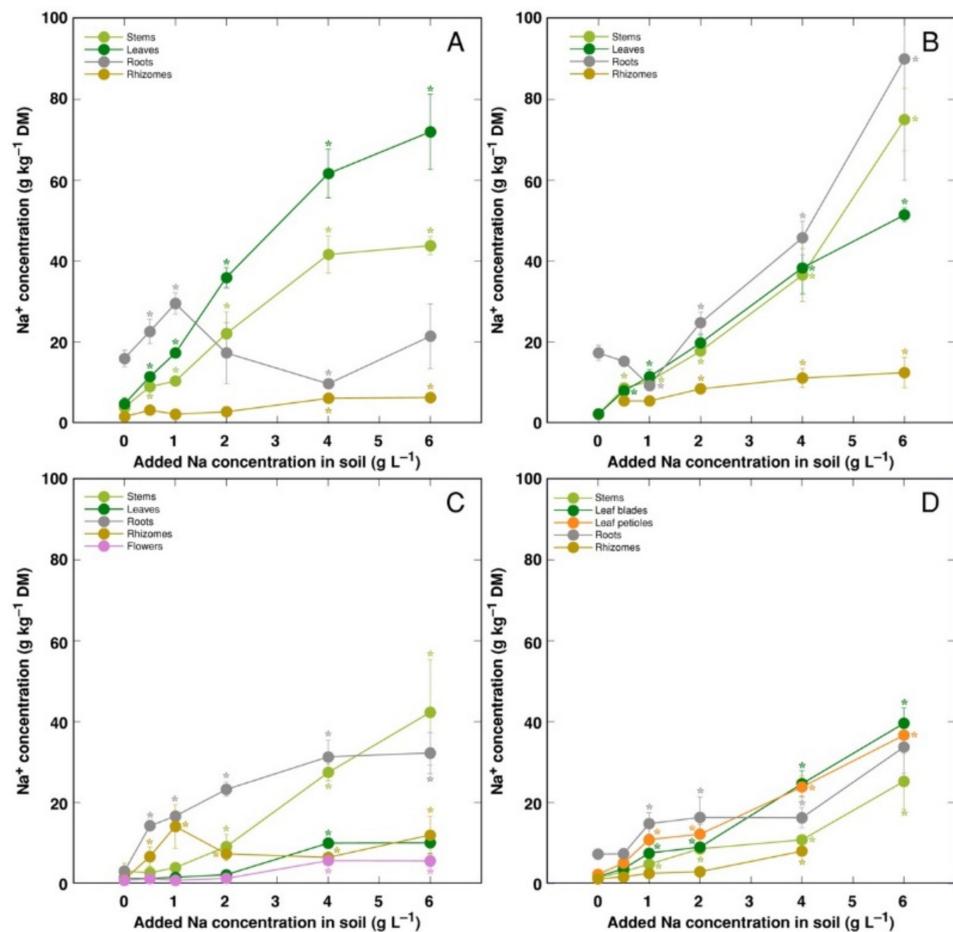


Figure 5. Changes in Na⁺ concentration under the effect of increasing soil salinity in different parts of *Calystegia sepium* accessions CSe1 (A), CSe2 (B), CSe3 (C) and *Calystegia soldanella* (D). Na⁺ was added in a form of NaCl. Data are means from three replicates for each concentration \pm SE. Asterisks of the respective color indicate statistically significant differences ($p < 0.05$) from control.

K⁺ concentration increased in stems and decreased in leaves for CSe1 and CSe2 plants under the effect of salinity (Figure 6A,B). However, leaf K⁺ concentration increased for CSe3 and CSo (Figure 6C,D). In roots, K⁺ concentration was not significantly affected by salinity for CSe1, but it decreased for all other model plants. In rhizomes, K⁺ concentration increased for CSe1 and decreased in CSe3, and it was not affected for CSe2 and CSo.

For control plants, the molar concentration ratio of K⁺ to Na⁺ was the lowest for CSe1, and it increased equally in CSe2 and CSo, with the highest value in CSe3 (Figure 7). The K⁺:Na⁺ ratio decreased with increasing soil salinity in all plant genotypes and parts, but the character of decrease was highly variable. In general, a less-pronounced decrease was evident for CSo and especially CSe3. In leaves and flowers of CSe3, the K⁺:Na⁺ ratio at first even increased at low salinity (Figure 7C). For all model plants, the lowest K⁺:Na⁺ ratio was for roots.

Electrical conductivity was measured as an indication of the total concentration of electrolytically active ions in plant tissues, and, in general, it increased with increasing soil salinity, but to a different degree (Figure 8). The lowest electrolytical activity and, relatively, the least increase of the activity by salinity was evident in rhizomes for all genotypes, and flowers for CSe3. Electrical conductivity was similar in stems and leaves for all model plants except CSo, where its level was lower in stems.

