

Article

Association Analysis of Genetic Variants of Sodium Taurocholate Co-Transporting Polypeptide NTCP Gene (*SLC10A1*) and HBV Infection Status in a Cohort of Egyptian Patients

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Abstract: Background: Single nucleotide polymorphisms (SNPs) in the *SLC10A1* gene, coding for a functional receptor of hepatitis B virus (HBV), sodium taurocholate co-transporting polypeptide (NTCP), may influence the susceptibility, outcome, and disease course of HBV infection in some populations. Aim: to determine the prevalence of SNPs of the NTCP gene, rs2296651 and rs943277, and their relationship with chronic HBV infection in a group of Egyptian patients. Methods: One hundred and thirty seven patients with HBV and 65 healthy controls were enrolled, and the patients were divided into two groups; group I chronic HBV infection (68 patients with normal ALT and minimal or no liver necroinflammation or fibrosis) and group II chronic hepatitis B (69 patients with elevated ALT and moderate or severe liver necroinflammation). They were subjected to full history taking, clinical examination, laboratory investigations, abdominal ultrasound, and liver stiffness measurement using both Echosens[®] Fibroscan and acoustic radiation force impulse (ARFI). A real time PCR TaqMan 5' allelic discrimination assay was applied to detect the SNPs in the NTCP gene, rs2296651 and rs943277. Results: On studying the rs2296651 variant, all controls and patients had genotype GG without any significant association with HBV infection or disease progression. However, the rs943277 variant in all controls and 98% of patients had genotype GA, except for two chronic HBV infection patients who had genotype AA, but no significant difference between patients and controls was found. The non-invasive methods for liver fibrosis assessment ARFI, AST/platelet's ratio (APRI), and fibrosis-4 score (FIB-4) could predict the stages of fibrosis in agreement with Fibroscan with AUCOR 0.8, 0.79, and 0.76, respectively. Conclusion: These findings may suggest that there is no relation between these SNPs of the NTCP gene and the susceptibility or chronicity of HBV infection in the Egyptian population. We also suggest that the use of the non-invasive methods for liver fibrosis assessment, ARFI, FIB-4, and APRI, may decrease the need for liver biopsies in the prediction of significant hepatic fibrosis in chronic HBV patients.

Keywords: functional receptor; hepatitis B virus; polymorphism; sodium taurocholate co-transporting polypeptide; hepatic fibrosis; Egypt

1. Introduction

Although effective hepatitis B virus (HBV) vaccines are in use worldwide, infection with HBV is still a serious global health problem with significant morbidity and mortality. In 2017, the World Health Organization (WHO) estimated that approximately 2 billion people have evidence of past or present infection with HBV [1]. A prevalence of 1.4% in Egypt is reported [2]. The number of HBV-related deaths due to complications of chronic HBV infection, such as liver cirrhosis and/or hepatocellular carcinoma (HCC), increased by 33% between 1990 and 2013 [3].

Different genome-wide association studies (GWAS) have identified genetic variants that are related to susceptibility to HBV infection and its complications like NTCP and human leucocytic antigen (HLA) genetic variants. One of the recent GWAS studies is the study done by Huang et al. that genotyped the 583,383 autosomal SNPs of 15,352 participants seropositive for HBV core antibodies in Taiwan Biobank and examined their associations with chronic HBV infection. They concluded that HLA class II variants are associated with chronicity after HBV acquisition [4].

Factors that can affect the persistence or clearance of HBV infection and its variable clinical outcomes include viral, environmental, and host factors [5]. Studying these factors allows an understanding of the viral life cycle, the natural history of the disease, and the development of new therapeutic agents aimed at curing HBV. One of the host factors is the genetic variations of *NTCP* receptor genes. HBV enters the hepatocytes via an entry receptor, sodium taurocholate co-transporting polypeptide (NTCP, also known as solute carrier family 10 member 1), which specifically interacts with the pre-S1 region of HBV [6]. This co-transporter, encoded by the *NTCP* (*SLC10A1*) gene, is highly expressed on the sinusoidal membranes, and plays a crucial role in bile duct enterohepatic circulation and regulating functions of the hepatocytes [7].

The relationship between *NTCP* polymorphisms and HBV infection, liver cirrhosis, or HCC is controversial, with some studies concluding that *NTCP* polymorphisms have been associated with resistance to HBV infection [8,9], while others have demonstrated that it promotes HBV infection [10–12]. Accordingly, it remains to be explored whether *NTCP* polymorphisms influence the susceptibility to HBV infection and the occurrence of liver cirrhosis or HCC.

The rs2296651 SNP is a non-synonymous G/A single nucleotide transition substitution, missense variant in the fourth exon of *SLC10A1* gene, located on Chr.14 [13]. The rs943277 SNP is a G/A single nucleotide transition substitution in the first intron of the *SLC10A1* gene. It has been reported that variants in the intron might also influence gene expression via regulating a transcription enhancer or silencer [14]. The prevalence of rs2296651 or rs943277 SNPs of the *NTCP/SLC10A1* gene is controversial in different geographic areas. To the best of our knowledge, there are no studies investigating the prevalence of rs2296651 or rs943277 SNPs of the *NTCP* gene in Egypt. Consequently, the present study was conducted in a group of Egyptian patients to determine the prevalence of these SNPs, and to explore if there is a possible relation between them and HBV infection status.

2. Materials and Methods

2.1. Study Subjects

The present study comprised 137 patients with HBV infection and 65 age-matched healthy volunteers as a control group recruited from Cairo Fatemic Hospital, Ministry of Health and Population (MOHP), and Cairo University Center for Hepatic Fibrosis (CUC-HF), Cairo University, between January 2019 and September 2019. All participants were Egyptians.

