

Supplementary Information

Table S1. Detected polymyxin ion clusters obtained by nanoESI-HR MS in negative mode.

Compound	Chemical formula	Theoretical mass	m/z	Delta (ppm)
[PxB ₃ +Br] ⁻	C ₅₅ H ₉₆ O ₁₃ N ₁₆ Br	1267.65316	1267.65166	-1.19
[PxB ₃ +2Br+H] ⁻	C ₅₅ H ₉₇ O ₁₃ N ₁₆ Br ₂	1347.57933	1347.57566	-2.72
[PxB ₃ +3Br+2H] ⁻	C ₅₅ H ₉₈ O ₁₃ N ₁₆ Br ₃	1427.50549	1427.50214	-2.35
[PxB ₃ +4Br+3H] ⁻	C ₅₅ H ₉₉ O ₁₃ N ₁₆ Br ₄	1507.43165	1507.42861	-2.02
[PxB ₃ +5Br+4H] ⁻	C ₅₅ H ₁₀₀ O ₁₃ N ₁₆ Br ₅	1587.35781	1587.35352	-2.70
[PxB ₃ +2Br+Na] ⁻	C ₅₅ H ₉₆ O ₁₃ N ₁₆ Br ₂ Na	1369.56078	1369.55790	-2.10
[PxB ₃ +3Br+2Na] ⁻	C ₅₅ H ₉₆ O ₁₃ N ₁₆ Br ₃ Na ₂	1471.46889	1471.46340	-3.73
[PxB ₃ +4Br+3Na] ⁻	C ₅₅ H ₉₆ O ₁₃ N ₁₆ Br ₄ Na ₃	1573.37699	1573.37640	-0.37
[PxB ₃ +Br] ⁻	C ₅₅ H ₉₆ O ₁₃ N ₁₆ Br	1267.65316	1267.65352	0.28
[PxB ₃ +TFA] ⁻	C ₅₇ H ₉₆ O ₁₅ N ₁₆ F ₃	1301.71986	1301.72247	2.00
[PxB ₃ +2TFA+H] ⁻	C ₅₉ H ₉₇ O ₁₇ N ₁₆ F ₆	1415.71273	1415.70990	-2.00
[PxB ₃ +3TFA+2H] ⁻	C ₆₁ H ₉₈ O ₁₉ N ₁₆ F ₉	1529.70559	1529.70270	-1.89
[PxB ₃ +4TFA+3H] ⁻	C ₆₃ H ₉₉ O ₂₁ N ₁₆ F ₁₂	1643.69846	1643.69595	-1.52
[PxB ₃ +5TFA+4H] ⁻	C ₆₅ H ₁₀₀ O ₂₃ N ₁₆ F ₁₅	1757.69132	1757.68656	-2.71
[PxB ₃ +Cl] ⁻	C ₅₅ H ₉₆ O ₁₃ N ₁₆ Cl	1223.70368	1223.70517	1.22
[PxB ₃ +2Cl+H] ⁻	C ₅₅ H ₉₇ O ₁₃ N ₁₆ Cl ₂	1259.68036	1259.67870	-1.31
[PxB ₃ +3Cl+2H] ⁻	C ₅₅ H ₉₈ O ₁₃ N ₁₆ Cl ₃	1295.65703	1295.65562	-1.09
[PxB ₃ +4Cl+3H] ⁻	C ₅₅ H ₉₉ O ₁₃ N ₁₆ Cl ₄	1331.63371	1331.63487	0.87
[PxB ₃ +5Cl+4H] ⁻	C ₅₅ H ₁₀₀ O ₁₃ N ₁₆ Cl ₅	1367.61039	1367.61386	2.54

Table S2: Detected polymyxin, RR4, or dusquetide ion clusters obtained by a quadrupole-resolution instrument tuned to detect negative ion clusters.

Compound	Chemical formula	Theoretical mass	m/z	Delta (ppm)
[PxB ₃ +Br] ⁻	C ₅₅ H ₉₆ O ₁₃ N ₁₆ Br	1267.65316	1268.23	455.05
[PxB ₃ +2Br+H] ⁻	C ₅₅ H ₉₇ O ₁₃ N ₁₆ Br ₂	1347.57933	1348.37	586.73
[PxB ₃ +3Br+2H] ⁻	C ₅₅ H ₉₈ O ₁₃ N ₁₆ Br ₃	1427.50549	1430.37	2006.65
[PxB ₃ +4Br+3H] ⁻	C ₅₅ H ₉₉ O ₁₃ N ₁₆ Br ₄	1507.43165	1511.97	3010.65
[PxB ₃ +TFA] ⁻	C ₅₇ H ₉₆ O ₁₅ N ₁₆ F ₃	1301.71986	1302.45	560.90
[PxB ₃ +Cl] ⁻	C ₅₅ H ₉₆ O ₁₃ N ₁₆ Cl	1223.70368	1224.34	519.00
[RR4+Br] ⁻	C ₄₄ H ₇₆ N ₁₆ O ₇ Br	1019.52663	1019.7	170.05
[RR4+2Br+H] ⁻	C ₄₄ H ₇₇ N ₁₆ O ₇ Br ₂	1099.45279	1099.9	406.76
[RR4+3Br+2H] ⁻	C ₄₄ H ₇₈ N ₁₆ O ₇ Br ₃	1179.37895	1179.9	441.80
[RR4+4Br+3H] ⁻	C ₄₄ H ₇₉ N ₁₆ O ₇ Br ₄	1259.30511	1263.1	3013.48
[Dusquetide+Br] ⁻	C ₂₅ H ₄₇ N ₉ O ₅ Br	632.28835	632.63	540.34
[Dusquetide+2Br+H] ⁻	C ₂₅ H ₄₈ N ₉ O ₅ Br ₂	712.21451	712.60	541.26
[Dusquetide+TFA] ⁻	C ₂₇ H ₄₇ N ₉ O ₇ F ₃	666.35506	666.73	562.67
[Dusquetide+2TFA+2H] ⁻	C ₂₉ H ₄₈ N ₉ O ₉ F ₆	780.34792	780.79	566.52
[Dusquetide+Br+TFA+2H] ⁻	C ₂₇ H ₄₈ N ₉ O ₇ F ₃ Br	746.28122	746.72	587.96

Table S3: The ESI-MS characterisation (positive mode) was used for the synthetic polymyxins B3, RR4, and dusquetide peptides obtained in a Waters ESI-MS Micromass ZQ 4000 spectrometer.

Compound	Chemical formula	Theoretical mass	m/z	Delta (ppm)
[PxB ₃ +H] ⁺	C ₅₅ H ₉₇ O ₁₃ N ₁₆	1189.74210	1190.11	309.23
[PxB ₃ +2H] ²⁺	C ₅₅ H ₉₈ O ₁₃ N ₁₆	595.37497	595.68	512.33
[RR4+H] ⁺	C ₄₄ H ₇₇ O ₇ N ₁₆	941.61611	941.8	6567.32
[RR4+2H] ²⁺	C ₄₄ H ₇₈ O ₇ N ₁₆	471.31197	471.1	-449.74
[RR4+3H] ³⁺	C ₄₄ H ₇₉ O ₇ N ₁₆	314.54392	314.6	178.29
[Dusquetide+H] ⁺	C ₂₅ H ₄₈ N ₉ O ₅	554.37784	554.64	472.89