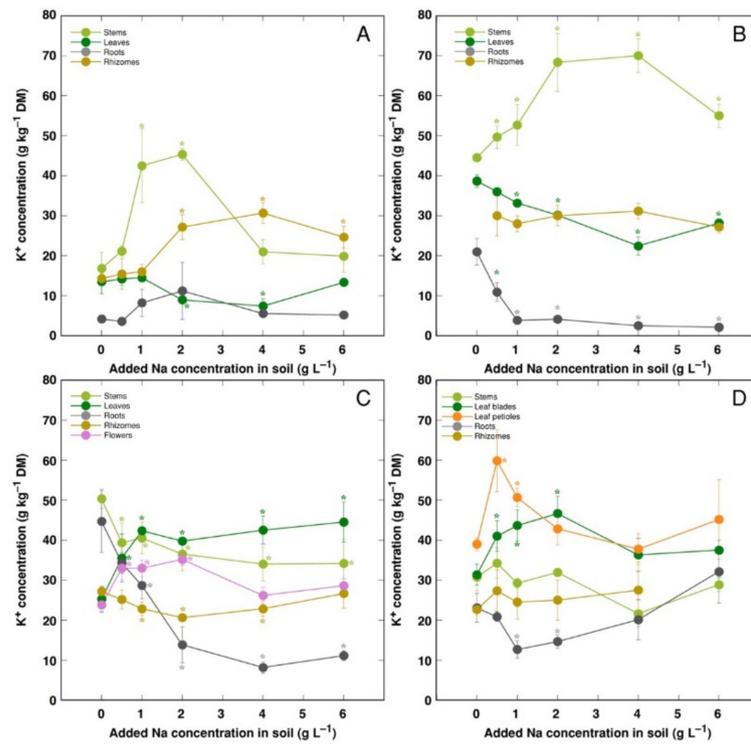


Figure 6. Changes in K^+ concentration under the effect of increasing soil salinity in different parts of *Calystegia sepium* accessions CSe1 (A), CSe2 (B), CSe3 (C) and *Calystegia soldanella* (D). Na^+ was added in a form of NaCl. Data are means from three replicates for each concentration \pm SE. Asterisks of the respective color indicate statistically significant differences ($p < 0.05$) from control.

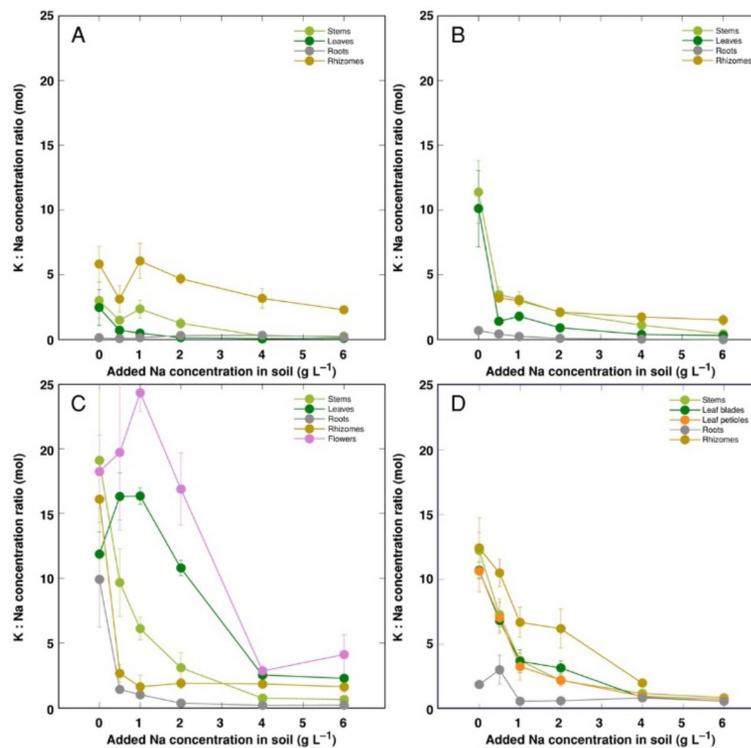


Figure 7. Changes in $Na^+ : K^+$ concentration ratio under the effect of increasing soil salinity in different parts of *Calystegia sepium* accessions CSe1 (A), CSe2 (B), CSe3 (C) and *Calystegia soldanella* (D). Na^+ was added in a form of NaCl. Data are means from three replicates for each concentration \pm SE. Asterisks of the respective color indicate statistically significant differences ($p < 0.05$) from control.