All patients were HBV treatment naïve with positive HBsAg for more than 6 months, detectable HBV DNA, and negative HCV, HIV, and autoimmune antibodies. All control subjects were HBV uninfected with negative serum HBsAg, HBeAg, anti-HBe, and anti-HBc with negative or positive anti-HBs in line with the history of hepatitis B vaccination. All control subjects had no history of hepatitis or evidence of hepatic fibrosis by Transient Elastography (Fibroscan[®], manufactured by Echosens[™] (Paris, France)).

The patients were divided into two clinical subgroups:

- Group I: chronic HBV infection ($n = 68$) characterized by normal ALT according to traditional cut-off values (ULN approximately 40 IU/L) and minimal or no liver necro-inflammation or fibrosis [15].
- Group II: chronic hepatitis B ($n = 69$) characterized by elevated ALT and moderate or severe liver necro-inflammation and accelerated progression of fibrosis [15].

The concept of the study was clearly explained to all participants who then provided informed consent. The study protocol conformed to the ethical guidelines of the declaration of Helsinki and was approved by National Committee for combating Viral Hepatitis (NCCVH) ethical committee.

2.2. Methods

All patients were subjected to full history taking, proper physical examination, basic laboratory investigations, hepatitis markers by EIA (HBsAg, HBeAg, anti-HBe, and HCV Antibodies), quantitative HBV DNA by real time PCR (with a lower limit of detection of 16 IU/mL), abdominal ultrasonography, and assessment of liver fibrosis by the following non-invasive methods:

- Indirect serum indices:

APRI was calculated using the formula $(AST/\text{upper limit of normal} \times 100)/\text{platelet count}$ [16]. FIB-4 was calculated using the formula $(\text{Age [years]} \times AST/\text{platelet count} \times ALT)$ [17]. AAR index was calculated using the formula $(AST/ALT \text{ ratio})$ [18].

- Liver stiffness measurement by:

(I) Transient elastography (TE) using a FibroScan[®] device (Echosens, Paris, France) [19]

(II) Acoustic Radiation Force Impulse (ARFI) elastography: using a Siemens ACUSON S3000 Ultrasound System (Siemens AG, Erlangen, Germany) [20]

Both Transient elastography (TE) and ARFI elastography were conducted at the Cairo University Center for Hepatic Fibrosis (CUC-HF); funded by the Science and Technology Development Fund (STDF) (5274 Center of excellence).

Since non-invasive measurements became widely available and, according to EASL guidelines for the management of HBV, Transient elastography (TE) is an acceptable tool for fibrosis assessment in the setting of chronic hepatitis B infection, and due to the limitations of using liver biopsy, tissue elastography was considered the standard diagnostic tool for the assessment of liver fibrosis [21].

All participants were subjected to determination of the *SLC10A1* gene SNPs rs2296651 and rs943277 by real time PCR TaqMan 5'.

Allelic discrimination assays in both patients and control subjects were conducted as described in [22]. Two milliliters of blood were put into a vacutainer containing ethylene diamine tetra-acetic acid (EDTA) for DNA extraction and analysis of *SLC10A1* gene polymorphisms. Total genomic DNA of patients and controls was extracted using Qiagen extraction kit (Cat. No.51104, QIAGEN, Valencia, CA 91355, USA). To detect the variants in this gene both SNPs rs2296651 and rs943277 kit of Cat. No: 4362691 and 4351379 (Applied Biosystems, Thermo Fisher Scientific, Foster City, CA 94404, USA) were used for real time PCR assay. The kit contained both forward and reverse primers in addition to probes for both wild and minor type alleles. The TaqMan universal PCR master mix contained AmpliTaq Gold[®] DNA polymerase, dNTPs, optimized buffer component and was purchased from Applied Biosystem.

Genotyping of the SNPs was carried out using the TaqMan genotyping assay on the Applied Biosystems 7300 Real-Time PCR system (Applied Biosystems, Thermo Fisher Scientific, Foster City, CA 94404, USA). Each Custom TaqMan[®] SNP Genotyping Assay consisted of a single tube containing two primers for amplifying the polymorphic sequence of interest, and two TaqMan[®] MGB probes for distinguishing between the two alleles. Each TaqMan MGB probe contained a reporter dye at the 5' end of each probe (VIC[®] dye is linked to the 5' end of the Allele 1 probe; FAM[™] dye is linked to the 5' end of the Allele 2 probe) and a non-fluorescent quencher (NFQ) at the 3' end of each probe.

During PCR, each TaqMan MGB probe anneals specifically to its complementary sequence between the forward and reverse primer sites. When the oligonucleotide probe is intact, the proximity of the reporter dye to the quencher dye results in quenching of the reporter fluorescence primarily by Förster-type energy transfer [23,24]. AmpliTaq Gold[®] DNA polymerase extends the primers bound to the template DNA. AmpliTaq Gold DNA polymerase cleaves only probes that are hybridized to the target. Cleavage separates the reporter dye from the quencher dye, which results in increased fluorescence by the reporter. The increase in the fluorescence signal occurs when probes that have hybridized to the complementary sequence are cleaved. Thus, the fluorescence signal generated by PCR amplification indicates which alleles are present in the sample. So, a substantial increase in VIC-dye fluorescence only indicates homozygosity for Allele 1, while a substantial increase in FAM-dye fluorescence only indicates homozygosity for Allele 2. An increase in both VIC- and FAM-dye fluorescence indicates Allele 1-Allele 2 heterozygosity. The rs2296651 and rs943277 genotyping assays were read at the PCR end point. The GG genotype was considered as the wild genotype, GA as the heterozygous genotype, and AA as the homogenous minor genotype.

