

Review

Modulating Inflammation-Mediated Diseases via Natural Phenolic Compounds Loaded in Nanocarrier Systems

Tojofaniry Fabien Rakotondrabe ^{1,2,3,4}, Min-Xia Fan ^{1,2,3}, Felix Wambua Muema ^{1,2,3,4} and Ming-Quan Guo ^{1,2,3,*}

¹ CAS Key Laboratory of Plant Germplasm Enhancement and Specialty Agriculture, Wuhan Botanical Garden, Chinese Academy of Sciences, Wuhan 430074, China

² Sino-Africa Joint Research Center, Chinese Academy of Sciences, Wuhan 430074, China

³ Innovation Academy for Drug Discovery and Development, Chinese Academy of Sciences, Shanghai 201203, China

⁴ University of Chinese Academy of Sciences, Beijing 100049, China

* Correspondence: guomq@wbcas.cn; Tel.: +86-027-87700850

Abstract: The global increase and prevalence of inflammatory-mediated diseases have been a great menace to human welfare. Several works have demonstrated the anti-inflammatory potentials of natural polyphenolic compounds, including flavonoid derivatives (EGCG, rutin, apigenin, naringenin) and phenolic acids (GA, CA, etc.), among others (resveratrol, curcumin, etc.). In order to improve the stability and bioavailability of these natural polyphenolic compounds, their recent loading applications in both organic (liposomes, micelles, dendrimers, etc.) and inorganic (mesoporous silica, heavy metals, etc.) nanocarrier technologies are being employed. A great number of studies have highlighted that, apart from improving their stability and bioavailability, nanocarrier systems also enhance their target delivery, while reducing drug toxicity and adverse effects. This review article, therefore, covers the recent advances in the drug delivery of anti-inflammatory agents loaded with natural polyphenolics by the application of both organic and inorganic nanocarriers. Even though nanocarrier technology offers a variety of possible anti-inflammatory advantages to naturally occurring polyphenols, the complexes' inherent properties and mechanisms of action have not yet been fully investigated. Thus, expanding the quest on novel natural polyphenolic-loaded delivery systems, together with the optimization of complexes' activity toward inflammation, will be a new direction of future efforts.

Keywords: polyphenolic; nanocarriers; drug delivery; bioavailability; targets; anti-inflammatory

Citation: Rakotondrabe, T.F.; Fan, M.-X.; Muema, F.W.; Guo, M.-Q. Modulating Inflammation-Mediated Diseases via Natural Phenolic Compounds Loaded in Nanocarrier Systems. *Pharmaceutics* **2023**, *15*, 699. <https://doi.org/10.3390/pharmaceutics15020699>

Academic Editors: Rosa Direito and Maria-Eduardo Figueira

Received: 31 December 2022

Revised: 9 February 2023

Accepted: 16 February 2023

Published: 19 February 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Inflammation is the stereotypical organism response actuated by the immune system to encounter and adapt deleterious tissue homeostasis caused by tissue injury, infections, or chemical agents [1,2]. Typically, inflammation acts for two main finalities, either it can be a defensive-adaptive reaction that re-establishes the organism's homeostasis or it can be harmful when it last and pertains to destabilizing the physiological circumstances within the organism [3]. Based on the foreign stimulus and the tissue response efficiency, the inflammation can be classified as acute or chronic. Acute inflammation occurs during a short period, generally within a few weeks, until tissues complete the healing process and reach their equilibrium [4]. On the other hand, chronic inflammation lasts for a longer time, ranging from months to years. This second type of inflammation arises from unsolved acute inflammation or is commonly de novo via bypassing the initial inflammation step. Recent uncoverings have attributed the important roles of the inflammatory phenomenon in the early stage of degenerative disorders pathology, which progressively turn to chronic diseases, including arthritis, neurodegenerative conditions, skin inflammations, inflammatory bowel diseases, metabolic syndromes, and cardiovascular diseases

(CVD). According to the World Health Organization (WHO), these inflammatory-based chronic diseases have increased worldwide and present great menaces to human life, as they lead to high rates of mortality ($\geq 60\%$) [2].

So far, a plethora of synthetic anti-inflammatory drugs, including steroids, non-steroidal anti-inflammatory drugs (NSAIDs), and glucocorticoids, have been developed and clinically used for the treatment of these diseases. However, these drugs have mostly expensive and prompt short- or long-term side effects, in addition to their efficacy [5]. A large number of scientific investigations have recently proved that natural phenolic compounds, encompassing flavonoids, phenolic acids, and stilbenoids, displayed promising effects in managing these types of inflammatory-mediated diseases. In addition to their great therapeutic potential, these compounds are inexpensive to produce and present fewer side effects and toxicity. Phenolic curative effects are explained by a variety of mechanisms, which consist mainly in silencing inflammation development. However, the physicochemical (size, structures, polarity) and poor pharmacokinetics properties (absorption, distribution, metabolism, excretion, toxicity: ADMET) of these compounds sometimes refrain from the mass production of phenolic-based marketable pharmaceuticals. Additionally, their ultimate therapeutic effects can be hampered by the low bioavailability to the organisms, as well as the abject targeting efficacy, due to instabilities and/or low final concentrations delivered to the targets [6,7].

Within this framework, drug delivery systems (DDS) have arisen during the last five decades to enhance pharmacokinetics properties, increase target site carriage, and stabilize these chemicals to improve their therapeutic activities. Among them, nanocarrier delivery systems (NDS) have recently demonstrated promising results for attaining the above-cited objectives in a more controlled manner [8]. Phenolic-loaded NDS consists of encapsulating, entrapping, or decorating phenolics with one or a variety of materials to form a final complex of the nanoscale range. Regarding the nature of the carrier materials, NDS could be classified as organic, inorganic, or hybrid. Organic NDS consists of biocompatible carbon-based vesicles, such as liposomes, solid lipids, micelle, polymeric, and hydrogel nanocarriers. However, those formed with inorganic-based elements, such as metals, constitute the second group. Hybrid NDS types are synthesized when these organic and inorganic materials are compounded to form nanoparticle complexes.

This present review paper synthesizes and discusses the recent progress of the common phenolic-loaded NDS in the management of inflammatory-based diseases. Although nanocarrier technology offers a variety of advantages in polyphenols delivery, the synthesized complexes' mechanisms of action have not yet been systematically explored. Additionally, many of the demonstrated potential distinctive phenolic compounds research is limited to their coarse form uses. Hence, broadening the application of nanocarrier systems to a variety of bioactive phenolics, along with the optimization of complexes to inflammatory target efficiency, will be a new focus of future endeavors.

2. Natural Phenolic Compounds in Inflammatory Mitigation and Their Challenges

Flavonoids and phenolic acids are the most abundant phenolic compounds [9,10]. Amongst flavonoids, EGCG (catechin), rutin (flavonol), naringenin (flavonone), and apigenin (flavone) are the most abounding (Figure 1) [11]. However, ellagic acid and caffeic acid are among the most prevalent phenolic acids [12]. In addition to the antioxidant effects of phenolic acids to promote health improvements, their anti-inflammatory roles add more benefits for the management of different impairments. So far, much research has demonstrated the ability of these types of phenolics to mediate various types of inflammatory-based diseases. *In vitro* studies have demonstrated their ability to hinder the differentiation of Th1 and Th17 in the pathogenesis of rheumatoid arthritis. Additionally, they could downregulate pro-inflammatory cytokines and chemokines production via their interference with IL-1 β signaling pathways or with laminin receptor (67LR) [13–15]. In neuroinflammation, a lot of evidence has demonstrated that flavonoids can protect neurons towards activated microglial cells, mediate the production of inflammatory markers,

and regulate pro-inflammatory signaling pathways (MAPK, JAK/STAT) [16–18]. The beneficial intestinal inflammatory mediation of flavonoids was also evidenced. They can hinder eicosanoids (COX, LOX, and leukotrienes) synthesis and overexpression, downregulate the production and hiring of immune cells, which reduces the releases of pro-inflammatory effectors, and modulate the associated signaling pathways [19,20]. Along with the several proofs supporting the beneficial effects of flavonoids on carbohydrates and lipid homeostasis in the organism, they have been also demonstrated to reduce the production of inflammatory molecules and improve the activation of the peroxisome proliferator-activated receptor gamma (PPAR- γ) [21–23].

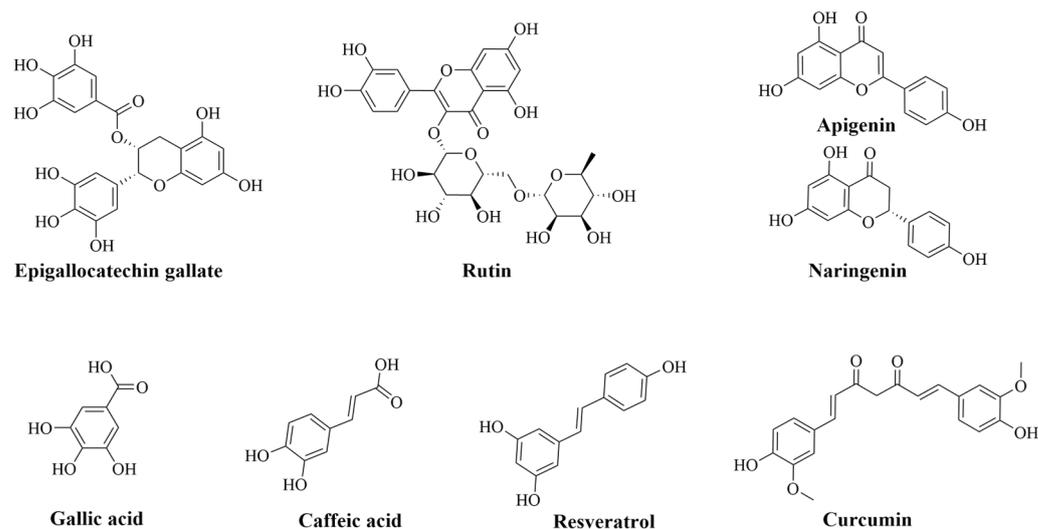


Figure 1. The common natural phenolic compounds studied in this review.

In addition to the above-mentioned phenolic classes, there are also other popular efficient phenolic compounds, such as curcumin and resveratrol. These compounds are extensively reported to express favorable pharmacological actions in managing diverse inflammatory-based diseases. Many references have defined their efficacies towards RA by decreasing the levels of inflammatory markers and the associated matrix metalloproteinases (MMP), hindering activation of cytokines produced by T cells, and regulating the balance of Th17/Treg [24,25]. Regarding their effects on neurodegenerative diseases, it has been supported that they can inhibit the overactivation of microglial cells, downregulate the neuroinflammatory markers, and help to clear the aggregated β -amyloid [26,27]. Their effects in IBD were promising and explained by their ability to obstruct neutrophils cells infiltrations, downregulate eicosanoids and iNOS overproductions, and lower T-cell inflammatory responses [28–31]. By blockading the NF-KB pathway via upregulation of SIRT-1 and AMPK functions, resveratrol can attenuate the inflammatory cascade in metabolic disorders [32].

Despite these various positive effects of phenolic compounds on human health, they have their limitations and challenges. First of all, they are easy to degrade, due to the auto-oxidation, epimerization, hydrolysis, and crystallization during the storage period or processing [33]. Various factors may affect these instabilities, involving solvent dissolution and alkalinity, the surrounding temperature and oxygen availability, and the interaction with oxygen and other molecules [34]. Secondly, their bioavailability (ADMET) to human organisms is very poor, due to their physicochemical features. We can cite their poor water dispersibility, intestinal uptakes via the transport mechanisms, and the different metabolic reaction changes within the gastrointestinal tract throughout the digestion [6,7]. As most of the promising pharmacological potentialities of these compounds have been evaluated *in vitro* and *in vivo*, their bioefficacy in real conditions should be accompanied by a delivery system that helps their release at the targeted site of action [35]. The nanocarrier

system loaded with phenolic compounds is one of the most utilized strategies to overcome these challenges, which, in parallel, improves their physical appearances.

3. Nanocarriers Systems Used for Loading Phenolic Compounds

The recourse of nanocarrier systems in drug delivery has gained much interest since its initiation around the mid of 19th century. Physicists started to develop the conjugation of drugs to polymers in 1955, and during the first two decades, the formulation of liposomes-based drugs engaged many researchers [36]. The discovery of some liposome limitations in specific targets and the advancement of the technology led to the development of new strategies, such as the colloidal-based nanoparticles, which were rapidly approved by the U.S. Food and Drug Administration (FDA) [37,38]. Currently, a variety of nanocarrier encapsulations have been exploited, which could be classified into organic and inorganic nanocarriers. Among the organic types, we could cite solid lipid nanoparticles, lipid nanoparticles, polymeric nanoparticles, and hydrogels. However, mesoporous silica and metallic nanocarriers are frequently used as inorganic nanocarriers.

3.1. Organic Nanocarriers

Solid lipid nanoparticles (SLNPs) are formed of a colloidal system prepared by the dispersion of a solid lipid matrix, such as fatty acids, some glycerides, and steroids, in the presence of surfactants for stabilization and having a final size ranging from 50–10,000 nm [39]. The loaded compounds are incorporated within the solid lipid central core. This nanocarrier type is suitable for a wide range of administration routes and significantly ensures the stability of the loaded compound from degradation [40].

Liposomes are formed of a lipid-based vesicles shell made with amphiphilic molecules (phospholipids) and sometimes stabilized with niosomes, with a final size ranging from 250 nm to 2500 nm [41,42]. The features of liposomes can be handled by varying the type, amount, and size of phospholipids, as well as their mode of production. In general, producers add cholesterol to enhance their rigidity and stability [43]. The crucial advantage of liposomes concerns their capability to monitor the encapsulated drug release at the targeted site of action with higher molecule-loading ability, biodegradability, and low toxicity [44].

Polymeric-based nanocarriers, also known as sub-micron solid particles, are colloidal vesicles built with biodegradable polymers, whether natural (proteins or polysaccharides) or synthetic, with a final size of 10 nm to 1000 nm. Depending on the manufacturing techniques and the encapsulation mode, polymeric nanocarriers could be nanospheres, nanocapsules, dendrimers, or micelles [45]. The loaded drug is dispersed evenly within the polymer matrix in the nanosphere when they are entrapped in small cavities made by the polymeric membrane in nanocapsules [46]. Polymeric micelles are constituted by assembling amphiphilic copolymer structures, which can be di-block or tri-block, in which the loaded drugs are incorporated within the internal hydrophobic core of the micelles [46]. Beyond other nanocarriers, the drug release control can be monitored well with an extended circulating time in the organism when using this type of delivery system. It is also recognized to be more stable within the blood flow [41,47].

Hydrogels nanocarriers consist of colloid hyper-branched polymers that are capable of absorbing large amounts of fluids and can swell in an aqueous milieu. The loaded drugs are entrapped within the mesh cross-links of the gel awaiting their release after the hydrogel reaches equilibrium and are triggered by adequate stimuli (temperature, pH, ionic strength, and light) [48]. The crucial benefits of using hydrogel are the prominent increase of the drug bioavailability, in a controlled manner, and its great variety of administration routes [40]. The schematic representation of these types of organic nanocarriers is represented in Figure 2.

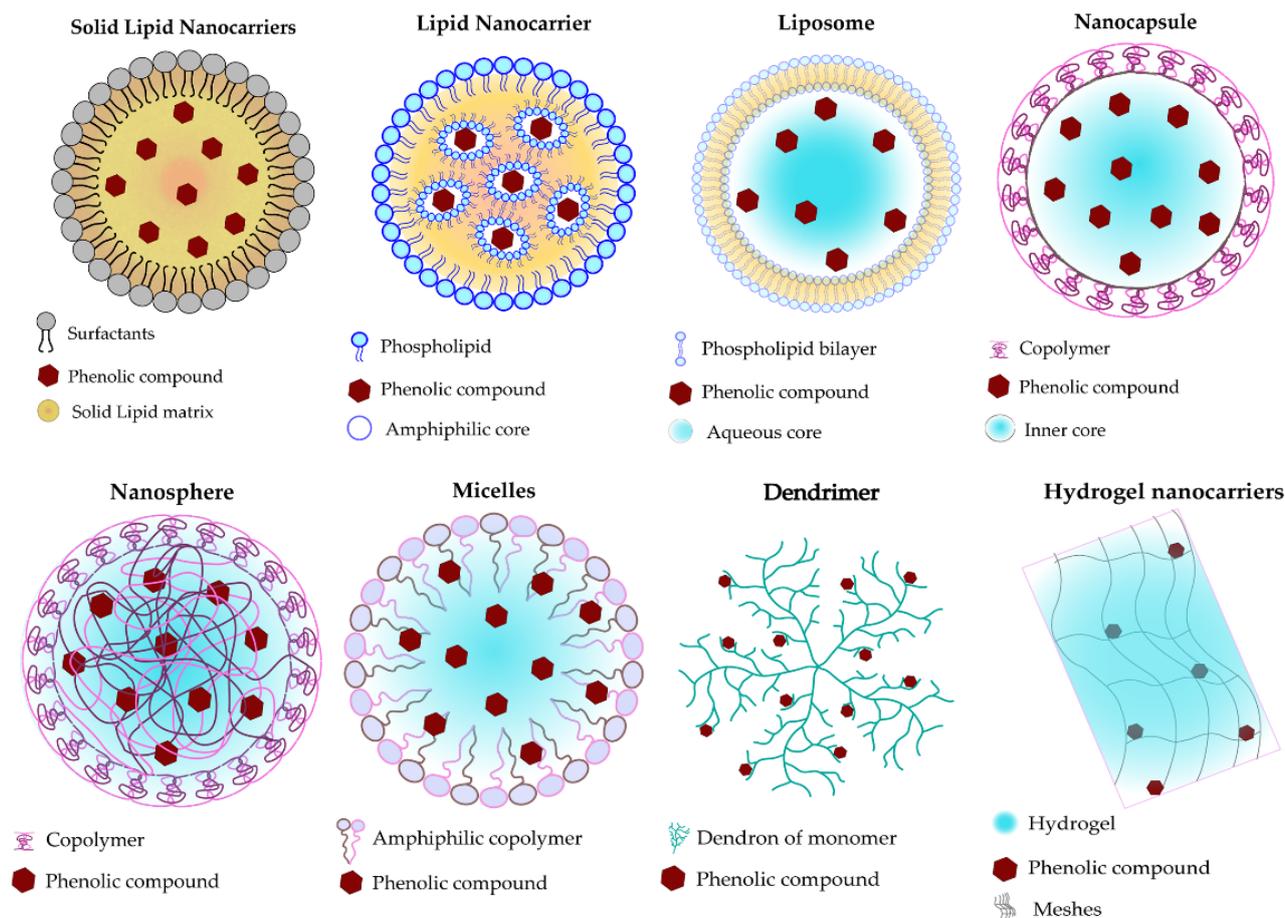


Figure 2. Schematic representation of organic nanocarriers used for loading polyphenolics.

3.2. Inorganic Nanocarriers

Mesoporous silica has received a lot of interest in drug delivery, due to its biocompatibility, high specific surface area, and porous structure, which can load an important amount of drugs [49]. All types of drugs, lipophilic or hydrophilic, can be charged within the controllable size pores (20 nm to 50 nm) [50]. The mesoporous silica is initially obtained from melting inorganic silicon dioxide (SiO_2) powders at high temperatures before leaching out the metal oxide to obtain the porosities [51].

Metallic nanoparticles are tiny particles of 1 nm to 20 nm made with stable metals, such as gold, silver, palladium, zinc, and titanium, whose surfaces are easily functionalized and facilitate the conjugation with active drugs through various interactions modes (covalent bond, hydrogen bonds, and electrostatic) (Figure 3). The drugs are then loaded at the coated surface of the metals to ensure their optimal release at the target site. Such nanoparticles could be synthesized from a bulk material size reduction approach, such as lithography and laser ablation, or via atom assembling techniques, such as flame spraying, laser pyrolysis, and microemulsion [52]. Depending on the delivery purposes, metallic nanoparticles can be coated with non-ionic or charged coatings with diverse functions (polymers, surfactants, etc.) [53,54]. The tiny size of metallic nanoparticles allows them to link smoothly with the organism's biomolecules, which warrants the specific target area delivery and fewer side effects [55]. It is of note that the functionalization or conjugation of nanocarrier systems are made with targeting ligands, peptides, proteins, or antibodies to promote the recognition of the complex to the targeted cells receptors and achieve active targeting [56,57].