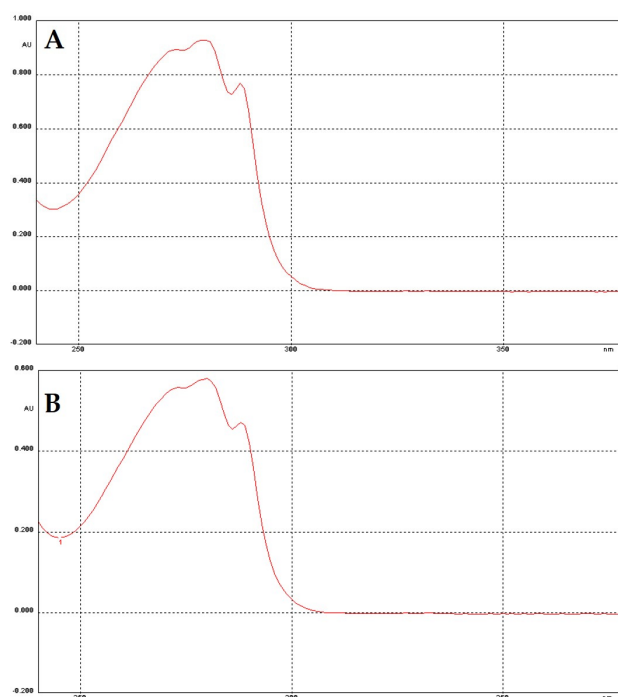


Figure S1: Absorption spectrum of tryptophan. A) H-Trp(OAll)·HCl; B) RR4 heptapeptide after cleavage with TFA/TES/Br₂ (82.5:15:2.5, v/v/v) for 45 minutes.

The following images correspond to the study of the isotopic distribution of the different species obtained by ESI-HR MS of synthetic PxB_3 :

PxB_3 after cleavage with HBr (a detailed study of Figure 7A)

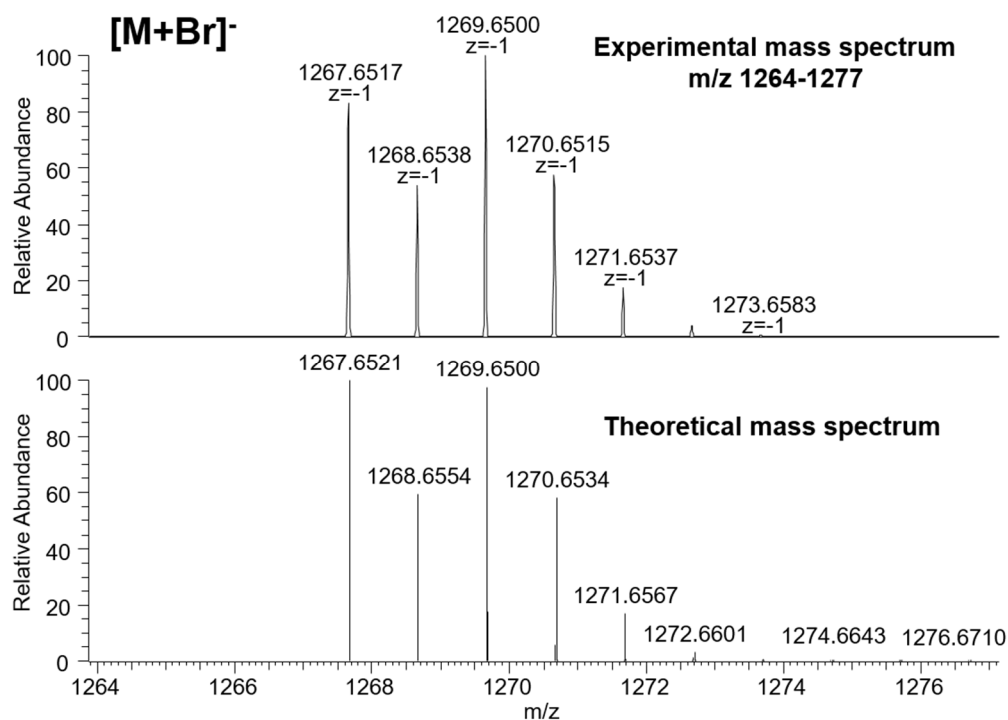


Figure S2: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + Br]^-$, $C_{55}H_{96}O_{13}N_{16}Br$.

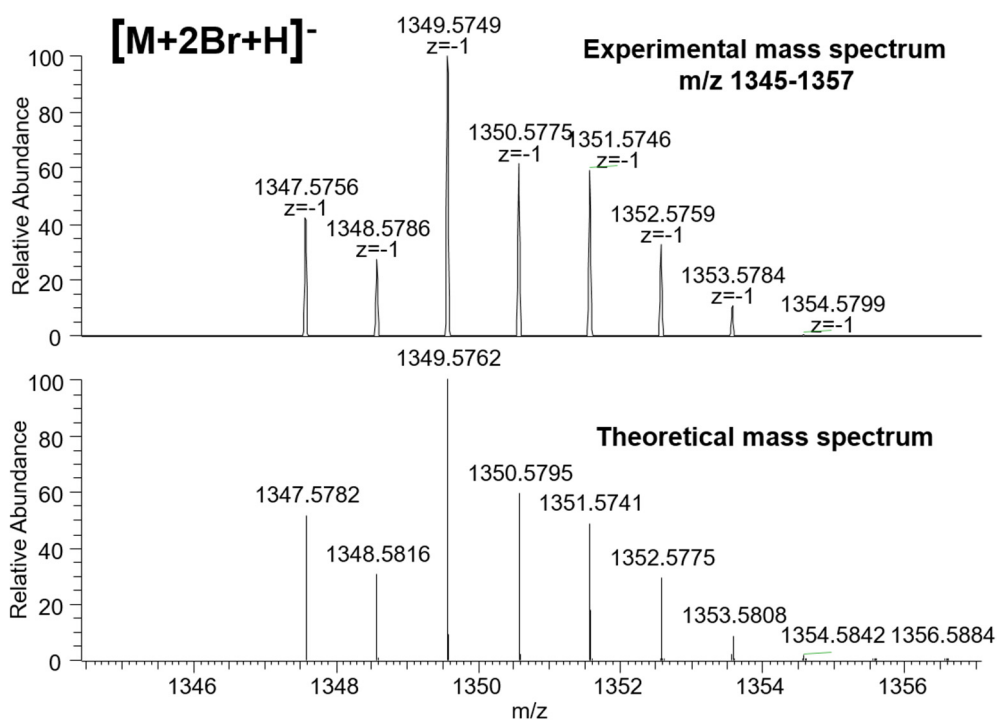


Figure S3: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 2Br + H]^-$, $C_{55}H_{97}O_{13}N_{16}Br_2$.

