

## **SUPPLEMENTARY MATERIAL**

### **SUPRAMOLECULAR ARRANGEMENT OF DOXYCYCLINE WITH SULFOBUTYLETHER- $\beta$ -CYCLODEXTRIN: IMPACT ON NANOSTRUCTURATION WITH CHITOSAN, DRUG DEGRADATION AND ANTIMICROBIAL POTENCY**

Renata de Carvalho Feitosa <sup>a</sup>, Juliana Souza Ribeiro Costa <sup>a</sup>, Marcelo van Vliet Lima <sup>a</sup>, Elina Sawa Akioka Ishikawa <sup>a</sup>, Karina Cogo Müller <sup>a</sup>, Fernando Bonin Okasaki <sup>b</sup>, Edvaldo Sabadini <sup>b</sup>, Claudia Garnero <sup>c</sup>, Marcela Raquel Longhi <sup>c</sup>, Vladimir Lavayen <sup>d</sup>, Arnóbio Antônio da Silva-Júnior <sup>e</sup> and Laura Oliveira-Nascimento <sup>\*a</sup>.

<sup>a</sup> Faculty of Pharmaceutical Sciences, University of Campinas (UNICAMP), Campinas - SP, Brazil.

<sup>b</sup> Department of Physical Chemistry, Institute of Chemistry, University of Campinas (UNICAMP), Campinas - SP, Brazil.

<sup>c</sup> Research and Pharmaceutical Technology Development Unit (UNITEFA, CONICET-UNC) and Department of Pharmacy, Faculty of Chemical Sciences, National University of Córdoba, Córdoba, Argentina.

<sup>d</sup> Department of Inorganic Chemistry, Institute of Chemistry, Federal University of Rio Grande do Sul (UFRGS), Porto Alegre - RS, Brazil.

<sup>e</sup> Laboratory of Pharmaceutical Technology and Biotechnology, Department of Pharmacy, Federal University of Rio Grande do Norte (UFRN), Natal - RN, Brazil.

\*Correspondence: Laura Oliveira-Nascimento (lauraon@unicamp.br)

HPLC Method performance data:

Linearity: linear range between 4.88 – 97.62 µg/mL ( $r^2 = 0.9996$ ). The calibration curve data are in **Table S1** and the related statistical data are in **Table S2**.

Detection Limit (DL) and Quantification Limit (QL): DL and QL were calculated according to the following equations:

$$DL = \frac{3.3\sigma}{S}$$

$$QL = \frac{10\sigma}{S}$$

Where: S = slope of the calibration curve;  $\sigma$  = standard deviation of y-intercept of the regression line, as described by ICH [1]. The results are shown in **Table S3**.

Selectivity: complexation did not change Doxycycline retention time nor the major related substance peak in a significant manner (**Figure S1**). The filtrate of blank nanoparticles did not change the baseline.

Precision: repeatability (intra-day) analysis was carried out using three concentrations (low, medium and high) that contemplate the linear range of the analytical method, with three replicates at each level. The test was performed under the same operating conditions, same analyst, and same instrumentation, in a single analytical run. The relative standard deviation (RSD) was calculated, indicating the accuracy of the method. Intermediate precision (inter-day) was carried out under the same conditions of the previous analysis, but on a different day. The results are shown in **Table S3**.

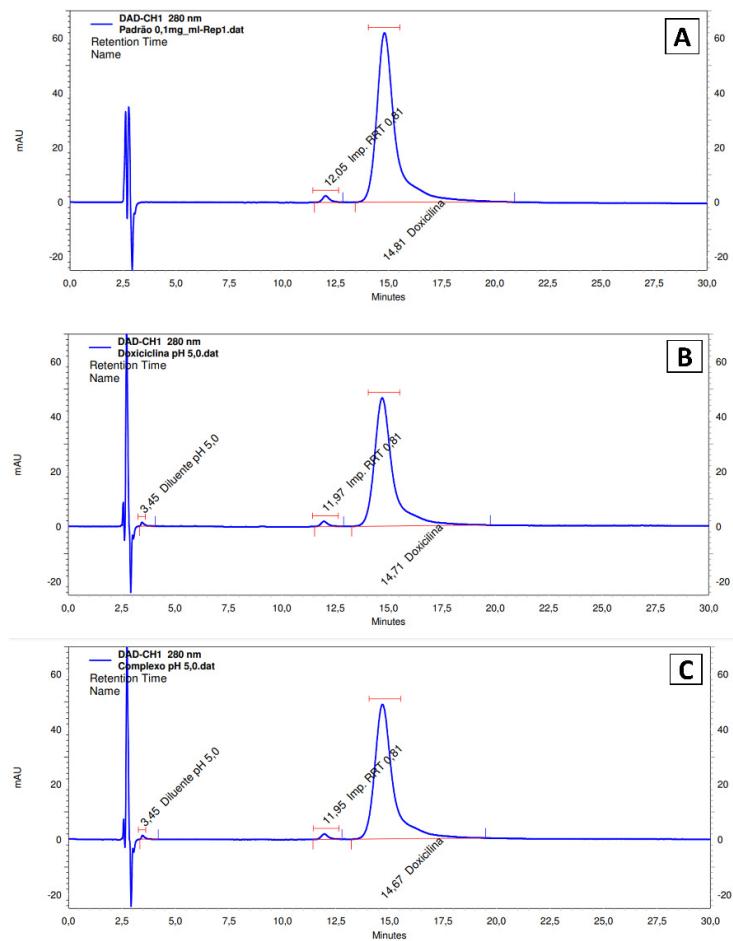
Accuracy: The accuracy was inferred because linearity, precision and selectivity have been established (as recommended by ICH [1]). In addition, a drug recovery test was performed in the dispersing medium containing the empty nanoparticles. The results obtained indicate that the diluent and the matrix do not have significant interference in the quantification of the drug. The assay was performed in triplicate using an intermediate concentration of the calibration curve (30 µg/mL), results are in **Table S3**.