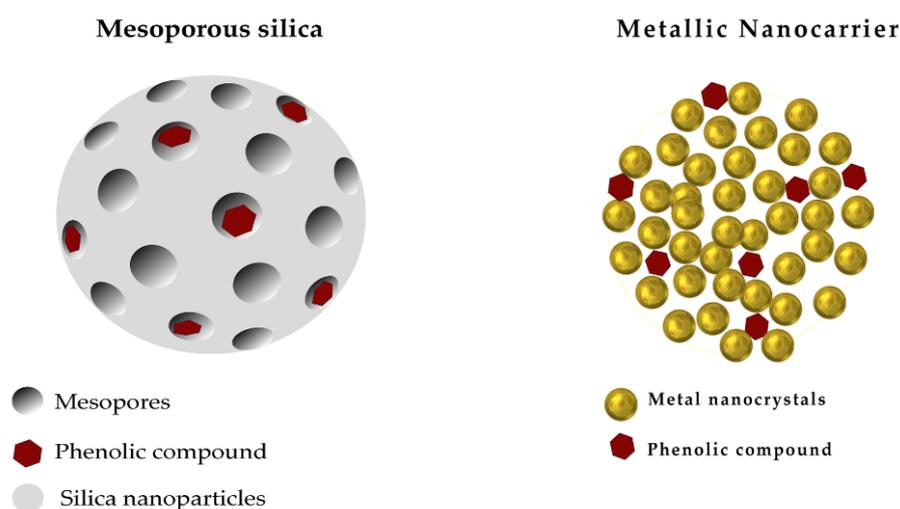


Figure 3. Schematic representation of inorganic nanocarriers used for loading polyphenolics.

4. Designed Polyphenolic-Loaded Nanocarriers for Inflammation-Mediated Diseases

4.1. Polyphenolic Nano Delivery for Rheumatoid Arthritis

Rheumatoid arthritis is one of the inflammation-mediated diseases and is characterized by continuous inflammation of the synovial joints, and it can result in lasting suffering, disability, and even early death [58]. The joints start swelling when the synovium is subjected to hyperplasia by an inner proliferation of macrophages and fibroblast cells. Additionally, this hypertrophied synovium invades the adjacent tissues, such as the cartilage and bone surfaces, that enhance inflammatory responses. Therefore, it has been reported that high amounts of pro-inflammatory mediators, such as TNF- α , IL-1, IL-6, and IL-17, are quantified in RA [59]. Additionally, this TNF production has been reported to be triggered by the autoantibodies ACPAs. However, these cytokines mitigate the main function of the cartilage-replenishing matrix and chondrocytes through nitric oxide (NO) stimulation, as well as other pro-inflammatory signal releases [60].

Recent investigations have improved the delivery of phenolic compounds for managing arthritis. Zheng et al. have improved the antiarthritic of EGCG by formulating optimized casein nanoparticles loaded with an EGCG glucosamine matrix (EGCG/GA/casein). This formulation dispersion was found to be highly stable, even after 12 months of storage, and showed excellent inhibitory effects on fibroblast-like synoviocytes arthritis cells in vitro. Similarly, the oral administration of EGC-NPs suppressed arthritis aggravation in collagen-induced arthritis (CIA) rats, which was explained by significant down-regulation of TNF- α , IL-1 β , IL-6, and IL-8 cytokines [61]. Another nanomicelle loaded with a combination of the antiarthritic drug 9-aminoacridine (9AA) and caffeic acid was also developed by conjugating caffeic acid with methoxy polyethylene glycol (mPEG) and ϵ -caprolactone (mPEG- β -PCL) to form 9AA-NMs. The intraperitoneal injection of 9AA-NMs reduced the severity of arthritis inflammation in CIA Wistar rats, which was due to the synergistic activities on the blocking NF- κ B pathway and triggering the orphan nuclear factor NR4A1 in arthritis [62].

In terms of flavonoids, a comparison of different solid lipid-based nanocarriers (stearic acid, stearic-lauric, or lecithin-chitosan) loaded with naringenin resulted in the effective activity of this encapsulation within stearic-lauric (Nar-SL). In addition to the extended releases, Nar-SL also diminished the inflammatory factors, together with the degradation of the joint in CFA-induced RA. The complementary anti-inflammatory action between naringenin and lauric acid was supposed to be the principal reason [63]. Another study by Mohanty's research group, in 2020, combined naringenin with phenethyl isothiocyanate (PEITC) in a liposome carrier (1,2-dipalmitoyl-sn-glycero-3-phosphocholine/cholesterol/1,2-stearoyl-sn-glycerol-3-phosphoethanolamine-020CN (DPPC/Chol/DSPE-

020CN)). The intraperitoneal injection of this combinatorial formulation for 3 weeks increased the anti-inflammatory cytokine IL-10 level and attenuated bone erosion. The immune cell infiltrations in Freund's complete adjuvant (FCA)-induced arthritis were also hindered. A synergistic enhancement of the antiarthritic action of naringenin and PEITC was determined from this preparation [64]. The same research group lately investigated the delivery optimization of naringenin encapsulated within a biodegradable polymer PLGA (NAR-PLGA-NPs). A sustained release of the loaded compound in the intestine was discovered when stabilizing NAR-PLGA-NPs with poloxamer and sodium dioxyolate. In the meantime, a significant increase of the anti-inflammatory marker IL-10, as well as the attenuation of RF and C-reactive protein (CRP) releases, were discovered in arthritic-induced rats [65]. A nanocrystal strategy through the wet milling of naringenin with a copoloxamer (F127) was also applied and resulted in bioavailability improvement and synovial damage, as well. Oral administration of nanocrystal has increased the cellular uptake and diffusion in CIA-induced RA model [66].

Metallic nanocarriers, loaded with resveratrol, using ruthenium as the core product and coated with PLGA-dextran sulfate (QRu-PLGA-RES-DS), were developed to improve the bioefficacy of resveratrol in RA. The self-assembly of the formulation has enhanced the bioavailability and circulating time of resveratrol, while triggering macrophage polarization to the M2 phenotype. Meanwhile, M1 macrophage infiltration at the synovia was reduced, which signifies the antiarthritic strength of this elaborate core-shell structure [67]. A mixed micelle, built of a mixture of poloxamer 188 and poloxamer 407 coated with poly-lactic acid (PLA), was also designed to load resveratrol for intra-articular injection. One week of the co-micellar injection on CFA-induced RA was enough to reduce the synovium swelling, which correlated with the reduction of TNF- α levels, as well as the cartilage replenishment [68]. Combinatorial application of methotrexate and resveratrol (MTX-RSV) nanoemulsion loaded in gel (carbopol 940) showed a greater inhibition of inflammation ($78.76 \pm 4.16\%$) and anti-arthritic effects *in vivo* than those loaded individually [69]. Likewise, the oral administration of resveratrol loaded in hydrogel (cellulose aerogel) was demonstrated to exert a great impact on arthritis model management. In addition to the bioavailability amelioration of resveratrol, this delivery system also helped to induce the anti-inflammatory effects by downregulating the P38 pathway and activating SIRT-1 expressions [70].

When loaded alone in solid-liquid nanocarriers, curcumin showed promising potential in ameliorating the synovial inflammation of CFA-induced models by reducing immune modulation and inflammatory cascade [71]. However, a co-encapsulated formulation of resveratrol and curcumin in lipid nanoparticles showed pronounced antioedematogenic efficiency and cartilage damage protection than the single-loaded standard in CFA-induced models. The lipid core of the product was formed of grape seed oil, and the emulsifier was sorbitan monostearate and polysorbate 80, in which an equal amount of the two phenolic standards were incorporated [72]. On the other hand, a HAS-based developed nanoparticle was recently developed to co-deliver curcumin and prednisolone. This latter exerted a more extended circulating time of curcumin before release, due to the attachment of the loaded drug in the albumin mesh. Furthermore, the synergistic anti-inflammation effect of curcumin and prednisolone was discovered with an apparent accumulation at the targeted site of action [73]. Another curcumin delivery carrier using gel nanoemulsion was developed for its anti-arthritic effect. The nanoemulsion increased the permeability of the carrier and resulted in a significant decrease of inflammatory markers, notably TNF- α and IL-1 β , while its gelation within carbopol-980 enhanced the topical anti-arthritis efficacy on CFA-induced model paw [74]. The incorporation of curcumin in polymeric nanocarrier, based on carboxymethyl cellulose acetate butyrate (CUR-CMCAB) resulted in a promising release time and anti-arthritic efficiency when taken orally by CFA-induced rats. These were explained by the complete dispersal of curcumin to its amorphous form in the polymeric shell and the improvements seen from the different

behavior testing along the treatment [75]. In 2018, Fan et al. entrapped curcumin in hyaluronic acid to form nanomicelle particles, in which the intraperitoneal injection significantly decreased the swelling of CFA-induced arthritis models. A reduced level of VEGF expression and inflammatory cytokines was determined following the treatment. In parallel, it improved the lubrication of joint cartilage in arthritis [76]. Kang and coworkers formulated an acid-triggered micelle particle that releases instantly the loaded curcumin once in an acidic environment. In monosodium iodoacetate (MIA)-induced osteoarthritis, the injection of this micelle solution blocked pro-inflammatory factors expressions, such as IL-1 β and TNF- β . In addition, this latter protected the degradation of the cartilage in the synovium [77]. Figure 4, below, illustrates some examples of applications of phenolic-loaded nanocarriers applied in rheumatoid arthritis.

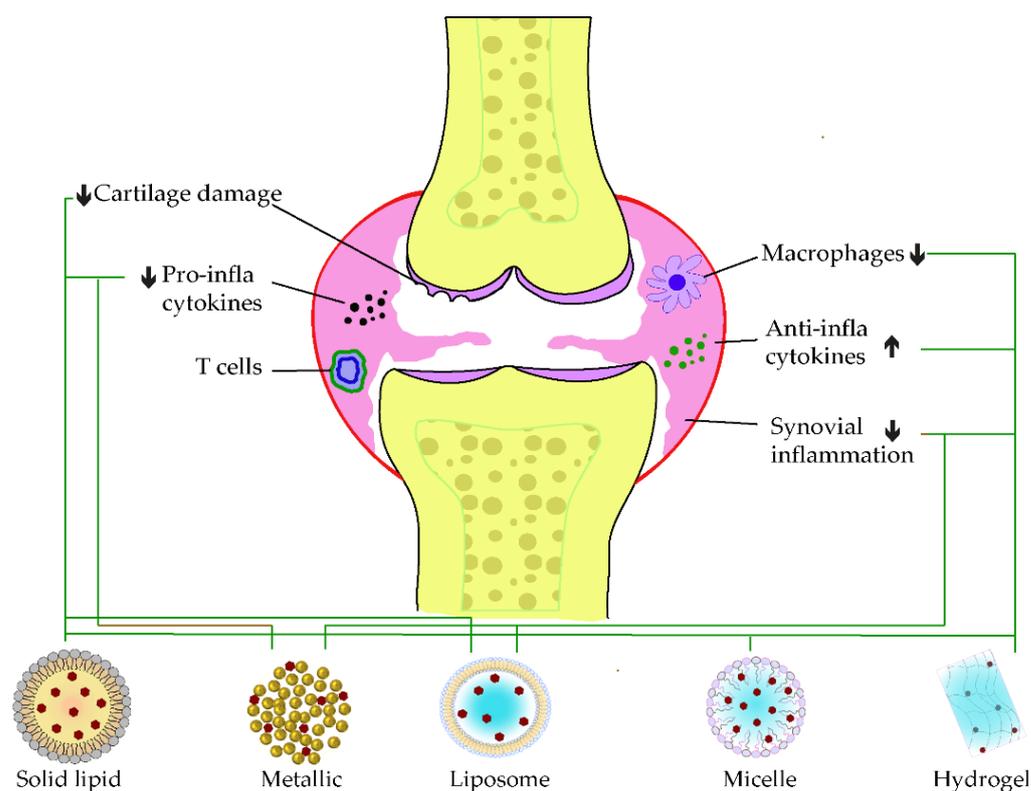


Figure 4. Examples of nanocarrier delivery of polyphenolics and their targets in Rheumatoid arthritis. The black arrows up signify the increasing while the down ones indicate the decreasing effects. The black dots in the figure consist of pro-inflammatory cytokines, the red dots are anti-inflammatory cytokines, and the yellow dots represent the bone marrow. The bold lines match the nanocarriers and their respective targets.

4.2. Polyphenolic Nano Delivery for Neurodegenerative Disease

Neurodegenerative disease is among the crucial health-threatening issues affecting learning abilities and memory, especially for old people. It is commonly manifested in the form of Alzheimer's and Parkinson's diseases. In addition to the abnormal aggregation of neuronal proteins, recent findings have demonstrated that inflammation plays a critical role in the pathogenesis of this disease [78]. Alzheimer's disease (AD) is characterized by the extraneuronal aggregations of amyloid- β ($A\beta$) proteins, intraneuronal formation of neurofibrillary tangles from hyperphosphorylation of tubulin-associated unit (tau) protein, and neuronal damages [79]. Parkinson's disease (PD) mostly impacts the motor system. The hallmark of this disease consists of a midbrain gradual loss of dopaminergic neurons, accompanied by the intracellular aggregation of α -synuclein (α -syn), also known

as Lewy bodies (LB) [80]. Likewise, neuro-inflammation caused by microglia plays a pivotal function in the pathogenesis and development of PD.

A great number of delivery systems loaded with phenolic compounds have been pioneered for improving their neuroprotection efficiency and target deliverance. It has been reported that nanoparticles formed with EGCG in PLVA-PEG-PLA (nanoEGCG), when administrated orally, not only improve the bioavailability of EGCG, but also mitigate the release of A β plaques, as well as the levels of A β ₁₋₄₂, in Alzheimer-induced rats. Additionally, this nanoEGCG was attested to reduce the expression of amyloid precursor proteins, acetylcholinesterase, and glycogen synthase kinase-3 beta (GSK-3 β), while the levels of 3-phosphoinositide-dependent protein kinase-1 were found to be rising [81]. Another work of Cano and coworkers reported that polymeric nanocarriers built with co-entrapped EGCG and ascorbic acid within PEGylated-PLGA can smoothly pass the blood-brain barrier (BBB) and enhance the neuro-inflammatory effects of EGCG, together with synaptogenesis in APP/PS1 Alzheimer mice. Similarly, the aggregation of A β plaques and A β ₁₋₄₂ levels declined [82]. Given the effectivity of PEGylated-PLGA encapsulation, recent work has mended EGCG loading by co-encapsulation with shRNA, a β -site APP cleaving enzyme antisense (BACE1-AS), and the RGV29 peptide. This nanocarrier formulation sustained the circulation time of the loaded drugs in the bloodstream and facilitated their ability to pass across the BBB. Their neuroprotection effects at the targeted site, the brain, were promising by reducing A β levels and mitigating BACE1 function [83]. In PD, Li et al., 2020, formulated diverse DSPE-PEG B6-based micelles coated with EGCG (B6ME-NPs). To ease their traceability throughout the deliverance, they incorporated SPIONS in the core shell of the particle. The intravenous administration of these formulations has improved the EGCG stability and delivery to the brain by B6 contribution in overcoming the BBB and inhibited noticeably the aggregation of α -syn agglomeration in vitro [84]. Selenoprotein analogs (Tet-1) conjugation of selenium nanoparticles coated with EGCG (Tet-1-EGCG@Se) has also well-enhanced the inhibition of A β aggregation and fibrillation in AD. The Tet-1 functionalization was found to facilitate the cellular uptake of the formulation, while Tet-1-EGCG@Se reversed the fibrillation of A β agglomerate to its native form [85]. A recent investigation showed the effective action of EGCG-Se nanoparticles in neuroprotection via reducing the microglia inflammation in LPS-induced PC12 cells (in vitro) and in the targeted site of a spinal cord injury rat model (in vivo) [86].

Rutin loaded in solid lipid nanoparticles has also shown great stability and circulation time in the blood flow. In addition, this nanoparticle can well-infiltrate the BBB, which suggests its efficient brain target delivery [87]. Similarly, oral administration of phospholipidic nanocarrier, incorporated with rutin, exhibited eminent brain concentration and attenuated neurological, as well as ischemic, damage in middle cerebral artery occlusion (MCAO) rats [88]. A metallic nanocarrier loaded with rutin and coated by the traceable red congo facilitated the localization of A β plaques by imaging. Additionally, this theranostic nanoparticle significantly attenuated the cytotoxicity of the A β and the release of oxidative products in AD [89]. Other antiparkinson's flavonoids, such as apigenin, were encapsulated in the phospholipid nanocarrier before nasal administration and proved to be delivered well within the brain, while upregulating dopamine releases in haloperidol-induced Parkinson's models [90]. Naringenin nanoemulsion in capryol, tween 20, and water were also highlighted to mitigate amyloidogenesis in A β -induced SH-SY5Y by reducing the expression of APP, BACE, and phosphorylated tau in vitro [91]. Likewise, the nasal administration of co-loaded nanoemulsion of naringenin and Vit E reversed the MDA levels in 6-OHDA-induced PD rats [92]. A polymeric nanocarrier system using modified polycaprolactone was also used to encapsulate naringenin for the treatment of cerebral stroke. It was established that this nanoformulation can impede the release of pro-inflammatory factors and markers in deprived oxygen glucose-induced mesenchyme stem cells (MSC), which suggests their effectiveness in MSC-based ischemic treatment [93].

Some investigations have demonstrated the effects of encapsulating phenolic acids for neurodegenerative disease treatment. For example, by encapsulating gallic acid in chitosan nanoparticles, its circulation time after oral administration was prolonged. Additionally, it downregulated the secretion of pro-inflammatory factors, reduced infarction size, and alleviated neuronal impairments in ischemic rats [94].

For its part, resveratrol has been entrapped in lipid core nanoparticles to improve its therapeutic target delivery. The results established that the said formulation enhanced the neuroprotective activity in AD by attenuating the releases of neuroinflammatory markers and blocking kinases pathways (JNK, GSK-3 β) [95,96]. Another type of encapsulation in PLA-PAS80 of resveratrol also showed promising neuroprotection effects in MPTP-induced mice [97]. However, Loureiro et al. optimized an ameliorated solid lipid nanocarriers functionalized with an anti-transferrin monoclonal antibody (OX26 mAb), which was defined as more efficient in BBB crossing and amyloidogenesis modulation [98]. Recent works have also evidenced the therapeutic enhancement of selenium nanoparticles loaded with resveratrol. These formulations not only reduced the A β aggregation in AD, but also impeded GSK-3 β -mediated tau hyperphosphorylation and neuroinflammatory signaling pathways (STAT3, NF- κ B, MAPK, and Akt) in AlCl₃-AD rat models [99,100].

In 2014, an investigation on curcumin loaded in lipid-PEG conjugated with a monoclonal antibody in AD brain patients exhibited a high uptake of the said formulation and good clearance of A β aggregates [101]. Similar effects were demonstrated when conjugating curcumin-loaded polymeric nanocarrier (PLGA-CU) with glycopeptide (g7), brain targeting peptide (CRT), or serum albumin (BSA). It was discovered that these latter could enhance the BBB crossing and enhanced the phagocytosis of A β peptides via the modulation of macrophage polymerization [102–104]. On the other hand, solid lipid nanocarrier has shown better anti-amyloid and anti-inflammatory effects in AD models of 5XFAD mice, due to its better brain deliverance and glial astrocytes clearance [105]. Curcumin loaded in selenium nanoparticles covered with PLGA was also established to diminish extracellular A β peptides burden at the targeted AD lesion of transgenic mice (5XFAD) [106]. A co-loaded strategy of curcumin with piperine in a lipid nanocarrier built of glyceryl monooleate (GMO) and surfactants was also developed. This formulation has shown interesting attenuation of the α -syn fibrillation and toxicity induced by rotenone. Furthermore, it activated the anti-apoptotic of neuronal cells and the autophagy of fibril peptides, which ameliorates the neuronal abnormalities in PD models. The co-loaded drugs were stated to work synergistically once released to the PD target site [107]. The recent applications of phenolic-loaded nanocarriers in neurodegenerative diseases are synthesized in Table 1.

Table 1. Application of phenolic-loaded nanocarriers in neurodegenerative diseases.

