

Supplementary Materials: Oligoarginine Peptide Conjugated to BSA Improves Cell Penetration of Gold Nanorods and Nanoprisms for Biomedical Applications

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1. BSA functionalization by r_8 and AuNPs coating by BSA- r_8

For this study, a D-octaarginine peptide (r_8 , Figure S1), with a Br terminal was synthesized and used to functionalize the BSA protein.

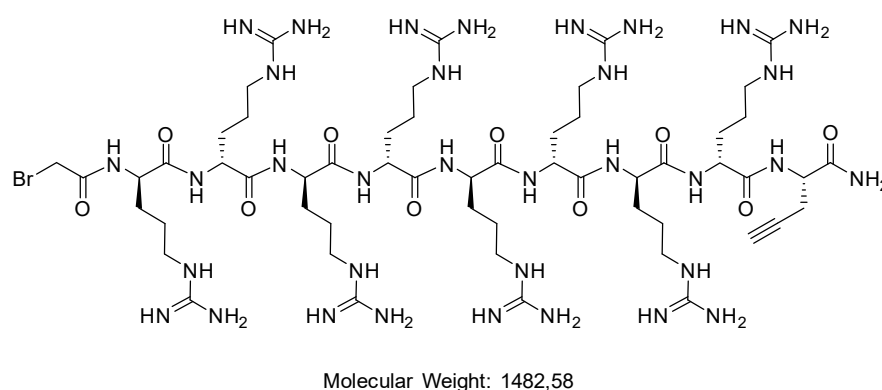


Figure S1. Structure of the Br-CH₂- r_8 peptide.

The peptide was obtained with high purity (> 95%), and a characteristic mass spectrum, as shown in Figure S2.

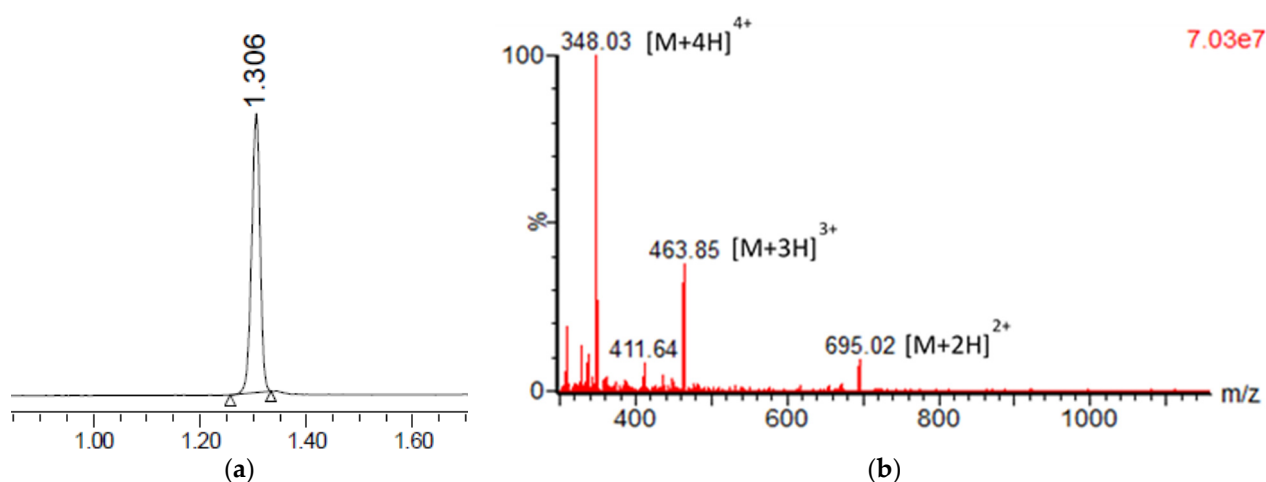


Figure S2. UPLC trace (a) and MS spectra (b) of Br- r_8 (BrCH₂CO- r_8). UPLC chromatogram was recorded at 220 nm in a 2 min linear gradient from 0 to 30% of MeCN (0.036% TFA) in H₂O (0.045% TFA). M_{cal} : 1387.48; M_{found} : 695.02 [M+2H]²⁺; 463.85 [M+3H]³⁺; 348.03 85 [M+4H]⁴⁺.

