

Review

New and Old Horizons for an Ancient Drug: Pharmacokinetics, Pharmacodynamics, and Clinical Perspectives of Dimethyl Fumarate

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Abstract: (1) Background: In their 60-year history, dimethyl fumarate and other salts of fumaric acid have been used for the treatment of psoriasis and other immune-mediated diseases for their immune-modulating properties. Over the years, new mechanisms of action have been discovered for this evergreen drug that remains a first-line treatment for several different inflammatory diseases. Due to its pleiotropic effects, this molecule is still of great interest in varied conditions, not exclusively inflammatory diseases. (2) Methods: The PubMed database was searched using combinations of the following keywords: dimethyl fumarate, pharmacokinetics, pharmacodynamics, adverse effects, psoriasis, multiple sclerosis, and clinical indications. This article reviews and updates the pharmacokinetics, mechanisms of action, and clinical indications of dimethyl fumarate. (3) Conclusions: The pharmacology of dimethyl fumarate is complex, fascinating, and not fully known. Progressive insights into the molecule's mechanisms of action will make it possible to maximize its clinical efficacy, reduce concerns about adverse effects, and find other possible areas of application.

Keywords: dimethyl fumarate; pharmacodynamics; pharmacokinetics; adverse effects; psoriasis; multiple sclerosis; immune-mediated diseases



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1. Introduction

Characterized by high hygroscopicity and fungicidal power, dimethyl fumarate (dimethyl (2E)-but-2-enedioate—DMF) was once used as a dehumidifying agent and was placed in special bags, sometimes called 'silica gel', inside furniture or shoe boxes and leather bags to prevent deterioration during transport and storage in high humidity environments. DMF is the methyl ester of fumaric acid, whose name derives from the earth smoke plant *Fumaria officinalis*. The first description of a medical use dates to 1959, when the German biochemist Schweckendiek, who suffered from psoriasis, highlighted the benefits of fumaric acid esters (FAEs) on his inflammatory skin disease [1]. In 1994, a combination of FAEs containing DMF and salts of monoethyl fumarate was licensed for the treatment of psoriasis by the German Drug Administration with the name of Fumaderm[®]. The combination was empirically determined, as well as its dosing regimen. Since then, the action and contribution of the different FAEs to their overall therapeutic effect in psoriasis has been studied more in detail, showing that DMF is the main active agent via its metabolic transformation to monomethyl fumarate (MMF) [2]. Today, DMF is still a mainstay in the treatment of psoriasis, and over time its clinical indication has expanded to multiple sclerosis (MS). Moreover, during the current SARS-CoV-2 pandemic, DMF is experiencing renewed interest for the treatment of inflammatory immune-mediated diseases, not being a conventional immunosuppressant but rather a molecule with pleiotropic effects.

and proteolysis of Nrf2. In fact, KEAP1 serves as an adaptor for Cullin3 (Cul3)-based E3 ligase that regulates the proteosomal degradation of Nrf2. In the case of oxidation or alkylation of the highly reactive cysteine residues of KEAP1, as occurs in the presence of increased oxidative or electrophilic agents, the protein complex destabilizes and Nrf2 is no longer degraded [9]. Subsequently, Nrf2 moves into the nucleus as a heterodimer with small Maf proteins and acts as a transcription factor by binding the ARE sequence in the promoter of different target genes that encode for cytoprotective proteins, including heme oxygenase-1 (HO-1), quinoline oxidoreductase-1 (NQO1), catalase, superoxide dismutase (SOD), glutathione, and glutathione-dependent antioxidant enzymes [10]. In vitro studies have shown that DMF, as an electrophilic agent, oxidizes reactive thiols on KEAP1 and causes depletion of the intracellular antioxidant stocks of glutathione by forming conjugations with it, resulting in an enhanced activation of the Nrf2 pathway (Figure 2).

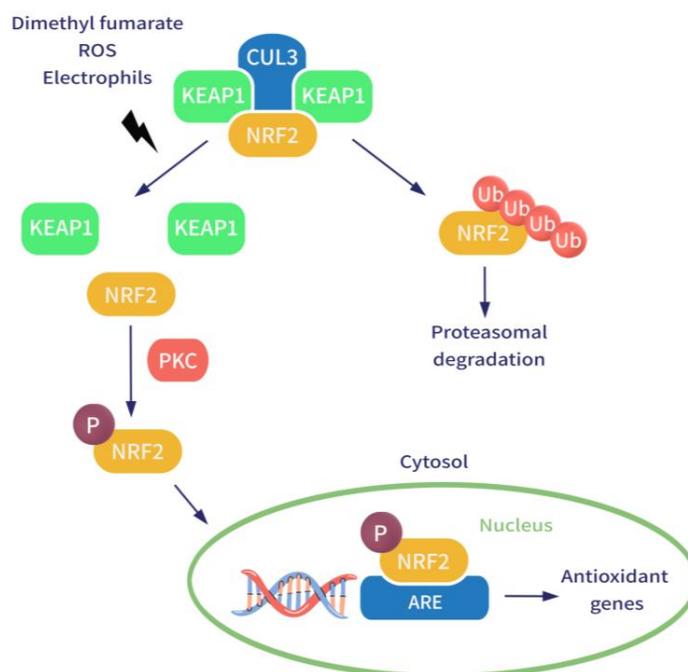


Figure 2. Effect of dimethyl fumarate on Nrf2 pathway. Abbreviations: NRF2, nuclear factor (erythroid-derived 2)-related factor; ARE, antioxidant response element; CUL3, Cullin3; KEAP1, Kelch-like ECH-associated protein; PKC, protein kinase C; Ub, ubiquitin; ROS, reactive oxygen species.

Therefore, the operating principle of DMF is related to the toxicological mechanism called “hormesis”, for which subtoxic amounts of potentially harmful oxidants or electrophiles can stimulate cell protection against toxic concentrations of the same or other molecules via the activation of the Nrf2 pathway. This phenomenon is analogous to the ischemic preconditioning, in which a sub-lethal ischemic insult has a protective effect on myocardial or brain cells against a subsequent ischemic challenge [11]. In vitro studies have shown that, even if to a lower degree, MMF also directly enhances Nrf2’s pathway but causes less intranuclear accumulation of the NFR2 protein compared to DMF [12]. Cuadrado et al. demonstrated that DMF could also stimulate Nrf2 activation by enhancing the PI3K/AKT/GSK-3 β pathway [13]. Glycogen synthase kinase-3 β is a serine-threonine kinase that phosphorylates critical residues of Nrf2, facilitating its ubiquitin-proteasome degradation. DMF modifies the phosphorylation status of GSK-3 β and reduces its activity in the hippocampus of laboratory mice, resulting in a downregulation of this proteolytic pathway targeting Nrf2. Thus, DMF could exert a double mechanism of activation of Nrf2 by disrupting KEAP1/NRF2 and by reducing Nrf2 degradation [14]. DMF also appears to modulate TAU levels of phosphorylation in mouse models. TAU proteins stabilize microtubules, and when hyperphosphorylated cause severe neurodegenerative disease called

taupathies, such as Alzheimer's disease. GSK-3 β regulates intracellular substrates involved in neuronal polarization, including collapsing response mediator protein 2 (CRMP2) and TAU, active players of microtubule organization. GSK-3 β hyperactivity has been linked to this group of diseases, and DMF appears a promising approach in their treatment [15]. Finally, Nrf2 exerts a modulation of the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) pathway within a regulatory feedback loop, leading to a repression in the activity of this potent transcription factor of pro-inflammatory genes [16].

