

Supplementary data

Polymorphisms Related to Iron Homeostasis Associate with Liver Disease in Chronic Hepatitis C

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Table S1. Characteristics of patients selected for the study.

Variable	Characteristics
Gender (Male/Female)	153/96
Age (yr)	19 - 78 (50)
Hemoglobin (g/dL)	9.6 – 18.4 (14.9)
ALT (IU/L)	14-852 (79)
AST (IU/L)	3-369 (59.5)
GGT (IU/L)	9-663 (70)
Bilirubin (mg/dL)	0.2-4.2 (0.7)
Iron (μ g/dL)	29-357 (146)
Transferrin saturation (%)	6-100 (40)
Ferritin (ng/mL)	7-3410 (218)
Inflammation grade (0-3)	2 (2/2)
Fibrosis grade (0-4)	2 (1/3)
Iron deposits grade (0-3)	0 (0/1)
Steatosis grade (0-3)	1 (0/2)
HCV genotype (n=131)	120 genotype 1 15 genotype 3 9 genotype 4
HCV RNA (kIU/mL) (n=89)	37-24047 (1390)
Median observation time [years]	6.1 (min-max 1.5-20)
Median follow up time [years]	6.3 (75% CI 2.9-9.3)
HCC yes/no	19/230

Quantitative variables are presented as minimal - maximal values (median). Histopathological data is shown as median values with percentiles (25th/ 75th).

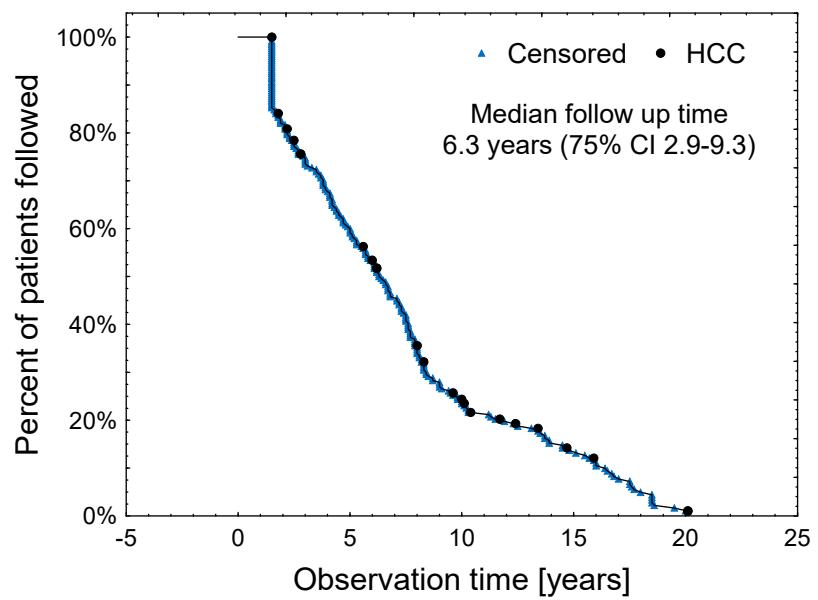


Figure S1. Reverse Kaplan-Meier curve showing patients follow up time.

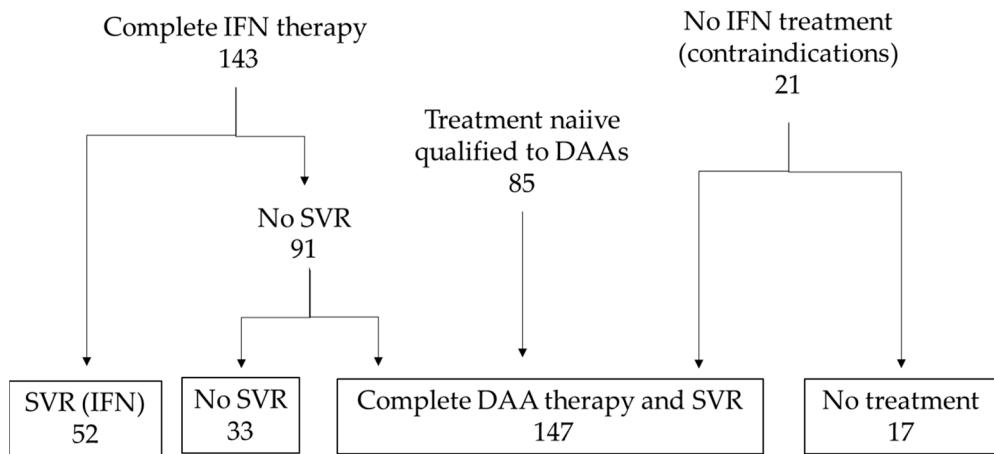


Figure S2. Summary of the therapeutic outcome for patients included in the study.

Table S2. Primers used for quantitative gene expression analysis.