2.3. Statistical Analysis

Epi-calc 2000 was used to calculate the sample size of this study. Assuming 80% power and 0.05 level of significance. The calculation was based on the results of NTCP SNPs studies carried out in different ethnic populations [25,26] and HBV mono-infection prevalence in Egypt (1.4%, low prevalence) [27].

Data were coded and entered using the statistical package SPSS (Statistical Package for the Social Sciences) version 23 and GraphPad Prism 7. Data were summarized using median, minimum, and maximum in quantitative data and using frequency (count) and relative frequency (percentage) for categorical data. Comparisons between quantitative variables were done using the non-parametric Kruskal–Wallis and Mann–Whitney tests. For comparing categorical data, a chi-square test was performed. An exact test was used instead when the expected frequency was less than five. Genotype and allele frequencies were compared between every two groups using chi-square tests. The odds ratio (OR) with 95% confidence intervals was calculated. The receiver operating characteristic (ROC) curve was used for the prediction of cut-off values. *p*-values less than 0.05 were considered statistically significant.

3. Results

The basic clinical and demographic data of the patients and controls are described in Table 1. Age and sex were well-matched between controls and patients, though ages were significantly higher in the group of patients with more advanced liver disease (*p* = 0.002 and 0.01 respectively). Among patients, a history of undergoing surgical operations or dental procedures were the main risk factors for the acquisition of HBV infection.

Regarding laboratory data, serum hepatic transaminases and total bilirubin were significantly higher in the patients group compared to the control group and increased with the progression of liver disease (*p* < 0.0001), while platelet count and albumin were significantly lower in the patients than in controls, and decreased with the progression of liver disease; HBV DNA levels were significantly higher in the CHB patients group

compared to the chronic HBV patients' group ($p < 0.0001$) (Table 2). From HBV markers, HBeAg was negative in 95% of patients.

Table 1. The basic characteristics of the patients.

Variable		Healthy Control (HC) $n = 65$	Patients Group I Chronic HBV Infection $n = 68$	Patients Group II Chronic Hepatitis B (CHB) $n = 69$	P1 Value	P2 Value
Age (years)	mean \pm SD	40 \pm 11	37	43	N.S	0.002
	min-max	23-75	22-62	23-68		
Sex	Male	39 (60%)	41 (60.3%)	55 (79.7%)	N.S	0.01
	Female	26 (40%)	27 (39.7%)	14 (20.3%)		
Residency	Rural	8 (12.3%)	11 (16.2%)	22 (31.9%)	0.01	N.S
	Urban	57 (87.7%)	57 (83.8%)	47 (68.1%)		
Comorbidities	DM	3 (4.6%)	5 (7.4%)	6 (8.7%)	N.S	N.S
	HTN	4 (6.2%)	3 (4.4%)	4 (5.8%)		
Manifestation of CLD	No	58 (89.2%)	59 (86.8%)	59 (85.5%)	-	0.02
	Yes	—	0 (0.0%)	6 (8.7%)		
BMI (kg/m^2)		28 \pm 3.5	27 \pm 3.4	27 \pm 2.6	0.04	N.S

Table 2. Laboratory data of the patients and controls.

Variable	HC $n = 65$	Patients Group I Chronic HBV Infection $n = 68$	Group II CHB $n = 69$	P1 Value	P2 Value
Hb (13-18 g/dL)	13 \pm 1.4	14 \pm 1.5	13 \pm 1.6	0.19 N.S	0.19 N.S
TLC ($4-11 \times 10^3/\text{mm}^3$)	6.2 \pm 1.8	6.5 \pm 1.6	6.1 \pm 1.7	0.3 N.S	0.3 N.S
PLT ($150-450 \times 10^3/\text{mm}^3$)	294 \pm 83	258 \pm 76	186 \pm 85	<0.0001	<0.0001
INR	1 \pm 0.06	1.1 \pm 0.1	1.1 \pm 0.18	<0.0001	<0.0001
ALT (up to 41 IU/L)	27 \pm 6.3	30 (11-78)	39 (13-114)	<0.0001	<0.0001
AST (up to 40 IU/L)	22 \pm 5.6	28 (10-50)	37 (16-152)	<0.0001	<0.0001
T. Bilirubin (0.5-1.2 mg/dL)	0.64 \pm 0.19	0.7 (0.3-1.4)	0.8 (0.2-3.1)	<0.0001	<0.0001
Albumin (3.4-5.4 g/dL)	4.5 \pm 0.58	4.3 (3.5-5.7)	4 (2.9-5.5)	<0.0001	<0.0001
AFP (up to 10 ng/mL)	—	4 (0.4-20)	4.8 (0-59)	0.7 N.S	0.7 N.S
Creat (0.6-1.2 mg/dL)	0.62 \pm 0.2	0.8 (0.3-3.9)	0.8 (0.3-2)	0.0004	N.S
HBV PCR ($\times 10^3$ IU/mL)	—	1.446 (0.198-5298.090)	5.030 (0.074-170,000)	<0.0001	<0.0001
HBeAg	+Ve	—	1 (1.5%)	-	0.03
	-Ve	—	67 (98.5%)	64 (92.8%)	

Among all studied patient groups, the degree of fibrosis was absent or mild in 49.64% (47 patients F0, 6 patients F0-F1, 15 patients F1), and significant in 50.36% (32 patients F2-3, 17 patients F3, 11 patients F3-F4, 9 patients F4) using Fibroscan (Echosens®).