3.2. Effects on the Hydroxycarboxylic Acid Receptor 2 (HCAR2)

The cytoprotective effect of fumaric acid esters, especially on nervous tissue, can also occur due to the agonist action that DMF and, even more so, MMF exert on the HCAR2. This is a G protein-coupled receptor for nicotinic acid (niacin) expressed in many immune cells such as neutrophils, dendritic cells, macrophages, and microglia that, independent from Nrf2, activates a potent anti-inflammatory signaling. The HCAR2 downstream pathway leads to NF- κ B inhibition, and subsequently modulates the phenotype of microglia in the brain by promoting a switch towards an anti-inflammatory pattern of cytokine secretion and by normalizing the glutamatergic synaptic transmission [17]. This mechanism has shown a therapeutic effect on some experimental models of neurological diseases such as autoimmune encephalomyelitis, where DMF and MMF reduced neutrophil adhesion, migration, and recruitment to the central nervous system (CNS) [18]. The effects are not limited to the CNS but also involve other immune cells: HCAR2 activation on macrophages has been linked to an anti-atherogenic activity. HCAR2 also promotes the secretion of prostaglandins D2 and E2, whose effect of vasodilatation is responsible for mediating niacin and DMF-induced flushing, which is one of the most common adverse effects of the drug [19].

3.3. Modulation of GSH

Reactive oxygen species (ROS) have a critical role in the pathogenesis of neurological inflammatory disease and psoriasis. ROS induce transendothelial leukocyte migration with subsequent myelin and axonal damage. ROS production was found to be increased in the psoriatic plaque. GSH represents a main intracellular antioxidant system. The initial depletion of GSH caused by fumarates is not as severe to cause programmed cell death, but rather it promotes antioxidant mechanisms of compensation not only by boosting glutathione and glutathione-related enzymes *de novo* via the Nrf2 pathway, but also by enhancing GSH recycling, as demonstrated by Hoffman et al. [20]. GSH levels' modulation caused by DMF could contribute to the inhibition of NF- κ B in an Nrf2-independent manner [21].

3.4. Inhibition of NF- κ B

The effect on NF- κ B can be explained not only by the mechanisms described above, but indeed the inhibition of this important pro-inflammatory transcription factor results also from the interaction of DMF with the p65 protein, also known as the nuclear factor NF- κ B p65 subunit. This molecule is an essential post-translational regulator of NF- κ B, whose phosphorylation and acetylation are required for its activation. DMF, acting as an electrophilic agent, induces covalent modification of cysteine residues on p65, resulting in the blockade of the nuclear translocation and phosphorylation of this protein. This DMF-mediated mechanism of inhibition of NF- κ B has been demonstrated by Kastrati et al. in breast cancer cells and, added to the activation of the Nrf2 cytoprotective pathway, may suggest an anti-tumor activity of DMF [22].

3.5. Effects on the Immune System

In vitro studies by Treumer et al. demonstrated a proapoptotic action of DMF on human purified T-lymphocytes in a concentration- and time-dependent manner [23]. Apoptosis is the consequence of GSH depletion induced by DMF that, in some target cells, may

reach the threshold for induction of cell death. The mechanism was found to be more intense on activated T cells when compared to resting T cells. Indeed, during DMF therapy lymphopenia represents one of the major adverse effects, and complete blood count needs to be monitored periodically. However, this side effect only affects a minority of patients, thus it is possible that only in some predisposed individuals DMF reaches high tissue concentrations, which perhaps combined with other intercurrent lymphocytotoxic stimuli induces massive T cell death. In addition to the possible effect on leukocyte count, fumarates modulate the inflammatory profile of T cells by inducing a shift from the Th1 to Th2 lymphocyte phenotype and cytokine pattern. Th1-activated lymphocytes with their secretion of interferon γ (IFN- γ) and interleukin (IL)-17 play an important role in the pathogenesis of both psoriasis and MS. In vitro studies by de Jong et al. demonstrated that MMF increased the production of cytokines related to Th2 lineage such as IL-4 and IL-5 in stimulated T cells, without suppressing the production of Th1 cytokines [24]. Another investigation has shown that DMF reduces the differentiation of T naïve cells' into reactive Th1 and Th17 lymphocytes by inhibiting dendritic cell maturation. The inhibition of the pro-inflammatory cytokines downstream of the NF- κ B pathway has an important role in rebalancing these two branches of the adaptive response. As a result of this anti-inflammatory activity, DMF caused inhibition of the tumor necrosis factor (TNF)- α -induced expression of the adhesion molecule, as did E-selectin and VCAM-1 in an experimental model of endothelial cell cultures, leading to the reduction in leukocyte recruitment [25]. The psoriatic plaque is characterized by a skin accumulation of lymphocytes due to up-regulation of endothelial cell adhesion molecules, and this could explain one possible mechanism of action of DMF in this inflammatory disease [26].

3.6. Regulation of Iron Metabolism in the Brain

Iron is an essential element for myelination [27], and disorders of iron metabolism have been recently incriminated as possible triggers of neurodegenerative diseases. Extracellular unbound iron accumulation in the brain is an age-related phenomenon, but elevated iron levels in microglia can be considered a hallmark of pathological processes [28]. In physiological conditions, microglia cells store iron-bound ferritin and release it to oligodendrocytes to support myelination [29]. In MS, there is a pathological unbound iron release by damaged oligodendrocytes in the extracellular space that induces the production of free radicals and accumulates in macrophages and microglia. The subsequent activation of the inflammatory cascade results in neuron damage and disruption of the myelination process [30]. Iron accumulation in macrophages and microglia around the rim of the demyelinated plaques in MS can be highlighted by magnetic resonance imaging (MRI) [31]. Evidence in primary murine microglia shows that DMF acts as a regulator of the brain's iron metabolism by upregulating specific proteins (such as T-cell immunoglobulin mucin domain 2 protein (TIM-2) and transferrin receptor 1) that improve the uptake of iron in microglial cells and the transfer of ferritin to oligodendrocytes and promote a shift of the microglia towards an anti-inflammatory and neuroprotective phenotype [32].

3.7. Antimicrobial Effects and Modulation of Gut Microbiota

Fumaric acid esters, as organic acids, are commonly used as antimicrobials in foods [33]. Although it has never been used in humans for its antibiotic activity, DMF exerts an antimicrobial activity towards different microorganisms. An antibacterial effect against *Escherichia coli* has been demonstrated in vitro by Wang et al. [34] and against *Clostridium perfringens* by Rumah et al. [35]. DMF in vivo also seems to modulate the gut microbiome profile. This is possible, maybe not only due to its antimicrobial effect, but also because some intestinal bacteria are able to metabolize fumaric acid esters with a competitive advantage [36]. A treatment of 12 weeks with oral DMF in patients with MS was not associated with significant alterations in the gut microbiota composition but showed a near-normalization of the low abundance of butyrate-producing *Faecalibacterium* seen in MS patients [37]. Butyrate is an important nutrient for the homeostasis of the gut barrier and

the intestinal immune system, exerting a beneficial anti-inflammatory effect in intestinal and extraintestinal disease. Thus, a direct effect on the gut microbiota could account for the therapeutic action of DMF.