**Table S1.** Calibration curve data.

<i>Drug concentration (ug/mL)</i>	<i>Peak area (mAU)</i>	<i>Average peak area (mAU)</i>	<i>RSD (%)</i>
4.881	380373		
	396550	385533	2.48
	379676		
9.762	891925		
	930012	919510	2.62
	936594		
24.405	2563578		
	2505054	2538378	1.19
	2546503		
48.810	5281628		
	5447629	5378534	1.61
	5406346		
97.620	11206014		
	11263016	11258907	0.45
	11307691		

**Table S2.** Statistical data related to the calibration curve (excel)

Regression statistics							
multiple R	0.999808099						
R-squared	0.999616235						
Adjusted R-squared	0.999586715						
Standard error	83772.98214						
Observation	15						
ANOVA							
	df	SS	MS	F	Significance F		
Regression	1	2.3764E+14	2.3764E+14	33861.92	1.35869E-23		
Residual	13	91232862967	7017912536				
Total	14	2.37731E+14					
	Coefficients	Standard error	t Stat	P-value	Lower 95%	Upper 95%	Lower 95.0%
Intercept	-259187.3129	32063.25649	-8.0836241	2E-06	-328455.7672	-189918.8585	-328455.7672
Variable X 1	117409.0704	638.0370361	184.016074	1.36E-23	116030.6752	118787.4656	116030.6752
							118787.4656
Residue results							
Observation	Predicted Y	Residual	standardized residual	Observation	Predicted Y	Residual	standardized residual
1	313886.3599	66486.64013	0.823612165	8	2606181.051	-101127.0509	-1.252724895
2	313886.3599	82663.64013	1.024006921	9	2606181.051	-59678.05086	-0.739269853
3	313886.3599	65789.64013	0.814977984	10	5471549.415	-189921.4146	-2.352676976
4	886960.0326	4964.967385	0.061504199	11	5471549.415	-23920.41459	-0.296317342
5	886960.0326	43051.96739	0.533312016	12	5471549.415	-65203.41459	-0.807716037
6	886960.0326	49633.96739	0.614847423	13	11202286.14	3727.857933	0.04617934
7	2606181.051	-42603.05086	-0.527751002				



**Figure S1.** High-performance liquid chromatography (HPLC) chromatograms of (A) doxycycline standard, (B) doxycycline at pH 5.0 and (C) drug complex with sulfobutylether- $\beta$ -cyclodextrin at pH 5.0. The concentration of all samples was 0.1 mg/mL.