3.1. NTCP rs2296651 Variants among the Studied Groups

All studied subjects had genotype GG of SNP rs2296651 of the NTCP gene. Accordingly, it did not show a statistically significant difference between patients and controls.

3.2. NTCP rs943277 Variants among the Studied Groups

The heterogeneous mutant genotype GA of SNP rs943277 of NTCP gene was the most prevalent genotype among the controls (100%) and patients (98.5%), with only two patients from the chronic HBV infection group having the homogenous mutant genotype AA. None of the genotypes showed a statistically significant difference neither among patients' groups nor between different patient groups and controls (Tables 3 and 4).

Table 3. NTCP rs943277 variant among the studied groups.

Variable		HC <i>n</i> = 65	Group I Chronic HBV Infection <i>n</i> = 68	Group II CHB <i>n</i> = 69	<i>p</i> Value
rs943277 variant	GG	0 (0.0%)	0 (0.0%)	0 (0.0%)	0.13
	GA	65 (100%)	66 (97%)	69 (100%)	
	AA	0 (0.0%)	2 (3%)	0 (0.0%)	
	Allele G	65 (50%)	66 (48.52%)	69 (50%)	0.91
	Allele A	65 (50%)	70 (51.47%)	69 (50%)	

Table 4. NTCP rs943277 variant among patient groups versus controls.

		Group I against HC		Group II against HC	
		<i>p</i> Value	OR (95%CI)	<i>p</i> Value	OR (95%CI)
rs943277 variant	GG	REF	REF	REF	REF
	GA + AA	0.49	Infin	>0.99	1
	Allele G	0.9	1.06	>0.99	1
	Allele A		(0.665–1.72)		(0.619–1.61)

4. Discussion

Single nucleotide polymorphisms (SNPs) in the *SLC10A1* gene, coding for sodium taurocholate co-transporting polypeptide (NTCP) functional receptor of HBV, has been associated with the natural history of HBV infection in some populations.

Based on the genome Aggregation Database (genomAD), the minor allele frequency of rs2269951 SNP is A = 0.005 in the global population, A = 0.095 in East Asian, A = 0.002 in African, A = 0.002 in American, and A = 0.00005 in European populations (SNPs with a minor allele frequency of 0.05 (5%) or greater are considered a common variant in this population) [13]. According to a 1000 genomes phase 3 combined population database, the minor allele frequency of rs943277 SNP is A = 0.078 in the global population, A = 0.003 in East Asians, A = 0.03 in South Asians, A = 0.007 in Africans, A = 0.25 in Americans, and A = 0.174 in Europeans [28].

However, the genetic mutants of *NTCP* and their relation to HBV infection have not yet been studied in the Egyptian population. The fundamental target of the present study was to determine the prevalence of SNPs of the *NTCP* gene, rs2296651 and rs943277, in Egyptians, to explore the possible association between these SNPs and HBV susceptibility in Egyptian patients, and to correlate their presence, if any, to disease progression.

Surprisingly, we detected only wild type GG genotype in all our studied Egyptian participants, and none of them presented the mutant GA or AA genotypes, with no difference between groups. Our results were similar to those of other populations such as Moroccan [29], Spanish Caucasian [30], Polish Caucasian [31], European American, and African American [32] populations, where the rs2296651 mutant genotypes were not detected in any of their participants.

In contrast to our results, several previous studies in the Chinese Han population showed that rs2296651 mutant variants were inversely correlated with HBV susceptibility, where GA and AA genotypes had lower frequencies in patients with CHB compared with healthy controls and were demonstrated to be protective, reducing the risk of liver cirrhosis, hepatocellular carcinoma, and liver failure in CHB patients [9,25,33–35]. Some studies conducted in other Asian populations, such as Taiwanese [8,36], Vietnamese [37], Korean [38], and Taiwanese [22] populations, reported that the *SLC10A1* A allele consistently decreased HBV infection risk compared with the G allele. The possible protective role of rs2296651 polymorphism was also demonstrated in a meta-analysis that extracted eight studies, including 14,591 chronically HBV-infected patients and 12,396 healthy controls from different populations (Chinese, Taiwanese, and Moroccan). It concluded that the mutant variant

was inversely associated with the risk of HBV infection (OR = 0.593, $p = 0.028$), where the A allele and GA genotype frequencies were lower in the CHB group compared with HCs group [26].

However, there were conflicting data in Asian populations that the rs2296651 mutant variant may be associated with increased susceptibility to HBV infection [9] or may not be a risk factor for HBV infection at all [10,11,39]. These rather contradictory results suggest that the rs2296651 variant varies among different ethnic populations and may be specific to Asian populations, and that it might display its advantage in conferring resistance to HBV infection.

On the other hand, our study population presented the heterogeneous mutant GA genotype of rs943277 SNP of *NTCP* gene, except for only two patients from the chronic HBV infection group that had the homogenous mutant AA genotype, with no statistically significant difference found between the studied groups.

SNP rs943277 of the *NTCP* gene was studied in a group of Chinese Hans (1232 HC and 2453 CHB patients) by Wang et al. in 2017 [26] and found that the wild GG genotype was the most prevalent among HC (99.3%) and CHB patients (98.4%). While the heterogeneous mutant GA genotype was 0.7% and 1.6% in HC and CHB patients, respectively, with only one CHB patient showing the homogenous mutant AA genotype, this difference in GA genotype was statistically significantly, being higher in CHB patients than in HC ($p = 0.014$), and was revealed to be a potential hazardous mutant, facilitating susceptibility to HBV infection.