Phenolic Compounds	Type of Nanocarriers (Size)	Targets	Mechanism of Action	Ref.
EGCG	Hybrid nanocarriers: EGCG@Se-Tet-1 coated	PC12 and NIH/3T3 cells	Disrupts A β aggregation and mitigates A β fibrillation. Protects PC12 cells from damages	[85]
EGCG	Polymeric nanocarriers: PEGylated PLGA (124 nm)	Alzheimer mice model (APP/PS1 mice)	Improves EGCG stability and efficacy by reducing neuroinflammation, A β plaque/peptide burden, and enhancing synaptogenesis	[82]
EGCG	Polymeric nanocarriers: PLA-PEG-PVA-EGCG (317 nm)	AlCl ₃ -induced neurobehavioral deficit model	Attenuates neurobehavioral impairments, reduces the generation of A β plaque, neurofibrillary tangles, level of A β ₁₋₄₂ , and Ache activity	[81]
EGCG	Micelle nanocarriers: PEG-PLGA-shRNA-RVG29 (152 nm)	BACE1, APP/PS1 mice	Reduces A β and BACE1 protein expression and enhances synaptogenesis, memory, and learning processes	[83]
EGCG	Selenium nanocarrier: Selenium (91 nm)	LPS-induced PC12 cells and SCI rats	Reduces the levels of inflammatory cytokines, improves locomotor capacity, and diminishes injury region	[86]

EGCG	Micelle nanocarriers –Hybrid (B6ME-NPs with SPIONS)	SH-SY5Y neuroblastoma cells	Improves the delivery to PD lesions and inhibits α -syn aggregation	[84]
Rutin	Phospholipid nanocarrier: RU-RPLC (<100 nm)	MCAO induced rats	Reduces toxicity and infiltrates well in BBB	[88]
Rutin	Metallic nanoparticle: Iron oxide-congo red-MNPS (oleic acid coated)	A β -induced SH-SY5Y cells	Inhibits A β -induced cytotoxicity and reduces the production of NO and ROS	[89]
Rutin	Solid lipid nanocarrier 100 nm	<i>Rattus norvegicus</i> rats	Infiltrates well in BBB	[87]
Apigenin	Lipid nanocarrier	Haloperidol-induced Parkinson's model	Upregulates dopamine level	[90]
Naringenin	Nanoemulsion: Capryol 90-tween 20 (113 nm)	A β - induced SH-SY5Y cells	Decreases A β levels, APP, BACE, and tau expressions	[91]
Naringenin	Nanoemulsion: Capryol 90-tween 80-Vit E (38 nm)	6-OHDA-induced Parkinson's disease in rats	Reverses PD symptoms in rats	[92]
Naringenin	Polymeric nanocarrier: PCL-gelatin-coated (192 nm)	OGD-induced mesenchymal stem cells	Reduces the levels of proinflammatory cytokines and biomarkers	[93]
Gallic acid	Lipid nanocarrier: GA-NPs	MCAO rats	Presents neuroprotection	[94]
Resveratrol	Lipid nanocarrier: RSV LNC (242 nm)	A β ₁₋₄₂ -induced rats	Rescues deleterious effect of A β ₁₋₄₂	[96]
Resveratrol	Lipid core nanocapsule: RSV-LNC (242 nm)	Hypocampal culture	Modulates neuroinflammation	[95]
Resveratrol	Polymeric nanocarrier: PLA-PS80	MPTP-induced C57BL/6 mice	Presents neuroprotection	[97]
Resveratrol	Solid lipid nanocarrier: RSV-SLN-OX26 (254 nm)	Endothelial human brain	Enhances A β -aggregation clearance, and blocks A β fibrillation	[98]
Resveratrol	Res-selenium-peptide: TGN-Res@SeNPs	AIC13-AD model mice	Decreases A β aggregation in the hippocampus, and downregulates neuroinflammation	[99]
Resveratrol	RSV-Se nanoparticles (RSV-SeNPs)	AIC13-AD rat model	Attenuates the mitochondrial dysfunction, downregulates inflammatory pathways: STAT3, GSK-3B, and triggers PI3K	[100]
Curcumin	Polymeric nanocarrier: PLGA-CU-g7 (204 nm)	Hippocampal cell cultures	Reduces the aggregation of A β	[102]
Curcumin	Solid lipid nanocarrier: SLNP-CU (Ni)	5XFAD mice	Improves neuroprotection and reduces astrocytic and microglia immune responses in AD-mice	[105]
Curcumin	Polymeric nanocarrier PLGA-NPs-CRT (128 nm)	(APP/PS1dE9) AD mouse model	Diminishes A β burden, astrogliosis, microgliosis, and memory impairment	[103]
Curcumin	Polymeric Se-NP-PLGA (160 nm)	(5XFAD) model	Diminishes A β burden and reduces memory impairment	[106]
Curcumin	Nanoliposomes Lip-PEG-CU-mAb (153 nm)	AD brain patients	Interrupts A β ₁₋₄₂ aggregations	[101]
Curcumin	Polymeric nanocarrier: Chit-BSA (143 nm)	hCMEC/D3 BBB cell model	Reduces NF- κ B signaling and improves A β ₄₂ uptakes	[104]
Curcumin	Lipid nanocarrier: Cur-Pip-GMO-based NPs (93 nm)	Rat cell line, a cellular model of PD,	Inhibits α -syn aggregation and stimulates autophagy	[107]

4.3. Polyphenolic Nano Delivery for Skin Inflammation and Wound

Most dermatological problems are issued by inflammation reactions. However, the inflammation of the skin represents chronic immune-inflammatory diseases involving psoriasis, dermatitis, and lupus erythematosus [108]. The psoriatic lesion is specified by epithelial hyperplasia, irregular differentiation of keratinocytes, infiltration of different inflammatory cells in the dermis, and vascularization [109]. The occurrence of these keratinocytes and immune cells enhances the release of pro-inflammatory factors around the lesions, which progressively amplify the psoriatic disease [110]. Atopic dermatitis, however, is associated with severe pruritus, persistent skin inflammation, and impaired skin barrier function [111]. Pathogenesis of this disease includes the upregulation of Th-2-driven inflammation, immunoglobulin E (IgE), and T-cells expressions [112].

The application of a nanodelivery system to ameliorate the efficacy of phenolic compounds has gained researchers' attention during the last decade. Diverse strategies have been applied and achieved more advancement in the exploration of the enhanced method to deliver these kinds of compounds. With regards to EGCG, when entrapped in chitosan polymeric nanoparticles (CHI-EGCG-NPs), it has shown a four-fold effectivity in reducing inflammation responses and the proliferation of cultured keratinocytes. The topical application of this formulation on the imiquimod (IMQ)-induced skin model resulted in a significant decrease in immune cell penetration, as well as diverse psoriasis-related inflammatory markers [113]. Similarly, in dermatitis, the encapsulation of this catechin derivative in PEG-PLGA ameliorated the blockading of necroptosis (RIP 1, RIP 3, and MLKL expressions) in the 2,4 dinitrochlorobenzene (DNCB)-induced skin model, while the topical application resulted in noticeable downregulation of inflammatory cytokines releases. In the meantime, the synthesis of MAPK pathways (p-p38, ERK1, ERK2) in the epidermal layers of the dermatitis was also suppressed [114]. Co-loading of EGCG with Vit C in chitosan was established to quicken the wound healing of streptozocin-induced mice when administered intraperitoneally. The promotion of collagen deposition and angiogenesis, together with the blockage of immune cell infiltrations in the wounds, was speculated as the main synergistic effects of the co-delivered drugs [115]. A hydrogel nanocarrier entrapping EGCG was formulated for wound dressing. When applied to wounded rats, the hydrogel patch showed better wound healing efficacy than the commercial Neuskin[®] tape, a mechanism which was determined as the regulation of inflammatory and growth factors, together with the promotion of collagen disposition [116].

A carbopol-based hydrogel nanocarrier incorporated with rutin nanocrystals was also developed to increase the efficiency of rutin in wound healing. The transdermal application of this preparation showed the protection of epidermal tissues induced by UV radiation [117]. An optimized ethosome formulation of rutin in lipid and ethanol has been also proven to protect against skin disorders, through their enhanced anti-inflammatory activity on keratinocytes cells and patient volunteers [118]. In the same way, optimal apigenin-loaded ethosome expressed a significant reduction of COX-2 levels in the UV-induced skin mice model [119]. Another work, conducted by Pleguezuelos et al., established a liposome formulation loaded with naringenin and tested for biocompatibility in 3T3 fibroblast cells. In vivo assessment demonstrated a better delivery of naringenin in the epidermis of TPA-induced skin mice, as well as great inflammatory-reducing effectivity [120]. A chitosan-coated liposome of naringenin was recently developed and proved to ameliorate the wound of experimental rats. The synergistic effects of chitosan and naringenin were stated as the principal therapeutical mechanisms of the optimized product in wound dressing [121].

Delivery trials of phenolic acids for skin inflammation have also been conducted. For illustration, the encapsulation of gallic acid in a polymeric nanocarrier composed of a lipid, glycosome, and poloxamer has increased its deliverance in the skin. More importantly, the topical treatment of TPA-induced skin mice exhibited better anti-inflammatory effects, as explained by the impediment of leukocyte infiltration and activation [122]. Co-encapsulation of gallic acid with rutin in polymeric nanovesicle also exhibited positive

effects in psoriasis treatment. It acts by blockading the hyperproliferation of keratinocytes, as well as protecting from psoriasis-related inflammation [123]. Recent works have established the effectiveness of hydrogel embodiment with gallic acid in wound healing. Topical application of such gallic acid nanovesicle speeds up wound regeneration and demonstrated to downregulate the expression of inflammatory cytokines [124,125]. Caffeic acid conjugated on nanofiber nanovesicle, formed with PCL and CH, ameliorated its delivery on fibroblast neonatal cell line (NHDF-neo) [126]. While conjugated on PLGA nanofiber, caffeic acid expressed better biocompatibility and wound dressing actions in the human fibroblast model [127].

Resveratrol was co-loaded with quercetin in lipid nanoparticles (oleic acid-S75) to improve their cellular intakes. When applied topically to a TPA-induced wound in mice, it rapidly regenerated the skin lesion by lowering the permeation of inflammatory cells [128]. An optimized co-loading of resveratrol and omega-3 (ω 3) in lipid nanosystem has strengthened their anti-inflammatory potential in skin lesion diseases. The said formulation displayed longer circulation time and better inhibition of COX-2 and NO production in LPS-induced cells [129]. The incorporation of resveratrol in peptide hydrogel also boosted its wound-healing effectiveness by hindering the release of macrophage cytokines and promoting collagen restructuring [130]. Likewise, the hybrid composite encapsulation of curcumin by loading into polymeric (MPEG-PCL) or micelles (PEG-PCL, or PEG-PLA) vesicles before incorporation in hydrogel carriers (chitosan or dextran) resulted in sustained releases with a rapid wound dressing in vivo. Applications of these formulated hydrogels not only triggered the proliferation of fibroblasts, but also enhanced collagen agglomeration and angiogenesis in the tissue lesion [131–133]. This system also had positive therapeutic effects in the treatment of psoriasis, in which the pre-encapsulation in nanocarrier was supported to enhance the deliverance of curcumin to the targeted site [134]. Adhering curcumin on electrospun nanofibers was also developed for wound healing. Dai and coworkers developed curcumin loaded in gelatin nanofiber and discovered its advantages in inhibiting macrophages, as well as their cytokines releases [135]. Moreover, the encapsulation of silver nanoparticles functionalized with curcumin in chitosan was proven to be efficient in reducing wound lesions, without secondary irritation effects in the experimental model [136]. Lee and his team have developed a lipid nanocarrier co-loaded with epidermal growth factor and curcumin for treating diabetic chronic wounds. Their finding suggested that, in addition to the potential antioxidant of the formulation, it also helped in triggering the migration of fibroblasts and keratinocytes for wound closure [137]. Similar fibroblast migration was also determined when applying silica-based nanoparticles (CU-Si Np) loaded with curcumin on dermal fibroblast cells (HDF fibroblast) [138]. The detailed application of these different nanocarriers system loaded with phenolics and their mechanism of actions toward skin inflammation are summarized in Table 2.

Table 2. Application of phenolic-loaded nanocarriers in skin inflammation.

Phenolic Compounds	Type of Nanocarriers (Size)	Targets	Mechanism of Action	Ref.
EGCG	Polymeric nanocarrier: PEG-PLGA (176 nm)	DNCB-induced dermatitis model	Reduces ear and skin thickness, mitigates the inflammatory cytokines releases (TNF- α , IFN- γ , IL-4, IL-6, and IL-17A), blockades necroptosis (RIP1, RIP3, and MLKL expressions), and regulates MAPK pathways (p-p38, ERK1, and ERK2)	[114]
EGCG	Polymeric nanocarrier: CHI-EGCG-NPs (211 nm)	Cultured keratinocytes and IMQ-induced model	Reduces inflammatory responses and modulates psoriasis-related inflammatory cytokines	[113]
EGCG	Hydrogel nanocarrier: HG-Ag-EGCG (217 nm)	Subcutaneous wound in Wistar rats	Accelerates wound properties (modulates growth factors and cytokines)	[116]

EGCG	Polymeric nanocarrier: EGCG-Vit C-Gelatin, chitosan (200 nm)	Wound on STZ-induced diabetic mice	Promotes wound healing by raising collagen accumulation, promoting angiogenesis, and reducing inflammatory cell infiltrations	[115]
Rutin	Hydrogel nanocarrier: NC-RU-gel (447 nm)	UV-induced BALB/c mice	Increases the skin permeability of rutin	[117]
Rutin	Ethosomal nanocarrier: ETOH-PL90G-H ₂ O (112 nm)	Keratinocyte cells (NCTC2544) and volunteer patients	Improves anti-inflammatory effect	[118]
Apigenin	Ethosomal nanocarrier (36 nm)	UV-induced skin inflammation	Reduces COX-2 levels	[119]
Naringenin	Liposome: NAR-Polysorbate 80-Lipoid® (100 nm)	3T3 fibroblasts and TPA-induced mice	Reduces skin inflammation	[120]
Naringenin	Liposome: chitosan-coated naringenin nanoemulsion (105 nm)	Abrasion wound in rat model	Controls the delivery and ameliorates the wounds construction and skin regeneration	[121]
Gallic acid	Polymeric nanocarrier: Tween 80-chitosan (330 nm)	HaCaT cell line	Reduces keratinocyte proliferation and exerts protein protection in vitro.	[123]
Gallic acid	Hydrogel nanocarrier	Total skin defect model	Enhance wound healing by reducing the expression of IL-6, IL-1 β , and TNF- α	[125]
Gallic acid	Hydrogel nanocarrier: GH/GGA	Skin wounded mice	Speeds up the wound healing by scavenging the ROS and promoting tissue regeneration	[124]
Gallic acid	Polymeric nanocarrier: liposome, glycosome GA polyxomer (70 nm)	TPA induced mice	Improves the skin target delivery and blockades leukocytes infiltration	[122]
Caffeic acid	Nanofiber nanocarrier: PLGA	In vitro scratch assay	Presents better wound healing properties on human fibroblast	[127]
Caffeic acid	Nanofiber nanocarrier: Chitosan-PCL/CCA	NHDF-neo cell line	Improves cell attachment	[126]
Resveratrol	Lipid nanocarrier: Oleic acid-S75 (79 nm)	TPA induced mice	Neutralizes the inflammatory response	[128]
Resveratrol	Lipid nanocarrier: DSPC/DOPE/ ω 3 (156 nm)	RAW 264.7 cell line	Inhibits COX and NO productions	[129]
Resveratrol	Peptide-hydrogel nanocarrier: Fmoc-FFGGRGD	Rat skin damage model	Inhibits macrophage production of pro-inflammatory cytokines	[130]
Curcumin	Hydrogel nanocarriers: PLGA NPS in hydrogel (150 nm)	IMQ-induced-C57/BL6 mice	Improves anti-psoriasis activity	[134]
Curcumin	Gelatin nanofiber mats	Rat wounded model	Improves the wounds by increasing fibroblast proliferation and migration, inhibiting macrophages, and reducing pro-inflammatory cytokines	[135]
Curcumin	Nanocrystal coated nanocarrier: Ch/CNC (Ag NPx/Cury)	Injured rat model	Accelerates wound closure and repairs tissues	[136]
Curcumin	Silica nanocarrier: CU-Si-Nps (36 nm)	HDF fibroblast cell	Enhances fibroblast migrations	[138]
Curcumin	Nanostructured lipid carrier: EGF-Cur-NLC (331 nm)	Punched wound on the skin	Accelerates wound closure	[137]
Curcumin	Hydrogel nanocarrier: PEG-PLA in dextran hydrogel (65 nm)	BALB/c mice	Accelerates angiogenesis, fibroblast accumulation, and wound healing	[133]

Curcumin	Hydrogel nanocarrier: MPEG-PCL in CCS-OA hydrogel (nano: 40 nm)	Injured tissue	Improves re-epithelialization of the injury	[131]
Curcumin	Hydrogel nanocarrier: PEG-PCL-PEG in hydrogel (micelle: 26 nm)	Wound model	Enhances cutaneous repair, and increases collagen content and wound maturity	[132]

4.4. Polyphenolic Nano Delivery in Inflammatory Bowel Disease

Inflammatory bowel disease (IBD) is a relapsing gastrointestinal tract (GIT) chronic inflammatory that comprehends ulcerative colitis and Crohn's disease [139]. IBD is featured by an irregular immune response. Its pathogenesis is linked with the upregulation of NF- κ B pathways, along with the spectacular recruitment of epithelial and immune cells in the lamina propria [140]. Therefore, a rise in the transcription process increases the synthesis of inflammatory mediators involving interleukins (IL-1 β , IL-6, IL-12, IL-23) and tumor necrosis factors (TNF- α). In IBD the highly expressed cytokine by T cells is the interferon- γ (IFN γ) that mainly contributes to the blood vessel's vasculatures, apart from its immunomodulatory action [141]. However, the IL-13 cytokine, expressed by the natural killer cells (NK cells), is in charge of the epithelial barrier interruption [142]

In recent years, investigations on the delivery systems of phenolics for the treatment of inflammatory bowel diseases have emerged. An optimized polymeric nanosystem for the delivery of small interfering RNAs (siRNAs) complexed with EGCG was developed. The intrarectal administration of the obtained supramolecular showed significant decreases of TNF- α , prolyl hydroxylase 2 (PDH2), and inflammatory cell infiltration in the intestine tissue of a dextran sulfate sodium (DSS)-induced model [143]. In another study, ovalbumin was used to encapsulate EGCG, and it revealed its better cellular uptake. More importantly, the developed formula displayed noticeable suppression of pro-inflammatory markers expressions, such as TNF- α , IL-6, and IL-12, in vitro, while the intravenous injections ameliorated colitis in the DSS-induced model [144].

An advanced technique, by realizing pH response nanovector for delivering rutin to the inflamed intestine, was also established recently. It was constituted by polyglycerol as dendron and dodecyl sulfobetaine as surfactants. The brutal alteration of the environment from the gastric lumen to the mid-alkaline milieu of the intestine triggers the smooth release of rutin from the backbone shell [145]. Metallic-based nanoparticles coated with hyaluronic acid (HA) have also been established to load apigenin for targeting colitis. The oral gavage of API-Mn(II)@HA NPs displayed a better restoration of the damaged epithelial colon barrier, reduced inflammatory markers, and enhanced therapeutic effects against DSS-induced colitis mice [146].

Priyadarshi and coworkers, for their part, conjugated poly(amidoamine) dendrimers (PAMAM) with gallic acid before treating NIH 3T3 fibroblast cells. Firstly, a sustained release of gallic acid was remarked during the payload, appended with higher cellular uptake. In addition, the in vitro evaluations showed the downregulation of inflammatory markers NG- κ B, IL-1 β , and IL-6, when treated with the said nanocarrier [147]. Similar effects were found when using gallic acid as a stabilizer of tungsten-based polyxomerates nanocluster (W-POM NCs) in the DSS-induced colitis model [148]. Co-encapsulation of caffeic acid and the anti-inflammatory drug piceatannol in albumin resulted in a greater modulation of IBD-related inflammation. More specifically, a significant drop in HIF-1 α levels and p65 was observed in colitis tissue [149].

The silk fibroin nanoparticle was thoroughly used to entrap the stilbene resveratrol for its delivery. In their work, Lozano et al. loaded resveratrol in cocoon source fibroin (RL-FNPs) by adsorption to apply for IBD. Their result emphasized that better anti-inflammatory impacts were found on TNBS model colitis treated with intracolonic injections of RL-FNPs than those treated with coarse resveratrol. The levels of chemokines such as CINC-1, MCP-1, and ICAM-1 dropped in the presence of RL-FNPs, while the peptide amounts in charge of repairing the epithelial barriers increased the mucins, TFF-3, and

villin. A recent study used β -lactoglobulin for the encapsulation of this stilbene, which prolonged its release time, when taken orally, and showed greater clinical effects in the IBD mice model. Histological assessment supported that the BLG-RES acts in promoting the expressions of the anti-inflammatory cytokines IL-10 [150]. Polymeric nanocarrier, formed with PLGA, galactosamine, and tween 80, was also applied to enhance the oral delivery of resveratrol. This formulation not only increased the intestinal crossing of the loaded drug, but also displayed promising anti-inflammatory effects in RAW 264.7 macrophages [151].