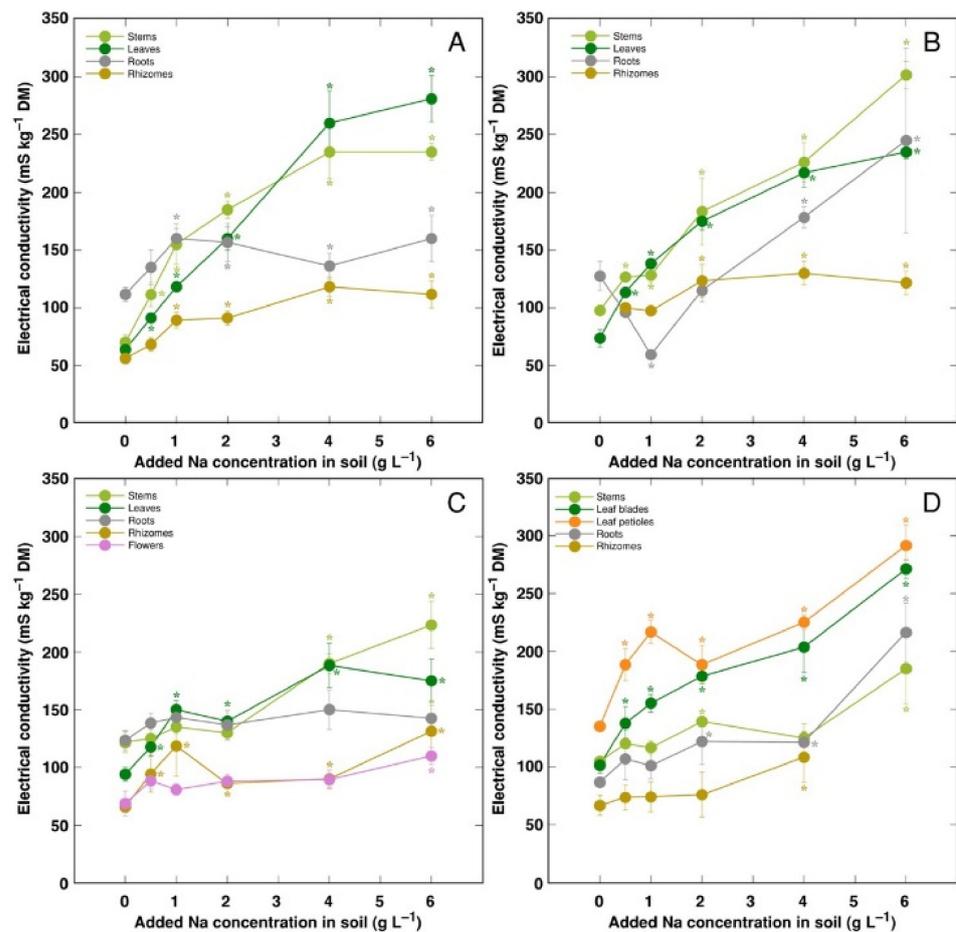


Figure 8. Changes in electrical conductivity under the effect of increasing soil salinity in different parts of *Calystegia sepium* accessions CSe1 (A), CSe2 (B), CSe3 (C) and *Calystegia soldanella* (D). Na⁺ was added in a form of NaCl. Data are means from three replicates for each concentration \pm SE. Asterisks of the respective color indicate statistically significant differences ($p < 0.05$) from control.

The trend of changes in the total osmotic value was similar to that for electrical conductivity; however, differences between plant organs were less pronounced (Figure 9). However, distribution of the osmotic value not associated with Na⁺, K⁺ and Cl⁻ (non-ionic osmotic value), under the effect of increasing soil salinity, showed both genotype- and organ-specific differences (Figure 10). Concentration of non-ionic osmolytes increased as a result of increased soil salinity in stems, leaves, roots and rhizomes of CSe1 (Figure 10A), stems and leaves of CSe2 (Figure 10B), all plant parts except roots of CSe3 (Figure 10C), and in leaf petioles of CSo, with a significant increase in all plant parts at the highest salinity (Figure 10D).