With reference to the basic data results of our study, the mean age of our patients was 44 ± 13 years, which matches a previous Egyptian study conducted by Ismail et al. in 2017, who showed a similar HBV infection risk peak among those aged 35–44 [2]. There was a significant increase in age of patients with a higher degree of liver disease progression, which is consistent with Li et al.'s study in 2017, which found that the older the age, the higher the stage of fibrosis [40]. This could be explained by the longer duration of exposure to HBV infection [41]. Male predominance was obvious in our study (70%), matching another Egyptian study by Maklad et al. in 2018 (with a male: female ratio of about 5:1), which may be due to more exposure to risk factors of viral acquisition among males [42].

Fortuitously, our study showed interesting findings regarding the results of non-invasive methods for hepatic fibrosis measurement, including ARFI, APRI, and FIB-4 compared to Fibroscan. The non-invasive serum biomarkers for predicting liver fibrosis APRI and FIB-4 were significantly higher in the CHB patient group compared to the chronic HBV infection group, while AAR did not show a significant difference between both groups. Zhang et al. in 2016 [43] also reported that the mean values of FIB-4 and APRI in patients with HBV infection were significantly higher for each successive fibrosis stage ($p < 0.05$), but there were no differences between successive fibrosis stages for AAR. To discriminate between significant and non-significant fibrosis by serum APRI, we found the cut-off value of 0.34 to have sensitivity of 74% and specificity of 71%, with AUC 0.79. This is in accordance with the findings of Yue et al. in 2019 [44] who proposed 0.35 as a cut off for significant fibrosis ($F \geq 2$) in patients with CHB with sensitivity 78% and specificity 72%. For serum FIB-4, we found the cut off value of 0.83 to have a sensitivity of 74% and specificity of 60% with AUC 0.76. The findings suggested by Ucar et al. in 2013 [45] were 0.68 AUC of FIB-4 for a cut-off value of 1.08 with sensitivity 70.7% and specificity 62.5% in CHB patients.

In addition, our study showed that mean values of the LSM by ARFI elastography for predicting liver fibrosis were significantly higher in the CHB patient group compared to the chronic HBV infection group. Another study by Tseng and colleagues in 2018 [46] concluded that the value of the ARFI measurement increased with the severity of liver fibrosis and found a significant correlation between fibrosis stage and ARFI ($p = 0.0001$), APRI ($p = 0.012$), and FIB-4 ($p = 0.004$). However, LSM should be interpreted with caution among patients with elevated ALT and should not be used in patients with extensive liver necro-inflammation (ALT levels $>10 \times$ ULN). Transient elastography also has limited use in

hepatic congestion, cholestasis, obese patients, and patients with ascites [21]. Furthermore, in our work, ARFI achieved AUC = 0.8 with p value < 0.0001 for cut-off value 1.3 m/s in the prediction of significant fibrosis with sensitivity 80% and specificity 74%. These results agree with the results of previous studies conducted by Friedrich-Rust et al. in 2012 [47] who proposed 1.34 m/s as a cut-off for the diagnosis of significant fibrosis, with AUC = 0.75, sensitivity 81.0% and specificity 64.2%.

This study had some limitations. Firstly, one limitation of our study is the sample size. Clearly 202 participants are not enough to generalize the results over our population or ethnic group. However, from this number of participants a clear idea was formed about the prevalence of the studied NTCP SNPs and whether they are related to HBV infection in our population. Secondly, although the study of limited number of NTCP SNPs here is considered another pitfall which can miss other important mutant sites, rs2296651 was one of the most studied NTCP SNPs in patients with HBV infection in the literature. It would be better to conduct whole NTCP gene sequencing to detect the possible SNPs related to HBV infection and disease. The third limitation is the distribution of the minor allele of rs943277 in our study which is not consistent with the Hardy–Weinberg equilibrium in healthy control and patient groups. However, the prevalence of rs943277 in different populations and its role in HBV infection is still poorly understood due to limited baseline research on this SNP. Further comprehensive studies using large scale multicenter populations of varying ethnic origins, with different outcomes of infection and therapeutic schedules, are needed to determine the prevalence of NTCP polymorphisms and their exact role in the pathogenesis of HBV infection.

5. Conclusions

Based on the results of the current study, we can conclude that the rs2296651 variant within the *NTCP* gene is not prevalent in the studied group of the Egyptian population, with a subsequent absence of relation to susceptibility to HBV infection or disease progression which could be related to our ethnic group. Genotype GA of the rs943277 variant within the *NTCP* gene was the prevalent genotype among our studied group of Egyptian healthy controls and 98% of patients, but no association was found between this variant and HBV infection susceptibility or disease progression.

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Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Ethics Committee of Cairo Fatemic Hospital and Faculty of Medicine, Cairo University (protocol code I-170318, date of approval July 2018).

Informed Consent Statement: Written informed consent has been obtained from all subjects involved in the study to use their clinical, investigational data for research and subsequent publication of research results.

Data Availability Statement: All data we have already presented in our manuscript, the supporting data will be found at data bank such as Medline, PubMed, Scopus, and web of sciences.