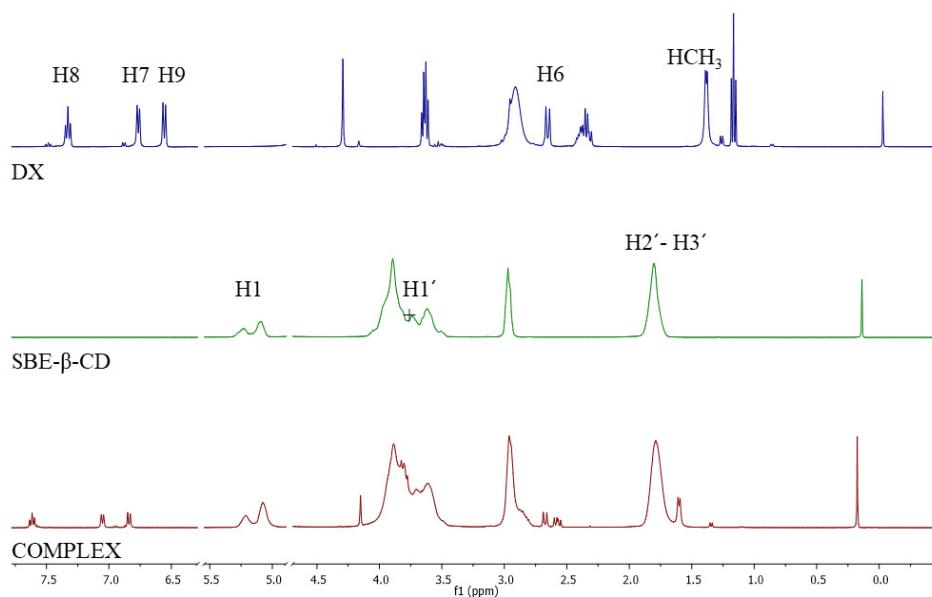
**Table S3.** High Performance Liquid Chromatography method performance data.

Parameter	Result
Selectivity	no interference detectable
Calibration Curve	$y = 117409x - 259187$
Linearity ( $\mu\text{g/mL}$ )	4.88 – 97.62
Coefficient of Determination ( $R^2$ )	0.9996
DL ( $\mu\text{g/mL}$ )	1.045
QL ( $\mu\text{g/mL}$ )	3.168
Accuracy (% Recovery) *	$101.48 \pm 2.52$
Precision intra-day (repeatability % RSD)	High level (4.9 $\mu\text{g/mL}$ ): 2.48 Medium level (24.4 $\mu\text{g/mL}$ ): 1.19 Low level (97.6 $\mu\text{g/mL}$ ): 0.45
Precision inter-day (intermediate precision % RSD)	High level (5.2 $\mu\text{g/mL}$ ): 1.12 Medium level (25.8 $\mu\text{g/mL}$ ): 2.75 Low level (103.3 $\mu\text{g/mL}$ ): 0.80

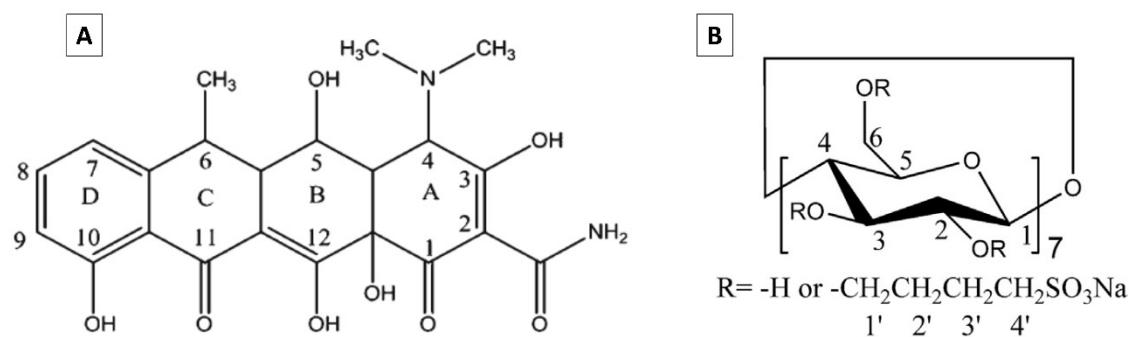
DL: Detection Limit; QL: Quantification Limit. \* Accuracy inferred by the recovery experiment.

**Table S4.**  $^1\text{H}$  NMR chemical shifts ( $\delta$ ) of doxycycline hydulate (DX), sulfobutyl- $\beta$ -cyclodextrin (SBE- $\beta$ -CD), and the complexes obtained by freeze-drying.

	Assignment	$\delta_{\text{free}}$	Complex ( $\Delta\delta = \delta_{\text{system}} - \delta_{\text{free}}$ )
DX	H6	2.65	0.02
	H7	6.76	0.29
	H8	7.33	0.29
	H9	6.56	0.28
	HCH <sub>3</sub>	1.38	0.22
	NH <sub>2</sub>	7.49	No signal
SBE- $\beta$ -CD	H1	5.23, 5.09	-0.02, -0.01
	H1'	3.74	-0.03
	H2	3.62	Overlap
	H3	4.05	No signal
	H2' - H3'	1.80	-0.01
	H4'	2.97	Overlap
	H5	3.89	0



**Figure S2.** Nuclear magnetic resonance spectra of doxycycline (DX), sulfobutylether- $\beta$ -cyclodextrin (SBE- $\beta$ -CD) and complex.