Homogenization within high pressure was utilized to stabilize the polymeric D- α -tocopherol PEG 1000 succinate curcumin (TPGS-stabilized curcumin) for its rectal delivery against ulcerative colitis. At first glance, the kinetic profile of the prepared nanoparticle was seven-fold superior to the free curcumin. Furthermore, tremendous anti-inflammatory effects were remarked during the reparation of colitis damages, which were explained by the synergistic effects of the TPGS and the nanosized nature of curcumin [152]. A pH-responsive CAP multilayer core-shell nanoparticle system (CAP₁AG₄CH₃@CUNCs) to improve the delivery of curcumin nanocrystals was also developed for ulcerative colitis. The release of the curcumin nanoparticle was sustained, depending on the layer numbers and the colon pH surrounding 7.2. More than ten layers were necessary to ensure the optimal target delivery. Moreover, pronounced re-epithelization of mucosa leading to the hindrance of macrophage infiltrations was found after treating the colitis model with that of a multilayer curcumin-loaded core-shell nanoparticle [153]. In Figure 5, some examples of phenolic-loaded nanocarriers with their targets in the management of inflammatory bowel disease are illustrated.

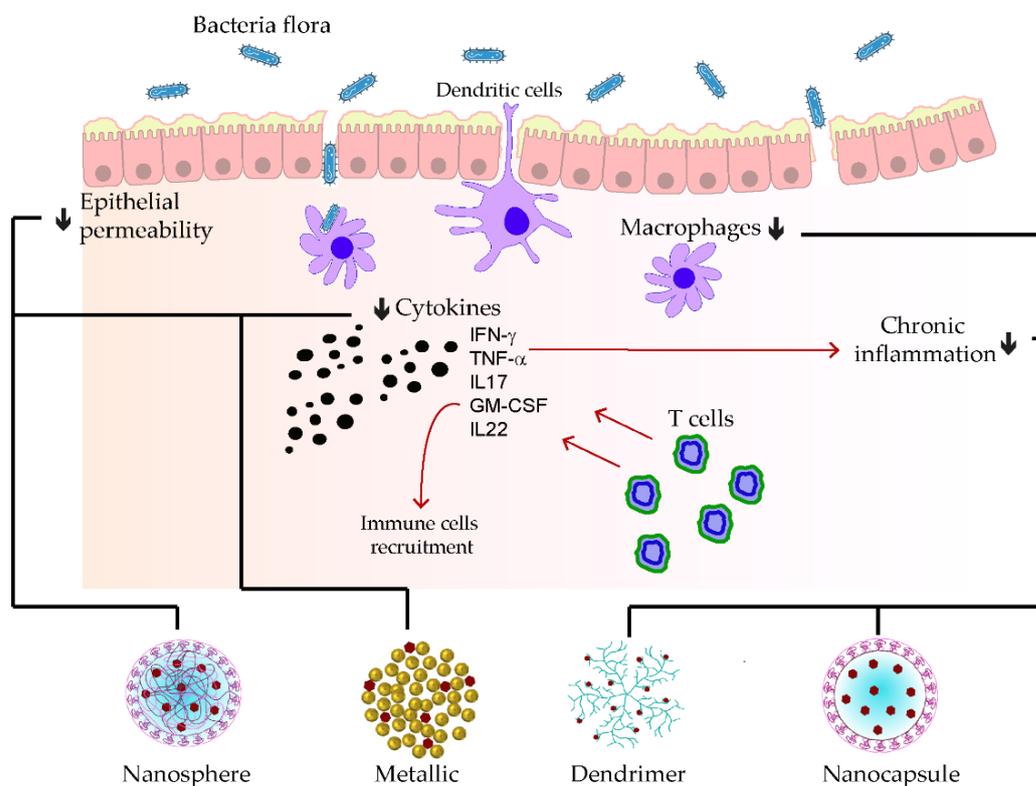


Figure 5. Examples of nanocarrier delivery of polyphenolics and their targets in inflammatory bowel disease. The black arrows up signify the increasing while the down ones indicate the decreasing effects. The red arrows stand for promotions. The black dots consist of inflammatory cytokines. The bold lines match the nanocarriers and their respective targets.

4.5. Polyphenolic Nano Delivery for Metabolic Disorder

Metabolic disorders, mainly involving diabetes and obesity, are among the life-threatening illnesses of human beings nowadays [154,155]. Diabetes is manifested by the selective degradation of the insulin producer, pancreatic beta cells, or by insulin resistance or secretion deficiency. When these pancreatic beta cells are inflamed, inflammatory cells are recruited and infiltrate within the pancreatic islets. However, these cells produce inflammatory mediators involving the main cytokines IFN γ , TNF- α , and IL-1 β , of which, overexpressions exacerbate the evolution of the pancreatic β -cells inflammation [156]. On the other hand, these pro-inflammatory cytokines also contribute to the reduction of insulin sensitivity [157]. Insulin resistance strongly impacts lipid metabolism and contributes to the onset of obesity, in general, along with adipose tissue hyperinflammation [158]. In obese people, the production of inflammatory mediators is promoted by resident and infiltrated immune cells in the adipose tissue through their MCP-1 stimulations [159].

The application of nanocarrier delivery has also received much interest in the management of metabolic diseases. As mentioned in Section 4.3, EGCG has been delivered to quicken the regeneration of impaired wounds in diabetic conditions [115,116]. Moreover, EGCG loaded in protein nanocarriers with a shell composed of β -lactoglobulin was developed to prevent metabolic syndrome. The administration of the formed nano-complex affects the levels of triglycerides and glycemia in high-fat-fed mice. The insulin sensitivity was also found to be ameliorated, which suggests its potential to reverse the severity of diabetes and obesity [160]. Similar therapeutic outcomes were determined while encapsulating EGCG in a hybrid formulation composed of PLGA in a hydrogel. This latter not only merely sustained EGCG releases, but also improved the regulations of LDL and HDL cholesterol in the HFD mice model [161].

Site-specific target delivery of rutin within a polymeric carrier composed of ethylene glycol-bis(succinic acid N-hydroxysuccinimide ester) tagged with argpyrimidine ligand (ARG-EG-RU) was also established and assessed with streptozotocin (STZ)-induced diabetic mice. The administration of ARG-EG-Ru considerably diminished the blood glucose levels and GHb in the STZ-induced models, while their insulin releases increased. A decrease in hyperlipidemia was also remarked after the treatment [162]. Amjadi et al. used a nanophytosomes system based on lecithin to entrap rutin. Their application in the STZ model resulted in a considerable alleviation of hyperglycemia and hyperlipidemia. In parallel, quick restorations of diabetic-damaged organs, such as the pancreas, liver, and kidney, were also discovered [163]. Co-application of rutin with selenium nanoparticles succeeded to improve diabetic nephropathy when administrated orally. It showed relevant cutback of inflammatory markers levels and JAK-2/STAT-3 signaling expressions. Adversely, the levels of SIRT-1 and Nrf-2 were raising, which suggested the renoprotective effects of this combination therapy [164]. With regards to apigenin, its delivery by encapsulation in polymeric nanocarrier (PEGlated-PLGA) for managing pancreatitis in a diabetic model has been promising. In addition to its ability to inhibit PSC growth by promoting apoptosis, the formulation also alleviated the overexpression of PSC-related inflammatory mRNAs (IL-6 mRNA, fibronectin mRNA, etc.) [165]. In a similar way, naringenin loaded in PLGA (N-PLGA) minimized the amount of glycated hemoglobin and triglyceride levels in STZ-induced diabetic mice after two doses of injections [166]. This compound has also been proven to be better delivered when encapsulated within nanoliposome in the treatment of non-alcoholic fatty acid disease (NAFLD), one of the obesity hallmarks [167].

Recent studies have synthesized phenolic acid delivered through nanocomposites for mitigating metabolic disorders. In 2018, Zhang et al. developed PEGlated hydroxyapatite functionalized with gallic acid and insulin for oral administration purposes. The said product presented a longer circulation time in the fluidic system and effective deliverance of insulin at the targeted site. It also lowered the glucose level of STZ-induced diabetic rats after 7 h ingestion [168]. Gelation of the gallic acid solution was made through con-

volution and supplemented with Konjac glucomannan. The composite hydrogel was further applied to diabetic wounds, which resulted in a considerable re-epithelization of the lesions and on-site collagen aggregation. In addition, an immunofluorescent study supported a diminution of inflammatory markers and COX-2 levels in the recovering injury [169]. A synthesized caffeic acid-loaded nanoliposomal vesicle formed of sodium deoxycholate (SDC) was developed for attenuating pancreatitis. In addition to its improved bioavailability and stability, its oral pretreatment counteracted the biochemical dysregulation, caused by L-ornithine in the pancreatitis model, through the modulation of the Nrf2 and NF- κ B signaling pathways [170]. Phospholipid entrapment of caffeic acid (CA-PC) also exerted the amelioration of lipid serum profile of HFD rats, following 4 weeks of oral administration. The therapy not only reduced the amount of LDL-C and VLDL-C, but also rose the HDL level, which implies a favorable regulation of hyperlipidemia [171].

When entrapping resveratrol in PLGA through the emulsion method, beneficial therapeutic effects were evidenced, especially in alleviating hyperlipidemia. On oleic acid (OA)-induced Hep G2 cells, *in vitro* study showed that treatment with RSV-PLGA-NPs enhanced lipolysis of the high-fat cells, which resulted in the clearance of triglyceride accumulations [172]. Additionally, a PEGylated phenylalanine encapsulation of resveratrol (RES-PEG-PPhe) downregulated interestingly the level of blood glucose and iNOS overexpression in the STZ-induced diabetic rat model. It also triggered insulin production in the model [173]. Other delivery techniques based on lipid shell (dipalmitoylphosphatidylcholine and cholesterol) revealed a considerable increase in insulin degree and expression in STZ-induced β -T cells and further diminished the glucose levels in diabetic models. Similarly, the upregulation of SNARE proteins (Snap23, Stx4, and Vamp2) expressions in the insulin resistance of the STZ-induced model was demonstrated. The delivery system consisted of an oral administration of resveratrol encapsulated in lecithin-palm oil carrier [174]. The same diabetic model was used for the evaluation of gold nanocarrier efficiency in delivering resveratrol. Surprisingly, AuNPs ameliorated diabetic retinopathy by lowering the levels of adhered intracellular molecules (ICAM-1), TNF- α , and mRNA expressions and hindered the ERK1/2 signaling pathway [175]. The ligand coating of polymeric nanoparticles made of a lipid layer covered with DSPE-PEG5000-peptide (L-Rnano) was delivered properly to the targeted adipose stromal cells (ASCs) of the HFD rat model. The vein injection of the formulated products resulted in a successful diminution of fat and glucose level in the model, while the insulin expressions and obesity-related inflammation were regulated [176].

Regarding curcumin, its co-encapsulation with a coenzyme Q10 in PLGA improved its bioavailability. When administering CoQ10, both diabetic inflammation and lipid metabolism were extenuated in STZ-induced models. Precisely, the synergistic effects of CoQ10 and curcumin in reducing CP, IL-6, and TNF- α , as well as the downregulation of cholesterol were determined [177]. Additionally, curcumin loaded in amphiphilic copolymer (PLGA-PEG-NPs), stabilized by CTAB, impaired the hepatic inflammation of diabetic rats. Orally administered CUR-PLGA-PEG-NPs displayed a lower regulation of COX-2, NF- κ B, and TGF- β in STZ-induced diabetic serum, while promoting PPAR γ expression [178]. Recent research performed a clinical trial delivery of this compound using an oral treatment of an optimized nanomicelle formulation for 12 weeks on patients with metabolic syndromes. The obtained result showed that the treatment has reduced noticeably the plasma triglyceride concentration and HOMA-b index. However, other metabolic syndromes indexes, such as anthropometric features, FBS, and HbA1c, have not been affected [179]. Table 3 synthesizes the current application of nanocarriers loaded with phenolic compounds and their actions towards their respective targets in metabolic disorders management.

Table 3. Application of phenolic-loaded nanocarriers in metabolic disorders.

Phenolic Compounds	Type of Nanocarriers (Size)	Targets	Mechanism of Action	Ref.
EGCG	Polymeric nanocarrier: EGCG-PLGA-in hydrogel (112 nm)	SC model	Increases delivery and reduces cholesterol, LDL-cholesterol while increasing HDL	[161]
EGCG	Hybrid nanocarrier HG-AG-EGCG	HFD-induced T2DM C57BL/6	Improves wound healing in diabetes by suppressing related inflammation	[116]
EGCG	Protein nanocarrier: EGCG- β -Lg (22 nm)	HFD obese mice	Lowers triglycerides amount in the model and improved glycemic homeostasis, as well as insulin sensitivity	[160]
Rutin	Lipid nanocarrier: Lecithin nanophytosome: (72 nm)	STZ-induced diabetic rats	Mitigates hyperglycemia and hyperlipidemia, reduces the induced damage of the kidney, liver, and pancreas in rats	[163]
Rutin	Polymeric nanocarrier: ARG-EG-RU (68 nm)	STZ-induced diabetic rats	Reduces glucose, GHb, and lipid levels, while increasing insulin amount	[162]
Rutin	Selenium nanocarrier: RU+Se-NPs	Sprague-Dawley rats	Upregulates SIRT-1, Nrf-2, and HO-1 and downregulates JAK-2/STAT3 pathways, as well as inflammatory markers	[164]
Apigenin	Polymeric nanocarrier: PEGylated-PLGA (160 nm)	Cholecystokinin-induced C57/BL6 mice	Inhibits PSC growth, promotes PSC apoptosis, reduce the expression of PSC-related inflammation	[165]
Naringenin	Lipid nanocarrier: NRG-Nano (98 nm)	Methionine choline-induced mice	Improved absorption and showed protection in fatty liver	[167]
Naringenin	Polymeric nanocarrier: N-PLGA (129 nm)	STZ-induced diabetic rat	Ameliorates diabetogenic (increases insulin level, and improves dyslipidemia)	[166]
Gallic acid	Hydrogel nanocarrier: GA-KGM	STZ-induced diabetic rats	Reduces the expression of IL-1 β , TNF- α , and COX-2	[169]
Gallic acid	Polymeric nanocarrier: HAP-PEG-GA-INS (396 nm)	STZ-induced T1D rats	Reduces blood glucose level in diabetic rats due to a higher delivery of insulin	[168]
Caffeic acid	Lipid nanocarrier: CAPE-loaded-NL (309 nm)	L-ornithine induced rat	Modulated Nrf2 and NF-kB signaling	[170]
Caffeic acid	Lipid nanocarrier: CA-PC (168 nm)	HFD-induced hyperlipidemic model	Maintain hepatocyte structure which promotes lipid absorption	[171]
Resveratrol	Lipid nanocarrier: RSV-LPs (215 nm)	Glucose/STZ-induced β -TC cell	Reduces glucose and increased insulin level	[180]
Resveratrol	Metallic nanocarrier: RES-Au-NPs (20 nm)	STZ-induced diabetic rats	Overcomes blood-retinal barrier, reduces VEGF-1, TNF- α , MCP-1, ICAM-1, IL-6, and blockades of ERK1/2 signaling pathway	[175]
Resveratrol	Polymeric nanocarrier: RES/PEG-PPhe	Intestine of STZ-induced diabetic rats	Reduces glucose level and increases insulin level while alleviating intestine injury	[173]
Resveratrol	Solid lipid nanocarrier: SLN-RES (248 nm)	STZ-induced rats	Displays hypoglycemic activity, reduces Snap 23, Stx4 and Vamp2 in insulin resistance	[174]
Resveratrol	Polymeric nanocarrier: RSV-PLGA-NPs (176 nm)	OA-induced HepG2	Promotes lipolysis and mitigates hepatocellular proliferation	[172]
Resveratrol	Hybrid nanocarrier: L-Rnano (90 nm)	HFD-C57BL/6 J mice	Decreases fat mass and inflammation while improving glucose homeostasis	[176]

Curcumin	Polymeric nanocarrier: CU-PLA-PEG (117 nm)	STZ-induced rat	Reduces NF- κ B activation, COX-2, TGF- β , and PPAR γ expressions	[178]
Curcumin	Polymeric nanocarrier: CoQ10-PLGA (115 nm)	STZ-induced rat	Reduces CRP, IL-6, TNF- α , triglyceride, and total cholesterol levels in plasma.	[177]
Curcumin	Micelle nanocarrier: C3- CU-GRAS (12 nm)	MetS patients for 12 weeks	Reduces the level of triglyceride and HOMA-b index	[179]

4.6. Polyphenolic Nano Delivery for Cardiovascular Disease

Cardiovascular disease is one of the silent health burdens that enclose a cluster of disorders attaining the blood vessels and heart. Three main impairments cause this disease, involving atherosclerosis, arterial hypertension, and coronary artery diseases [181]. Inflammation plays a substantial role in the pathophysiology of each of these disorders. In atherosclerosis, the damaged arterial vasculature walls are the agglomeration points of lipid mediators. These lipids activate immune cells that trigger a continuous secretion of pro-inflammatory factors and adhesive molecules (VCAM and ICAM). These latter provoke the sustained recruitment of immune cells in the endothelium and enhance the formation of atherosclerotic plaques [182]. In coronary artery diseases, diverse mechanisms, such as immune complex-mediated and cell-mediated inflammation, contribute to its growth [181]. In addition to the depiction of high rates of inflammatory markers in patients with arterial hypertension, many investigations have also evidenced an abnormal ratio between MMPS and TIMPS [183].

To maintain the wellness of the cardiovascular system, the application of nanoscale delivery systems has recently gained the focus of researchers to improve polyphenolic bioavailability, as well as their therapeutic efficacy. In 2014, Hong et al. formulated a polymeric nanocarrier composed of aspartic acid and chitosan self-assemblage (EGCG-CS-PAA) for atherosclerosis therapy. The said formulation showed PH-responsive releases of EGCG, and its oral administration resulted in a considerable decrease of lipid deposition throughout the artery wall which was comparable to simvastatin [184]. Another work functionalized the core-shell of the nanocarrier with a CD36 ligand named (Palmitoyl)-2-(5-keto-6-octene-dioyl) phosphatidylcholine (KodiA-PC) to heighten the delivery and macrophage uptake of EGCG. Mouse vein injections for 22 weeks of this formulation affected the macrophage releases of MCP-1, TNF- α , and IL-6, while diminishing the aorta epithelial lesions [185]. Lipid nanoparticle coated with chitosan (CSNLCE) was also used and proven to ameliorate the delivery, together with the uptake of EGCG to THP1-macrophages and inhibit their cholesterol contents as well as the expression of MCP-1 [186].

With rutin, a silver nanoparticle system (rutin@AgNPs) was established to interrupt the thromboembolism in the vascular system. The release of rutin together with its physiological compatibilities improved considerably, while it prolonged the coagulation times (PT and aPTT). The thrombogenesis formation was also inhibited by injection of this nanoparticle at low doses in vivo [187]. Oral administration of nano-lipid vesicle of this biflavonoid also impacted thrombogenesis and clotting formations in the ferric chloride (FC)-induced microvascular model [188]. Nanolipid encapsulation of naringenin decorated with folic acid ligands (FA-LNPs/Nrg) expressed higher delivery and anti-atherosclerotic effects in the ApoE $^{-/-}$ model after oral treatment for 3 months. This therapeutic advantage was explained by the ease of FA-LNPs/Nrg to pass across different transmembranes, infiltrate the targeted atherosclerotic cells, and reduce atherosclerotic plaque burdens [189]. When co-encapsulating the lipid vesicle with indocyanine green (ICG) and decorated with VCAM-1 molecule to form a theranostic (V-Nar/ICG/LN), other beneficial information arose that supported the effectiveness of the nanoparticle to suppress mRNA expression of inflamed vasculature of different organs [189].

Phenolic acid-loaded nanocarrier systems also exhibited enhanced beneficial effects in CVD. He et al. developed gallic acid-loaded polymeric nanovesicles made with PTMC (poly(1,3-trimethylene carbonate)) to impede the cytotoxicity of the clinically eluting stent drug used in severe atherosclerosis. Their finding supported that the formulated product improved vein endothelial cells (HUVEC) adhesions, while blockading artery smooth cells (HUASMC) development [190]. In addition, anti-platelet aggregation of gallic acid decorated dendrimer (GA-PAMAM) was also established [191].