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References

1. World Health Organization. *WHO Guidelines on Hepatitis B and C Testing*; World Health Organization: Geneva, Switzerland, 2017; Volume 204.
2. Ismail, S.A.; Cuadros, D.F.; Benova, L. Hepatitis B in Egypt: A cross-sectional analysis of prevalence and risk factors for active infection from a nationwide survey. *Liver Int.* **2017**, *37*, 1814–1822. [[CrossRef](#)] [[PubMed](#)]
3. Stanaway, J.D.; Flaxman, A.D.; Naghavi, M.; Fitzmaurice, C.; Vos, T.; Abubakar, I.; Abu-Raddad, L.J.; Assadi, R.; Bhala, N.; Cowie, B.; et al. The global burden of viral hepatitis from 1990 to 2013: Findings from the Global Burden of Disease Study 2013. *Lancet* **2016**, *388*, 1081–1088. [[PubMed](#)]
4. Huang, Y.-H.; Liao, S.-F.; Khor, S.-S.; Lin, Y.-J.; Chen, H.-Y.; Chang, Y.-H.; Huang, Y.-H.; Lu, S.-N.; Lee, H.-W.; Ko, W.-Y.; et al. Large-scale genome-wide association study identifies HLA class II variants associated with chronic HBV infection: A study from Taiwan Biobank. *Aliment. Pharmacol. Ther.* **2020**, *52*, 682–691. [[CrossRef](#)] [[PubMed](#)]
5. Huang, Y.H.; Liao, S.F.; Khor, S.S.; Lin, Y.J.; Chen, H.Y.; Chang, Y.H.; Huang, Y.H.; Lu, S.N.; Lee, H.W.; Ko, W.Y.; et al. New loci associated with chronic hepatitis B virus infection in Han Chinese. *Nat. Genet.* **2013**, *45*, 1499–1503.
6. Ni, Y.; Lempp, F.A.; Mehrle, S.; Nkongolo, S.; Kaufman, C.; Fälth, M.; Stindt, J.; Königer, C.; Nassal, M.; Kubitz, R.; et al. Hepatitis B and D viruses exploit sodium taurocholate co-transporting polypeptide for species-specific entry into hepatocytes. *Gastroenterology* **2014**, *146*, 1070–1083. [[CrossRef](#)] [[PubMed](#)]
7. Seeger, C.; Mason, W.S. Sodium-dependent taurocholic cotransporting polypeptide: A candidate receptor for human hepatitis B virus. *Gut* **2013**, *62*, 1093–1095. [[CrossRef](#)]
8. Hu, H.H.; Liu, J.; Lin, Y.L.; Luo, W.S.; Chu, Y.J.; Chang, C.L.; Jen, C.L.; Lee, M.H.; Lu, S.N.; Wang, L.Y.; et al. The rs2296651 (S267F) variant on NTCP (*SLC10A1*) is inversely associated with chronic hepatitis B and progression to cirrhosis and hepatocellular carcinoma in patients with chronic hepatitis B. *Gut* **2016**, *65*, 1514–1521. [[CrossRef](#)]
9. Peng, L.; Zhao, Q.; Li, Q.; Li, M.X.; Li, C.; Xu, T.; Jing, X.; Zhu, X.; Wang, Y.; Li, F.; et al. The p.Ser267Phe variant in *SLC10A1* is associated with resistance to chronic hepatitis B. *Hepatology* **2015**, *61*, 1251–1260. [[CrossRef](#)]
10. Li, N.; Zhang, P.; Yang, C.; Zhu, Q.; Li, Z.; Li, F.; Han, Q.; Wang, Y.; Lv, Y.; Wei, P.; et al. Association of genetic variation of sodium taurocholate cotransporting polypeptide with chronic Hepatitis B virus infection. *Genet. Test. Mol. Biomark.* **2014**, *18*, 425–429. [[CrossRef](#)]
11. Zhang, Y.; Li, Y.; Wu, M.T.; Cao, P.; Liu, X.; Ren, Q.; Zhai, Y.; Xie, B.; Hu, Y.; Hu, Z.; et al. Comprehensive assessment showed no associations of variants at the *SLC10A1* locus with susceptibility to persistent HBV infection among Southern Chinese. *Sci. Rep.* **2017**, *7*, 46490. [[CrossRef](#)]
12. Yang, J.; Yang, Y.Y.; Xia, M.; Wang, L.; Zhou, W.; Yang, Y.; Jiang, Y.; Wang, H.; Qian, J.; Jin, L.; et al. A genetic variant of the NTCP gene is associated with HBV infection status in a Chinese population. *BMC Cancer* **2016**, *16*, 211. [[CrossRef](#)]