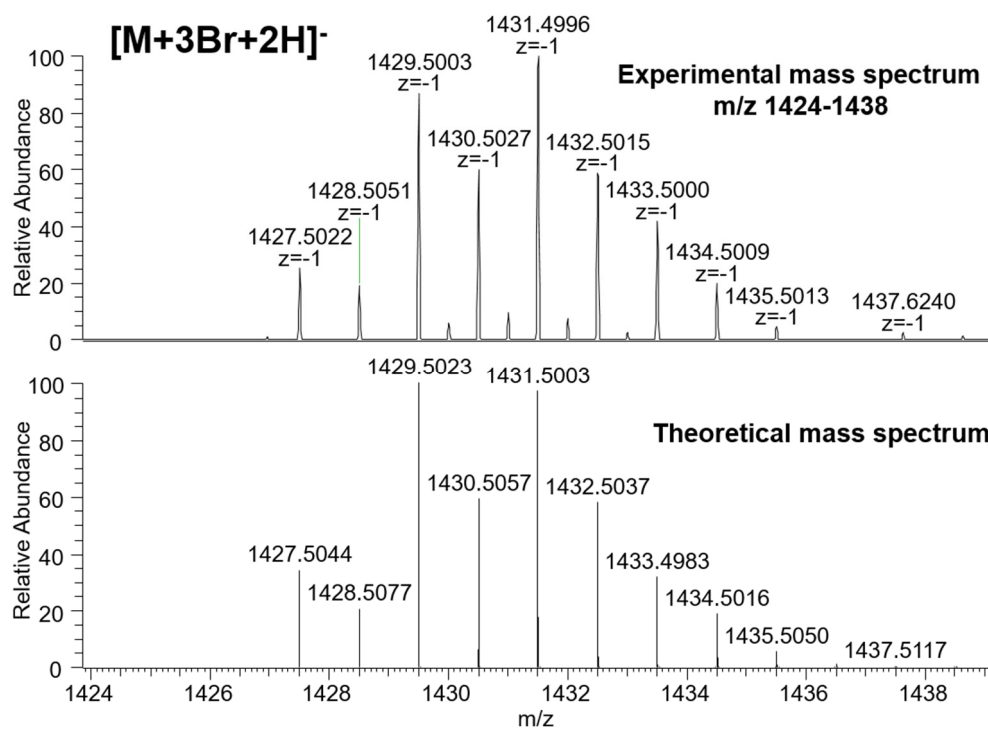


Figure S4: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 3\text{Br} + 2\text{H}]^-$, $\text{C}_{55}\text{H}_{98}\text{O}_{13}\text{N}_{16}\text{Br}_3$.

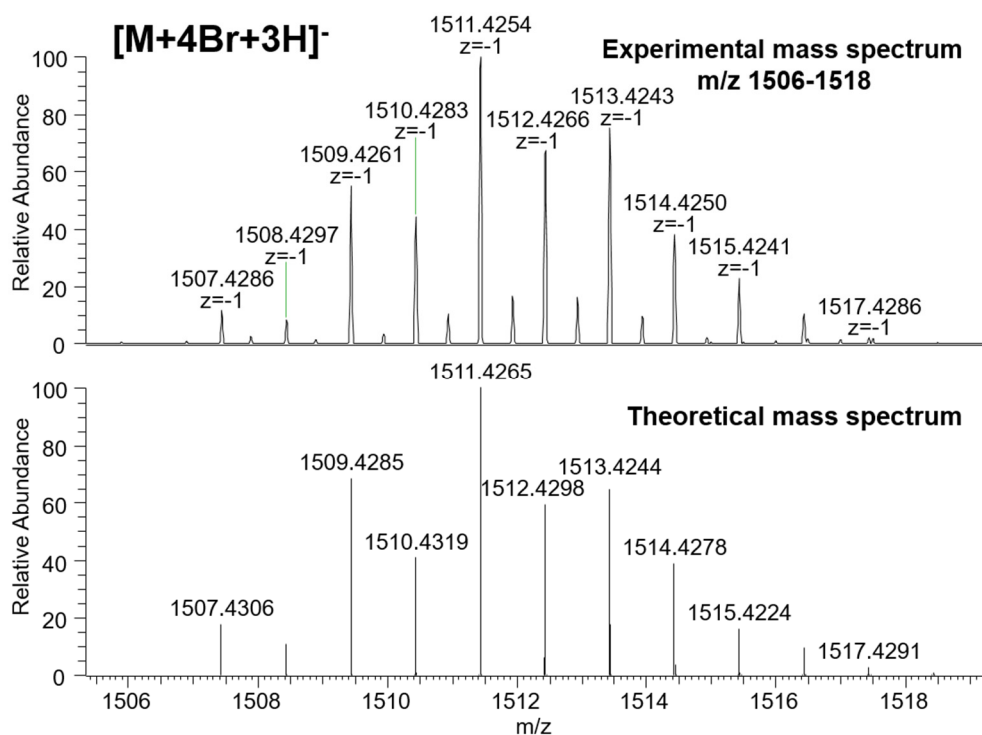


Figure S5: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 4\text{Br} + 3\text{H}]^-$, $\text{C}_{55}\text{H}_{99}\text{O}_{13}\text{N}_{16}\text{Br}_4$.

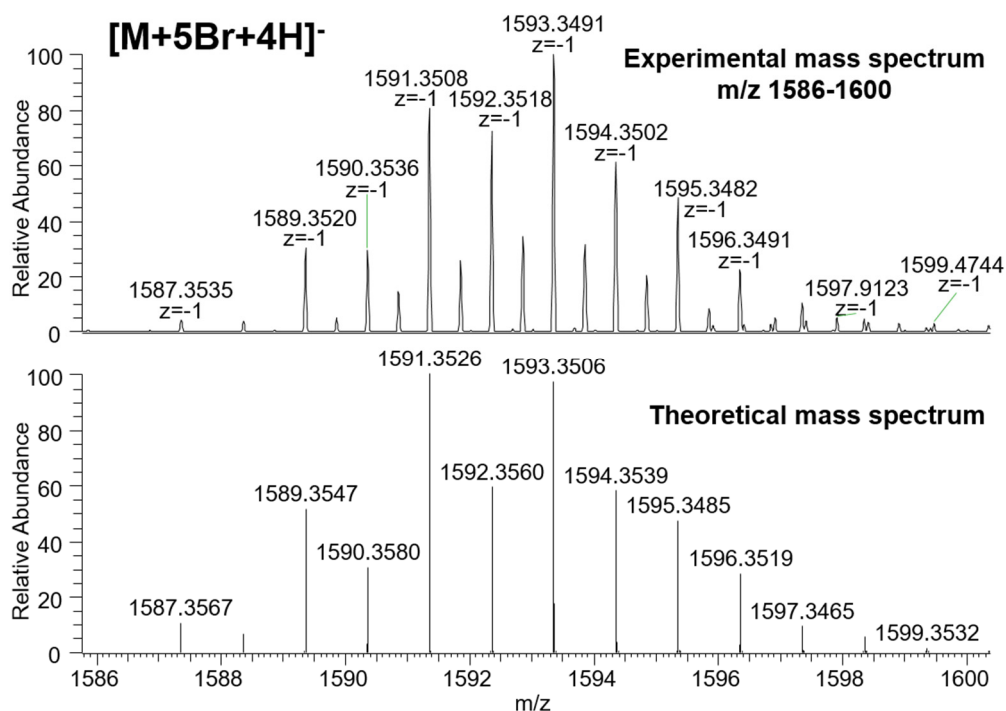


Figure S6: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 5Br + 4H]^-$, $C_{55}H_{100}O_{13}N_{16}Br_5$.