To surmount the limitation of coarse resveratrol application in CVD treatment, Zou et al. synthesized a polymeric carrier prepared with PEG-L-Leucine for myocardial protection. This strategy prolonged the release time of resveratrol in the cardiomyocyte cells, along with satisfactory protection on ischemia reperfusion-injury in MI/RI model [192]. In the same manner, oral treatment of resveratrol loaded in PLGA prevented myocardial injury marker releases (troponin, LDH, AST), inhibited the expression of inflammatory cytokines, and upregulated eNOS expressions [193].

A mesoporous silica nanocarrier was established to embed curcumin compound before treating the myocardial defect induced by doxorubicin. In addition to the delivery improvement of curcumin, mesoporous nanoparticles also enhanced its anti-cardiotoxicity effects by reducing MDA levels and promoting the production of GSH, CAT, as well as SOD in cardiac tissue [194]. Synergistic anti-atherosclerotic effects were observed when co-encapsulating curcumin with atorvastatin calcium (Ato) in E-selectin-binding (Esb) ligand-coated liposome (T-AC-Lipo). The decoration with Esb ligands improved the cell delivery of the nanoparticle while curcumin and Ato worked interactively in reducing foam cell generation from monocytes and in blockading adhesions of molecules, such as ICAM-1 and E-selectin [195]. Some of these applications of phenolic-loaded nanocarriers and their respective targets for the mitigation of atherosclerosis are shown in Figure 6.

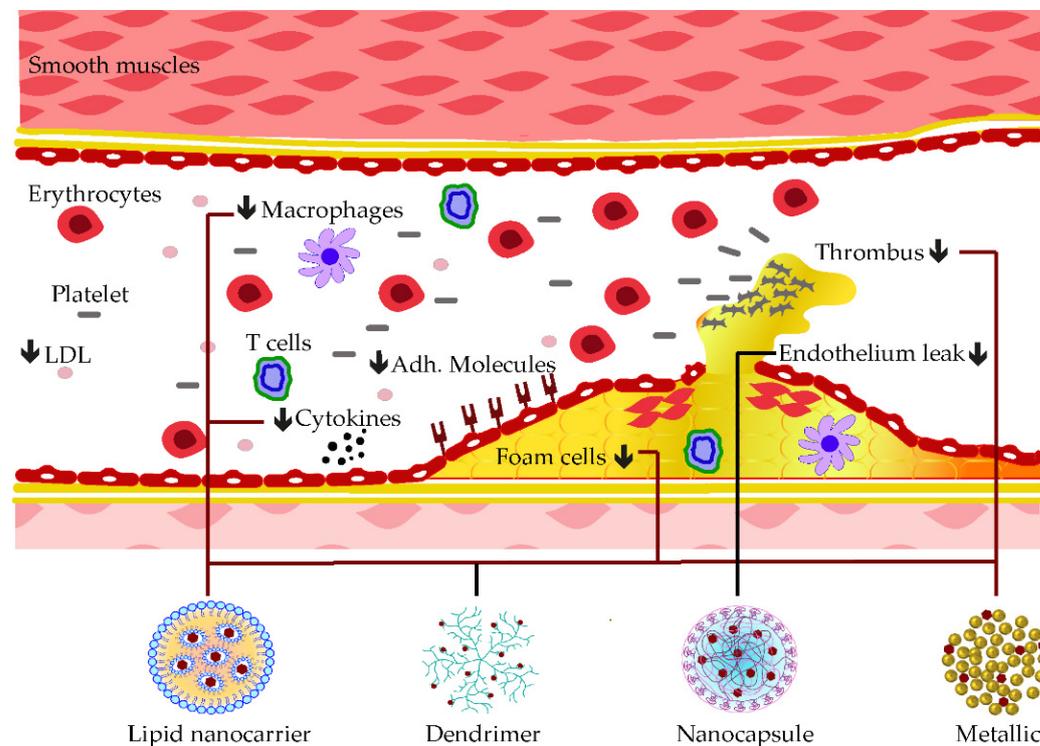


Figure 6. Examples of nanocarrier delivery of polyphenolics and their targets in Atherosclerosis. The black arrows down signify the decreasing effects. The black dots consist of inflammatory cytokines. The bold lines match the nanocarriers and their respective targets.

5. Conclusions and Future Perspectives

Many natural phenolic compounds have been proven to possess beneficial effects in the management of inflammation-mediated diseases, but their clinical applications are limited, due to their organism bioavailability and low target deliverance. Nanocarrier delivery systems have recently been widely used for improving the efficacy of diverse phenolic compounds. This strategy not only enhances the circulation time of the loaded phenolic in the body system, but also improves their deliverance to the expected site of actions. Our present review has synthesized the recent applications of nanocarriers loaded with common phenolics (EGCG, rutin, naringenin, apigenin, gallic acid, caffeic acid, resveratrol, and curcumin) for their effects in inflammatory-mediated diseases. Their physicochemical features, as well as their mode of action, have been discussed. Hybrid nanoparticle encapsulations were discovered to exhibit considerable advantages, due to their great variety of administration and high drug capacity loading. Furthermore, nanoparticle surface functionalization with biological molecules, such as peptides or/and nucleic acids, increased their effectiveness, in terms of target delivery and recognition. Co-delivery of different phenolics, or with approved drugs expressed as well better actions through their synergistic functions. However, side effects and nanotoxicology should be considered, as nanocarriers can cause allosteric sequestration on the cell membranes of vasculature, kidney, and liver. Sometimes, they may trigger immune defenses, which quicken their fast elimination by phagocytoses. Regarding the active targeting of functionalized nanocarriers, in some cases, knowing the specific receptors of the targeted cells remains challenging, which hampers the fast recognition and reachability of the formulated nanoparticles. So far, there is no explicit legislation for the standardization of medications formulated with nanocarriers, which holds their clinical extrapolation back. Consequently, given that the library of promising anti-inflammatory phenolic compounds has never ceased growing, future efforts should concentrate more on developing powerful and efficient strategies, while designing nanocarriers to specific target receptors through precision medicine. Harnessing the accrescent development of technologies, as well as the artificial intelligences in nanocarriers engineering and development (nanorobotics, pharmacocytes, and simulations) also invigorate the advancement of nanomedicine. Finally, establishing standardized processing methods and preclinical application guidelines by the regulatory authorities helps in overcoming the clinical extrapolation barriers. Considering all this will give more allowance for the complexes' inherent properties, modes of action, and the clinical translation.

Author Contributions: Conceptualization, M.-Q.G. and T.F.R.; investigation, T.F.R.; writing—original draft preparation, T.F.R.; writing—review and editing, T.F.R., F.W.M., M.-X.F. and M.-Q.G.; supervision, M.-Q.G. and M.-X.F. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. He, Y.; Yue, Y.; Zheng, X.; Zhang, K.; Chen, S.; Du, Z. Curcumin, Inflammation, and Chronic Diseases: How Are They Linked? *Molecules* **2015**, *20*, 9183–9213. <https://doi.org/10.3390/molecules20059183>.
2. Placha, D.; Jampilek, J. Chronic Inflammatory Diseases, Anti-Inflammatory Agents and Their Delivery Nanosystems. *Pharmaceutics* **2021**, *13*, 64. <https://doi.org/10.3390/pharmaceutics13010064>.
3. Jogpal, V.; Sanduja, M.; Dutt, R.; Garg, V.; Tinku Advancement of Nanomedicines in Chronic Inflammatory Disorders. *Inflammopharmacology* **2022**, *30*, 355–368. <https://doi.org/10.1007/s10787-022-00927-x>.

4. Cronkite, D.A.; Strutt, T.M. The Regulation of Inflammation by Innate and Adaptive Lymphocytes. *J. Immunol. Res.* **2018**, *2018*, e1467538. <https://doi.org/10.1155/2018/1467538>.
5. Conte, R.; Marturano, V.; Peluso, G.; Calarco, A.; Cerruti, P. Recent Advances in Nanoparticle-Mediated Delivery of Anti-Inflammatory Phytocompounds. *Int. J. Mol. Sci.* **2017**, *18*, 709. <https://doi.org/10.3390/ijms18040709>.
6. Tang, D.-W.; Yu, S.-H.; Ho, Y.-C.; Huang, B.-Q.; Tsai, G.-J.; Hsieh, H.-Y.; Sung, H.-W.; Mi, F.-L. Characterization of Tea Catechins-Loaded Nanoparticles Prepared from Chitosan and an Edible Polypeptide. *Food Hydrocoll.* **2013**, *30*, 33–41. <https://doi.org/10.1016/j.foodhyd.2012.04.014>.
7. Garavand, F.; Jalai-Jivan, M.; Assadpour, E.; Jafari, S.M. Encapsulation of Phenolic Compounds within Nano/Microemulsion Systems: A Review. *Food Chem.* **2021**, *364*, 130376. <https://doi.org/10.1016/j.foodchem.2021.130376>.
8. Patra, J.K.; Das, G.; Fraceto, L.F.; Campos, E.V.R.; Rodriguez-Torres, M.D.P.; Acosta-Torres, L.S.; Diaz-Torres, L.A.; Grillo, R.; Swamy, M.K.; Sharma, S.; et al. Nano Based Drug Delivery Systems: Recent Developments and Future Prospects. *J. Nanobiotechnology* **2018**, *16*, 71. <https://doi.org/10.1186/s12951-018-0392-8>.
9. Rashmi, H.B.; Negi, P.S. Phenolic Acids from Vegetables: A Review on Processing Stability and Health Benefits. *Food Res. Int.* **2020**, *136*, 109298. <https://doi.org/10.1016/j.foodres.2020.109298>.
10. Albuquerque, B.R.; Heleno, S.A.; Oliveira, M.B.P.P.; Barros, L.; Ferreira, I.C.F.R. Phenolic Compounds: Current Industrial Applications, Limitations and Future Challenges. *Food Funct.* **2021**, *12*, 14–29. <https://doi.org/10.1039/D0FO02324H>.
11. Tao, H.; Li, L.; He, Y.; Zhang, X.; Zhao, Y.; Wang, Q.; Hong, G. Flavonoids in Vegetables: Improvement of Dietary Flavonoids by Metabolic Engineering to Promote Health. *Crit. Rev. Food Sci. Nutr.* **2022**, *11*, 1–15. <https://doi.org/10.1080/10408398.2022.2131726>.
12. Călinoiu, L.F.; Vodnar, D.C. Whole Grains and Phenolic Acids: A Review on Bioactivity, Functionality, Health Benefits and Bioavailability. *Nutrients* **2018**, *10*, 1615. <https://doi.org/10.3390/nu10111615>.
13. Fechtner, S.; Singh, A.; Chourasia, M.; Ahmed, S. Molecular Insights into the Differences in Anti-Inflammatory Activities of Green Tea Catechins on IL-1 β Signaling in Rheumatoid Arthritis Synovial Fibroblasts. *Toxicol. Appl. Pharmacol.* **2017**, *329*, 112–120. <https://doi.org/10.1016/j.taap.2017.05.016>.
14. Miao, Z.; Zhao, Y.; Chen, M.; He, C. Using Flavonoids as a Therapeutic Intervention against Rheumatoid Arthritis: The Known and Unknown. *Pharm. Res. Mod. Chin. Med.* **2022**, *3*, 100014. <https://doi.org/10.1016/j.prmcm.2021.100014>.
15. Lin, Y.; Luo, T.; Weng, A.; Huang, X.; Yao, Y.; Fu, Z.; Li, Y.; Liu, A.; Li, X.; Chen, D.; et al. Gallic Acid Alleviates Gouty Arthritis by Inhibiting NLRP3 Inflammasome Activation and Pyroptosis Through Enhancing Nrf2 Signaling. *Front. Immunol.* **2020**, *11*, 580593.
16. Ide, K.; Matsuoka, N.; Yamada, H.; Furushima, D.; Kawakami, K. Effects of Tea Catechins on Alzheimer’s Disease: Recent Updates and Perspectives. *Molecules* **2018**, *23*, 2357. <https://doi.org/10.3390/molecules23092357>.
17. Devi, S.; Kumar, V.; Singh, S.K.; Dubey, A.K.; Kim, J.-J. Flavonoids: Potential Candidates for the Treatment of Neurodegenerative Disorders. *Biomedicines* **2021**, *9*, 99. <https://doi.org/10.3390/biomedicines9020099>.
18. Caruso, G.; Godos, J.; Privitera, A.; Lanza, G.; Castellano, S.; Chillemi, A.; Bruni, O.; Ferri, R.; Caraci, F.; Grosso, G. Phenolic Acids and Prevention of Cognitive Decline: Polyphenols with a Neuroprotective Role in Cognitive Disorders and Alzheimer’s Disease. *Nutrients* **2022**, *14*, 819. <https://doi.org/10.3390/nu14040819>.
19. Fan, F.-Y.; Sang, L.-X.; Jiang, M. Catechins and Their Therapeutic Benefits to Inflammatory Bowel Disease. *Molecules* **2017**, *22*, 484. <https://doi.org/10.3390/molecules22030484>.
20. Hoensch, H.P.; Weigmann, B. Regulation of the Intestinal Immune System by Flavonoids and Its Utility in Chronic Inflammatory Bowel Disease. *World J. Gastroenterol.* **2018**, *24*, 877–881. <https://doi.org/10.3748/wjg.v24.i8.877>.
21. Ginwala, R.; Bhavsar, R.; Chigbu, D.G.I.; Jain, P.; Khan, Z.K. Potential Role of Flavonoids in Treating Chronic Inflammatory Diseases with a Special Focus on the Anti-Inflammatory Activity of Apigenin. *Antioxidants* **2019**, *8*, 35. <https://doi.org/10.3390/antiox8020035>.
22. Dinda, B.; Dinda, M.; Roy, A.; Dinda, S. Dietary Plant Flavonoids in Prevention of Obesity and Diabetes. *Adv. Protein Chem. Struct. Biol.* **2020**, *120*, 159–235. <https://doi.org/10.1016/bs.apcsb.2019.08.006>.
23. Vinayagam, R.; Jayachandran, M.; Xu, B. Antidiabetic Effects of Simple Phenolic Acids: A Comprehensive Review. *Phyto. Res.* **2016**, *30*, 184–199. <https://doi.org/10.1002/ptr.5528>.
24. Pourhabibi-Zarandi, F.; Shojaei-Zarghani, S.; Rafraf, M. Curcumin and Rheumatoid Arthritis: A Systematic Review of Literature. *Int. J. Clin. Pract.* **2021**, *75*, e14280. <https://doi.org/10.1111/ijcp.14280>.
25. Karimi, A.; Azar, P.S.; Kadkhodayi, M.; Tandorost, A.; Vajdi, M.; Shoorei, H.; Farhangi, M.A. A Comprehensive Insight into Effects of Resveratrol on Molecular Mechanism in Rheumatoid Arthritis: A Literature Systematic Review. *Int. J. Rheum. Dis.* **2022**, *25*, 827–843. <https://doi.org/10.1111/1756-185X.14356>.
26. Eghbaliferiz, S.; Farhadi, F.; Barreto, G.E.; Majeed, M.; Sahebkar, A. Effects of Curcumin on Neurological Diseases: Focus on Astrocytes. *Pharmacol. Rep.* **2020**, *72*, 769–782. <https://doi.org/10.1007/s43440-020-00112-3>.
27. Huang, J.; Huang, N.; Xu, S.; Luo, Y.; Li, Y.; Jin, H.; Yu, C.; Shi, J.; Jin, F. Signaling Mechanisms Underlying Inhibition of Neuroinflammation by Resveratrol in Neurodegenerative Diseases. *J. Nutr. Biochem.* **2021**, *88*, 108552. <https://doi.org/10.1016/j.jnutbio.2020.108552>.
28. Fallahi, F.; Borran, S.; Ashrafzadeh, M.; Zarrabi, A.; Pourhanifeh, M.H.; Khaksary Mahabady, M.; Sahebkar, A.; Mirzaei, H. Curcumin and Inflammatory Bowel Diseases: From in Vitro Studies to Clinical Trials. *Mol. Immunol.* **2021**, *130*, 20–30. <https://doi.org/10.1016/j.molimm.2020.11.016>.

29. Karthikeyan, A.; Young, K.N.; Moniruzzaman, M.; Beyene, A.M.; Do, K.; Kalaiselvi, S.; Min, T. Curcumin and Its Modified Formulations on Inflammatory Bowel Disease (IBD): The Story So Far and Future Outlook. *Pharmaceutics* **2021**, *13*, 484. <https://doi.org/10.3390/pharmaceutics13040484>.
30. Nunes, S.; Danesi, F.; Del Rio, D.; Silva, P. Resveratrol and Inflammatory Bowel Disease: The Evidence so Far. *Nutr. Res. Rev.* **2018**, *31*, 85–97. <https://doi.org/10.1017/S095442241700021X>.
31. Gowd, V.; Kanika; Jori, C.; Chaudhary, A.A.; Rudayni, H.A.; Rashid, S.; Khan, R. Resveratrol and Resveratrol Nano-Delivery Systems in the Treatment of Inflammatory Bowel Disease. *J. Nutr. Biochem.* **2022**, *109*, 109101. <https://doi.org/10.1016/j.jnutbio.2022.109101>.
32. Meng, X.; Zhou, J.; Zhao, C.-N.; Gan, R.-Y.; Li, H.-B. Health Benefits and Molecular Mechanisms of Resveratrol: A Narrative Review. *Foods* **2020**, *9*, 340. <https://doi.org/10.3390/foods9030340>.
33. Thakur, L.; Ghodasra, U.; Patel, N.; Dabhi, M. Novel Approaches for Stability Improvement in Natural Medicines. *Phcog. Rev.* **2011**, *5*, 48. <https://doi.org/10.4103/0973-7847.79099>.
34. Krupkova, O.; Ferguson, S.J.; Wuertz-Kozak, K. Stability of (–)-Epigallocatechin Gallate and Its Activity in Liquid Formulations and Delivery Systems. *J. Nutr. Biochem.* **2016**, *37*, 1–12. <https://doi.org/10.1016/j.jnutbio.2016.01.002>.
35. Kyriakoudi, A.; Spanidi, E.; Mourtzinis, I.; Gardikis, K. Innovative Delivery Systems Loaded with Plant Bioactive Ingredients: Formulation Approaches and Applications. *Plants* **2021**, *10*, 1238. <https://doi.org/10.3390/plants10061238>.
36. Fendler, J.H.; Romero, A. Liposomes as Drug Carriers. *Life Sci.* **1977**, *20*, 1109–1120. [https://doi.org/10.1016/0024-3205\(77\)90481-7](https://doi.org/10.1016/0024-3205(77)90481-7).
37. Couvreur, P.; Vauthier, C. Polyalkylcyanoacrylate Nanoparticles as Drug Carrier: Present State and Perspectives. *J. Control. Release* **1991**, *17*, 187–198. [https://doi.org/10.1016/0168-3659\(91\)90058-L](https://doi.org/10.1016/0168-3659(91)90058-L).
38. Harrington, K.J.; Lewanski, C.; Northcote, A.D.; Whittaker, J.; Peters, A.M.; Vile, R.G.; Stewart, J.S.W. Phase II Study of Pegylated Liposomal Doxorubicin (Caelyx™) as Induction Chemotherapy for Patients with Squamous Cell Cancer of the Head and Neck. *Eur. J. Cancer* **2001**, *37*, 2015–2022. [https://doi.org/10.1016/S0959-8049\(01\)00216-7](https://doi.org/10.1016/S0959-8049(01)00216-7).
39. Mehnert, W.; Mäder, K. Solid Lipid Nanoparticles: Production, Characterization and Applications. *Adv. Drug Deliv. Rev.* **2001**, *47*, 165–196. [https://doi.org/10.1016/S0169-409X\(01\)00105-3](https://doi.org/10.1016/S0169-409X(01)00105-3).
40. Sarfraz, M.; Qamar, S.; Rehman, M.U.; Tahir, M.A.; Ijaz, M.; Ahsan, A.; Asim, M.H.; Nazir, I. Nano-Formulation Based Intravesical Drug Delivery Systems: An Overview of Versatile Approaches to Improve Urinary Bladder Diseases. *Pharmaceutics* **2022**, *14*, 1909. <https://doi.org/10.3390/pharmaceutics14091909>.
41. Vanti, G. Recent Strategies in Nanodelivery Systems for Natural Products: A Review. *Environ. Chem. Lett.* **2021**, *19*, 4311–4326. <https://doi.org/10.1007/s10311-021-01276-x>.
42. Ajeeshkumar, K.K.; Aneesh, P.A.; Raju, N.; Suseela, M.; Ravishankar, C.N.; Benjakul, S. Advancements in Liposome Technology: Preparation Techniques and Applications in Food, Functional Foods, and Bioactive Delivery: A Review. *Compr. Rev. Food Sci. Food Saf.* **2021**, *20*, 1280–1306. <https://doi.org/10.1111/1541-4337.12725>.
43. Filipczak, N.; Pan, J.; Yalamarty, S.S.K.; Torchilin, V.P. Recent Advancements in Liposome Technology. *Adv. Drug Deliv. Rev.* **2020**, *156*, 4–22. <https://doi.org/10.1016/j.addr.2020.06.022>.
44. Figueroa-Robles, A.; Antunes-Ricardo, M.; Guajardo-Flores, D. Encapsulation of Phenolic Compounds with Liposomal Improvement in the Cosmetic Industry. *Int. J. Pharm.* **2021**, *593*, 120125. <https://doi.org/10.1016/j.ijpharm.2020.120125>.
45. Nishimoto-Sauceda, D.; Romero-Robles, L.E.; Antunes-Ricardo, M. Biopolymer Nanoparticles: A Strategy to Enhance Stability, Bioavailability, and Biological Effects of Phenolic Compounds as Functional Ingredients. *J. Sci. Food Agric.* **2022**, *102*, 41–52. <https://doi.org/10.1002/jsfa.11512>.
46. Faridi Esfanjani, A.; Jafari, S.M. Biopolymer Nano-Particles and Natural Nano-Carriers for Nano-Encapsulation of Phenolic Compounds. *Colloids Surf. B Biointerfaces.* **2016**, *146*, 532–543. <https://doi.org/10.1016/j.colsurfb.2016.06.053>.
47. Kedar, U.; Phutane, P.; Shidhaye, S.; Kadam, V. Advances in Polymeric Micelles for Drug Delivery and Tumor Targeting. *Nanomedicine.* **2010**, *6*, 714–729. <https://doi.org/10.1016/j.nano.2010.05.005>.
48. Chai, Q.; Jiao, Y.; Yu, X. Hydrogels for Biomedical Applications: Their Characteristics and the Mechanisms behind Them. *Gels* **2017**, *3*, 6. <https://doi.org/10.3390/gels3010006>.
49. Wang, Y.; Zhao, Q.; Han, N.; Bai, L.; Li, J.; Liu, J.; Che, E.; Hu, L.; Zhang, Q.; Jiang, T.; et al. Mesoporous Silica Nanoparticles in Drug Delivery and Biomedical Applications. *Nanomedicine* **2015**, *11*, 313–327. <https://doi.org/10.1016/j.nano.2014.09.014>.
50. Li, Y.; Li, N.; Pan, W.; Yu, Z.; Yang, L.; Tang, B. Hollow Mesoporous Silica Nanoparticles with Tunable Structures for Controlled Drug Delivery. *ACS Appl. Mater. Interfaces* **2017**, *9*, 2123–2129. <https://doi.org/10.1021/acsami.6b13876>.
51. Liu, S.; Chen, X.; Ai, W.; Wei, C. A New Method to Prepare Mesoporous Silica from Coal Gasification Fine Slag and Its Application in Methylene Blue Adsorption. *J. Clean. Prod.* **2019**, *212*, 1062–1071. <https://doi.org/10.1016/j.jclepro.2018.12.060>.
52. Chandrakala, V.; Aruna, V.; Angajala, G. Review on Metal Nanoparticles as Nanocarriers: Current Challenges and Perspectives in Drug Delivery Systems. *Emergent. Mater.* **2022**, *5*, 1593–1615. <https://doi.org/10.1007/s42247-021-00335-x>.
53. Soliman, M.G.; Pelaz, B.; Parak, W.J.; del Pino, P. Phase Transfer and Polymer Coating Methods toward Improving the Stability of Metallic Nanoparticles for Biological Applications. *Chem. Mater.* **2015**, *27*, 990–997. <https://doi.org/10.1021/cm5043167>.
54. Jurašin, D.D.; Čurlin, M.; Capjak, I.; Crnković, T.; Lovrić, M.; Babić, M.; Horák, D.; Vinković Vrček, I.; Gajović, S. Surface Coating Affects Behavior of Metallic Nanoparticles in a Biological Environment. *Beilstein J. Nanotechnol.* **2016**, *7*, 246–262. <https://doi.org/10.3762/bjnano.7.23>.