13. rs2296651 RefSNP Report—dbSNP—NCBI. Available online: <https://www.ncbi.nlm.nih.gov/snp/rs2296651> (accessed on 20 January 2020).
14. Tokuhira, S.; Yamada, R.; Chang, X.; Suzuki, A.; Kochi, Y.; Sawada, T.; Suzuki, M.; Nagasaki, M.; Ohtsuki, M.; Ono, M.; et al. An intronic SNP in a RUNX1 binding site of *SLC22A4*, encoding an organic cation transporter, is associated with rheumatoid arthritis. *Nat. Genet.* **2003**, *35*, 341–348. [[CrossRef](#)]
15. Papatheodoridis, G.; Buti, M.; Cornberg, M.; Janssen, H.; Mutimer, D.; Pol, S.; Raimondo, G.; Dusheiko, G.; Lok, A.; Marcellin, P. EASL clinical practice guidelines: Management of chronic hepatitis B virus infection. *J. Hepatol.* **2012**, *57*, 167–185.
16. Wai, C.T.; Greenon, J.K.; Fontana, R.J.; Kalbfleisch, J.D.; Marrero, J.A.; Conjeevaram, H.S.; Lok, A.S.F. A simple noninvasive index can predict both significant fibrosis and cirrhosis in patients with chronic hepatitis C. *Hepatology* **2003**, *38*, 518–526. [[CrossRef](#)] [[PubMed](#)]
17. Vallet-Pichard, A.; Mallet, V.; Nalpas, B.; Verkarre, V.; Nalpas, A.; Dhalluin-Venier, V.; Fontaine, H.; Pol, S. FIB-4: An inexpensive and accurate marker of fibrosis in HCV infection. Comparison with liver biopsy and FibroTest. *Hepatology* **2007**, *46*, 32–36. [[CrossRef](#)] [[PubMed](#)]
18. Kim, S.U.; Park, J.Y.; Kim, D.Y.; Ahn, S.H.; Choi, E.H.; Seok, J.Y.; Lee, J.M.; Park, Y.N.; Chon, C.Y.; Han, K.H. Non-invasive assessment of changes in liver fibrosis via liver stiffness measurement in patients with chronic hepatitis B: Impact of antiviral treatment on fibrosis regression. *Hepatol. Int.* **2010**, *4*, 673–680. [[CrossRef](#)] [[PubMed](#)]
19. Castera, L.; Forns, X.; Alberti, A. Non-invasive evaluation of liver fibrosis using transient elastography. *J. Hepatol.* **2008**, *48*, 835–847. [[CrossRef](#)] [[PubMed](#)]
20. Fierbinteanu-Braticевич, C.; Andronescu, D.; Usvat, R.; Cretoiu, D.; Baicus, C.; Marinocchi, G. Acoustic radiation force imaging sonoelastography for noninvasive staging of liver fibrosis. *World J. Gastroenterol.* **2009**, *15*, 5525–5532. [[CrossRef](#)]
21. Castera, L.; Yuen Chan, H.L.; Arrese, M.; Afdhal, N.; Bedossa, P.; Friedrich-Rust, M.; Han, K.-H.; Pinzani, M. EASL-ALEH Clinical Practice Guidelines: Non-invasive tests for evaluation of liver disease severity and prognosis. *J. Hepatol.* **2015**, *63*, 237–264.
22. Chuaypen, N.; Tuyapala, N.; Pinjaroen, N.; Payungporn, S.; Tangkijvanich, P. Association of NTCP polymorphisms with clinical outcome of hepatitis B infection in Thai individuals. *BMC Med. Genet.* **2019**, *20*, 87. [[CrossRef](#)]
23. Förster, T. Zwischenmolekulare Energiewanderung und Fluoreszenz. *Ann. Phys.* **1948**, *437*, 55–75. [[CrossRef](#)]
24. Lakowicz, J.R. Energy Transfer. In *Principles of Fluorescence Spectroscopy*; Springer: New York, NY, USA, 1983; pp. 367–394.
25. Wu, W.; Zeng, Y.; Lin, J.; Wu, Y.; Chen, T.; Xun, Z.; Ou, Q. Genetic variants in NTCP exon gene are associated with HBV infection status in a Chinese Han population. *Hepatol. Res.* **2018**, *48*, 364–372. [[CrossRef](#)] [[PubMed](#)]