PxB_3 after purification in presence of 0.1 % TFA (a detailed study of Figure 7B):

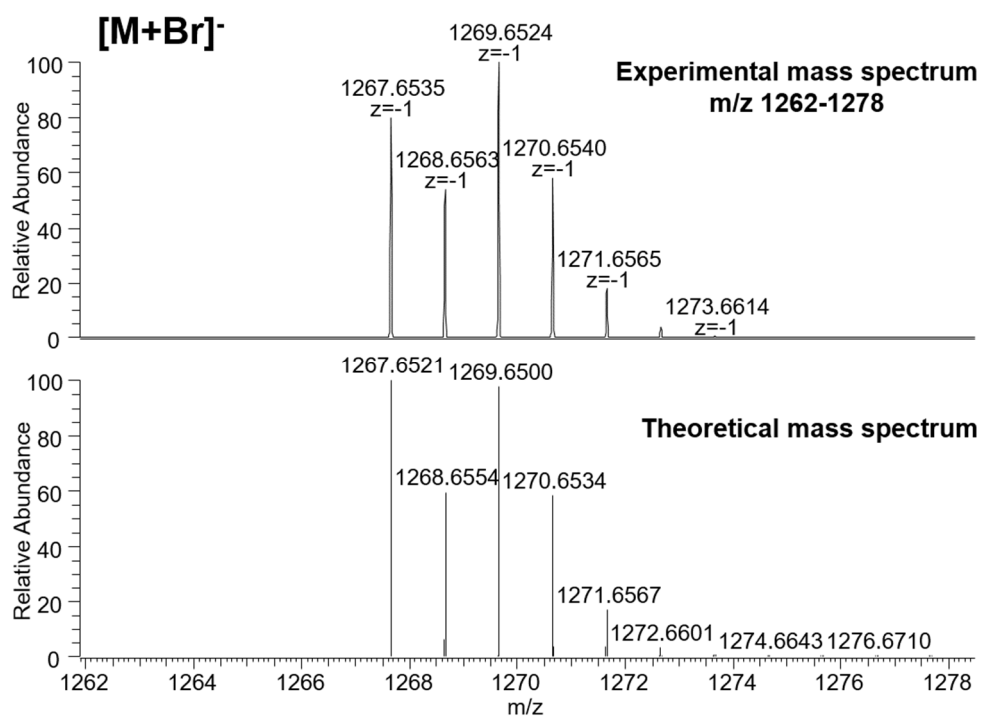


Figure S7: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + Br]^-$, $C_{55}H_{96}O_{13}N_{16}Br$.

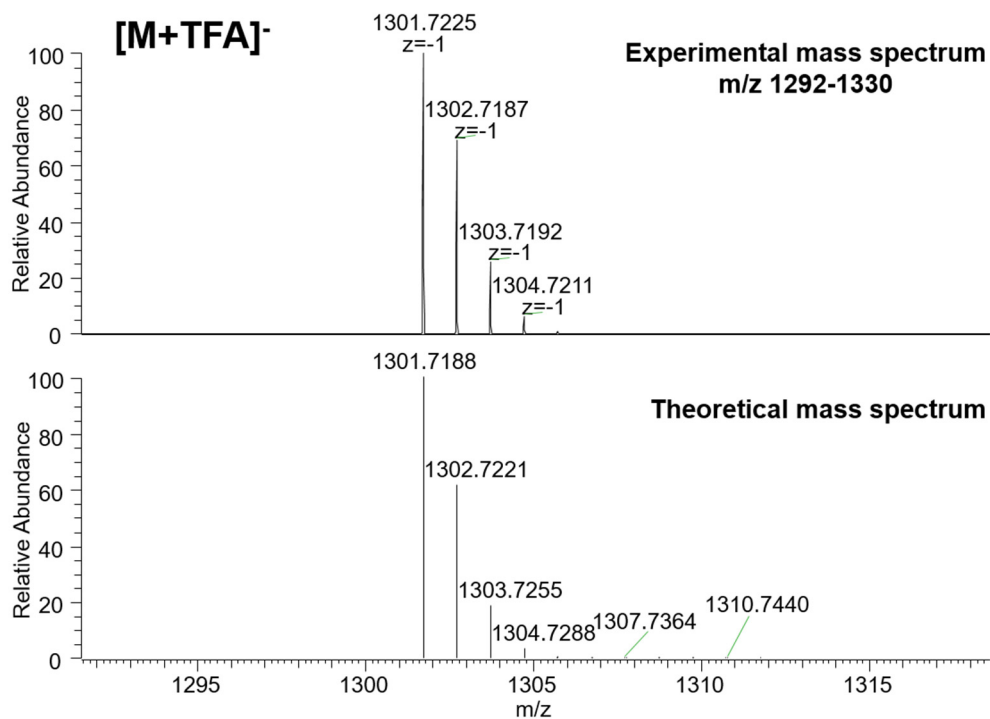


Figure S8: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + \text{TFA}]^-$, $\text{C}_{57}\text{H}_{96}\text{O}_{15}\text{N}_{16}\text{F}_3$.

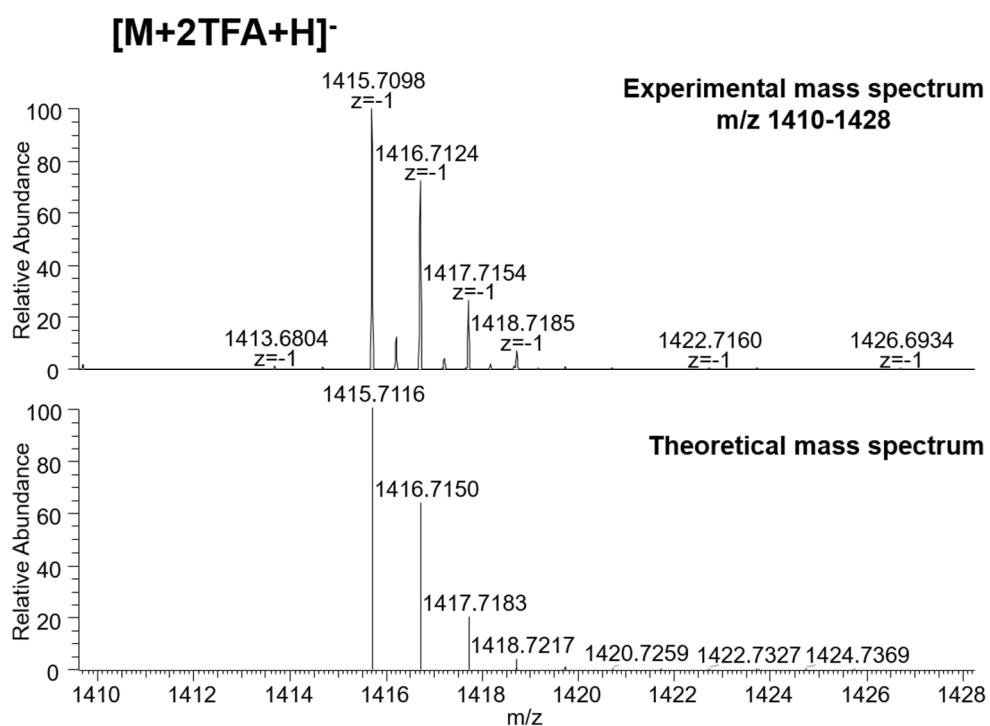


Figure S9: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 2\text{TFA} + \text{H}]^-$, $\text{C}_{59}\text{H}_{97}\text{O}_{17}\text{N}_{16}\text{F}_6$.