55. Neha Desai; Momin, M.; Khan, T.; Gharat, S.; Ningthoujam, R.S.; Omri, A. Metallic Nanoparticles as Drug Delivery System for the Treatment of Cancer. *Expert Opin. Drug Deliv.* **2021**, *18*, 1261–1290. <https://doi.org/10.1080/17425247.2021.1912008>.
56. Nicolas, J.; Mura, S.; Brambilla, D.; Mackiewicz, N.; Couvreur, P. Design, Functionalization Strategies and Biomedical Applications of Targeted Biodegradable/Biocompatible Polymer-Based Nanocarriers for Drug Delivery. *Chem. Soc. Rev.* **2013**, *42*, 1147–1235. <https://doi.org/10.1039/C2CS35265F>.
57. Li, Y.; Champion, J.A. Self-Assembling Nanocarriers from Engineered Proteins: Design, Functionalization, and Application for Drug Delivery. *Adv. Drug Deliv. Rev.* **2022**, *189*, 114462. <https://doi.org/10.1016/j.addr.2022.114462>.
58. Jiang, Q.; Wang, X.; Huang, E.; Wang, Q.; Wen, C.; Yang, G.; Lu, L.; Cui, D. Inflammasome and Its Therapeutic Targeting in Rheumatoid Arthritis. *Front. Immunol.* **2022**, *12*, 816839.
59. Kennedy, A.; Fearon, U.; Veale, D.J.; Godson, C. Macrophages in Synovial Inflammation. *Front. Immunol.* **2011**, *2*, 52. <https://doi.org/10.3389/fimmu.2011.00052>.
60. Macfarlane, E.; Seibel, M.J.; Zhou, H. Arthritis and the Role of Endogenous Glucocorticoids. *Bone Res.* **2020**, *8*, 33. <https://doi.org/10.1038/s41413-020-00112-2>.
61. Zheng, Y.; Xiao, L.; Yu, C.; Jin, P.; Qin, D.; Xu, Y.; Yin, J.; Liu, Z.; Du, Q. Enhanced Antiarthritic Efficacy by Nanoparticles of (–)-Epigallocatechin Gallate–Glucosamine–Casein. *J. Agric. Food Chem.* **2019**, *67*, 6476–6486. <https://doi.org/10.1021/acs.jafc.9b02075>.
62. Vyawahare, A.; Prakash, R.; Jori, C.; Ali, A.; Raza, S.; Khan, R. Caffeic Acid Modified Nanomicelles Inhibit Articular Cartilage Deterioration and Reduce Disease Severity in Experimental Inflammatory Arthritis. *ACS Nano* **2022**, *16*, 18579–18591. <https://doi.org/10.1021/acsnano.2c07027>.
63. Munir, A.; Muhammad, F.; Zaheer, Y.; Ali, A.; Iqbal, M.; Rehman, M.; Munir, M.U.; Akhtar, B.; Webster, T.J.; Sharif, A.; et al. Synthesis of Naringenin Loaded Lipid Based Nanocarriers and Their In-Vivo Therapeutic Potential in a Rheumatoid Arthritis Model. *J. Drug Deliv. Sci. Technol.* **2021**, *66*, 102854. <https://doi.org/10.1016/j.jddst.2021.102854>.
64. Mohanty, S.; Sahoo, A.K.; Konkimalla, V.B.; Pal, A.; Si, S.C. Naringin in Combination with Isothiocyanates as Liposomal Formulations Potentiates the Anti-Inflammatory Activity in Different Acute and Chronic Animal Models of Rheumatoid Arthritis. *ACS Omega* **2020**, *5*, 28319–28332. <https://doi.org/10.1021/acsomega.0c04300>.
65. Mohanty, S.; Konkimalla, V.B.; Pal, A.; Sharma, T.; Si, S.C. Naringin as Sustained Delivery Nanoparticles Ameliorates the Anti-Inflammatory Activity in a Freund's Complete Adjuvant-Induced Arthritis Model. *ACS Omega* **2021**, *6*, 28630–28641. <https://doi.org/10.1021/acsomega.1c03066>.
66. Zhang, G.; Sun, G.; Guan, H.; Li, M.; Liu, Y.; Tian, B.; He, Z.; Fu, Q. Naringenin Nanocrystals for Improving Anti-Rheumatoid Arthritis Activity. *Asian J. Pharm. Sci.* **2021**, *16*, 816–825. <https://doi.org/10.1016/j.ajps.2021.09.001>.
67. Chen, X.; Zhu, X.; Ma, L.; Lin, A.; Gong, Y.; Yuan, G.; Liu, J. A Core–Shell Structure QRu–PLGA–RES–DS NP Nanocomposite with Photothermal Response-Induced M2 Macrophage Polarization for Rheumatoid Arthritis Therapy. *Nanoscale* **2019**, *11*, 18209–18223. <https://doi.org/10.1039/C9NR05922A>.
68. Kamel, R.; Abbas, H.; Shaffie, N.M. Development and Evaluation of PLA-Coated Co-Micellar Nanosystem of Resveratrol for the Intra-Articular Treatment of Arthritis. *Int. J. Pharm.* **2019**, *569*, 118560. <https://doi.org/10.1016/j.ijpharm.2019.118560>.
69. Poonia, N.; Lather, V.; Kaur, B.; Kirthanashri, S.V.; Pandita, D. Optimization and Development of Methotrexate- and Resveratrol-Loaded Nanoemulsion Formulation Using Box–Behnken Design for Rheumatoid Arthritis. *Assay Drug Dev. Technol.* **2020**, *18*, 356–368. <https://doi.org/10.1089/adt.2020.989>.
70. Cui, N.; Xu, Z.; Zhao, X.; Yuan, M.; Pan, L.; Lu, T.; Du, A.; Qin, L. In Vivo Effect of Resveratrol-Cellulose Aerogel Drug Delivery System to Relieve Inflammation on Sports Osteoarthritis. *Gels* **2022**, *8*, 544. <https://doi.org/10.3390/gels8090544>.
71. Arora, R.; Kuhad, A.; Kaur, I.P.; Chopra, K. Curcumin Loaded Solid Lipid Nanoparticles Ameliorate Adjuvant-Induced Arthritis in Rats. *Eur. J. Pain* **2015**, *19*, 940–952. <https://doi.org/10.1002/ejp.620>.
72. Coradini, K.; Friedrich, R.B.; Fonseca, F.N.; Vencato, M.S.; Andrade, D.F.; Oliveira, C.M.; Battistel, A.P.; Guterres, S.S.; da Rocha, M.I.U.M.; Pohlmann, A.R.; et al. A Novel Approach to Arthritis Treatment Based on Resveratrol and Curcumin Co-Encapsulated in Lipid-Core Nanocapsules: In Vivo Studies. *Eur. J. Pharm. Sci.* **2015**, *78*, 163–170. <https://doi.org/10.1016/j.ejps.2015.07.012>.
73. Yan, F.; Li, H.; Zhong, Z.; Zhou, M.; Lin, Y.; Tang, C.; Li, C. Co-Delivery of Prednisolone and Curcumin in Human Serum Albumin Nanoparticles for Effective Treatment of Rheumatoid Arthritis. *Int. J. Nanomed.* **2019**, *14*, 9113–9125. <https://doi.org/10.2147/IJN.S219413>.
74. Naz, Z.; Ahmad, F.J. Curcumin-Loaded Colloidal Carrier System: Formulation Optimization, Mechanistic Insight, Ex Vivo and in Vivo Evaluation. *Int. J. Nanomed.* **2015**, *10*, 4293–4307. <https://doi.org/10.2147/IJN.S82788>.
75. Dewangan, A.K.; Perumal, Y.; Pavurala, N.; Chopra, K.; Mazumder, S. Preparation, Characterization and Anti-Inflammatory Effects of Curcumin Loaded Carboxymethyl Cellulose Acetate Butyrate Nanoparticles on Adjuvant Induced Arthritis in Rats. *J. Drug Deliv. Sci. Tech.* **2017**, *41*, 269–279. <https://doi.org/10.1016/j.jddst.2017.07.022>.
76. Fan, Z.; Li, J.; Liu, J.; Jiao, H.; Liu, B. Anti-Inflammation and Joint Lubrication Dual Effects of a Novel Hyaluronic Acid/Curcumin Nanomicelle Improve the Efficacy of Rheumatoid Arthritis Therapy. *ACS Appl. Mater. Interfaces* **2018**, *10*, 23595–23604. <https://doi.org/10.1021/acsmi.8b06236>.
77. Kang, C.; Jung, E.; Hyeon, H.; Seon, S.; Lee, D. Acid-Activatable Polymeric Curcumin Nanoparticles as Therapeutic Agents for Osteoarthritis. *Nanomedicine* **2020**, *23*, 102104. <https://doi.org/10.1016/j.nano.2019.102104>.
78. Chamani, S.; Bianconi, V.; Tasbandi, A.; Pirro, M.; Barreto, G.E.; Jamialahmadi, T.; Sahebkar, A. Resolution of Inflammation in Neurodegenerative Diseases: The Role of Resolvins. *Mediat. Inflamm.* **2020**, *2020*, e3267172. <https://doi.org/10.1155/2020/3267172>.

79. Wang, R.-X.; Zhou, M.; Ma, H.-L.; Qiao, Y.-B.; Li, Q.-S. The Role of Chronic Inflammation in Various Diseases and Anti-Inflammatory Therapies Containing Natural Products. *Chem. Med. Chem.* **2021**, *16*, 1576–1592. <https://doi.org/10.1002/cmdc.202000996>.
80. Forloni, G.; La Vitola, P.; Cerovic, M.; Balducci, C. Inflammation and Parkinson's Disease Pathogenesis: Mechanisms and Therapeutic Insight. *Prog. Mol. Biol. Transl. Sci.* **2021**, *177*, 175–202. <https://doi.org/10.1016/bs.pmbts.2020.11.001>.
81. Singh, N.A.; Bhardwaj, V.; Ravi, C.; Ramesh, N.; Mandal, A.K.A.; Khan, Z.A. EGCG Nanoparticles Attenuate Aluminum Chloride Induced Neurobehavioral Deficits, Beta Amyloid and Tau Pathology in a Rat Model of Alzheimer's Disease. *Front. Aging Neurosci.* **2018**, *10*, 244. <https://doi.org/10.3389/fnagi.2018.00244>.
82. Cano, A.; Ettcheto, M.; Chang, J.-H.; Barroso, E.; Espina, M.; Kühne, B.A.; Barenys, M.; Auladell, C.; Folch, J.; Souto, E.B.; et al. Dual-Drug Loaded Nanoparticles of Epigallocatechin-3-Gallate (EGCG)/Ascorbic Acid Enhance Therapeutic Efficacy of EGCG in a APP^{swe}/PS1^{dE9} Alzheimer's Disease Mice Model. *J. Control. Release* **2019**, *301*, 62–75. <https://doi.org/10.1016/j.jconrel.2019.03.010>.
83. Lv, L.; Yang, F.; Li, H.; Yuan, J. Brain-Targeted Co-Delivery of β -Amyloid Converting Enzyme 1 ShRNA and Epigallocatechin-3-Gallate by Multifunctional Nanocarriers for Alzheimer's Disease Treatment. *IUBMB Life* **2020**, *72*, 1819–1829. <https://doi.org/10.1002/iub.2330>.
84. Li, Y.; Chen, Z.; Lu, Z.; Yang, Q.; Liu, L.; Jiang, Z.; Zhang, L.; Zhang, X.; Qing, H. "Cell-Addictive" Dual-Target Traceable Nanodrug for Parkinson's Disease Treatment via Flotillins Pathway. *Theranostics* **2018**, *8*, 5469–5481. <https://doi.org/10.7150/thno.28295>.
85. Zhang, J.; Zhou, X.; Yu, Q.; Yang, L.; Sun, D.; Zhou, Y.; Liu, J. Epigallocatechin-3-Gallate (EGCG)-Stabilized Selenium Nanoparticles Coated with Tet-1 Peptide To Reduce Amyloid- β Aggregation and Cytotoxicity. *ACS Appl. Mater. Interfaces* **2014**, *6*, 8475–8487. <https://doi.org/10.1021/am501341u>.
86. Wang, Y.; Luo, W.; Lin, F.; Liu, W.; Gu, R. Epigallocatechin-3-Gallate Selenium Nanoparticles for Neuroprotection by Scavenging Reactive Oxygen Species and Reducing Inflammation. *Front. Bioeng. Biotechnol.* **2022**, *10*, 989602.
87. Pandian, S.R.K.; Pavada, P.; Vellaisamy, S.; Ravishankar, V.; Palanisamy, P.; Sundar, L.M.; Chandramohan, V.; Sankaranarayanan, M.; Panneerselvam, T.; Kunjiappan, S. Formulation and Evaluation of Rutin-Loaded Solid Lipid Nanoparticles for the Treatment of Brain Tumor. *Naunyn Schmiedebergs Arch. Pharmacol.* **2021**, *394*, 735–749. <https://doi.org/10.1007/s00210-020-02015-9>.
88. Ahmad, H.; Arya, A.; Agrawal, S.; Mall, P.; Samuel, S.S.; Sharma, K.; Singh, P.K.; Singh, S.K.; Valicherla, G.R.; Mitra, K.; et al. Rutin Phospholipid Complexes Confer Neuro-Protection in Ischemic-Stroke Rats. *RSC Adv.* **2016**, *6*, 96445–96454. <https://doi.org/10.1039/C6RA17874J>.
89. Hu, B.; Dai, F.; Fan, Z.; Ma, G.; Tang, Q.; Zhang, X. Nanotheranostics: Congo Red/Rutin-MNPs with Enhanced Magnetic Resonance Imaging and H₂O₂-Responsive Therapy of Alzheimer's Disease in APP^{swe}/PS1^{dE9} Transgenic Mice. *Adv. Mat.* **2015**, *27*, 5499–5505. <https://doi.org/10.1002/adma.201502227>.
90. Rahman, M.; Kumar, V. Apigenin Loaded Phospholipid Based Nanoemulsion in Therapeutics of Parkinson's Disease via Attenuation of Oxidative Stress and Upregulation of Dopamine. *J. Neur. Sci.* **2019**, *405*, 247. <https://doi.org/10.1016/j.jns.2019.10.1269>.
91. Md, S.; Gan, S.Y.; Haw, Y.H.; Ho, C.L.; Wong, S.; Choudhury, H. In Vitro Neuroprotective Effects of Naringenin Nanoemulsion against β -Amyloid Toxicity through the Regulation of Amyloidogenesis and Tau Phosphorylation. *Int. J. Biol. Macromol.* **2018**, *118*, 1211–1219. <https://doi.org/10.1016/j.ijbiomac.2018.06.190>.
92. Gaba, B.; Khan, T.; Haider, M.F.; Alam, T.; Baboota, S.; Parvez, S.; Ali, J. Vitamin E Loaded Naringenin Nanoemulsion via Intranasal Delivery for the Management of Oxidative Stress in a 6-OHDA Parkinson's Disease Model. *Biomed. Res. Int.* **2019**, *2019*, e2382563. <https://doi.org/10.1155/2019/2382563>.
93. Ahmad, A.; Fauzia, E.; Kumar, M.; Mishra, R.K.; Kumar, A.; Khan, M.A.; Raza, S.S.; Khan, R. Gelatin-Coated Polycaprolactone Nanoparticle-Mediated Naringenin Delivery Rescue Human Mesenchymal Stem Cells from Oxygen Glucose Deprivation-Induced Inflammatory Stress. *ACS Biomater. Sci. Eng.* **2019**, *5*, 683–695. <https://doi.org/10.1021/acsbomaterials.8b01081>.
94. Zhao, Y.; Li, D.; Zhu, Z.; Sun, Y. Improved Neuroprotective Effects of Gallic Acid-Loaded Chitosan Nanoparticles Against Ischemic Stroke. *Rejuv. Res.* **2020**, *23*, 284–292. <https://doi.org/10.1089/rej.2019.2230>.
95. Frozza, R.L.; Bernardi, A.; Hoppe, J.B.; Meneghetti, A.B.; Battastini, A.M.O.; Pohlmann, A.R.; Guterres, S.S.; Salbego, C. Lipid-Core Nanocapsules Improve the Effects of Resveratrol Against A β -Induced Neuroinflammation. *J. Biomed. Nanotech.* **2013**, *9*, 2086–2104. <https://doi.org/10.1166/jbn.2013.1709>.
96. Frozza, R.L.; Bernardi, A.; Hoppe, J.B.; Meneghetti, A.B.; Matté, A.; Battastini, A.M.O.; Pohlmann, A.R.; Guterres, S.S.; Salbego, C. Neuroprotective Effects of Resveratrol Against A β Administration in Rats Are Improved by Lipid-Core Nanocapsules. *Mol. Neurobiol.* **2013**, *47*, 1066–1080. <https://doi.org/10.1007/s12035-013-8401-2>.
97. da Rocha Lindner, G.; Bonfanti Santos, D.; Colle, D.; Gasnhar Moreira, E.L.; Daniel Prediger, R.; Farina, M.; Khalil, N.M.; Mara Mainardes, R. Improved Neuroprotective Effects of Resveratrol-Loaded Polysorbate 80-Coated Poly(Lactide) Nanoparticles in MPTP-Induced Parkinsonism. *Nanomedicine* **2015**, *10*, 1127–1138. <https://doi.org/10.2217/nnm.14.165>.
98. Loureiro, J.A.; Andrade, S.; Duarte, A.; Neves, A.R.; Queiroz, J.F.; Nunes, C.; Sevin, E.; Fenart, L.; Gosselet, F.; Coelho, M.A.N.; et al. Resveratrol and Grape Extract-Loaded Solid Lipid Nanoparticles for the Treatment of Alzheimer's Disease. *Molecules* **2017**, *22*, 277. <https://doi.org/10.3390/molecules22020277>.