2. BSA functionalization by A565 and r_8

The r_8 was synthesized with a Br terminal and BSA was functionalized following a 2-Iminothiolane strategy on the Br terminal of the peptide. Uniprot (P02769, Bos Taurus) for BSA sequence was consulted to know the theoretical number of Arg in a BSA mole-

cule, resulting in 26 Arg/BSA molecule. For amino acid analysis, 1.5×10^{-3} mM of BSA were added, equivalent to 3.9×10^{-2} mM of arginine. Therefore, the arginine excess was calculated subtracting the quantified amount of Arg (Table S1) from the theoretical amount of Arg in the sample ($0.07 \text{ mM} - 0.039 \text{ mM} = 0.031 \text{ mM}$). The arginine excess is translated to 3.64×10^{-3} mM of r_8 , as follows:

$$r_8 \text{ amount} = \frac{\text{Arginine excess}}{\frac{\text{molar mass of arginine}}{\text{molar mass of } r_8}} = \frac{0.031 \text{ mM}}{\left(\frac{1482.6 \text{ g/mol}}{174.2 \text{ g/mol}} \right)} = 3.64 \times 10^{-3} \text{ mM } r_8$$

The ratio r_8 /BSA was calculated as 2.4 ($3.64 \times 10^{-3} \text{ mM } r_8 / 1.5 \times 10^{-3} \text{ mM BSA}$). The BSA- r_8 was subsequently labeled by an Atto-565 NHS-ester red fluorophore with a DOL of 0.66 mol Atto/mol BSA for the detection on fluorescence techniques, as previously described.

Table S1. Amino acid analysis of BSA- r_8 after acid digestion, using Y-aminobutyric acid as standard (aaba).

Peak Name	RT (min)	Amount (mM)	Number of residues in protein
asp	21.326	0.081	54
ser	23.120	0.053	31
glu	23.638	0.134	79
gly	25.041	0.031	17
his	25.328	0.039	17
arg	27.133	0.07	26
thr	27.500	0.072	34
ala	28.118	0.098	48
pro	29.263	0.06	28
aaba	30.504	0.103	0
tyr	32.417	0.023	21
val	33.292	0.067	38
lys	35.959	0.112	60
ile	37.030	0.029	15
leu	37.668	0.121	64
phe	39.453	0.052	30

3. TEM images of AuNRs and AuNPrs

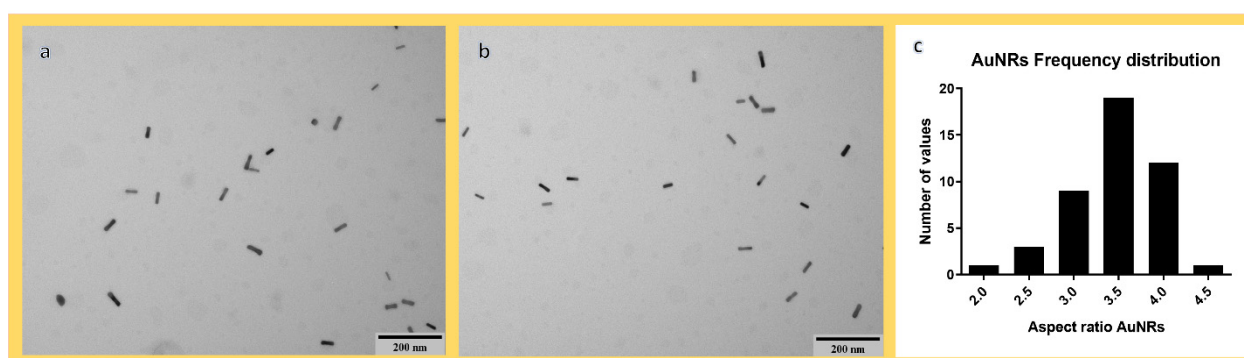


Figure S3. Microscopy characterization of AuNRs: **a**, **b**, representative TEM images, scale 200 nm, **c**, Aspect ratio frequency distribution (length/width), statistics of at least 50 AuNRs. Average population of 95% AuNRs.