**Figure S3.** Molecular structure of doxycycline and sulfobutylether- $\beta$ -cyclodextrin.

**Table S5:** Main vibrational modes of infrared spectra (4000 – 650 cm<sup>-1</sup>) of hyclare doxycycline (DX), sulfobutylether-β-cyclodextrin (SBE-β-CD), physical mixture (1:4 molar ratio), and freeze-dried complex (1:4 molar ratio).

DX (cm <sup>-1</sup> )	SBE-β-CD (cm <sup>-1</sup> )	Mixture (cm <sup>-1</sup> )	Complex (cm <sup>-1</sup> )	Assignment
3440(83) <sup>*</sup>	3619(66) <sup>*</sup>	3607(51) <sup>*</sup>		v(O-H), CD partial Hbond
3372(42)	3548(109)	3545(86)	3556(128) <sup>*</sup>	v(O-H), CD partial Hbond
3331(33)	3416(172)	3428(157)		v(H-O-H), retained in the interstices among CD
3285(179)		3287(81)	3401(186)	v(H-O-H)
3266(224)			3267(197)	v(OH)
2934 <sub>(w)</sub>	2934 <sub>(w)</sub>	2939 <sub>(w)</sub>	3227(114)	v <sub>as</sub> (CH <sub>2</sub> )
2880 <sub>(vw)</sub>		2880 <sub>(vw)</sub>	2880 <sub>(vw)</sub>	v <sub>s</sub> (CH <sub>2</sub> )
1710 <sub>(sh)</sub>	1664 <sub>(w)</sub>			δ(H-O-H) bound to CD
1643 <sub>(m)</sub>		1656 <sub>(w,vw)</sub>	1651 <sub>(vw)</sub>	v(C=O), DX
1610 <sub>(m)</sub>				v(C=C), DX
1556 <sub>(m)</sub>	1544 <sub>(vw)</sub>			
1452 <sub>(m,vw)</sub>	1456 <sub>(m)</sub>	1453 <sub>(m,vw)</sub>	1451 <sub>(vw)</sub>	φOCH, φHCH
		1422 <sub>(m,vw)</sub>	1420 <sub>(vw)</sub>	φOCH, φCCH
1415 <sub>(m,vw)</sub>		1411 <sub>(m,vw)</sub>		
1366 <sub>(m,vw)</sub>	1362 <sub>(m)</sub>	1364 <sub>(m,vw)</sub>	1365 <sub>(vw)</sub>	φCCH, φOCH, φCOH δ(C-H), CD/in plane bending of CH, enolic COH, skeletal CCC
1330 <sub>(vw)</sub>	1330 <sub>(m)</sub>			Complex modes CH <sub>2</sub> OH, CD
	1301 <sub>(m)</sub>			v(S=O)
1261 <sub>(vw)</sub>				δ <sub>in-plane</sub> (C-H)
1240 <sub>(sh,m)</sub>				Complex modes CH <sub>2</sub> OH, CD
1222 <sub>(m)</sub>				
1196 <sub>(sh,m)</sub>		1202 <sub>(sh,m)</sub>	1198 <sub>(sh,m)</sub>	
1153 <sub>(s)</sub>		1156 <sub>(m)</sub>	1156 <sub>(m)</sub>	vCO,vCC v(C-O-C) glucosidic/CH overtone stretching
1125 <sub>(w)</sub>				v(S=O)
1041 <sub>(w)</sub>				v(C-C)
1044				v(C-N)/DX
1027 <sub>(m)</sub>		1029 <sub>(w)</sub>	1029 <sub>(w)</sub>	δ <sub>in-plane</sub> (C-H) DX /vCC, δOCH, δCCH, δCCO
		1026 <sub>(w)</sub>		vCC, δOCH, δCCH, δCCO
946 <sub>(vw)</sub>				Skeletal vibration a-1,4 linkage
934 <sub>(vw)</sub>				

---

858 <sub>(vw)</sub>	$\delta$ <i>Out-of-plane</i> (=C-H) DX
788 <sub>(vw)</sub>	

<sup>a</sup> Relative intensity: w = weak; s = strong; sh = shoulder; m = medium; vw = very weak; v = stretching mode;  $\delta$  = bending mode; mm = multiple;  $\varphi$  = stretching mode guest ring. The values among brackets are the FWHM numbers to come from deconvolution process on the complex 1:4.

---

#### References:

- [1] International Council for Harmonisation. ICH Harmonised Guideline: Validation of analytical procedures Q2(R2). Available online: <https://www.ich.org/page/quality-guidelines> (accessed on 30 March 2023).