4. Adverse Effects

4.1. Gastrointestinal Adverse Effects

Given the pharmacokinetics of DMF, it is known that only a small fraction of the molecule enters the systemic circulation after oral administration, and it is possible that this small amount is responsible for the hermetic dose response in the Nrf2 pathway that is observed in neurons or other tissues exposed to a low concentration of the drug. The gut barrier, on the other hand, is exposed to a high concentration of the drug, which would cause large oxidative stress and GSH depletion. This could explain the high frequency of gastrointestinal (GI) adverse effects in patients treated with DMF and the warning not to consume alcohol with the drug to improve tolerability. In fact, combining several harmful stimuli at the same time would exponentially amplify the toxic effect on the GI epithelium, while taking the drug with food (particularly a high-fat meal) reduces the incidence and severity of GI adverse effects. Diarrhea is the most common adverse reaction, followed by abdominal distension and pain, nausea, vomiting, dyspepsia, and constipation. These side effects usually occur in the first 2 to 3 months of treatment, after which a gradual tolerance develops. They are usually mild to moderate, although diarrhea could become so severe to affect the absorption of other medicinal and oral contraceptives. There are no apparent dose-related effects and no risk factors for the occurrence could be identified, however, slow dose titration when initiating therapy is useful to reduce GI discomfort. Furthermore, specific symptom-directed therapies are useful to manage intolerable side effects (Table 1) [38]. Although there are experimental studies that show anti-inflammatory responses in immune-mediated colitis, severe gastrointestinal disorders remain a contraindication in the use of the drug.

Table 1. Table with a list of the most frequent gastrointestinal adverse effects caused by DMF and possible therapeutic options.

Adverse Effects	Possible Therapeutic Options
Diarrhea	<ul style="list-style-type: none"> • Loperamide
Abdominal pain	<ul style="list-style-type: none"> • Proton pump inhibitors • Bismuth subsalicylates • H₂ blockers
Vomiting	<ul style="list-style-type: none"> • Ondansetron • Promethazine
Nausea	<ul style="list-style-type: none"> • Ondansetron • Antiacids • Promethazine • Bismuth subsalicylates

4.2. Hematological Disorders

DMF may decrease leukocyte and lymphocyte counts. Notably, the risk of DMF-associated lymphopenia increases with age. In adults older than 55 years this side effect is more frequent and more severe, with grade 2 (500–799 cells/ μ L) and 3 (200–500 cells/ μ L) lymphopenia developing in more than 40% of cases in this age group. No cases of lymphopenia were observed in pediatric populations [39]. Patients with MS switched to DMF from natalizumab and those with a lower baseline lymphocyte count have a higher risk of developing this side effect. Thus, prior to initiating DMF therapy it is mandatory to perform a complete blood count (CBC), and the treatment should not be initiated if leukopenia below $3.0 \times 10^9/L$ or lymphopenia below $1.0 \times 10^9/L$ are reported. Blood

monitoring should be performed after one month from the start and then every 3 months. If the lymphocyte count falls below $1.0 \times 10^9/L$ but is $\geq 0.7 \times 10^9/L$, blood monitoring should be performed monthly until levels return to $1.0 \times 10^9/L$ or higher for two consecutive blood tests. If leukocyte count decreases below $3.0 \times 10^9/L$ or lymphocytes below $0.7 \times 10^9/L$ the treatment must be stopped immediately, and scrupulous monitoring of the count should be performed until it has returned to the normal range. Recovery after drug withdrawal is usually rapid (2–4 months) but it may be prolonged (≥ 6 months), especially in patients who reached lower lymphocyte count at DMF interruption. Moreover, this collateral effect was found to be more persistent in MS patients with longer disease duration and previous exposure to other immunomodulating treatments [40]. Patients who present severe and prolonged lymphopenia are at increased risk of developing progressive multiple leukoencephalopathy (PML) [41], even if it has also been reported in patients without severe lymphocytopenia [42].

4.3. Progressive Multiple Leukoencephalopathy

Progressive multiple leukoencephalopathy (PML) is a demyelinating disease caused by the John Cunningham virus (JCV), an opportunistic pathogen. JCV is a type of polyomavirus that infects approximately 70 to 90% of the general population [43] asymptotically. In the case of immune deficiency such as HIV infections, chemotherapy, patients with transplants, and therapy with immunosuppressants, the virus can cause potentially fatal nervous system injuries. The most frequent symptoms are weakness, cognitive impairment, personality changes, and visual and speech disorders. The incidence of PML in patients with MS during treatment with DMF is 0.02 per 1000 patients, and the risk is higher in patients with persistent severe lymphopenia and older age, although cases have occurred in young patients with normal absolute lymphocyte count (ALC), suggesting that changes in specific subsets might be more important than total ALC [44]. In patients with psoriasis there are no specific data, although in a large, population-based study including subjects with psoriatic arthritis, psoriasis, rheumatoid arthritis, and other rheumatic diseases, the incidence of PML was 0.2 per 100,000 persons without HIV or cancer. Most of the cases occurred in patients previously treated with rituximab and infliximab, although a causal role for biologic agents remains uncertain [45]. The diagnosis of PML requires a high index of clinical suspicion, especially in patients with a pre-existent neurological disease such as MS who could present natural fluctuations in the activity of the disease. A polymerase chain reaction (PCR) on cerebrospinal fluid (CSF) and an MRI are essential to corroborate the suspect and interrupt potentially harmful therapies [46].

4.4. Fanconi Syndrome

Fumaric ester acids can cause drug-induced Fanconi syndrome (FS). This is a dysfunction of proximal renal tubules that can lead to inadequate reabsorption of glucose, bicarbonate, amino acids, and phosphate. Patients who develop FS typically present generalized weakness, myalgia, proteinuria and glycosuria, osteomalacia, and pathological bone fractures. The mechanism by which DMF appears to cause tubular damage, although unclear, appears to be related to mitochondrial damage, as shown by electron microscopy studies [47]. Although FS is an infrequent side effect, renal function (e.g., creatinine, blood urea nitrogen, and urinalysis) should be checked prior to initiation of treatment and every 3 months thereafter.

4.5. Flushing

Differently from FS, flushing is a very common adverse effect of DMF. HCAR2 is thought to be involved in the flushing reaction. When MMF binds to HCAR2, it induces prostanoid-forming enzymes in different cell types. Prostaglandin-induced vasodilatation may be mitigated by pretreatment with acetylsalicylic acid and slow titration of the dose [48].

5. Indications for DMF

5.1. Psoriasis

DMF is approved for moderate to severe chronic plaque psoriasis (CPP) as the first-line systemic therapy before starting a biological therapy. The first fumarate Fumaderm[®] (a mixture comprising 60% DMF and calcium, zinc, and magnesium salts of monoethyl fumarate) has been available in Germany since 1994, and more recently a new formulation containing only DMF (Skilarence[®]) was approved for psoriasis in 2008 by the European Medicines Agency. The therapeutic dose of the drug ranges between 30 and 720 mg. An Italian real-life multicentric retrospective (30-month period) study published in 2021 including 103 psoriatic patients showed interesting benefit/risk ratio results for a first-line therapy: in particular, 23% of patients reached PAS75 at week 12, while more than 40%, or almost 80% considering the patients who were continuing the treatment, reached it at week 26 [49]. Side effects, mainly mild to moderate transitory gastrointestinal symptoms, were the main cause of treatment discontinuation. Slow titration of the dose was needed for a good tolerability of the drug, and when side effects appeared after a temporary dose reduction a subsequent further increase was often better tolerated by many patients. Most of the dropouts occurred in the first 5 months of treatment. With reference to variables associated with DMF effectiveness, patients who achieved higher values of PASI at T12 had a lower BMI and were taking a higher daily dose than those who did not. At the same time, treatment discontinuation was inversely related to the median daily dose and the BMI. Moreover, the study highlights the safety of the molecule in oncological patients, as a consistent number of patients treated had an oncological history (18.4%) and none of them had recurrence of malignancy during the follow-up. Data from either prospective interventional (BRIDGE) or non-interventional (DIMESKIN 1, SKILL) studies among patients with moderate-to-severe psoriasis showed that DMF also provides a positive efficacy profile in all four body regions included in the Psoriasis Area and Severity Index assessment (head and neck, trunk, and upper and lower extremities) and a particularly interesting profile (strong efficacy) in the head and neck region. These findings may be of special interest to patients with scalp psoriasis, which has always been considered one of the most difficult-to-treat areas. Patient-reported outcomes (quality of life and pruritus) also improved during the 24 weeks of DMF treatment [50].