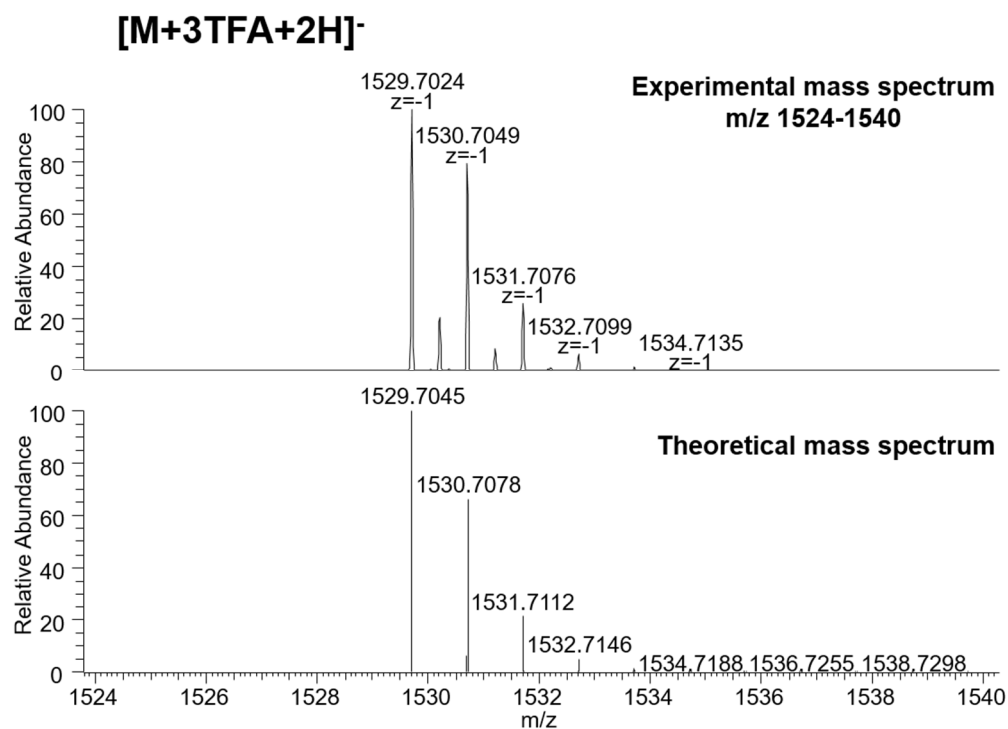


Figure S10: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 3\text{TFA} + 2\text{H}]^-$, $\text{C}_{61}\text{H}_{98}\text{O}_{19}\text{N}_{16}\text{F}_9$.

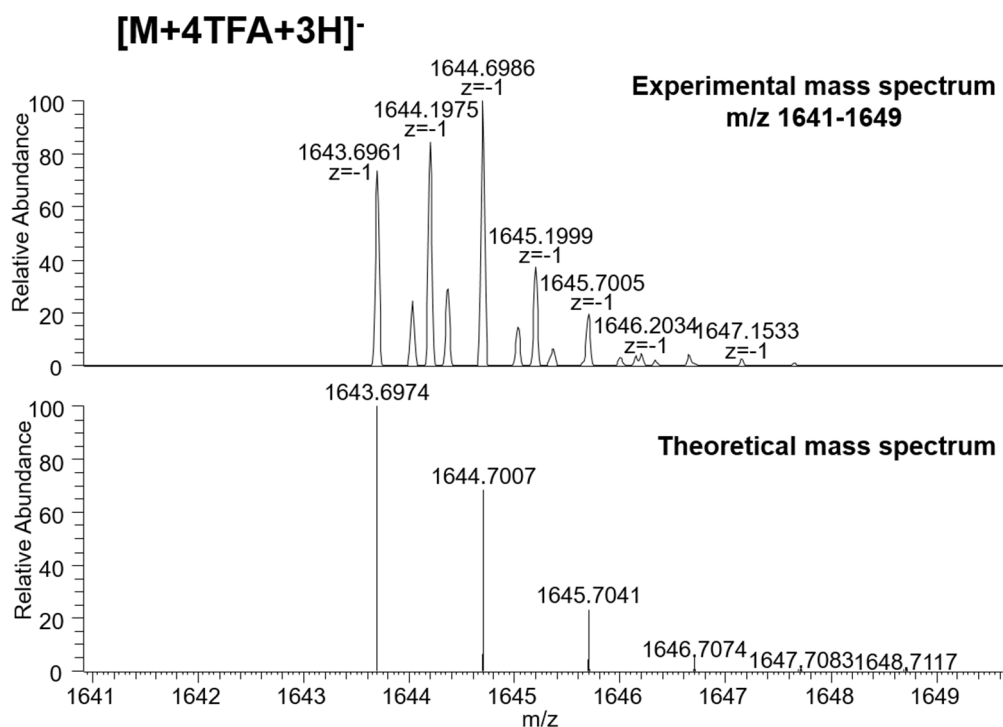


Figure S11: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 4\text{TFA} + 3\text{H}]^-$, $\text{C}_{63}\text{H}_{99}\text{O}_{21}\text{N}_{16}\text{F}_{12}$.

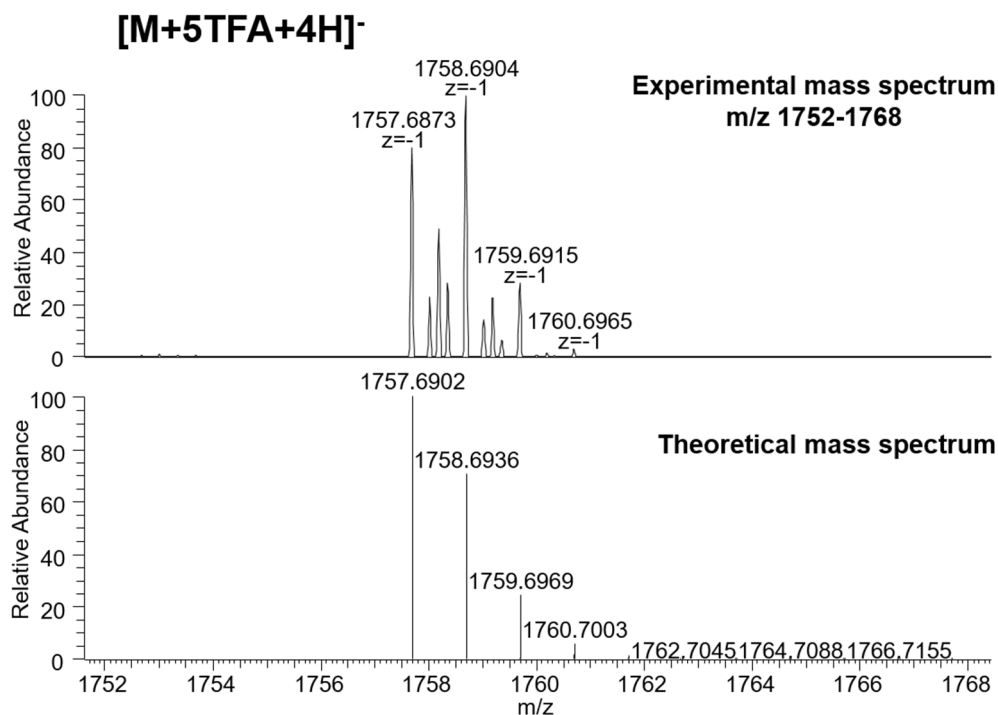


Figure S12: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 5TFA + 4H]^-$, $C_{65}H_{100}O_{23}N_{16}F_{15}$.

PxB₃ after lyophilization with HCl 8 mM (a detailed study of Figure 7C):