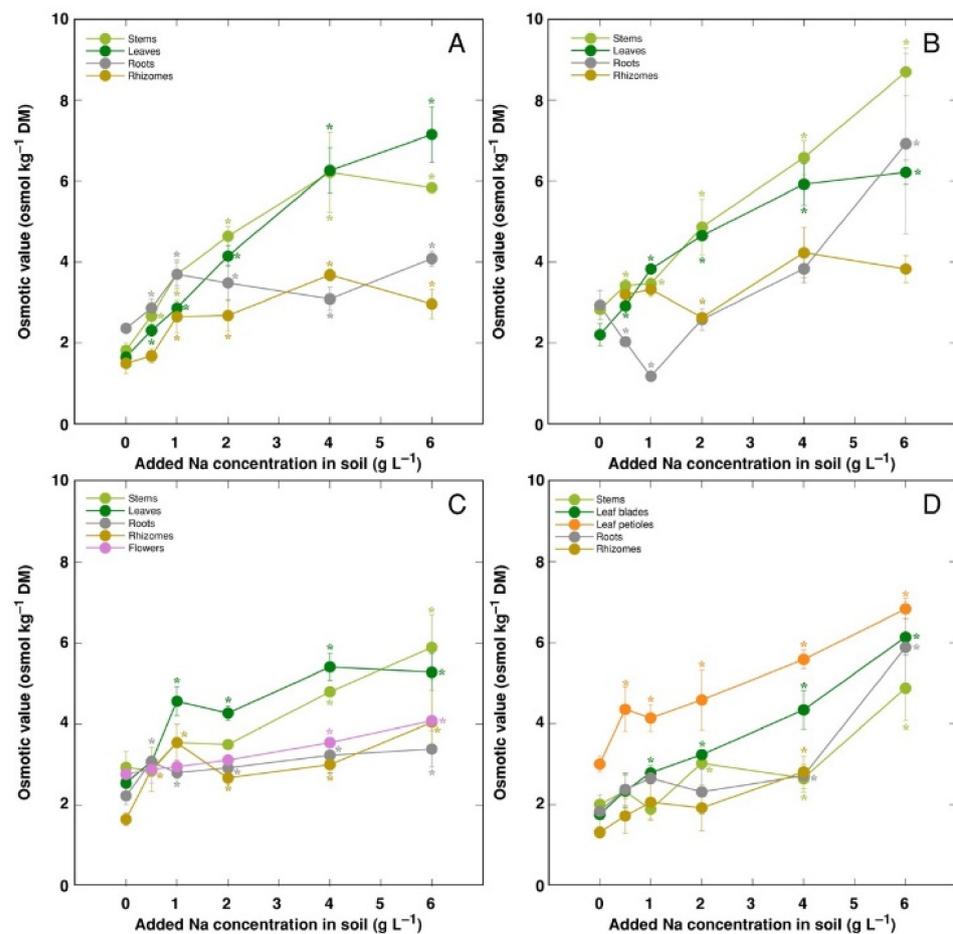


Figure 9. Changes in total osmotic value under the effect of increasing soil salinity in different parts of *Calystegia sepium* accessions CSe1 (A), CSe2 (B), CSe3 (C) and *Calystegia soldanella* (D). Na⁺ was added in a form of NaCl. Data are means from three replicates for each concentration \pm SE. Asterisks of the respective color indicate statistically significant differences ($p < 0.05$) from control.

There was a tight correlation between Na⁺ + K⁺ concentration and osmotic value (Figure 11A), Na⁺ + K⁺ concentration and electrical conductivity (Figure 11B) as well as electrical conductivity and osmotic value (Figure 11C) for all model plants ($p < 0.0001$). However, a shifted distribution of relationship between Na⁺ + K⁺ concentration and osmotic value was evident, reaching relatively higher osmotic activity at a lower ion concentration, indicating a higher contribution of non-ionic osmolytes in the total osmotic value (Figure 11A).

Multivariate analysis indicated relatively high variability of responses to increasing soil salinity between the tested model plants (Figure 12). In respect to morphological parameters (number of leaves, number of stems and rhizomes, length of stems and rhizomes), plants of coastal accession of *C. sepium* (CSe1) as well as plants of CSe2 growing in control conditions or at low salinity were very different from the others, but a certain similarity was evident between CSe1 and CSe2 and other genotypes at higher salinity (Figure 12A). The morphological parameters of CSe3 fully overlapped with these of CSo. Changes in biomass partitioning due to salinity treatment showed some overlap between CSe1 and CSe2, which was associated with increased partitioning in rhizomes by salinity (Figure 12B). The other two genotypes clearly separated from the previous two, with some overlap between CSe3 plants in control conditions or low salinity with CSo at high salinity. At the level of ion accumulation, Cse1 had some similarity with SCo, and these genotypes separated from the other two, which had similar responses (Figure 12C).