Furthermore, DMF, due to its mechanism of action, could be considered an interesting therapeutic choice for patients suffering from psoriasis with chronic infections (e.g., HIV and TBC), whose frailty could be addressed by restoring a balance between the Th1/Th2 pathways [51,52]. In times of economic constraints, a study from the UK, performing a cost-effectiveness analysis of treatment sequences, confirmed the role of DMF prior to biological agents as a dominant treatment strategy for the treatment of moderate-to-severe psoriasis [53]. In conclusion, DMF appears as an effective, easy handling, and dynamic first-line therapy for CPP with a high cost-effectiveness ratio.

5.2. Multiple Sclerosis

The first observations that fumaric acid esters might have a role in the treatment of MS date back to 2006, when Fumaderm[®] was administered to 10 patients in an open-label pilot study with encouraging results on both clinical and radiological progression of the disease [54]. Since then, there have been many clinical trials with fumarates on this autoimmune neurological disease. Currently, BG-12 (marketed as Tecfidera[®]), an orally administered, enteric-coated micro tablet slow-release preparation of DMF, is indicated for the treatment of this disease in the relapsing remitting variant (RRMS). RRMS is the most common subtype, characterized by unpredictable flare-ups followed by periods of months to years of incomplete or complete recovery, termed remissions. This formulation of DMF demonstrated fewer GI-related side effects compared to the old fumarates and is effective in reducing the number of new demyelinating lesions and annual relapses compared to the placebo [55]. Larger phase III group studies later confirmed the significant efficacy and the favorable benefit–risk profile of DMF in patients with RRMS failure to interferon beta. Over their 2-year duration, these multicenter, placebo-controlled, double-blind clinical

trials proved that BG-12 reduced the number and the size of neurological lesions (observed via MRI) and significantly decreased the proportion of relapsing patients [56]. Although adverse effects such as nausea, vomiting, dyspepsia, diarrhea, and flushing were commonly reported, especially in the first weeks, a progressive improvement was observed during the treatment, and safety and tolerability of DMF were deemed acceptable in these trials [57].

6. Special Issues and Possible Future Indications

6.1. Pregnancy and Breastfeeding

DMF is not recommended in pregnant and breastfeeding women and should be used only if the benefit justifies the potential fetal risk. However, an interim analysis of 345 patients exposed to DMF during the first trimester in a prospective international registry performed by Hellwig et al. demonstrated that pregnancy outcomes (including gestational size, pregnancy loss, birth defects, and infant or maternal death) were not affected by the drug (with a mean exposure of 4.9 weeks) [58]. Moreover, due to its extremely short half-life, DMF can be considered a versatile drug to use in potential childbearing women, because the prompt interruption in the case of unexpected pregnancy could prevent possible fetal consequences. No information is available on the clinical use of dimethyl fumarate during breastfeeding. However, amounts of the active metabolite of dimethyl fumarate, monomethyl fumarate, appear to be low in breastmilk and would not be expected to cause any adverse effects in breastfed infants. Before any data were available, some authors recommend avoiding breastfeeding during dimethyl fumarate therapy [59].

6.2. SARS-CoV-2 Infection

Since the beginning of pandemic of SARS-CoV-2 in 2019, clinicians of all specialties must deal with the issue of treating inflammatory diseases with immunosuppressive drugs that could potentially predispose the patient to an increased risk of acquiring the infection or a more severe course. This is especially important when the patient is elderly, frail, or has multiple comorbidities [60]. The possibility to rely on a pleiotropic drug without pure immunosuppressive features such DMF is important in the clinician armamentarium. Moreover, DMF has demonstrated to exert a potent antiviral and anti-inflammatory activity through the activation of the NRF2-pathway, distinct from the type I interferon pathway, towards Herpes simplex 1 and 2, Vaccinia virus, Zika virus, and SARS-CoV-2 [61]. Experimental data from a small case series showed that MS patients treated with DMF showed developed self-limiting benign SARS-CoV-2 infection and could continue the therapy with DMF at the usual dosage [62]. An Italian multicenter retrospective study including 44 psoriatic patients who were taking DMF during 48 weeks of the pandemic showed that DMF was effective as monotherapy, and the patients who became positive to SARS-CoV-2 developed a mild form that did not require discontinuing the drug [63]. This protective effect could probably be related to the antioxidative and anti-inflammatory properties of DMF, which could counteract the cytokine storm caused by severe SARS-CoV-2 infections [64]. These data suggest that continuing DMF might be safe in patients with mild forms of SARS-CoV-2 infection and a normal lymphocyte count [65].

6.3. Cancer

Clinical and pre-clinical investigations have demonstrated that DMF exerts anti-tumorigenic properties in several types of cancer [66]. DMF was found to inhibit the proliferation of melanoma cells and tumor progression in mouse models, due to the inhibition of pro-metastatic matrix metalloproteinases and its proapoptotic action [67]. The suppression of the NF- κ B pathway, which is abnormally activated in neoplastic cells and promotes their survival, migration, invasion, and resistance to chemotherapy, appears to be a crucial property of DMF in counteracting various types of cancer, including breast cancer and cutaneous T-cell lymphoma (CTCL) [22]. Notably, DMF has proved promising results and a good tolerability in CTCL patients in a phase II clinical trial (NCT02546440) [68]. DMF has also demonstrated cytotoxic effects towards other cancer cell lines, such as colorectal cancer [69], cervical cancer [70], and adenocarcinoma of the

lung and pancreas [71]. This evidence suggests that DMF can be safely utilized in patients with a history of oncological disease.

6.4. Cardiovascular Diseases

The antioxidant and anti-inflammatory effects of DMF might play a role in the treatment and prevention of multiple cardiovascular diseases. DMF has conferred anti-atherosclerotic effects in patients with MS and psoriasis by improving the lipidic serum profile [72,73] and by elevating the levels of adiponectin [74]. Along with the hypolipidemic action, DMF seems to reduce the lipidic peroxidation in the arteries [75]. Whilst other studies suggest that FAE might be beneficial for patients with stroke, myocardial ischemia, hypertension, and aneurysms, more research is warranted to establish DMF's efficacy for these indications [76].

6.5. Solid Lipid Nanoparticles for DMF Administration

DMF has been demonstrated to be useful in RRMS treatment. Nevertheless, since DMF administration can be associated with flushing, GI events, and other more serious drawbacks, Esposito E. et al. [77] investigated an alternative method of DMF administration, a nanoparticle-based system, to minimize side effects. The authors described the preparation and characterization of DMF-containing solid lipid nanoparticles (SLN). Namely, SLN based on tristearin, tristearin SLN treated with polysorbate 80, and cationic SLN constituting tristearin in a mixture with dimethyldioctadecylammonium chloride were investigated. The effects of the presence of DMF and functionalization by polysorbate 80 and dimethyldioctadecylammonium chloride were studied on the morphology and dimensional distribution of SLN by photon correlation spectroscopy and cryogenic transmission electron microscopy. DMF released from SLN, studied by Franz cell, evidenced a Fickian dissolutive-type kinetic in the case of SLN treated with polysorbate 80. The presented data indicate that DMF can be conveniently and efficiently encapsulated in SLN with dimensional and morphological properties well suitable for clinical applications requiring different administration routes. Microencapsulation of DMF in SLN could allow a significant improvement in drug absorption and the avoidance of first-pass effects. The results reported by the authors represent a promising starting point for the further development of an SLN-based DMF formulation for the treatment of MS.