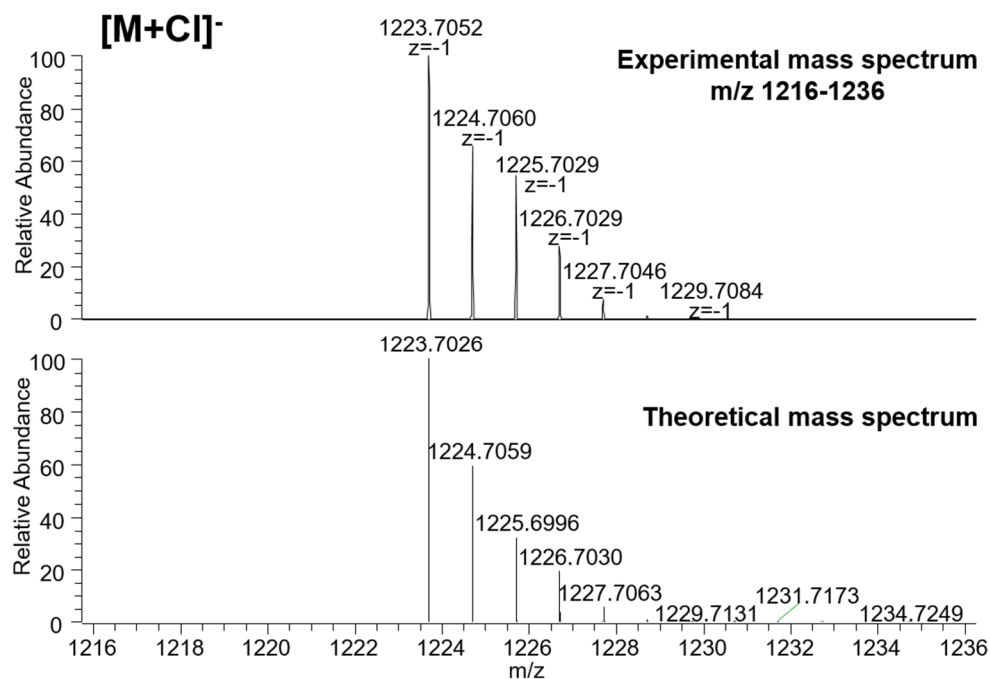


Figure S13: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + Cl]^-$, $C_{55}H_{96}O_{13}N_{16}Cl$.

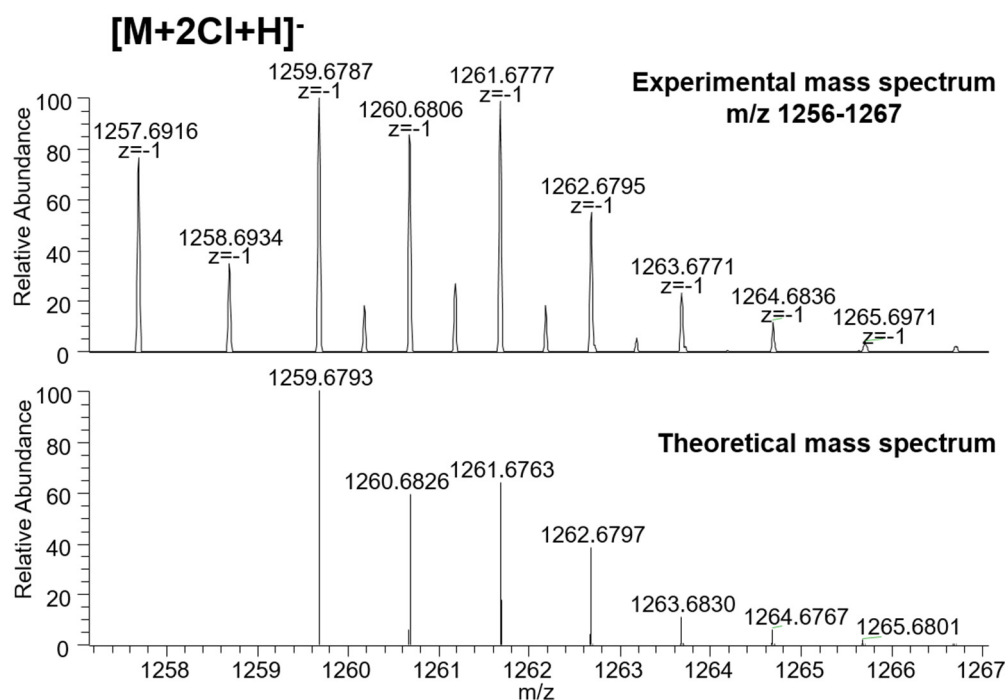


Figure S14: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 2Cl + H]^-$, $C_{55}H_{97}O_{13}N_{16}Cl_2$.

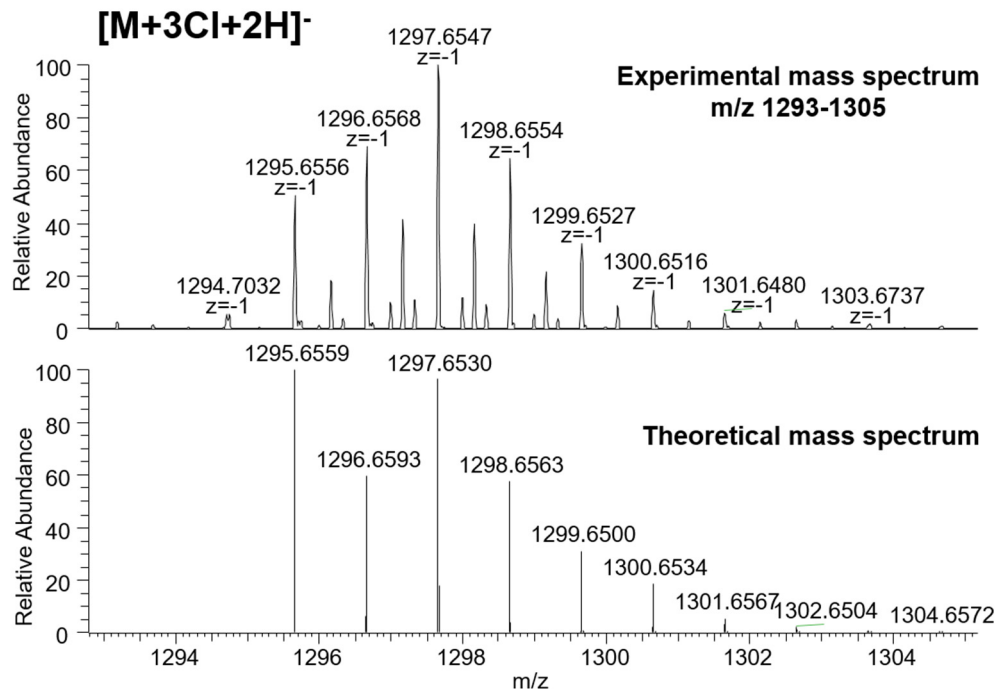


Figure S15: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 3Cl + 2H]^-$, $C_{55}H_{98}O_{13}N_{16}Cl_3$.

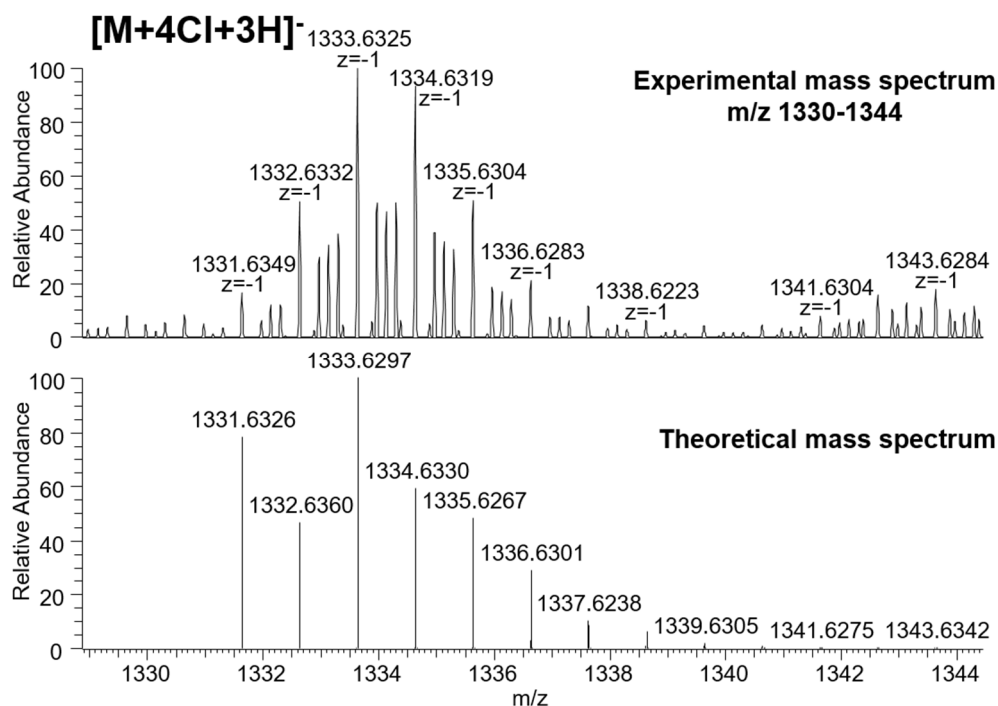


Figure S16: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 4Cl + 3H]^-$, $C_{55}H_{99}O_{13}N_{16}Cl_4$.