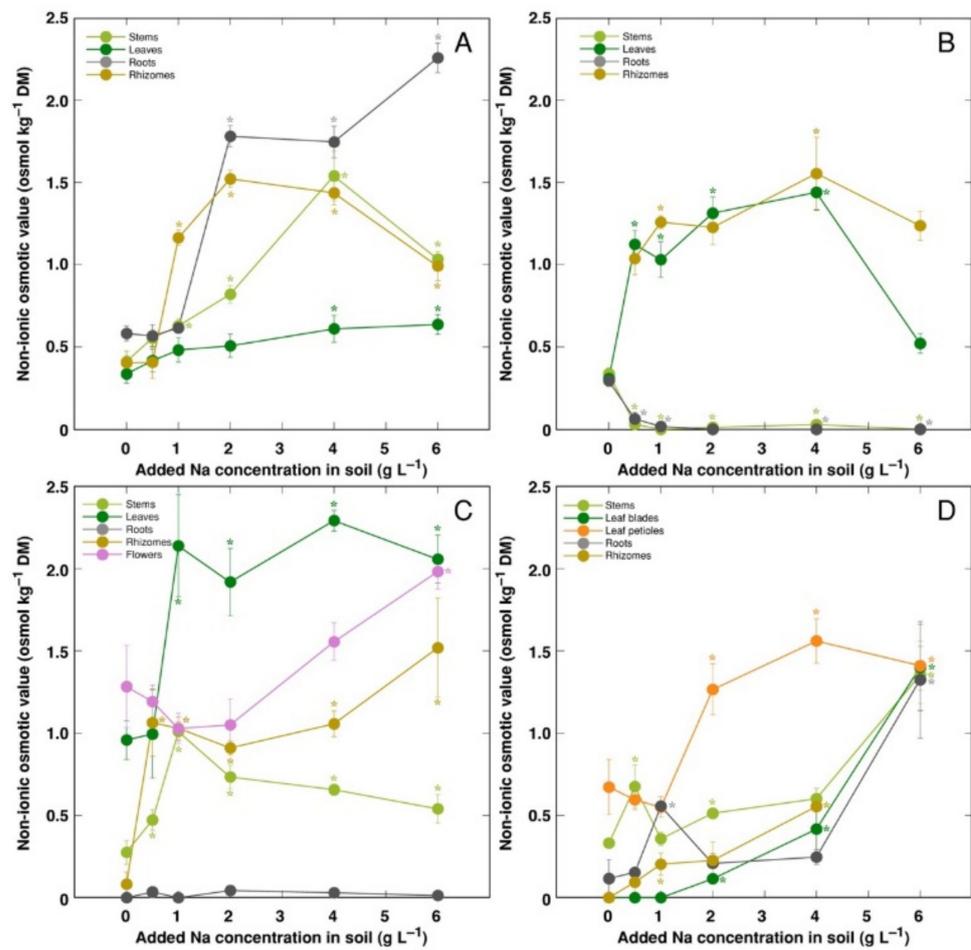


Figure 10. Changes in non-ionic osmotic value under the effect of increasing soil salinity in different parts of *Calystegia sepium* accessions CSe1 (A), CSe2 (B), CSe3 (C) and *Calystegia soldanella* (D). Na⁺ was added in a form of NaCl. Data are means from three replicates for each concentration ±SE. Asterisks of the respective color indicate statistically significant differences ($p < 0.05$) from control.

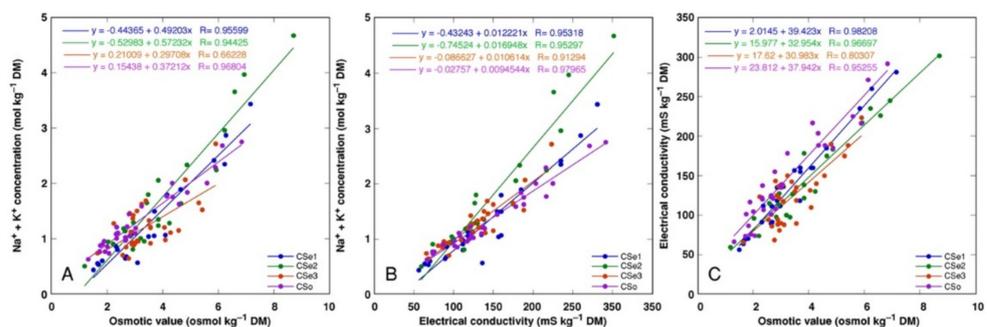


Figure 11. Correlation between Na⁺ + K⁺ concentration and osmotic value (A), Na⁺ + K⁺ concentration and electrical conductivity (B), and electrical conductivity and osmotic values (C) in different parts of *Calystegia sepium* accessions CSe1, CSe2, CSe3 and *Calystegia soldanella* (CSo).

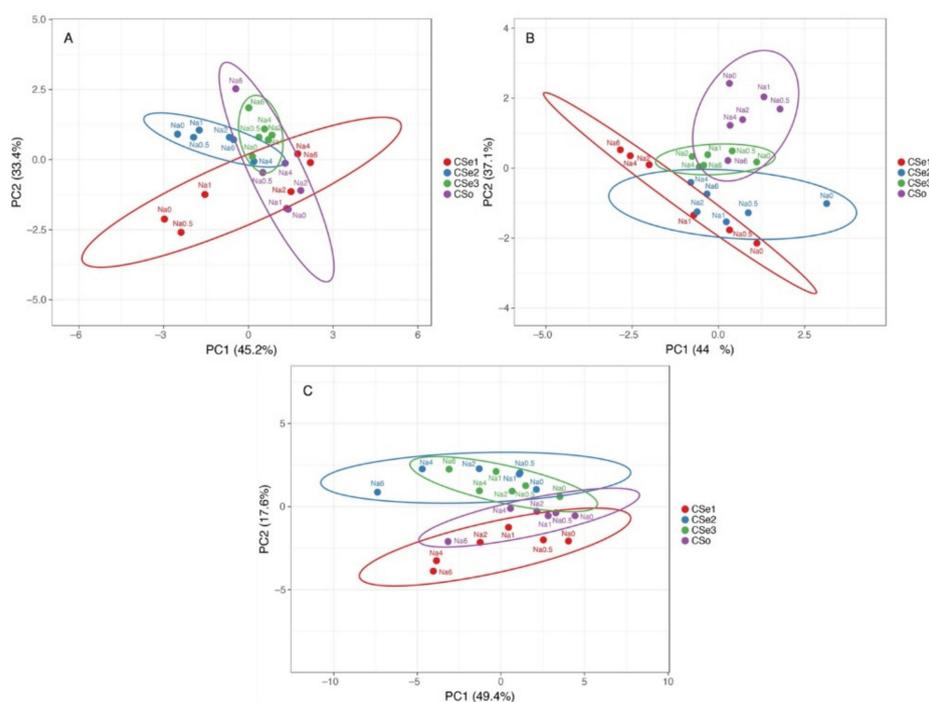


Figure 12. Principal component analysis on effect of increasing soil salinity on morphological parameters (A), biomass partitioning (B) and ion accumulation (C) in *Calystegia sepium* accessions CSe1, CSe2, CSe3 and *Calystegia soldanella* (CSo) plants. Prediction ellipses are such that with probability 0.95, a new observation from the same group will fall inside the ellipse. Unit variance scaling was applied to rows; singular value decomposition with imputation was used to calculate principal components.