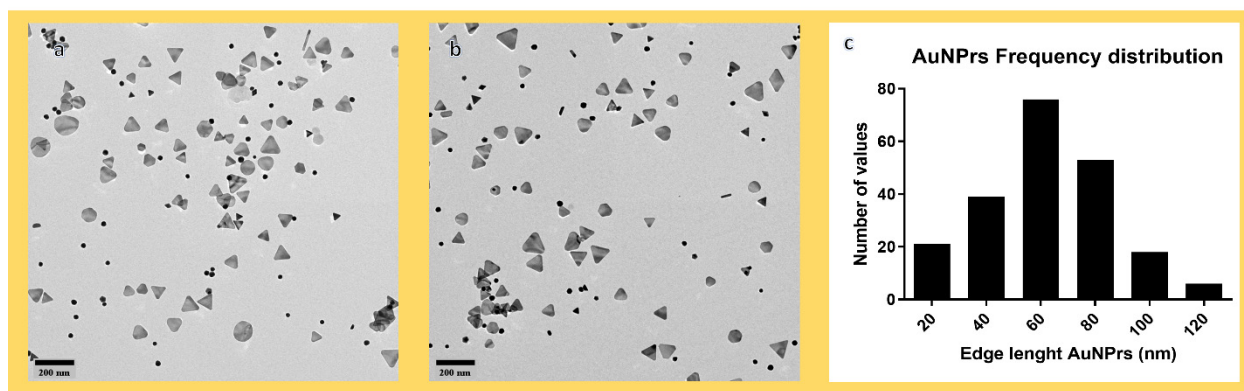


Figure S4. Microscopy characterization of AuNPs: **a, b.** representative TEM images, scale 200 nm, **c.** Edge length (nm) frequency distribution (length/width), statistics of at least 50 AuNPs. Average population of 78% AuNPs.

4. Stability of AuNR-BSA-rs and AuNPr-BSA-rs after 30 days

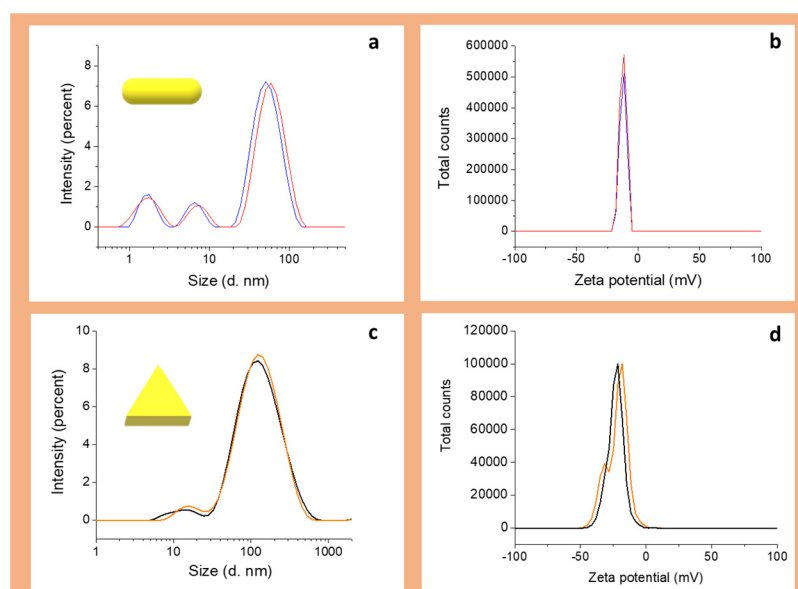


Figure S5. Characterization of BSA-rs capped AuNPs after 24 h, DLS and Zeta potential. **a, b.** AuNR-BSA-rs blue line $t = 0$, red line $t = 30$ days. **c, d.** AuNPr-BSA-rs orange line $t = 0$, black line $t = 30$ days. Acquired in PBS pH = 7.