7. Conclusions and Expert Opinion

DMF is a drug with a long history, which has proved to have pleiotropic pharmacological effects that could make it useful in multiple conditions including inflammatory, degenerative, neoplastic, and cardiovascular diseases. Since it is not a classical immunosuppressant, it is of particular interest in the current time of the SARS-CoV-2 pandemic. The lack of major pharmacokinetic interactions makes it an interesting drug in elderly and multi-comorbid patients. Moreover, the short half-life and recent observations on pregnancy suggest that it could be an interesting, safe option in fertile women. It has probably been underexploited in dermatology, especially in some countries such as Italy, due to concerns about its potentially serious adverse effects and a lack of familiarity with the dosage of the drug. However, good knowledge of its pharmacological features, slow titration of the dosage, and cautious monitoring of patients during therapy can maximize its therapeutic window. In a clinical setting dominated by increasingly expensive drugs, DMF could represent an inexpensive and effective option. Further understanding of its pharmacodynamics and large randomized trials in the future could better define its clinical indications and safety profile.

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Abbreviations

DMF, dimethyl fumarate; MMF, monomethyl fumarate; FAE, fumaric acid ester; GSH, glutathione; Nrf2, nuclear factor (erythroid-derived 2)-related factor 2; ARE, antioxidant response element; CPP, chronic plaque psoriasis; MS, multiple sclerosis; RRMS, relapsing remitting multiple sclerosis; KEAP1, Kelch-like ECH-associated protein 1; HCAR2, hydroxycarboxylic acid receptor 2; ROS, reactive oxygen species; GI, gastrointestinal; FS, Fanconi syndrome; PML, progressive multiple leukoencephalopathy; CBC, complete blood count; ALC, absolute lymphocyte count; MRI, magnetic resonance imaging; IFN, interferon; TNF, tumor necrosis factor; NF- κ B, nuclear factor kappa-light-enhancer of activated B cells; GSK-3 β , glycogen synthase kinase-3 β .

References

- Schweckendiek, W. Treatment of psoriasis vulgaris. *Med. Monatsschr.* **1959**, *13*, 103–104. [PubMed]
- Landeck, L.; Asadullah, K.; Amasuno, A.; Pau-Charles, I.; Mrowietz, U. Dimethyl fumarate (DMF) vs. monoethyl fumarate (MEF) salts for the treatment of plaque psoriasis: A review of clinical data. *Arch. Dermatol. Res.* **2018**, *310*, 475–483. [CrossRef] [PubMed]
- Werdenberg, D.; Joshi, R.; Wolfram, S.; Merkle, H.; Langguth, P. Presystemic metabolism and intestinal absorption of antipsoriatic fumaric acid esters. *Biopharm. Drug Dispos.* **2003**, *24*, 259–273. [CrossRef] [PubMed]
- Litjens, N.; Burggraaf, J.; Van Strijen, E.; Van Gulpen, C.; Mattie, H.; Schoemaker, R.C.; Van Dissel, J.T.; Thio, H.B.; Nibbering, P.H. Pharmacokinetics of oral fumarates in healthy subjects. *Br. J. Clin. Pharmacol.* **2004**, *58*, 429–432. [CrossRef]
- Linker, R.A.; Haghikia, A. Dimethyl fumarate in multiple sclerosis: Latest developments, evidence and place in therapy. *Ther. Adv. Chronic. Dis.* **2016**, *7*, 198–207. [CrossRef]
- Skilarence Summary of Products Characteristics. Almirall. May 2018. Available online: https://www.ema.europa.eu/en/documents/product-information/skilarence-epar-product-information_en.pdf (accessed on 29 November 2022).
- Zhao, H.; Eguchi, S.; Alam, A.; Ma, D. The role of nuclear factor-erythroid 2 related factor 2 (Nrf-2) in the protection against lung injury. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2017**, *312*, L155–L162. [CrossRef]
- Ruggieri, S.; Tortorella, C.; Gasperini, C. Pharmacology and clinical efficacy of dimethyl fumarate (BG-12) for treatment of relapsing-remitting multiple sclerosis. *Ther. Clin. Risk Manag.* **2014**, *10*, 229–239.
- Kobayashi, A.; Kang, M.-I.; Okawa, H.; Ohtsui, M.; Zenke, Y.; Chiba, T.; Igarashi, K.; Yamamoto, M. Oxidative stress sensor Keap1 functions as an adaptor for Cul3-based E3 ligase to regulate proteasomal degradation of Nrf2. *Mol. Cell. Biol.* **2004**, *24*, 7130–7139. [CrossRef]
- Itoh, K.; Chiba, T.; Takahashi, S.; Ishii, T.; Igarashi, K.; Katoh, Y.; Oyake, T.; Hayashi, N.; Satoh, K.; Hatayama, I.; et al. An Nrf2/small Maf heterodimer mediates the induction of phase II detoxifying enzyme genes through antioxidant response elements. *Biochem. Biophys. Res. Commun.* **1997**, *236*, 313–322. [CrossRef]
- Di Nuzzo, L.; Orlando, R.; Nascia, C.; Nicoletti, F. Molecular pharmacodynamics of new oral drugs used in the treatment of multiple sclerosis. *Drug Des. Devel. Ther.* **2014**, *8*, 555–568.
- Brennan, M.S.; Matos, M.F.; Li, B.; Hronowski, X.; Gao, B.; Juhasz, P.; Rhodes, K.J.; Scannevin, R.H. Dimethyl fumarate and monoethyl fumarate exhibit differential effects on KEAP1, NRF2 activation, and glutathione depletion in vitro. *PLoS ONE* **2015**, *10*, e0120254. [CrossRef] [PubMed]
- Cuadrado, A.; Kuügler, S.; Lastres-Becker, I. Pharmacological targeting of GSK-3 and NRF2 provides neuroprotection in a preclinical model of tauopathy. *Redox Biol.* **2018**, *14*, 522–534. [CrossRef] [PubMed]
- Piroli, G.G.; Manuel, A.M.; Patel, T.; Walla, M.D.; Shi, L.; Lanci, S.A.; Wang, J.; Galloway, A.; Ortinski, P.I.; Smith, D.S.; et al. Identification of Novel Protein Targets of Dimethyl Fumarate Modification in Neurons and Astrocytes Reveals Actions Independent of Nrf2 Stabilization. *Mol. Cell. Proteom.* **2019**, *18*, 504–519. [CrossRef] [PubMed]
- Rosito, M.; Testi, C.; Parisi, G.; Cortese, B.; Baiocco, P.; Di Angelantonio, S. Exploring the Use of Dimethyl Fumarate as Microglia Modulator for Neurodegenerative Diseases Treatment. *Antioxidants* **2020**, *9*, 700. [CrossRef] [PubMed]
- Cuadrado, A.; Martín-Moldes, Z.; Ye, J.; Lastres-Becker, I. Transcription factors NRF2 and NF- κ B are coordinated effectors of the Rho family, GTP-binding protein RAC1 during inflammation. *J. Biol. Chem.* **2014**, *289*, 15244–15258. [CrossRef] [PubMed]
- Parodi, B.; Rossi, S.; Morando, S.; Cordanò, C.; Bragoni, A.; Motta, C.; Usai, C.; Wipke, B.T.; Scannevin, R.H.; Mancardi, G.L.; et al. Fumarates modulate microglia activation through a novel HCAR2 signaling pathway and rescue synaptic dysregulation in inflamed CNS. *Acta Neuropathol.* **2015**, *130*, 279–295. [CrossRef]