4. Discussion

Usually, studies on plant tolerance mechanisms against salinity involve only single species. Due to differences in experimental conditions, results from these studies are difficult to compare directly. Another very large group of studies compare several halophytic species from a particular type of salt-affected habitat, with comparison of *Atriplex prostrata* and *Plantago coronopus* [29] or *Triglochin buchenauii*, *Bassia diffusa* and *Limonium linifolium* [30] as examples. Recently, taxonomically related plant species, including halophytes, have been compared in respect to their salinity tolerance, ion accumulation and maintenance of osmotic balance, in order to understand physiological responses crucial for differences in salinity tolerance in genetically allied species. Examples of this type of studies include but are not limited to species of *Plantago* [31,32], *Limonium* [33,34], *Silene* [35] and *Suaeda* [36]. However, studies comparing salinity responses of different accessions or cultivars within the same species are much less common [37–41].

In the present study, the three *C. sepium* accessions used as model plants for salinity tolerance and ion accumulation study were selected based on pronounced differences in their habitats (Table 1). The fourth model plant was coastal-specific taxonomically related species *C. soldanella*. It was initially hypothesized that *C. sepium* accession from a coastal salt-affected sandy beach (CSe1) will have characteristics similar to these in *C. soldanella* (CSo), but *C. sepium* from a dry grassland habitat (CSe3) will be more tolerant to salinity than *C. sepium* from a mesophytic habitat (CSe2). The first hypothesis was generally confirmed, as the relative changes in biomass accumulation of CSe1 and CSo were similar, but salinity tolerance of both plants was lower than that of the two other model plants (Figure 1B). In addition, CSe1 and CSo had similarities in ion accumulation characteristics (Figure 12C). However, the character of changes in biomass partitioning in response to salinity significantly differed between the two model plants (Figures 2 and 12B). Most importantly, salinity tolerance of the coastal accession of *C. sepium* and coastal-specific *C. soldanella* was less pronounced than that of the two inland

accessions of *C. sepium* (Figure 1). The second hypothesis was not confirmed, as CSe3 from a dry habitat was less salinity tolerant than CSe2 from a mesophytic habitat (Figure 1). Thus, the initial assumption for the relationship between salinity tolerance and habitat was not met, although there were significant differences between the tested accessions. In respect to total biomass accumulation, CSe2 from a mesophytic non-saline habitat had the highest absolute and relative salinity tolerance, but *C. soldanella* (CSo) had the lowest (Figure 1).

The results obtained in the present study are in apparent contradiction with several general assumptions concerning plant salinity tolerance and salinity responses that could be overgeneralizations or simplifications. First, it is often considered that salinity tolerance of a certain species or accession needs to be positively related to the actual salinity level in its habitat [42]. Second, it is widely regarded that any changes in plant growth due to increasing salinity are primarily related to the direct or indirect deleterious effect of substrate salt content or tissue Na⁺ accumulation on physiological processes, excluding the possibility of physiological regulation [43]. In respect to the first assumption, it can be valid if both glycophyte and halophyte species are considered, but for particular salt-adapted species, plants with higher salt resistance than the existing salinity level may grow in the particular habitat. It is necessary to remember that the salinity of the substrate is an extremely heterogeneous variable, subject to both spatial and temporal fluctuations [44,45]. It seems that *C. sepium* has a species-wide salinity tolerance; therefore, the accession CSe1 from a salt-affected coastal habitat does not represent an unique halophytic ecotype of *C. sepium*, as it was initially considered, as the accession CSe2 from a mesophytic inland habitat had higher salinity tolerance (Figure 1). In respect to the second assumption, responses to many single environmental factors have complex nature at physiological level, due to the fact that different signaling systems can be initiated, with various physiological responses having specific dose-response relationships, in addition to the direct or indirect deleterious effect on physiological processes [46]. In the present study, growth of *C. sepium* accession CSe3 was sensitive to low Na⁺ concentration, but further increase in salinity did not result in growth reduction (Figure 1). Similarly, the accession CSe2 even showed stimulation of leaf formation at high salinity, without further increase of their growth (Figure 3). Moreover, although growth of the accession CSe1 was most drastically inhibited by increasing salinity, biomass allocation to clonal underground structures, rhizomes, was stimulated (Figure 2A), showing a response similar to that in sand dune-adapted plant species as a result of sand accretion [47].