5. Flow cytometry of HeLa cells after treatment with AuNR-BSA-rs and AuNPr-BSA-rs

HeLa cells (1×10^4 cells/well) were seeded in pretreated 96 wells plates and allowed to attach at 37 °C for 24 h. Subsequently, the medium was removed, and a fresh medium was added. The treatments were added and allowed to incubate for 24 h, 1 nM AuNR-BSA-rs, 1 nM AuNPr-BSA-rs. After this time, the cells were washed, trypsinized and collected in a plastic tube for flow cytometry analysis in a Gallios beckman coulter flow cytometer. HOESCHT was added to the samples (5 μ L/mL) at least 10 min before the analysis. Dead cells are positive for the dye and identified in the upper region of the flow cytometry images.

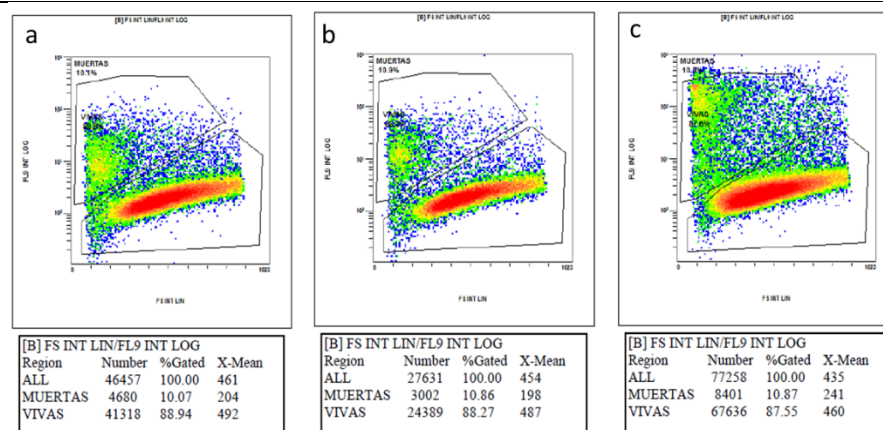


Figure S6. Flow cytometry of HeLa cells incubated by 24 h with: a. water, b. AuNR-BSA-rs 1 nM in AuNRs, c. AuNPr-BSA-rs 1 nM AuNPrs. Apoptotic cells are identified by positive fluorescence in the upper region by DAPI labeling.

For this study, death cells were identified by positive fluorescence in the upper region due to DAPI labeling. As shown in figure S7, no significative effect was observed at 24 h of treatment with 1nM AuNR-BSA-rs, 1 nM AuNPr-BSA-rs in the studied conditions, by comparing the treated cells with the control.