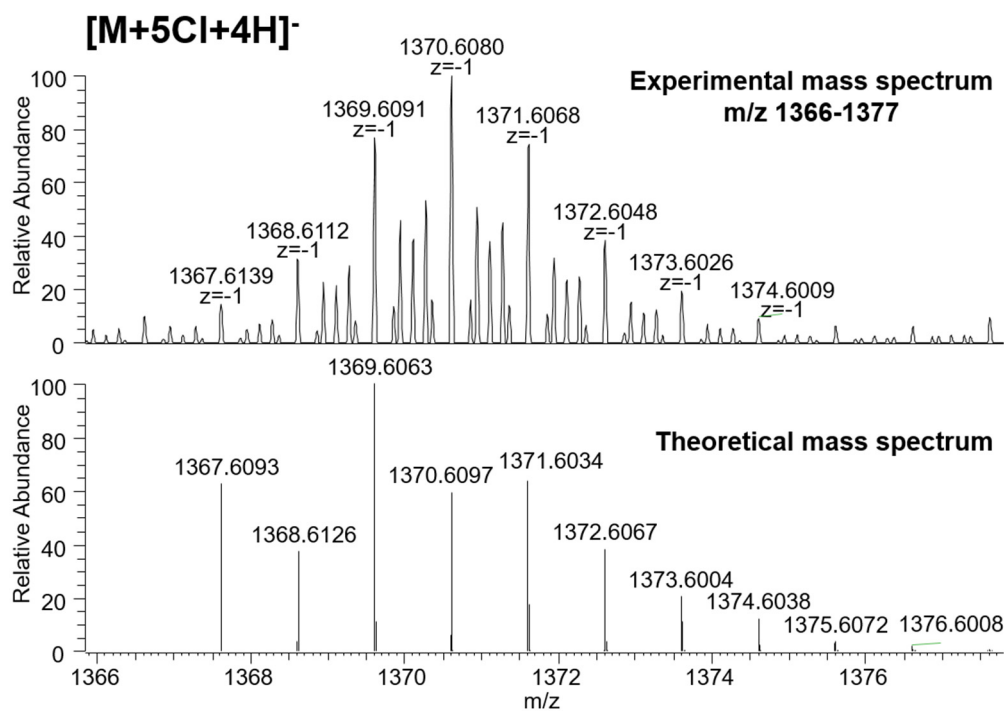


Figure S17: Experimental and theoretical isotopic distribution of the peak corresponding to $[M + 5Cl + 4H]^-$, $C_{55}H_{100}O_{13}N_{16}Cl_5$.

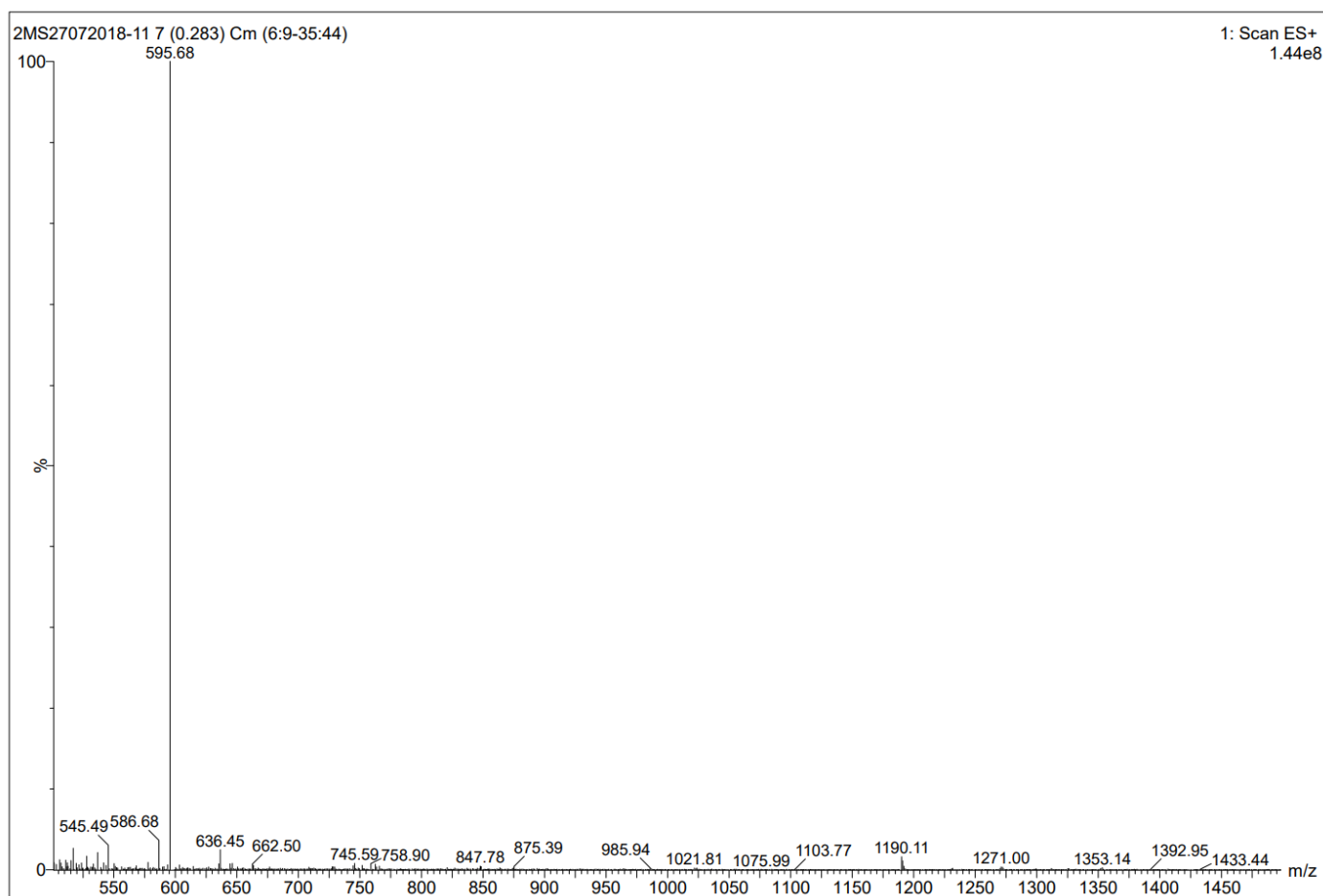


Figure S18: Spectra obtained by ESI-MS in positive mode for polymyxin B₃ peptide.