The most striking differences in physiological responses to salinity between accessions of *C. sepium* were found in respect to biomass partitioning. Being clonal species, *C. sepium* plants have a very high level of potential morphological plasticity [10]. In addition to long and branched rhizomes, which are present on both *C. sepium* and *C. soldanella*, the former species possesses an additional clonal growth system, represented by annual below-ground tubers on a distal part of the plagiotropic above-ground stems (runners), which are produced in autumn and readily penetrate soil [48]. Increased allocation to runners (stems) was evident for *C. sepium* plants in low nutrient conditions, but biomass allocation to rhizomes did not depend on nutrient availability [10]. In the present study, model plants showed prominent differences in biomass allocation patterns. Plants from coastal accession CSe1 allocated more biomass in rhizomes with increasing soil salinity, reaching more than 60% from the total biomass at the highest salinity (Figure 2A). Mesophytic accession CSe2 had similar features, but the rhizome share of the total biomass reached only 20% (Figure 2B). Surprisingly, the effect of increased allocation to rhizomes by salinity was not consistent in *C. soldanella*. From an ecological point of view, it is generally believed that clonal plants can escape unfavorable environmental conditions by placing new ramets in spots with more favorable conditions [49]. Biomass allocation to rhizomes could be seen as a manifestation of this escape response, as elongation of rhizomes makes it possible to reach soil areas with lower salinity, and this ability was most pronounced for the *C. sepium* accession from a salt-affected coastal habitat.

Apart from differences in morphological responses to salinity, *Calystegia* model plants in the present study also showed differences in ion accumulation, electrolytical activity

and osmotic values in plant parts. Organ specificity for accumulation of electrolytically and osmotically active ions was very high in both *Calystegium* species, with pronounced differences between accessions of *C. sepium*. Preferential accumulation of Na^+ in leaves is another characteristic feature of typical ion-accumulating halophytes. However, usually there is no correlation between a species tolerance to salinity and their Na^+ accumulation potential [50]. Only plants from the coastal accession of *C. sepium* (CSe1) accumulated Na^+ preferentially in leaves (Figure 5A), but the accumulation potential for Na^+ in leaves vs roots was relatively similar for *C. sepium* CSe2 (Figure 5B) and *C. soldanella* (Figure 5D). However, *C. sepium* CSe3 preferentially accumulated Na^+ in roots and stems (Figure 5C), showing typical salt excluder characteristics.

For coastal species growing in moderately saline soils, the Na^+ hyperaccumulation threshold range was proposed to be 18–30 g kg^{-1} [26]. According to this criterion, in the present study, *C. sepium* accession CSe1 reached the accumulation threshold in leaves at soil Na^+ concentration $>1 \text{ g L}^{-1}$ (Figure 5A), CSe2 at $>2 \text{ g Na}^+ \text{ L}^{-1}$ (Figure 5B), and *C. soldanella* only at $>3 \text{ g Na}^+ \text{ L}^{-1}$ (Figure 5C). However, *C. sepium* accession CSe3 did not reach the threshold concentration (Figure 5C). In natural conditions of coastal dunes, growing in sandy soil with relatively low Na^+ content, *C. soldanella* plants accumulated $7.1 \text{ g kg}^{-1} \text{ Na}^+$ in leaves [15]. In the present study, plants of coastal accession of *C. sepium* (CSe1), growing on seawater-affected saline sandy beach, accumulated $26.3 \text{ g kg}^{-1} \text{ Na}^+$ in leaves, but *C. sepium* plants from inland habitats had low levels of Na^+ (Table 2). Together with the results from the present experiments in controlled conditions, it shows that leaf Na^+ concentration for both *Calystegia* species is an indicator of substrate salinity.

K^+ is necessary for providing a certain ionic strength intensity inside plant tissues for supporting optimum protein interactions [51]; it also contributes to osmotic potential [52]. Due to a pronounced similarity in chemical properties between K^+ and Na^+ , Na^+ could contribute to both functions in Na^+ -accumulating halophytic species [53,54]. However, K^+ is retained in a cytoplasm, whereas Na^+ is sequestered in vacuole [55]. Differential localization in various leaf tissues of the two ions cannot be excluded: in addition to epidermis, where both ions are accumulated, K^+ can be found also in mesophyll cells [56]. As a result of increasing salinity, character of changes in K^+ concentration in *Calystegia* plants was organ-dependent and genotype-specific (Figure 6), and no general trend of changes in K^+ can be described. It is evident that, at the individual organ level, each model plant had a specific $\text{K}^+:\text{Na}^+$ concentration ratio, which decreased under increasing salinity, showing tight control of individual ion concentration (Figure 7). As based on a study in natural conditions comparing ion accumulation characteristics within and between coastal species, it was concluded that *C. sepium* is a tight regulator of electrical conductivity, proportionately adjusting both K^+ and Na^+ concentration to keep leaf tissue electrolytical activity as constant as possible, irrespective of soil concentration for these ions [26]. However, in controlled conditions, electrical conductivity gradually increased by increasing soil salinity both in stems and leaves of all model plants, but was relatively less affected in rhizomes and, to some extent, in roots (Figure 8). Consequently, the character of ion accumulation in *C. sepium*, irrespective of salinity level in the native habitat, and *C. soldanella* was at least partially similar to that typical for ion-accumulating halophytic species, accumulating high concentration of soluble ions in aboveground parts and excluding them from underground parts [54], but this characteristic was fully expressed only for coastal accession CSe1.