18. Chen, H.; Assmann, J.C.; Krenz, A.; Rahman, M.; Grimm, M.; Karsten, C.M.; Kohl, J.; Offermanns, S.; Wettschureck, N.; Schwaninger, M. Hydroxycarboxylic acid receptor 2 mediates dimethyl fumarate's protective effect in EAE. *J. Clin. Investig.* **2014**, *124*, 2188–2192. [[CrossRef](#)]
19. Hanson, J.; Gille, A.; Offermanns, S. Role of HCA₂ (GPR109A) in nicotinic acid and fumaric acid ester-induced effects on the skin. *Pharmacol. Ther.* **2012**, *136*, 1–7. [[CrossRef](#)]
20. Hoffmann, C.; Dietrich, M.; Herrmann, A.K.; Schacht, T.; Albrecht, P.; Methner, A. Dimethyl Fumarate Induces Glutathione Recycling by Upregulation of Glutathione Reductase. *Oxidative Med. Cell. Longev.* **2017**, *2017*, 6093903. [[CrossRef](#)]
21. Gillard, G.O.; Collette, B.; Anderson, J.; Chao, J.; Scannevin, R.H.; Huss, D.J.; Fontenot, J.D. DMF, but not other fumarates, inhibits NF- κ B activity in vitro in an Nrf2-independent manner. *J. Neuroimmunol.* **2015**, *283*, 74–85. [[CrossRef](#)]
22. Kastrati, I.; Siklos, M.I.; Calderon-Gierszal, E.L.; El-Shennawy, L.; Georgieva, G.; Thayer, E.N.; Thatcher, G.R.; Frasor, J. Dimethyl Fumarate Inhibits the Nuclear Factor κ B Pathway in Breast Cancer Cells by Covalent Modification of p65 Protein. *J. Biol. Chem.* **2016**, *291*, 3639–3647. [[CrossRef](#)] [[PubMed](#)]
23. Treumer, F.; Zhu, K.; Gläser, R.; Mrowietz, U. Dimethylfumarate is a potent inducer of apoptosis in human T cells. *J. Investig. Dermatol.* **2003**, *121*, 1383–1388. [[CrossRef](#)] [[PubMed](#)]
24. De Jong, R.; Bezemer, A.C.; Zomerdijk, T.P.; Pouw-Kraan, T.; Ottenhoff, T.H.; Nibbering, P.H. Selective stimulation of T helper 2 cytokine responses by the anti-psoriasis agent monomethylfumarate. *Eur. J. Immunol.* **1996**, *26*, 2067–2074. [[CrossRef](#)] [[PubMed](#)]
25. Vandermeeren, M.; Janssens, S.; Borgers, M.; Geysen, J. Dimethylfumarate is an inhibitor of cytokine-induced E-selectin, VCAM-1, and ICAM-1 expression in human endothelial cells. *Biochem. Biophys. Res. Commun.* **1997**, *234*, 19–23. [[CrossRef](#)] [[PubMed](#)]
26. Foster, C.A.; Dreyfuss, M.; Mandak, B.; Meingassner, J.G.; Naegeli, H.U.; Nussbaumer, A.; Oberer, L.; Scheel, G.; Swoboda, E.M. Pharmacological modulation of endothelial cell-associated adhesion molecule expression: Implications for future treatment of dermatological diseases. *J. Dermatol.* **1994**, *21*, 847–854. [[CrossRef](#)]
27. Connor, J.R.; Menzies, S.L. Relationship of iron to oligodendrocytes and myelination. *Glia* **1996**, *17*, 83–93. [[CrossRef](#)]
28. Hallgren, B.; Sourander, P. The effect of age on the non-haemin iron in the human brain. *J. Neurochem.* **1958**, *3*, 41–51. [[CrossRef](#)]
29. Todorich, B.; Zhang, X.; Connor, J.R. H-ferritin is the major source of iron for oligodendrocytes. *Glia* **2011**, *59*, 927–935. [[CrossRef](#)]
30. Craelius, W.; Migdal, M.W.; Luessenhop, C.P.; Sugar, A.; Mihalakis, I. Iron deposits surrounding multiple sclerosis plaques. *Arch. Pathol. Lab. Med.* **1982**, *106*, 397–399.
31. Haider, L.; Simeonidou, C.; Steinberger, G.; Hametner, S.; Grigoriadis, N.; Deretzi, G.; Kovacs, G.G.; Kutzelnigg, A.; Lassmann, H.; Frischer, J.M. Multiple sclerosis deep grey matter: The relation between demyelination, neurodegeneration, inflammation and iron. *J. Neurol. Neurosurg. Psychiatry* **2014**, *85*, 1386–1395. [[CrossRef](#)]
32. Kronenberg, J.; Pars, K.; Brieskorn, M.; Prajeeth, C.K.; Heckers, S.; Schwenkenbecher, P.; Skripuletz, T.; Pul, R.; Pavlou, A.; Stangel, M. Fumaric Acids Directly Influence Gene Expression of Neuroprotective Factors in Rodent Microglia. *Int. J. Mol. Sci.* **2019**, *20*, 325. [[CrossRef](#)] [[PubMed](#)]
33. Barnes, R.H.; Karatzas, K.A.G. Investigation into the antimicrobial activity of fumarate against *Listeria monocytogenes* and its mode of action under acidic conditions. *Int. J. Food Microbiol.* **2020**, *324*, 108614. [[CrossRef](#)] [[PubMed](#)]
34. Wang, H.H.; Sun, D.W.; Kuang, R. Inhibition of *Escherichia coli* by dimethyl fumarate. *Int. J. Food. Microbiol.* **2001**, *65*, 125–130. [[CrossRef](#)]
35. Rumah, K.R.; Vartanian, T.K.; Fischetti, V.A. Oral Multiple Sclerosis Drugs Inhibit the in vitro Growth of Epsilon Toxin Producing Gut Bacterium, *Clostridium perfringens*. *Front. Cell. Infect. Microbiol.* **2017**, *7*, 11. [[CrossRef](#)] [[PubMed](#)]
36. Jones, S.A.; Gibson, T.; Maltby, R.; Chowdhury, F.Z.; Stewart, V.; Cohen, P.S.; Conway, T. Anaerobic respiration of *Escherichia coli* in the mouse intestine. *Infect. Immun.* **2011**, *79*, 4218–4226. [[CrossRef](#)]
37. Storm-Larsen, C.; Myhr, K.M.; Farbu, E.; Midgard, R.; Nyquist, K.; Broch, L.; Berg-Hansen, P.; Bunes, A.; Holm, K.; Ueland, T.; et al. Gut microbiota composition during a 12-week intervention with delayed-release dimethyl fumarate in multiple sclerosis—A pilot trial. *Mult. Scler. J. Exp. Transl. Clin.* **2019**, *5*, 2055217319888767. [[CrossRef](#)]
38. Phillips, J.T.; Erwin, A.A.; Agrella, S.; Kremenchtzky, M.; Kramer, J.F.; Darkes, M.J.; Kendter, J.; Abourjaily, H.; Rana, J.; Fox, R.J. Consensus Management of Gastrointestinal Events Associated with Delayed-Release Dimethyl Fumarate: A Delphi Study. *Neurol. Ther.* **2015**, *4*, 137–146. [[CrossRef](#)]
39. Makhani, N.; Schreiner, T. Oral dimethyl fumarate in children with multiple sclerosis: A dual-center study. *Pediatr. Neurol.* **2016**, *57*, 101–104. [[CrossRef](#)]
40. Lucchini, M.; Prosperini, L.; Buscarinu, M.C.; Centonze, D.; Conte, A.; Cortese, A.; Elia, G.; Fantozzi, R.; Ferraro, E.; Gasperini, C.; et al. Predictors of lymphocyte count recovery after dimethyl fumarate-induced lymphopenia in people with multiple sclerosis. *J. Neurol.* **2021**, *268*, 2238–2245. [[CrossRef](#)]
41. Mills, E.A.; Ogronnik, M.A.; Plave, A.; Mao-Draayer, Y. Emerging Understanding of the Mechanism of Action for Dimethyl Fumarate in the Treatment of Multiple Sclerosis. *Front. Neurol.* **2018**, *9*, 5. [[CrossRef](#)]
42. Nieuwkamp, D.J.; Murk, J.L.; van Oosten, B.W.; Cremers, C.H.; Killestein, J.; Viveen, M.C.; Van Hecke, W.; Frijlink, D.W.; Wattjes, M.P. PML in Dutch MS Patients Consortium. PML in a patient without severe lymphocytopenia receiving dimethyl fumarate. *N. Engl. J. Med.* **2015**, *372*, 1474–1476. [[CrossRef](#)] [[PubMed](#)]
43. Shackelton, L.A.; Rambaut, A.; Pybus, O.G.; Holmes, E.C. JC Virus evolution and its association with human populations. *J. Virol.* **2006**, *80*, 9928–9933. [[CrossRef](#)]