99. Li, C.; Wang, N.; Zheng, G.; Yang, L. Oral Administration of Resveratrol-Selenium-Peptide Nanocomposites Alleviates Alzheimer's Disease-like Pathogenesis by Inhibiting A β Aggregation and Regulating Gut Microbiota. *ACS Appl. Mater. Interfaces* **2021**, *13*, 46406–46420. <https://doi.org/10.1021/acsami.1c14818>.
100. Abozaid, O.A.R.; Sallam, M.W.; El-Sonbaty, S.; Aziza, S.; Emad, B.; Ahmed, E.S.A. Resveratrol-Selenium Nanoparticles Alleviate Neuroinflammation and Neurotoxicity in a Rat Model of Alzheimer's Disease by Regulating Sirt1/MiRNA-134/GSK3 β Expression. *Biol. Trace Elem. Res.* **2022**, *200*, 5104–5114. <https://doi.org/10.1007/s12011-021-03073-7>.
101. Mourtas, S.; Lazar, A.N.; Markoutsas, E.; Duyckaerts, C.; Antimisariis, S.G. Multifunctional Nanoliposomes with Curcumin-Lipid Derivative and Brain Targeting Functionality with Potential Applications for Alzheimer Disease. *Eur. J. Med. Chem.* **2014**, *80*, 175–183. <https://doi.org/10.1016/j.ejmech.2014.04.050>.
102. Barbara, R.; Belletti, D.; Pederzoli, F.; Masoni, M.; Keller, J.; Ballestrazzi, A.; Vandelli, M.A.; Tosi, G.; Grabrucker, A.M. Novel Curcumin Loaded Nanoparticles Engineered for Blood-Brain Barrier Crossing and Able to Disrupt Abeta Aggregates. *Int. J. Pharm.* **2017**, *526*, 413–424. <https://doi.org/10.1016/j.ijpharm.2017.05.015>.
103. Huang, N.; Lu, S.; Liu, X.-G.; Zhu, J.; Wang, Y.-J.; Liu, R.-T. PLGA Nanoparticles Modified with a BBB-Penetrating Peptide Co-Delivering A β ; Generation Inhibitor and Curcumin Attenuate Memory Deficits and Neuropathology in Alzheimer's Disease Mice. *Oncotarget* **2017**, *8*, 81001–81013. <https://doi.org/10.18632/oncotarget.20944>.
104. Yang, R.; Zheng, Y.; Wang, Q.; Zhao, L. Curcumin-Loaded Chitosan-Bovine Serum Albumin Nanoparticles Potentially Enhanced A β 42 Phagocytosis and Modulated Macrophage Polarization in Alzheimer's Disease. *Nanoscale Res. Lett.* **2018**, *13*, 330. <https://doi.org/10.1186/s11671-018-2759-z>.
105. Maiti, P.; Paladugu, L.; Dunbar, G.L. Solid Lipid Curcumin Particles Provide Greater Anti-Amyloid, Anti-Inflammatory and Neuroprotective Effects than Curcumin in the 5xFAD Mouse Model of Alzheimer's Disease. *BMC Neurosci.* **2018**, *19*, 7. <https://doi.org/10.1186/s12868-018-0406-3>.
106. Huo, X.; Zhang, Y.; Jin, X.; Li, Y.; Zhang, L. A Novel Synthesis of Selenium Nanoparticles Encapsulated PLGA Nanospheres with Curcumin Molecules for the Inhibition of Amyloid β Aggregation in Alzheimer's Disease. *J Photochem. Photobiol. B* **2019**, *190*, 98–102. <https://doi.org/10.1016/j.jphotobiol.2018.11.008>.
107. Kundu, P.; Das, M.; Tripathy, K.; Sahoo, S.K. Delivery of Dual Drug Loaded Lipid Based Nanoparticles across the Blood-Brain Barrier Impart Enhanced Neuroprotection in a Rotenone Induced Mouse Model of Parkinson's Disease. *ACS Chem. Neurosci.* **2016**, *7*, 1658–1670. <https://doi.org/10.1021/acschemneuro.6b00207>.
108. Saalbach, A.; Kunz, M. Impact of Chronic Inflammation in Psoriasis on Bone Metabolism. *Front. Immunol.* **2022**, *13*, 925503.
109. Tashiro, T.; Sawada, Y. Psoriasis and Systemic Inflammatory Disorders. *Int. J. Mol. Sci.* **2022**, *23*, 4457. <https://doi.org/10.3390/ijms23084457>.
110. Yang, S.-C.; Alalawi, A.; Lin, Z.-C.; Lin, Y.-C.; Aljuffali, I.A.; Fang, J.-Y. Anti-Inflammatory MicroRNAs for Treating Inflammatory Skin Diseases. *Biomolecules* **2022**, *12*, 1072. <https://doi.org/10.3390/biom12081072>.
111. Abdel-Mottaleb, M.M.; Try, C.; Pellequer, Y.; Lamprecht, A. Nanomedicine Strategies for Targeting Skin Inflammation. *Nanomedicine* **2014**, *9*, 1727–1743. <https://doi.org/10.2217/nnm.14.74>.
112. Yang, G.; Seok, J.K.; Kang, H.C.; Cho, Y.-Y.; Lee, H.S.; Lee, J.Y. Skin Barrier Abnormalities and Immune Dysfunction in Atopic Dermatitis. *Int. J. Mol. Sci.* **2020**, *21*, 2867. <https://doi.org/10.3390/ijms21082867>.
113. Chamcheu, J.C.; Siddiqui, I.A.; Adhami, V.M.; Esnault, S.; Bharali, D.J.; Babatunde, A.S.; Adame, S.; Massey, R.J.; Wood, G.S.; Longley, B.J.; et al. Chitosan-Based Nanoformulated (-)-Epigallocatechin-3-Gallate (EGCG) Modulates Human Keratinocyte-Induced Responses and Alleviates Imiquimod-Induced Murine Psoriasiform Dermatitis. *Int. J. Nanomed.* **2018**, *13*, 4189–4206. <https://doi.org/10.2147/IJN.S165966>.
114. Han, M.; Wang, X.; Wang, J.; Lang, D.; Xia, X.; Jia, Y.; Chen, Y. Ameliorative Effects of Epigallocatechin-3-Gallate Nanoparticles on 2,4-Dinitrochlorobenzene Induced Atopic Dermatitis: A Potential Mechanism of Inflammation-Related Necroptosis. *Front. Nutr.* **2022**, *9*, 953646.
115. Sun, M.; Xie, Q.; Cai, X.; Liu, Z.; Wang, Y.; Dong, X.; Xu, Y. Preparation and Characterization of Epigallocatechin Gallate, Ascorbic Acid, Gelatin, Chitosan Nanoparticles and Their Beneficial Effect on Wound Healing of Diabetic Mice. *Int. J. Biol. Macromol.* **2020**, *148*, 777–784. <https://doi.org/10.1016/j.ijbiomac.2020.01.198>.
116. Kar, A.K.; Singh, A.; Dhiman, N.; Purohit, M.P.; Jagdale, P.; Kamthan, M.; Singh, D.; Kumar, M.; Ghosh, D.; Patnaik, S. Polymer-Assisted In Situ Synthesis of Silver Nanoparticles with Epigallocatechin Gallate (EGCG) Impregnated Wound Patch Potentiate Controlled Inflammatory Responses for Brisk Wound Healing. *Int. J. Nanomed.* **2019**, *14*, 9837–9854. <https://doi.org/10.2147/IJN.S228462>.
117. Li, J.; Ni, W.; Aisha, M.; Zhang, J.; Sun, M. A Rutin Nanocrystal Gel as an Effective Dermal Delivery System for Enhanced Anti-Photoaging Application. *Drug. Dev. Ind. Pharm.* **2021**, *47*, 429–439. <https://doi.org/10.1080/03639045.2021.1890113>.
118. Cristiano, M.C.; Barone, A.; Mancuso, A.; Torella, D.; Paolino, D. Rutin-Loaded Nanovesicles for Improved Stability and Enhanced Topical Efficacy of Natural Compound. *J. Funct. Biomater.* **2021**, *12*, 74. <https://doi.org/10.3390/jfb12040074>.
119. Shen, L.-N.; Zhang, Y.-T.; Wang, Q.; Xu, L.; Feng, N.-P. Enhanced in Vitro and in Vivo Skin Deposition of Apigenin Delivered Using Ethosomes. *Int. J. Pharm.* **2014**, *460*, 280–288. <https://doi.org/10.1016/j.ijpharm.2013.11.017>.
120. Pleguezuelos-Villa, M.; Mir-Palomo, S.; Díez-Sales, O.; Buso, M.A.O.V.; Sauri, A.R.; Nacher, A. A Novel Ultradeformable Liposomes of Naringin for Anti-Inflammatory Therapy. *Colloids Surf. B Biointerfaces* **2018**, *162*, 265–270. <https://doi.org/10.1016/j.colsurfb.2017.11.068>.

121. Akrawi, S.H.; Gorain, B.; Nair, A.B.; Choudhury, H.; Pandey, M.; Shah, J.N.; Venugopala, K.N. Development and Optimization of Naringenin-Loaded Chitosan-Coated Nanoemulsion for Topical Therapy in Wound Healing. *Pharmaceutics* **2020**, *12*, 893. <https://doi.org/10.3390/pharmaceutics12090893>.
122. Caddeo, C.; Manca, M.L.; Matos, M.; Gutierrez, G.; Díez-Sales, O.; Peris, J.E.; Usach, I.; Fernández-Busquets, X.; Fadda, A.M.; Manconi, M. Functional Response of Novel Bioprotective Poloxamer-Structured Vesicles on Inflamed Skin. *Nanomedicine* **2017**, *13*, 1127–1136. <https://doi.org/10.1016/j.nano.2016.12.017>.
123. Shandil, A.; Yadav, M.; Sharma, N.; Nagpal, K.; Jindal, D.K.; Deep, A.; Kumar, S. Targeting Keratinocyte Hyperproliferation, Inflammation, Oxidative Species and Microbial Infection by Biological Macromolecule-Based Chitosan Nanoparticle-Mediated Gallic Acid–Rutin Combination for the Treatment of Psoriasis. *Polym. Bull.* **2020**, *77*, 4713–4738. <https://doi.org/10.1007/s00289-019-02984-9>.
124. Thi, P.L.; Lee, Y.; Tran, D.L.; Thi, T.T.H.; Kang, J.I.; Park, K.M.; Park, K.D. In Situ Forming and Reactive Oxygen Species-Scavenging Gelatin Hydrogels for Enhancing Wound Healing Efficacy. *Acta Biomater.* **2020**, *103*, 142–152. <https://doi.org/10.1016/j.actbio.2019.12.009>.
125. Wang, X.-C.; Huang, H.-B.; Gong, W.; He, W.-Y.; Li, X.; Xu, Y.; Gong, X.-J.; Hu, J.-N. Resveratrol Triggered the Quick Self-Assembly of Gallic Acid into Therapeutic Hydrogels for Healing of Bacterially Infected Wounds. *Biomacromolecules* **2022**, *23*, 1680–1692. <https://doi.org/10.1021/acs.biomac.1c01616>.
126. Oh, G.-W.; Ko, S.-C.; Je, J.-Y.; Kim, Y.-M.; Oh, J.; Jung, W.-K. Fabrication, Characterization and Determination of Biological Activities of Poly(ϵ -Caprolactone)/Chitosan-Caffeic Acid Composite Fibrous Mat for Wound Dressing Application. *Int. J. Biol. Macromol.* **2016**, *93*, 1549–1558. <https://doi.org/10.1016/j.ijbiomac.2016.06.065>.
127. Kaya, S.; Yilmaz, D.E.; Akmayan, I.; Egri, O.; Arasoglu, T.; Derman, S. Caffeic Acid Phenethyl Ester Loaded Electrospun Nanofibers for Wound Dressing Application. *J. Pharm. Sci.* **2022**, *111*, 734–742. <https://doi.org/10.1016/j.xphs.2021.09.041>.
128. Caddeo, C.; Nacher, A.; Vassallo, A.; Armentano, M.F.; Pons, R.; Fernández-Busquets, X.; Carbone, C.; Valenti, D.; Fadda, A.M.; Manconi, M. Effect of Quercetin and Resveratrol Co-Incorporated in Liposomes against Inflammatory/Oxidative Response Associated with Skin Cancer. *Int. J. Pharm.* **2016**, *513*, 153–163. <https://doi.org/10.1016/j.ijpharm.2016.09.014>.
129. Caldas, A.R.; Catita, J.; Machado, R.; Ribeiro, A.; Cerqueira, F.; Horta, B.; Medeiros, R.; Lúcio, M.; Lopes, C.M. Omega-3- and Resveratrol-Loaded Lipid Nanosystems for Potential Use as Topical Formulations in Autoimmune, Inflammatory, and Cancerous Skin Diseases. *Pharmaceutics* **2021**, *13*, 1202. <https://doi.org/10.3390/pharmaceutics13081202>.
130. Zhao, C.-C.; Zhu, L.; Wu, Z.; Yang, R.; Xu, N.; Liang, L. Resveratrol-Loaded Peptide-Hydrogels Inhibit Scar Formation in Wound Healing through Suppressing Inflammation. *Regen. Biomater.* **2020**, *7*, 99–107. <https://doi.org/10.1093/rb/rbz041>.
131. Li, X.; Chen, S.; Zhang, B.; Li, M.; Diao, K.; Zhang, Z.; Li, J.; Xu, Y.; Wang, X.; Chen, H. In Situ Injectable Nano-Composite Hydrogel Composed of Curcumin, N,O-Carboxymethyl Chitosan and Oxidized Alginate for Wound Healing Application. *Int. J. Pharm.* **2012**, *437*, 110–119. <https://doi.org/10.1016/j.ijpharm.2012.08.001>.
132. Gong, C.; Wu, Q.; Wang, Y.; Zhang, D.; Luo, F.; Zhao, X.; Wei, Y.; Qian, Z. A Biodegradable Hydrogel System Containing Curcumin Encapsulated in Micelles for Cutaneous Wound Healing. *Biomaterials* **2013**, *34*, 6377–6387. <https://doi.org/10.1016/j.biomaterials.2013.05.005>.
133. Aliboland, M.; Mohammadi, M.; Taghdisi, S.M.; Abnous, K.; Ramezani, M. Synthesis and Preparation of Biodegradable Hybrid Dextran Hydrogel Incorporated with Biodegradable Curcumin Nanomicelles for Full Thickness Wound Healing. *Int. J. Pharm.* **2017**, *532*, 466–477. <https://doi.org/10.1016/j.ijpharm.2017.09.042>.
134. Sun, L.; Liu, Z.; Wang, L.; Cun, D.; Tong, H.H.Y.; Yan, R.; Chen, X.; Wang, R.; Zheng, Y. Enhanced Topical Penetration, System Exposure and Anti-Psoriasis Activity of Two Particle-Sized, Curcumin-Loaded PLGA Nanoparticles in Hydrogel. *J. Control. Release* **2017**, *254*, 44–54. <https://doi.org/10.1016/j.jconrel.2017.03.385>.
135. Dai, X.; Liu, J.; Zheng, H.; Wichmann, J.; Hopfner, U.; Sudhop, S.; Prein, C.; Shen, Y.; Machens, H.-G.; Schilling, A.F. Nano-Formulated Curcumin Accelerates Acute Wound Healing through Dkk-1-Mediated Fibroblast Mobilization and MCP-1-Mediated Anti-Inflammation. *NPG Asia Mater.* **2017**, *9*, e368. <https://doi.org/10.1038/am.2017.31>.
136. Bajpai, S.K.; Ahuja, S.; Chand, N.; Bajpai, M. Nano Cellulose Dispersed Chitosan Film with Ag NPs/Curcumin: An in Vivo Study on Albino Rats for Wound Dressing. *Int. J. Biol. Macromol.* **2017**, *104*, 1012–1019. <https://doi.org/10.1016/j.ijbiomac.2017.06.096>.
137. Lee, H.-J.; Jeong, M.; Na, Y.-G.; Kim, S.-J.; Lee, H.-K.; Cho, C.-W. An EGF- and Curcumin-Co-Encapsulated Nanostructured Lipid Carrier Accelerates Chronic-Wound Healing in Diabetic Rats. *Molecules* **2020**, *25*, 4610. <https://doi.org/10.3390/molecules25204610>.
138. Mirzahosseini-pour, M.; Khorsandi, K.; Hosseinzadeh, R.; Ghazaeian, M.; Shahidi, F.K. Antimicrobial Photodynamic and Wound Healing Activity of Curcumin Encapsulated in Silica Nanoparticles. *Photodiagnosis Photodyn. Ther.* **2020**, *29*, 101639. <https://doi.org/10.1016/j.pdpdt.2019.101639>.
139. Graham, D.B.; Xavier, R.J. Pathway Paradigms Revealed from the Genetics of Inflammatory Bowel Disease. *Nature* **2020**, *578*, 527–539. <https://doi.org/10.1038/s41586-020-2025-2>.
140. Papoutsopoulou, S.; Burkitt, M.D.; Bergey, F.; England, H.; Hough, R.; Schmidt, L.; Spiller, D.G.; White, M.H.R.; Paszek, P.; Jackson, D.A.; et al. Macrophage-Specific NF-KB Activation Dynamics Can Segregate Inflammatory Bowel Disease Patients. *Front. Immunol.* **2019**, *10*, 2168. <https://doi.org/10.3389/fimmu.2019.02168>.
141. Haep, L.; Britzen-Laurent, N.; Weber, T.G.; Naschberger, E.; Schaefer, A.; Kremmer, E.; Foersch, S.; Vieth, M.; Scheuer, W.; Wirtz, S.; et al. Interferon Gamma Counteracts the Angiogenic Switch and Induces Vascular Permeability in Dextran Sulfate Sodium Colitis in Mice. *Inflamm. Bowel. Dis.* **2015**, *21*, 2360–2371. <https://doi.org/10.1097/MIB.0000000000000490>.