Initially, it was proposed that accumulation of organic osmotica in a form of compatible solutes is the main mechanism for osmotic adjustment [52]. Thus, amino acid proline has been suggested as one of the most universal and main osmoticum, especially in response to salinity [57]. However, more recently, the actual role of compatible solutes in maintaining osmotic balance has been seriously questioned, arguing that the bulk osmotic adjustment is provided by inorganic osmotica [58]. However, relative contribution to ionic species vs organic osmolytes in plants has been rarely assessed. Instead, concentration of particular osmotically active metabolite has been measured, and increase in its concentration due to salinity has been suggested as a proof of the role in osmotic adjustment [33,59]. In the present study, an attempt was made to distinguish between ionic and non-ionic osmotica,

subtracting from the total osmotic value the part which depended on Na^+ , K^+ and Cl^- . As a result, the major contribution of these inorganic ions in osmotic pool for all model plants under increasing soil salinity was clearly evident, with contribution of non-ionic solutes being only up to 25% of the total osmotic value. Although, high organ specificity of accumulation of osmotically active ions vs compatible solutes was seen. It has been shown earlier that Na^+ and Cl^- concentration did not increase by NaCl treatment in stolons of clonal plant *Hydrocotyle vulgaris*, indicating that osmotic balance was maintained by an increase in organic osmolyte concentration [60]. Similarly, for example, in coastal accession of *C. sepium* (CSe1), Na^+ concentration did not increase in roots and rhizomes under increasing soil salinity (Figure 5A), but significant part of osmotic value in these organs were associated with increased value of non-ionic osmotica (Figure 10A). Similar phenomenon was evident for leaves, flowers, rhizomes and stems of CSe3 (Figures 5C and 10C). As it has been experimentally tested that *C. soldanella* plants accumulate a considerable concentration of proline [15], accumulation of proline in *Calystegia* species in response to salinity could be one of the reasons for the increase in non-ionic osmotic activity found in the present study. In addition, increase in sucrose concentration is shown to be a characteristic response to salinity, as in *Trifolium repens* [50].

In general, there was a very tight correlation between osmotic value in tissues and electrical conductivity, indicating that mainly electrolytically active soluble ions were responsible for ensuring osmotic potential. Leaves of CSe3 accumulated the highest level of non-ionic osmotica as a result of NaCl treatment, and this was also reflected by a shifted distribution of relationship between $\text{Na}^+ + \text{K}^+$ concentration and osmotic value, reaching relatively higher osmotic activity at lower ion concentration (Figure 11A). Interestingly, *C. sepium* accession CSe1 from a salt-affected habitat was the only model plant showing increased water content in roots due to enhanced substrate salinity (Figure 4A), and this coincided with a significant increase in non-ionic osmotic value (Figure 10A).

As a basis for variability in physiological responses to salinity between taxonomically related model plants, as between *C. sepium* accessions from different habitats, both genetic and epigenetic differences can be considered. From the genetic side, coastal-specific ecotypes of several plant species appearing both in coastal and inland habitats have been described [50,61–63]. However, apart from local genetic adaptation, epigenetic control may have played a role in showing different physiological responses between different accessions. The term “ecological stress memory”, implying also epigenetic modifications, has been used to discuss plant responses to climate variability [64,65]; however, there is no evidence on general heritability of epigenetic changes after stress exposure [66,67]. Although, response to herbivores and pathogens are examples of transgenerational epigenetic adaptation [68]. It can be suggested, however, that the species with high degree of clonal propagation in the life history of their populations could be an exception [69]. Therefore, vegetatively propagated *C. sepium* accessions from different habitats represent an excellent model system for further studies aimed at dissecting control of adaptation mechanisms of clonal plant species to environmental conditions, especially, at the level of epigenetic changes.

5. Conclusions

As based on morphological, biomass partitioning and ion accumulation characteristics, all *C. sepium* accessions from different habitats represented varied phenotypes, possibly associated with their genetic differences. Salinity tolerance of both *C. sepium* and *C. soldanella* was relatively high, but the tolerance of particular accessions did not depend on substrate salinity level in their natural habitats. *C. sepium* accession from a mesophytic non-saline habitat was only slightly negatively affected by increasing substrate salinity. However, coastal accession of *C. sepium* and coastal-specific species *C. soldanella* had some similarities in ion accumulation characteristics. Pattern of accumulation of inorganic and organic osmotica in plants grown in saline soil showed high genotype- and organ-specificity. *C. sepium* accessions from different habitats can be suggested as models for further studies aiming

at dissecting possible genetic, epigenetic and physiological mechanisms of adaptation to heterogeneous environmental conditions.

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