44. Jordan, A.L.; Yang, J.; Fisher, C.J.; Racke, M.K.; Mao-Draayer, Y. Progressive multifocal leukoencephalopathy in dimethyl fumarate-treated multiple sclerosis patients. *Mult. Scler.* **2022**, *28*, 7–15. [[CrossRef](#)]
45. Bharat, A.; Xie, F.; Baddley, J.W.; Beukelman, T.; Chen, L.; Calabrese, L.; Curtis, J.R. Incidence and risk factors for progressive multifocal leukoencephalopathy among patients with selected rheumatic diseases. *Arthritis. Care Res.* **2012**, *64*, 612–615. [[CrossRef](#)] [[PubMed](#)]
46. Vola, E.A.; Petracca, M.; Cocozza, S.; De Angelis, M.; Carotenuto, A.; Pontillo, G.; Morra, V.B.; Tedeschi, E.; Lanzillo, R. Possible progressive multifocal leukoencephalopathy and active multiple sclerosis under dimethyl fumarate: The central role of MRI in informing therapeutic decisions. *BMC Neurol.* **2021**, *21*, 146. [[CrossRef](#)] [[PubMed](#)]
47. Wan, E.R.; Siew, K.; Heptinstall, L.; Walsh, S.B. Fumaric acid ester-induced renal Fanconi syndrome: Evidence of mitochondrial toxicity. *Clin. Kidney J.* **2021**, *14*, 2085–2089. [[CrossRef](#)]
48. O’Gorman, J.; Russell, H.K.; Li, J.; Phillips, G.; Kurukulasuriya, N.C.; Viglietta, V. Effect of Aspirin Pretreatment or Slow Dose Titration on Flushing and Gastrointestinal Events in Healthy Volunteers Receiving Delayed-release Dimethyl Fumarate. *Clin. Ther.* **2015**, *37*, 1402–1419.e5. [[CrossRef](#)]
49. Corazza, M.; Odorici, G.; Conti, A.; Di Lernia, V.; Motolese, A.; Bardazzi, F.; Di Nuzzo, S.; Monti, A.; Arginelli, F.; Filippi, F.; et al. Dimethyl fumarate treatment for psoriasis in a real-life setting: A multicentric retrospective study. *Dermatol. Ther.* **2021**, *34*, e15066. [[CrossRef](#)]
50. Mrowietz, U.; Van De Kerkhof, P.; Schoenenberger, A.; Ryzhkova, A.; Pau-Charles, I.; Llamas-Velasco, M.; Daudén, E.; Carrascosa, J.M.; Cueva, P.; Salgado-Boquete, L.; et al. Efficacy of dimethyl fumarate treatment for moderate-to-severe plaque psoriasis: Presentation extracts from the 29th EADV virtual congress, 29–31 October 2020. *Expert. Rev. Clin. Immunol.* **2021**, *17*, 1–11. [[CrossRef](#)]
51. Bardazzi, F.; Magnano, M.; Campanati, A.; Loconsole, F.; Carpentieri, A.; Potenza, C.; Bernardini, N.; Di Lernia, V.; Carrera, C.; Raone, B.; et al. Biologic Therapies in HIV-infected Patients with Psoriasis: An Italian Experience. *Acta Derm. Venereol.* **2017**, *97*, 989–990. [[CrossRef](#)]
52. Gisondi, P.; Cazzaniga, S.; Chimenti, S.; Maccarone, M.; Picardo, M.; Girolomoni, G.; Naldi, L.; Psocare Study Group. Latent tuberculosis infection in patients with chronic plaque psoriasis: Evidence from the Italian Psocare Registry. *Br. J. Dermatol.* **2015**, *172*, 1613–1620, Erratum in: *Br. J. Dermatol.* **2017**, *176*, 1415–1416.. [[CrossRef](#)]
53. Naval, S.M.; Casas, L.R.; Spackman, E. Cost effectiveness of dimethyl fumarate (DMF) in the treatment of moderate to severe plaque psoriasis: A uk perspective. *Value Health* **2018**, *21*, S1eS481.
54. Schimrigk, S.; Brune, N.; Hellwig, K.; Lukas, C.; Bellenberg, B.; Rieks, M.; Hoffmann, V.; Pohlau, D.; Przuntek, H. Oral fumaric acid esters for the treatment of active multiple sclerosis: An open-label, baseline-controlled pilot study. *Eur. J. Neurol.* **2006**, *13*, 604–610. [[CrossRef](#)] [[PubMed](#)]
55. Kappos, L.; Gold, R.; Miller, D.H.; MacManus, D.G.; Havrdova, E.; Limmroth, V.; Polman, C.H.; Schmierer, K.; Yousry, T.A.; Yang, M.; et al. Efficacy and safety of oral fumarate in patients with relapsing-remitting multiple sclerosis: A multicentre, randomised, double-blind, placebo-controlled phase IIb study. *Lancet* **2008**, *372*, 1463–1472. [[CrossRef](#)] [[PubMed](#)]
56. Fernández, Ó.; Giovannoni, G.; Fox, R.J.; Gold, R.; Phillips, J.T.; Potts, J.; Okwuokenye, M.; Marantz, J.L. Efficacy and Safety of Delayed-release Dimethyl Fumarate for Relapsing-remitting Multiple Sclerosis in Prior Interferon Users: An Integrated Analysis of DEFINE and CONFIRM. *Clin. Ther.* **2017**, *39*, 1671–1679. [[CrossRef](#)] [[PubMed](#)]
57. Phillips, J.T.; Selmaj, K.; Gold, R.; Fox, R.J.; Havrdova, E.; Giovannoni, G.; Abourjaily, H.; Pace, A.; Novas, M.; Hotermans, C.; et al. Clinical Significance of Gastrointestinal and Flushing Events in Patients with Multiple Sclerosis Treated with Delayed-Release Dimethyl Fumarate. *Int. J. MS Care* **2015**, *17*, 236–243. [[CrossRef](#)]
58. Hellwig, K.; Rog, D.; McGuigan, C.; Houtchens, M.K.; Bruen, D.R.; Mokliatchouk, O.; Branco, F.; Peng, X.; Everage, N.J. Interim Analysis of Pregnancy Outcomes After Exposure to Dimethyl Fumarate in a Prospective International Registry. *Neurol. Neuroimmunol. Neuroinflamm.* **2021**, *9*, e1114. [[CrossRef](#)]
59. *Drugs and Lactation Database (LactMed)*; National Library of Medicine: Bethesda, MD, USA, 2006; Dimethyl Fumarate. [Updated 19 September 2022].
60. Campanati, A.; Brisigotti, V.; Diotallevi, F.; D’Agostino, G.M.; Paolinelli, M.; Radi, G.; Rizzetto, G.; Sapigni, C.; Tagliati, C.; Offidani, A. Active implications for dermatologists in ‘SARS-CoV-2 ERA’: Personal experience and review of literature. *J. Eur. Acad. Dermatol. Venereol.* **2020**, *34*, 1626–1632. [[CrossRef](#)]
61. Olagnier, D.; Farahani, E.; Thyrssted, J.; Blay-Cadanet, J.; Herengt, A.; Idorn, M.; Hait, A.; Hernaez, B.; Knudsen, A.; Iversen, M.B.; et al. SARS-CoV-2-mediated suppression of NRF2-signaling reveals potent antiviral and anti-inflammatory activity of 4-octyl-itaconate and dimethyl fumarate. *Nat. Commun.* **2020**, *11*, 4938. [[CrossRef](#)]
62. Mantero, V.; Abate, L.; Basilico, P.; Balgera, R.; Salmaggi, A.; Nourbakhsh, B.; Cordano, C. COVID-19 in dimethyl fumarate-treated patients with multiple sclerosis. *J. Neurol.* **2020**, *25*, 1–3. [[CrossRef](#)]
63. Esposito, M.; Campanati, A.; Giunta, A.; Calianno, G.; Bianchi, L.; Diotallevi, F.; Offidani, A.M.; Fargnoli, M.C. Dimethyl Fumarate’s Effectiveness and Safety in Psoriasis: A Real-Life Experience During the COVID-19 Pandemic. *Dermatol. Ther.* **2022**, *12*, 671–681. [[CrossRef](#)]
64. Timpani, C.A.; Rybalka, E. Calming the (cytokine) storm: Dimethyl fumarate as a therapeutic candidate for COVID-19. *Pharmaceutics* **2020**, *14*, 15. [[CrossRef](#)] [[PubMed](#)]