6. Internalization of AuNR-BSA-A565-rs on HeLa cells at 1h

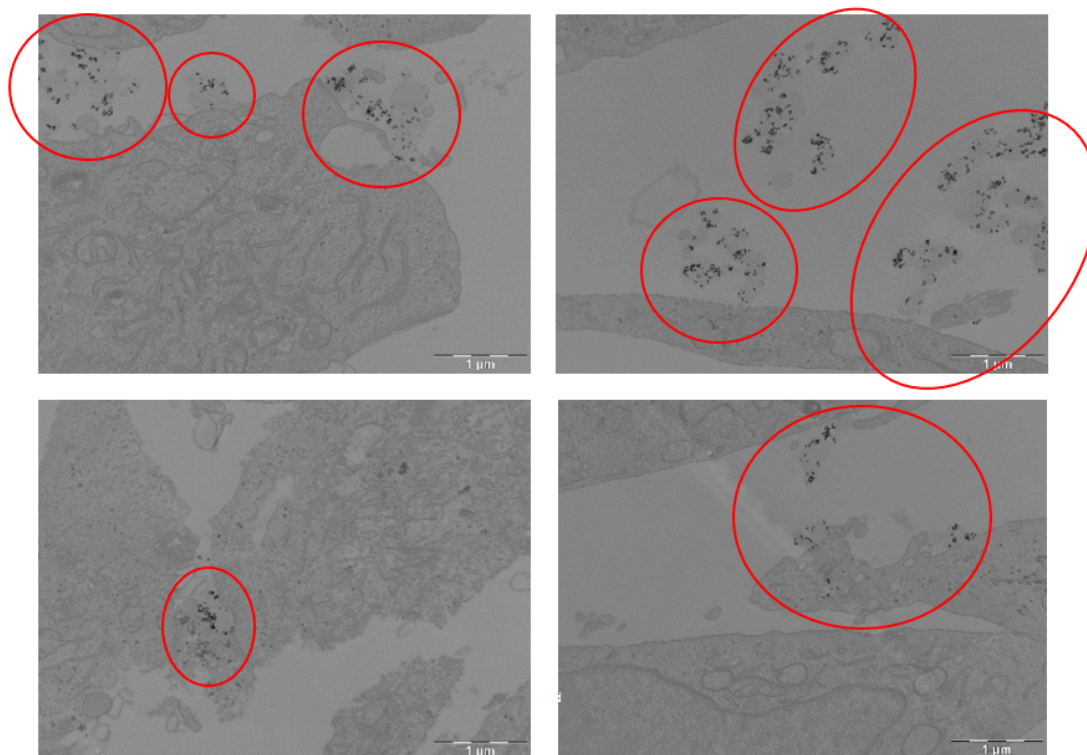


Figure S7. TEM images of HeLa cells after treatment by AuNR-BSA-rs incubated by 1h. Scale 1 μm. Red circles showing the AuNRs.

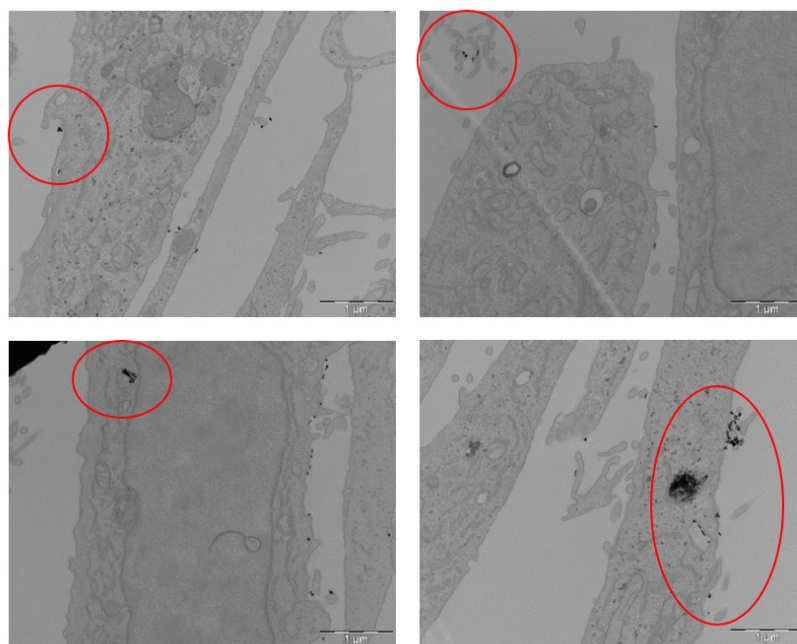


Figure S8. TEM images of HeLa cells after treatment by AuNPr-BSA-rs incubated by 1h. Scale 1 µm. Red circles showing the AuNPrs.