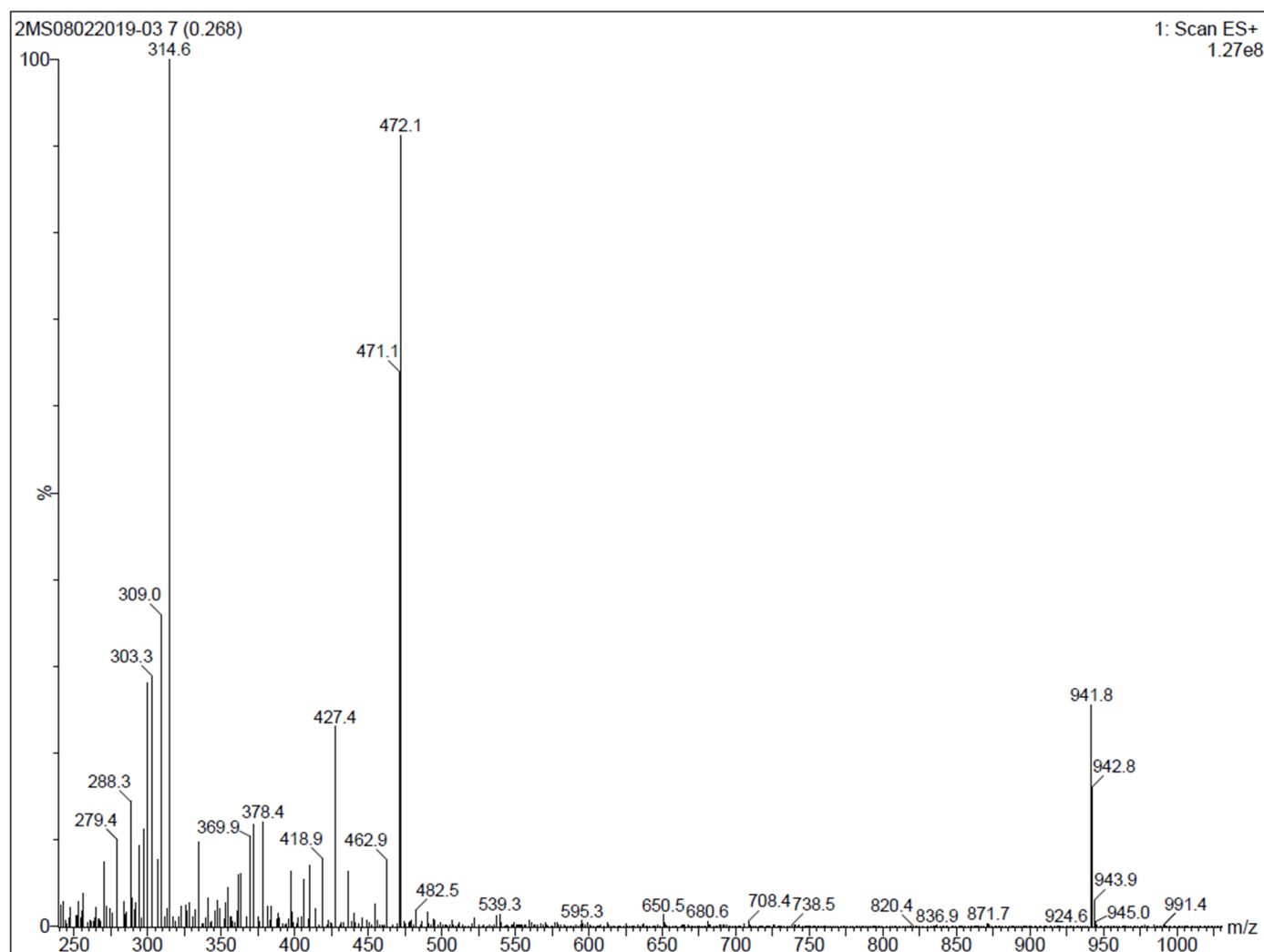


Figure S19: Spectra obtained by ESI-MS in positive mode for RR4 heptapeptide.

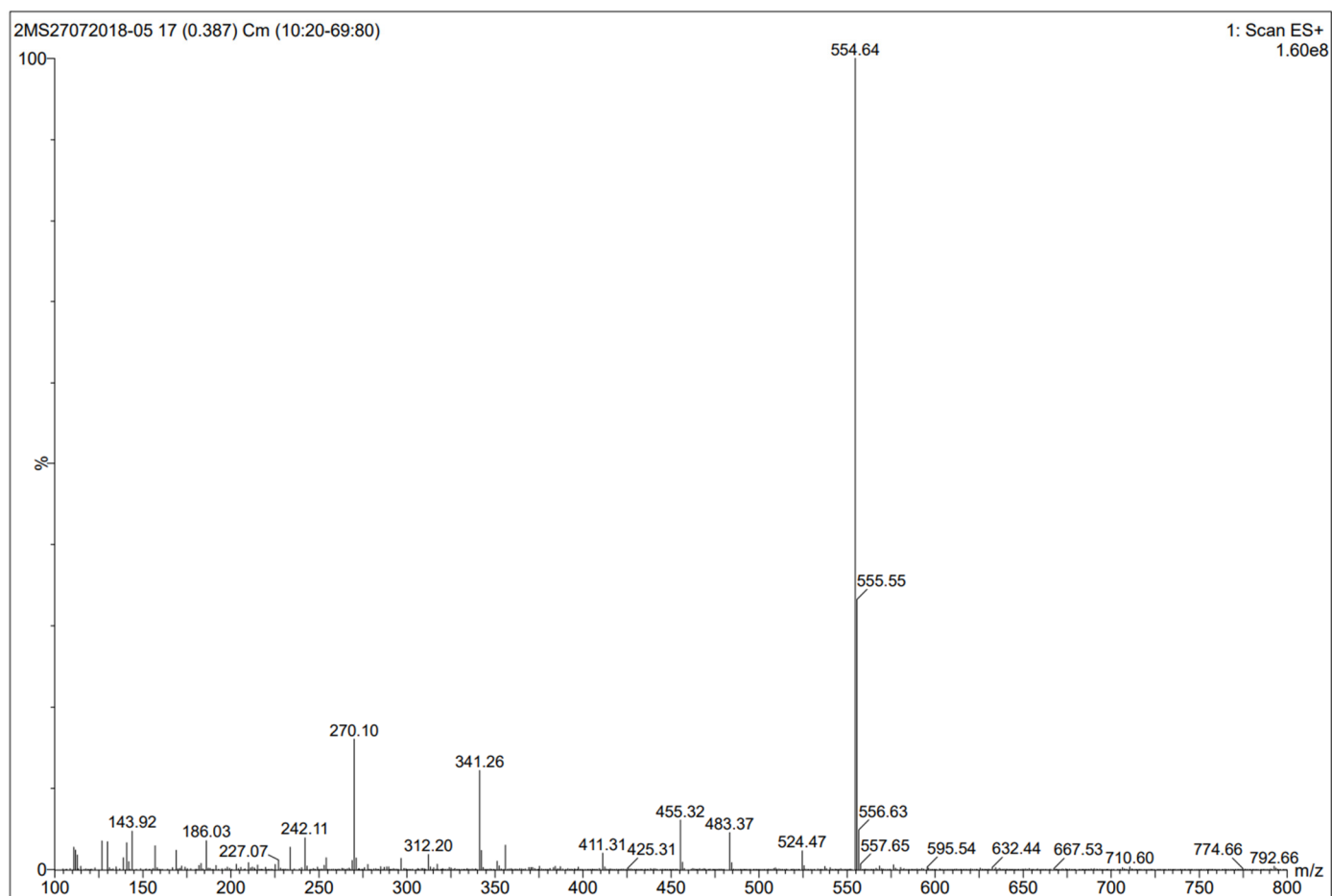


Figure S20: Spectra obtained by ESI-MS in positive mode for the dusquetide peptide.