26. Wang, P.; Mo, R.; Lai, R.; Xu, Y.; Lu, J.; Zhao, G.; Liu, Y.; Cao, Z.; Wang, X.; Li, Z.; et al. Genetic variations of NTCP are associated with susceptibility to HBV infection and related hepatocellular carcinoma. *Oncotarget* **2017**, *8*, 105407–105424. [CrossRef]
27. Zampino, R.; Boemio, A.; Sagnelli, C.; Alessio, L.; Adinolfi, L.E.; Sagnelli, E.; Coppola, N. Hepatitis B virus burden in developing countries. *World J. Gastroenterol.* **2015**, *21*, 11941–11953. [CrossRef]
28. rs943277 RefSNP Report—dbSNP—NCBI. Available online: https://www.ncbi.nlm.nih.gov/snp/rs943277#frequency_tab (accessed on 20 January 2020).
29. Ezzikouri, S.; Chihab, H.; Elhabazi, A.; Wakrim, L.; Benjelloun, S. Lack of Ser267Phe variant of sodium taurocholate cotransporting polypeptide among Moroccans regardless of hepatitis B virus infection status. *BMC Infect. Dis.* **2017**, *17*, 99. [CrossRef]
30. Casillas, R.; Tabernero, D.; Gregori, J.; Belmonte, I.; Cortese, M.F.; Gonzalez, C.; Riveiro-Barciela, M.; López, R.M.; Quer, J.; Esteban, R.; et al. Analysis of hepatitis B virus preS1 variability and prevalence of the rs2296651 polymorphism in a Spanish population. *World J. Gastroenterol.* **2018**, *24*, 680–692. [CrossRef]
31. Rybicka, M.; Woziwodzka, A.; Romanowski, T.; Sznarkowska, A.; Stalke, P.; Dęrczewski, M.; Bielawski, K.P. Host genetic background affects the course of infection and treatment response in patients with chronic hepatitis B. *J. Clin. Virol.* **2019**, *120*, 1–5. [CrossRef]
32. Ho, R.H.; Leake, B.F.; Roberts, R.L.; Lee, W.; Kim, R.B. Ethnicity-dependent Polymorphism in Na⁺-taurocholate Cotransporting Polypeptide (*SLC10A1*) Reveals a Domain Critical for Bile Acid Substrate Recognition. *J. Biol. Chem.* **2004**, *279*, 7213–7222. [CrossRef]
33. Wang, D.; Zhang, P.; Zhang, M. Predictors for advanced liver fibrosis in chronic hepatitis b virus infection with persistently normal or mildly elevated alanine aminotransferase. *Exp. Ther. Med.* **2017**, *14*, 5363–5370. [CrossRef] [PubMed]
34. An, P.; Zeng, Z.; Winkler, C.A. The loss-of-function S267F variant in HBV receptor NTCP reduces human risk for HBV infection and disease progression. *J. Infect. Dis.* **2018**, *218*, 1404–1410. [CrossRef] [PubMed]
35. Yang, F.; Wu, L.; Xu, W.; Liu, Y.; Zhen, L.; Ning, G.; Song, J.; Jiao, Q.; Zheng, Y.; Chen, T.; et al. Diverse effects of the ntcp p.ser267phe variant on disease progression during chronic hbv infection and on hbv pres1 variability. *Front. Cell Infect. Microbiol.* **2019**, *9*, 18. [CrossRef] [PubMed]
36. Nfor, O.N.; Wu, M.F.; Debnath, T.; Lee, C.T.; Lee, W.; Liu, W.H.; Tantoh, D.M.; Hsu, S.Y.; Liaw, Y.P. Hepatitis B virus infection in Taiwan: The role of NTCP rs2296651 variant in relation to sex. *J. Viral Hepat.* **2018**, *25*, 1116–1120. [CrossRef] [PubMed]
37. Binh, M.T.; Hoan, N.X.; Van Tong, H.; Sy, B.T.; Trung, N.T.; Bock, C.T.; Toan, N.L.; Bang, M.H.; Meyer, C.G.; Kremsner, P.G.; et al. NTCP S267F variant associates with decreased susceptibility to HBV and HDV infection and decelerated progression of related liver diseases. *Int. J. Infect. Dis.* **2019**, *80*, 147–152. [CrossRef]
38. Lee, H.W.; Park, H.J.; Jin, B.; Dezhbord, M.; Kim, D.Y.; Han, K.H.; Ryu, W.S.; Kim, S.; Ahn, S.H. Effect of S267F variant of NTCP on the patients with chronic hepatitis B. *Sci. Rep.* **2017**, *7*, 17634. [CrossRef]
39. Su, Z.; Li, Y.; Liao, Y.; Cai, B.; Chen, J.; Zhang, J.; Li, L.; Ying, B.; Tao, C.; Zhao, M.; et al. Polymorphisms in sodium taurocholate cotransporting polypeptide are not associated with hepatitis B virus clearance in Chinese Tibetans and Uygurs. *Infect. Genet. Evol.* **2016**, *41*, 128–134. [CrossRef]
40. Li, Q.; Lu, C.; Li, W.; Huang, Y.; Chen, L. The independent predictors of liver histological changes in chronic hepatitis B virus infection patients with persistently high-normal or low-normal alanine transaminase levels. *Discov. Med.* **2017**, *23*, 19–25.
41. Abdo, A.A.; Bzeizi, K.I.; Babatin, M.A.; Al Sohaibani, F.; Al Mana, H.; Alsaad, K.O.; AlGhamdi, H.; Al-Hamoudi, W.; AlSwat, K.; AlFaleh, F.Z.; et al. Predictors of significant fibrosis in chronic hepatitis b patients with low viremia. *J. Clin. Gastroenterol.* **2014**, *48*, e50–e56. [CrossRef]
42. Maklad, S.; Reyad, E.M.; William, E.A.; Abouzeid, A. Efficacy and Safety of Entecavir 0.5 mg in Treating Naive Chronic Hepatitis B Virus Patients in Egypt: Five Years of Real Life Experience. *Gastroenterol. Res.* **2018**, *11*, 138–144. [CrossRef]
43. Zhang, Z.; Wang, G.; Kang, K.; Wu, G.; Wang, P. The diagnostic accuracy and clinical utility of three noninvasive models for predicting liver fibrosis in patients with HBV infection. *PLoS ONE* **2016**, *11*, e0152757. [CrossRef] [PubMed]
44. Yue, W.; Li, Y.; Geng, J.; Wang, P.; Zhang, L. Aspartate aminotransferase to platelet ratio can reduce the need for transient elastography in Chinese patients with chronic hepatitis B. *Medicine* **2019**, *98*, e18038. [CrossRef]
45. Ucar, F.; Sezer, S.; Ginis, Z.; Ozturk, G.; Albayrak, A.; Basar, O.; Ekiz, F.; Coban, S.; Yuksel, O.; Armutcu, F.; et al. APRI, the FIB-4 score, and Forn's index have noninvasive diagnostic value for liver fibrosis in patients with chronic hepatitis B. *Eur. J. Gastroenterol. Hepatol.* **2013**, *25*, 1076–1081. [CrossRef]
46. Tseng, C.H.; Chang, C.Y.; Mo, L.R.; Lin, J.T.; Tai, C.M.; Perng, D.S.; Lin, C.W.; Hsu, Y.C. Acoustic radiation force impulse elastography with APRI and FIB-4 to identify significant liver fibrosis in chronic hepatitis B patients. *Ann. Hepatol.* **2018**, *17*, 789–794. [CrossRef] [PubMed]
47. Friedrich-Rust, M.; Buggisch, P.; De Knegt, R.J.; Dries, V.; Shi, Y.; Matschenz, K.; Schneider, M.D.; Herrmann, E.; Petersen, J.; Schulze, F.; et al. Acoustic radiation force impulse imaging for non-invasive assessment of liver fibrosis in chronic hepatitis B. *J. Viral Hepat.* **2013**, *20*, 240–247. [CrossRef]