65. Capone, F.; Ferraro, E.; Motolese, F.; Di Lazzaro, V. COVID-19 in multiple sclerosis patients treated with dimethyl fumarate. *J. Neurol.* **2021**, *268*, 3132–3134. [[CrossRef](#)]
66. Kourakis, S.; Timpani, C.A.; de Haan, J.B.; Gueven, N.; Fischer, D.; Rybalka, E. Dimethyl Fumarate and Its Esters: A Drug with Broad Clinical Utility? *Pharmaceutics* **2020**, *13*, 306. [[CrossRef](#)]
67. Loewe, R.; Valero, T.; Kremling, S.; Pratscher, B.; Kunstfeld, R.; Pehamberger, H.; Petzelbauer, P. Dimethylfumarate impairs melanoma growth and metastasis. *Cancer Res.* **2006**, *66*, 11888–11896. [[CrossRef](#)] [[PubMed](#)]
68. Nicolay, J.P.; Albrecht, J.D.; Assaf, C.; Dippel, E.; Stadler, R.; Wehkamp, U.; Wobser, M.; Guelow, K.; Goerdt, S.; Krammer, P.H. Dimethyl fumarate (DMF) therapy in CTCL: Results from a clinical phase II study. *Eur. J. Cancer* **2021**, *156*, S21–S22. [[CrossRef](#)] [[PubMed](#)]
69. Kaluzki, I.; Hailemariam-Jahn, T.; Doll, M.; Kaufmann, R.; Balermipas, P.; Zoöller, N.; Kippenberger, S.; Meissner, M.; Jahn, H. Doll Dimethylfumarate Inhibits Colorectal Carcinoma Cell Proliferation: Evidence for Cell Cycle Arrest, Apoptosis and Autophagy. *Cells* **2019**, *8*, 1329. [[CrossRef](#)]
70. Han, G.; Zhou, Q. Dimethylfumarate induces cell cycle arrest and apoptosis via regulating intracellular redox systems in HeLa cells. *Vitr. Cell. Dev. Biol. Anim.* **2016**, *52*, 1034–1041. [[CrossRef](#)]
71. Saidu, N.E.B.; Noeé, G.; Cerles, O.; Cabel, L.; Kavian-Tessler, N.; Chouzenoux, S. Dimethyl Fumarate Controls the NRF2/DJ-1 Axis in Cancer Cells: Therapeutic Applications. *Mol. Cancer Ther.* **2017**, *16*, 529. [[CrossRef](#)]
72. Schmieder, A.; Poppe, M.; Hametner, C.; Meyer-Schraml, H.; Schaarschmidt, M.-L.; Findeisen, P.; Benoit, S.; Bauer, B.; Schmid, S.; Goebeler, M.; et al. Impact of fumaric acid esters on cardiovascular risk factors and depression in psoriasis: A prospective pilot study. *Arch. Dermatol. Res.* **2015**, *307*, 413–424. [[CrossRef](#)]
73. Kan, S.B.; Staun-Ram, E.; Golan, D.; Miller, A. HDL-cholesterol elevation associated with fingolimod and dimethyl fumarate therapies in multiple sclerosis. *Mult. Scler. J. Exp. Transl. Clin.* **2019**, *5*, 2055217319882720.
74. Holzer, G.; Hoke, M.; Sabeti-Sandor, S.; Perkmann, T.; Rauscher, A.; Strassegger, B.; Radakovic, S.; Tanew, A. Disparate effects of adalimumab and fumaric acid esters on cardiovascular risk factors in psoriasis patients: Results from a prospective, randomized, observer-blinded head-to-head trial. *J. Eur. Acad. Dermatol. Venereol.* **2021**, *35*, 441–449. [[CrossRef](#)] [[PubMed](#)]
75. Nour, O.A.; Shehatou, G.S.; Rahim, M.A.; El-Awady, M.S.; Suddek, G.M. Antioxidant and anti-inflammatory effects of dimethyl fumarate in hypercholesterolemic rabbits. *Egypt. J. Basic. Appl. Sci.* **2017**, *4*, 153–159. [[CrossRef](#)]
76. Thomas, S.D.; Jha, N.K.; Sadek, B.; Ojha, S. Repurposing Dimethyl Fumarate for Cardiovascular Diseases: Pharmacological Effects, Molecular Mechanisms, and Therapeutic Promise. *Pharmaceutics* **2022**, *15*, 497. [[CrossRef](#)]
77. Esposito, E.; Cortesi, R.; Drechsler, M.; Fan, J.; Fu, B.M.; Calderan, L.; Mannucci, S.; Boschi, F.; Nastruzzi, C. Nanoformulations for dimethyl fumarate: Physicochemical characterization and in vitro/in vivo behavior. *Eur. J. Pharm. Biopharm.* **2017**, *115*, 285–296. [[CrossRef](#)]