142. Jovani, M.; Fiorino, G.; Danese, S. Anti-IL-13 in Inflammatory Bowel Disease: From the Bench to the Bedside. *Curr. Drug. Targets* **2013**, *14*, 1444–1452. <https://doi.org/10.2174/13894501113149990170>.
143. Shen, W.; Wang, Q.; Shen, Y.; Gao, X.; Li, L.; Yan, Y.; Wang, H.; Cheng, Y. Green Tea Catechin Dramatically Promotes RNAi Mediated by Low-Molecular-Weight Polymers. *ACS Cent. Sci.* **2018**, *4*, 1326–1333. <https://doi.org/10.1021/acscentsci.8b00363>.
144. Gou, S.; Chen, Q.; Liu, Y.; Zeng, L.; Song, H.; Xu, Z.; Kang, Y.; Li, C.; Xiao, B. Green Fabrication of Ovalbumin Nanoparticles as Natural Polyphenol Carriers for Ulcerative Colitis Therapy. *ACS Sustain. Chem. Eng.* **2018**, *6*, 12658–12667. <https://doi.org/10.1021/acssuschemeng.8b01613>.
145. Guo, Z.; Bai, G.; Zhan, X.; Zhuo, K.; Wang, J.; Wang, Y. Supramolecular Vector/Drug Coassemblies of Polyglycerol Dendrons and Rutin Enhance the PH Response. *Langmuir* **2022**, *38*, 3392–3402. <https://doi.org/10.1021/acs.langmuir.1c03131>.
146. Lv, F.; Zhang, Y.; Peng, Q.; Zhao, X.; Hu, D.; Wen, J.; Liu, K.; Li, R.; Wang, K.; Sun, J. Apigenin-Mn(II) Loaded Hyaluronic Acid Nanoparticles for Ulcerative Colitis Therapy in Mice. *Front. Chem.* **2022**, *10*, 969962. <https://doi.org/10.3389/fchem.2022.969962>.
147. Priyadarshi, K.; Shirsath, K.; Waghela, N.B.; Sharma, A.; Kumar, A.; Pathak, C. Surface Modified PAMAM Dendrimers with Gallic Acid Inhibit, Cell Proliferation, Cell Migration and Inflammatory Response to Augment Apoptotic Cell Death in Human Colon Carcinoma Cells. *J. Biomol. Struct. Dyn.* **2021**, *39*, 6853–6869. <https://doi.org/10.1080/07391102.2020.1802344>.
148. Wang, J.; Tao, Z.; Tian, T.; Qiu, J.; Qian, H.; Zha, Z.; Miao, Z.; Ma, Y.; Wang, H. Polyoxometalate Nanoclusters: A Potential Preventative and Therapeutic Drug for Inflammatory Bowel Disease. *Chem. Eng. J.* **2021**, *416*, 129137. <https://doi.org/10.1016/j.cej.2021.129137>.
149. Tambuwala, M.M.; Khan, M.N.; Thompson, P.; McCarron, P.A. Albumin Nano-Encapsulation of Caffeic Acid Phenethyl Ester and Piceatannol Potentiated Its Ability to Modulate HIF and NF-KB Pathways and Improves Therapeutic Outcome in Experimental Colitis. *Drug Deliv. Transl. Res.* **2019**, *9*, 14–24. <https://doi.org/10.1007/s13346-018-00597-9>.
150. Pujara, N.; Wong, K.Y.; Qu, Z.; Wang, R.; Moniruzzaman, Md.; Rewatkar, P.; Kumeria, T.; Ross, B.P.; McGuckin, M.; Papat, A. Oral Delivery of β -Lactoglobulin-Nanosphere-Encapsulated Resveratrol Alleviates Inflammation in Winnie Mice with Spontaneous Ulcerative Colitis. *Mol. Pharm.* **2021**, *18*, 627–640. <https://doi.org/10.1021/acs.molpharmaceut.0c00048>.
151. Siu, F.Y.; Ye, S.; Lin, H.; Li, S. Galactosylated PLGA Nanoparticles for the Oral Delivery of Resveratrol: Enhanced Bioavailability and in Vitro Anti-Inflammatory Activity. *Int. J. Nanomed.* **2018**, *13*, 4133–4144. <https://doi.org/10.2147/IJN.S164235>.
152. Rachmawati, H.; Pradana, A.T.; Safitri, D.; Adnyana, I.K. Multiple Functions of D- α -Tocopherol Polyethylene Glycol 1000 Succinate (TPGS) as Curcumin Nanoparticle Stabilizer: In Vivo Kinetic Profile and Anti-Ulcerative Colitis Analysis in Animal Model. *Pharmaceutics* **2017**, *9*, 24. <https://doi.org/10.3390/pharmaceutics9030024>.
153. Oshi, M.A.; Lee, J.; Naeem, M.; Hasan, N.; Kim, J.; Kim, H.J.; Lee, E.H.; Jung, Y.; Yoo, J.-W. Curcumin Nanocrystal/PH-Responsive Polyelectrolyte Multilayer Core-Shell Nanoparticles for Inflammation-Targeted Alleviation of Ulcerative Colitis. *Biomacromolecules* **2020**, *21*, 3571–3581. <https://doi.org/10.1021/acs.biomac.0c00589>.
154. Kelly, T.; Yang, W.; Chen, C.-S.; Reynolds, K.; He, J. Global Burden of Obesity in 2005 and Projections to 2030. *Int. J. Obes.* **2008**, *32*, 1431–1437. <https://doi.org/10.1038/ijo.2008.102>.
155. Tsalamandris, S.; Antonopoulos, A.S.; Oikonomou, E.; Papamikroulis, G.-A.; Vogiatzi, G.; Papaioannou, S.; Deftereos, S.; Tousoulis, D. The Role of Inflammation in Diabetes: Current Concepts and Future Perspectives. *Eur. Cardiol.* **2019**, *14*, 50–59. <https://doi.org/10.15420/ecr.2018.33.1>.
156. Feuerer, M.; Shen, Y.; Littman, D.R.; Benoist, C.; Mathis, D. How Punctual Ablation of Regulatory T Cells Unleashes an Auto-immune Lesion within the Pancreatic Islets. *Immunity* **2009**, *31*, 654–664. <https://doi.org/10.1016/j.immuni.2009.08.023>.
157. Laveti, D.; Kumar, M.; Hemalatha, R.; Sistla, R.; Naidu, V.G.M.; Talla, V.; Verma, V.; Kaur, N.; Nagpal, R. Anti-Inflammatory Treatments for Chronic Diseases: A Review. *Inflamm. Allergy Drug Targets*. **2013**, *12*, 349–361.
158. Divella, R.; Gadaleta Caldarola, G.; Mazzocca, A. Chronic Inflammation in Obesity and Cancer Cachexia. *J. Clin. Med.* **2022**, *11*, 2191. <https://doi.org/10.3390/jcm11082191>.
159. Shoelson, S.E.; Herrero, L.; Naaz, A. Obesity, Inflammation, and Insulin Resistance. *Gastroenterology* **2007**, *132*, 2169–2180. <https://doi.org/10.1053/j.gastro.2007.03.059>.
160. Zagury, Y.; Chen, S.; Edelman, R.; Karnieli, E.; Livney, Y.D. β -Lactoglobulin Delivery System for Enhancing EGCG Biological Efficacy in HFD Obesity Mice Model. *J. Funct. Foods* **2019**, *59*, 362–370. <https://doi.org/10.1016/j.jff.2019.05.053>.
161. Zhang, X.-Z.; Guan, J.; Cai, S.-L.; Du, Q.; Guo, M.-L. Polymeric In Situ Hydrogel Implant of Epigallocatechin Gallate (EGCG) for Prolonged and Improved Antihyperlipidemic and Anti-Obesity Activity: Preparation and Characterization. *J. Biomater. Tissue Eng.* **2015**, *5*, 813–817. <https://doi.org/10.1166/jbt.2015.1381>.
162. Bhattacharjee, A.; Chakraborti, A.S. Argpyrimidine-Tagged Rutin-Encapsulated Biocompatible (Ethylene Glycol Dimers) Nanoparticles: Application for Targeted Drug Delivery in Experimental Diabetes (Part 2). *Int. J. Pharm.* **2017**, *528*, 8–17. <https://doi.org/10.1016/j.ijpharm.2017.05.058>.
163. Amjadi, S.; Shahnaz, F.; Shokouhi, B.; Azarmi, Y.; Siah-Shadbad, M.; Ghanbarzadeh, S.; Kouhsoltani, M.; Ebrahimi, A.; Hamishehkar, H. Nanophytosomes for Enhancement of Rutin Efficacy in Oral Administration for Diabetes Treatment in Streptozotocin-Induced Diabetic Rats. *Int. J. Pharm.* **2021**, *610*, 121208. <https://doi.org/10.1016/j.ijpharm.2021.121208>.
164. Zaghoul, R.A.; Abdelghany, A.M.; Samra, Y.A. Rutin and Selenium Nanoparticles Protected against STZ-Induced Diabetic Nephropathy in Rats through Downregulating Jak-2/Stat3 Pathway and Upregulating Nrf-2/HO-1 Pathway. *Eur. J. Pharmacol.* **2022**, *933*, 175289. <https://doi.org/10.1016/j.ejphar.2022.175289>.

165. Feng, W.; Guo, H.; Xue, T.; Wang, X.; Tang, C.; Ying, B.; Gong, H.; Cui, G. Anti-Inflammation and Anti-Fibrosis with PEGylated, Apigenin Loaded PLGA Nanoparticles in Chronic Pancreatitis Disease. *RSC Adv.* **2015**, *5*, 83628–83635. <https://doi.org/10.1039/C5RA17686G>.
166. Maity, S.; Chakraborti, A.S. Formulation, Physico-Chemical Characterization and Antidiabetic Potential of Naringenin-Loaded Poly D, L Lactide-Co-Glycolide (N-PLGA) Nanoparticles. *Eur. Polymer. J.* **2020**, *134*, 109818. <https://doi.org/10.1016/j.eurpolymj.2020.109818>.
167. Chen, C.; Jie, X.; Ou, Y.; Cao, Y.; Xu, L.; Wang, Y.; Qi, R. Nanoliposome Improves Inhibitory Effects of Naringenin on Nonalcoholic Fatty Liver Disease in Mice. *Nanomedicine* **2017**, *12*, 1791–1800. <https://doi.org/10.2217/nnm-2017-0119>.
168. Zhang, Y.; Zhang, L.; Ban, Q.; Li, J.; Li, C.-H.; Guan, Y.-Q. Preparation and Characterization of Hydroxyapatite Nanoparticles Carrying Insulin and Gallic Acid for Insulin Oral Delivery. *Nanomedicine* **2018**, *14*, 353–364. <https://doi.org/10.1016/j.nano.2017.11.012>.
169. Hou, Y.; Huang, H.; Gong, W.; Wang, R.; He, W.; Wang, X.; Hu, J. Co-Assembling of Natural Drug-Food Homologous Molecule into Composite Hydrogel for Accelerating Diabetic Wound Healing. *Biomater. Adv.* **2022**, *140*, 213034. <https://doi.org/10.1016/j.bioadv.2022.213034>.
170. Shahin, N.N.; Shamma, R.N.; Ahmed, I.S. A Nano-Liposomal Formulation of Caffeic Acid Phenethyl Ester Modulates Nrf2 and NF- κ B Signaling and Alleviates Experimentally Induced Acute Pancreatitis in a Rat Model. *Antioxidants* **2022**, *11*, 1536. <https://doi.org/10.3390/antiox11081536>.
171. Mangrulkar, S.; Shah, P.; Navnage, S.; Mazumdar, P.; Chaple, D. Phytophospholipid Complex of Caffeic Acid: Development, In Vitro Characterization, and In Vivo Investigation of Antihyperlipidemic and Hepatoprotective Action in Rats. *AAPS Pharm. Sci. Tech.* **2021**, *22*, 28. <https://doi.org/10.1208/s12249-020-01887-7>.
172. Wan, S.; Zhang, L.; Quan, Y.; Wei, K. Resveratrol-Loaded PLGA Nanoparticles: Enhanced Stability, Solubility and Bioactivity of Resveratrol for Non-Alcoholic Fatty Liver Disease Therapy. *R. Soc. Open Sci.* **2018**, *5*, 181457. <https://doi.org/10.1098/rsos.181457>.
173. Wang, Z.; Yan, Y.; Wang, Y.; Tong, F. The Interaction between CSE/H2S and the INOS/NO-Mediated Resveratrol/Poly(Ethylene Glycol)-Poly(Phenylalanine) Complex Alleviates Intestinal Ischemia/Reperfusion Injuries in Diabetic Rats. *Biomed. Pharmacother.* **2019**, *112*, 108736. <https://doi.org/10.1016/j.biopha.2019.108736>.
174. Mohseni, R.; ArabSadeghabadi, Z.; Ziamajidi, N.; Abbasalipourkabir, R.; RezaeiFarimani, A. Oral Administration of Resveratrol-Loaded Solid Lipid Nanoparticle Improves Insulin Resistance Through Targeting Expression of SNARE Proteins in Adipose and Muscle Tissue in Rats with Type 2 Diabetes. *Nanoscale Res. Lett.* **2019**, *14*, 227. <https://doi.org/10.1186/s11671-019-3042-7>.
175. Dong, Y.; Wan, G.; Yan, P.; Qian, C.; Li, F.; Peng, G. Fabrication of Resveratrol Coated Gold Nanoparticles and Investigation of Their Effect on Diabetic Retinopathy in Streptozotocin Induced Diabetic Rats. *J. Photochem. Photobiol. B* **2019**, *195*, 51–57. <https://doi.org/10.1016/j.jphotobiol.2019.04.012>.
176. Zu, Y.; Zhao, L.; Hao, L.; Mechref, Y.; Zabet-Moghaddam, M.; Keyel, P.A.; Abbasi, M.; Wu, D.; Dawson, J.A.; Zhang, R.; et al. Browning White Adipose Tissue Using Adipose Stromal Cell-Targeted Resveratrol-Loaded Nanoparticles for Combating Obesity. *J. Control. Release* **2021**, *333*, 339–351. <https://doi.org/10.1016/j.jconrel.2021.03.022>.
177. Devadasu, V.R.; Wadsworth, R.M.; Kumar, M.N.V.R. Protective Effects of Nanoparticulate Coenzyme Q10 and Curcumin on Inflammatory Markers and Lipid Metabolism in Streptozotocin-Induced Diabetic Rats: A Possible Remedy to Diabetic Complications. *Drug Deliv. Transl. Res.* **2011**, *1*, 448–455. <https://doi.org/10.1007/s13346-011-0041-3>.
178. El-Naggar, M.E.; Al-Joufi, F.; Anwar, M.; Attia, M.F.; El-Bana, M.A. Curcumin-Loaded PLA-PEG Copolymer Nanoparticles for Treatment of Liver Inflammation in Streptozotocin-Induced Diabetic Rats. *Colloids Surf. B Biointerfaces* **2019**, *177*, 389–398. <https://doi.org/10.1016/j.colsurfb.2019.02.024>.
179. Bateni, Z.; Rahimi, H.R.; Hedayati, M.; Afsharian, S.; Goudarzi, R.; Sohrab, G. The Effects of Nano-Curcumin Supplementation on Glycemic Control, Blood Pressure, Lipid Profile, and Insulin Resistance in Patients with the Metabolic Syndrome: A Randomized, Double-Blind Clinical Trial. *Phytother. Res.* **2021**, *35*, 3945–3953. <https://doi.org/10.1002/ptr.7109>.
180. Yücel, Ç.; Karatoprak, G.Ş.; Aktaş, Y. Nanoliposomal Resveratrol as a Novel Approach to Treatment of Diabetes Mellitus. *J. Nanosci. Nanotechnol.* **2018**, *18*, 3856–3864. <https://doi.org/10.1166/jnn.2018.15247>.
181. Frańk, W.; Wojtasińska, A.; Lisińska, W.; Młynarska, E.; Franczyk, B.; Rysz, J. Pathophysiology of Cardiovascular Diseases: New Insights into Molecular Mechanisms of Atherosclerosis, Arterial Hypertension, and Coronary Artery Disease. *Biomedicines* **2022**, *10*, 1938. <https://doi.org/10.3390/biomedicines10081938>.
182. Kotlyarov, S.; Kotlyarova, A. Molecular Pharmacology of Inflammation Resolution in Atherosclerosis. *Int. J. Mol. Sci.* **2022**, *23*, 4808. <https://doi.org/10.3390/ijms23094808>.
183. Mirhafez, S.R.; Mohebati, M.; Feiz Disfani, M.; Saberi Karimian, M.; Ebrahimi, M.; Avani, A.; Eslami, S.; Pasdar, A.; Rooki, H.; Esmaeili, H.; et al. An Imbalance in Serum Concentrations of Inflammatory and Anti-Inflammatory Cytokines in Hypertension. *J. Am. Soc. Hypertens.* **2014**, *8*, 614–623. <https://doi.org/10.1016/j.jash.2014.05.007>.
184. Hong, Z.; Xu, Y.; Yin, J.-F.; Jin, J.; Jiang, Y.; Du, Q. Improving the Effectiveness of (–)-Epigallocatechin Gallate (EGCG) against Rabbit Atherosclerosis by EGCG-Loaded Nanoparticles Prepared from Chitosan and Polyaspartic Acid. *J. Agric. Food Chem.* **2014**, *62*, 12603–12609. <https://doi.org/10.1021/jf5044603n>.

185. Zhang, J.; Nie, S.; Zu, Y.; Abbasi, M.; Cao, J.; Li, C.; Wu, D.; Labib, S.; Brackee, G.; Shen, C.-L.; et al. Anti-Atherogenic Effects of CD36-Targeted Epigallocatechin Gallate-Loaded Nanoparticles. *J. Control. Release* **2019**, *303*, 263–273. <https://doi.org/10.1016/j.jconrel.2019.04.018>.
186. Zhang, J.; Nie, S.; Wang, S. Nanoencapsulation Enhances Epigallocatechin-3-Gallate Stability and Its Antiatherogenic Bioactivities in Macrophages. *J. Agric. Food Chem.* **2013**, *61*, 9200–9209. <https://doi.org/10.1021/jf4023004>.
187. Wu, H.; Su, M.; Jin, H.; Li, X.; Wang, P.; Chen, J.; Chen, J. Rutin-Loaded Silver Nanoparticles With Antithrombotic Function. *Front. Bioeng. Biotechnol.* **2020**, *8*, 598977. <https://doi.org/10.3389/fbioe.2020.598977>.
188. Chen, D.; Liu, Y.; Liu, P.; Zhou, Y.; Jiang, L.; Yuan, C.; Huang, M. Orally Delivered Rutin in Lipid-Based Nano-Formulation Exerts Strong Antithrombotic Effects by Protein Disulfide Isomerase Inhibition. *Drug Deliv.* **2022**, *29*, 1824–1835. <https://doi.org/10.1080/10717544.2022.2083726>.
189. Guo, M.; He, Z.; Jin, Z.; Huang, L.; Yuan, J.; Qin, S.; Wang, X.; Cao, L.; Song, X. Oral Nanoparticles Containing Naringenin Suppress Atherosclerotic Progression by Targeting Delivery to Plaque Macrophages. *Nano Res.* **2023**, *16*, 925–937. <https://doi.org/10.1007/s12274-022-4808-2>.
190. He, Y.; Wang, J.; Yan, W.; Huang, N. Gallic Acid and Gallic Acid-Loaded Coating Involved in Selective Regulation of Platelet, Endothelial and Smooth Muscle Cell Fate. *RSC Adv.* **2014**, *4*, 212–221. <https://doi.org/10.1039/C3RA44217A>.
191. Alfei, S.; Grazia Signorello, M.; Schito, A.; Catena, S.; Turrini, F. Reshaped as Polyester-Based Nanoparticles, Gallic Acid Inhibits Platelet Aggregation, Reactive Oxygen Species Production and Multi-Resistant Gram-Positive Bacteria with an Efficiency Never Obtained. *Nanoscale Adv.* **2019**, *1*, 4148–4157. <https://doi.org/10.1039/C9NA00441F>.
192. Zhou, H.; Shan, Y.; Tong, F.; Zhang, Y.; Tang, J.; Shen, R.; Chen, D. Resveratrol Nanoparticle Complex: Potential Therapeutic Applications in Myocardial Ischemia Reperfusion Injury. *J. Biomed. Nanotechnol.* **2020**, *16*, 382–389. <https://doi.org/10.1166/jbn.2020.2900>.
193. Sun, L.; Hu, Y.; Mishra, A.; Sreeharsha, N.; Moktan, J.B.; Kumar, P.; Wang, L. Protective Role of Poly(Lactic-co-glycolic) Acid Nanoparticle Loaded with Resveratrol against Isoproterenol-induced Myocardial Infarction. *BioFactors* **2020**, *46*, 421–431. <https://doi.org/10.1002/biof.1611>.
194. Yadav, Y.C.; Pattnaik, S.; Swain, K. Curcumin Loaded Mesoporous Silica Nanoparticles: Assessment of Bioavailability and Cardioprotective Effect. *Drug Dev. Ind. Pharm.* **2019**, *45*, 1889–1895. <https://doi.org/10.1080/03639045.2019.1672717>.
195. Li, X.; Xiao, H.; Lin, C.; Sun, W.; Wu, T.; Wang, J.; Chen, B.; Chen, X.; Cheng, D. Synergistic Effects of Liposomes Encapsulating Atorvastatin Calcium and Curcumin and Targeting Dysfunctional Endothelial Cells in Reducing Atherosclerosis. *Int. J. Nanomed.* **2019**, *14*, 649–665. <https://doi.org/10.2147/IJN.S189819>.

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.