Primer name	Primer sequence 5'-->3'	Reference
GUS_F	CGAGAGTGCTGGGAATAAA	[18]
GUS_R	CCTGGTTTCATTGGCAATCT	
HAMP_F	AGACACCCACTTCCCCATCT	[18]
HAMP_R	CACATCCCACACTTGATCG	
FPN1_F	CAGGGACTGAGTGGTCCAT	[18]
FPN1_R	ACCACATTTCGACGTAGCC	
HJV_F	GGCCTAGGAGACACGTGAAA	[20]
HJV_R	GAGGCTGGAAAAATTGGTGA	
TFR2_F	GACCCCTGCAGTGGGTGTACT	[20]
TFR2_F	CAGTCGCTCGTCTCTCTCCT	
HO-1_F	GAAAAGCACATCCAGGCAAT	[20]
HO-1_R	CTGCTGCAGGAACTGAGGAT	
ID1_F	TGTTCCATTTCCGTATCTGC	[20]
ID1_R	TGAAACAGAACATGGCAAAGC	
BMP6_F	CTTACGACAAGCAGCCCTTC	[20]
BMP6_R	CACGTGCACCTCACTCACTT	
CTLA4_F	CTGTGCGGCAACCTACATGA	this study
CTLA4_R	TGCAGATGTAGAGTCCGTG	
PD-1_F	GCCTGTGTTCTCTGTGGACT	this study
PD-1_R	ATGGTGGCATACTCCGTCTG	
Tim3_F	GGAGCCTGTCCTGTGTTGA	this study
Tim3_R	AGGGACACATCTCCTTTGCG	

Table S3. Genotype distribution within single-nucleotide polymorphisms selected for the study in chronic hepatitis C patients.

N o	Gene	dbSNP ID / mutation	SNP associated with	Consequence type	MAF (TOPMed)	Clinical significance	Ref.	Genotype	No of cases #	Freq.#
1	Hemochromatosis (<i>HFE</i>)	rs1800562 G>A / C282Y	serum transferrin and sFe in healthy individuals	missense variant (different aa sequence), 2KB upstream variant	A=0.053	Hereditary hemochromat osis; complications of diabetes, Alzheimer disease; familial	[6]	GG	233	0.94
								GA	16	0.06
								AA	0	0
2	Hemochromatosis (<i>HFE</i>)	rs1800730 A>T / S65C		missense variant, non coding transcript variant	T=0.009	porphyria cutanea tarda	[6]	AA	241	0.97
								AT	8	0.03
								TT	0	0
3		rs1799945 C>G / H63D			G=0.100		[6]	CC	172	0.7
								CG	68	0.3
								GG	9	0.04
4	Transferrin receptor 2 (<i>TFR2</i>)	rs7385804 A>C	sFe levels in healthy individuals	Intron variant	C=0.334	not reported in ClinVar	[7]	AA	71	0.3
		rs3778216 C>T	hepcidin expression; cellular iron	Intron variant	T=0.218	not reported in ClinVar	[34,36]	AC	128	0.5
								CC	50	0.2
5								CT	132	0.5
6	Histone deacetylase 3 (<i>HDAC3</i>)	rs976552 T>G	metabolism, inflammatory response	2KB Upstream variant	C=0.210	not reported in ClinVar		TT	104	0.4
								TG	13	0.1
								GG	162	0.7
7	Histone deacetylase 5 (<i>HDAC5</i>)	rs368328 A>G		Intron variant	G=0.328	not reported in ClinVar		AA	81	0.3
								AG	6	0.02
								GG	109	0.4
8	Transmembrane serine proteinase 2, matriptase-2 (<i>TMPRSS6</i>)	rs855791 C>T	sFe, TS; blood hepcidin mRNA and protein in urine in healthy individuals	missense variant	A=0.361	microcytic anemia, iron- refractory iron deficiency anemia	[8-9]	CC	29	0.4
								CT	128	0.5
								TT	13	0.1
9	Duodenal cytochrome b (<i>CYBRD1</i>)	rs884409 T>G	serum TS and ferritin concentration	2 KB upstream sequence variant	G=0.218	not reported in ClinVar	[20,40- 41]	TT	163	0.6
								TG	73	0.3
								GG	13	0.04

sFe -serum iron; TS – transferrin saturation; MAF (TOPMed) – minor allele frequency from TOPMed Programme; #this study

Table S4. Construction of logistic regression model for HCC occurrence.

Input variables	p*	Multivariate logistic regression model						
		Model no	Variables in the model		Training set		Cross-validated (10-fold)	
			AUC	AUC error	AUC	AUC error		
AST	<0.000001	1	AST	0.842	0.05	0.808	0.06	
sFe	0.00009	2	AST+sFe	0.850	0.05	0.827	0.05	
Minor <i>HDAC3</i> rs976552/ <i>CYBRD1</i> rs884409	0.0003	3	AST+sFe+ALT	0.883	0.07	0.738	0.11	
Liver inflammation grade	0.0003	4	AST+sFe+ALT+ Minor <i>HDAC3</i> rs976552/ <i>CYBRD1</i> rs884409	0.871	0.05	0.839	0.06	
TS	0.0009							
ALT	0.004							
Age	0.021							
Billirubin	0.023							
GGT	0.039							
sFerritin	0.047							

* p values for monovariate logistic regression analysis. **Model 3** was automatically constructed using backward stepwise regression with all the listed variables significant in the monovariate analysis as an input. **Model 1**, **Model 2** and **Model 4** were evaluated for comparison.

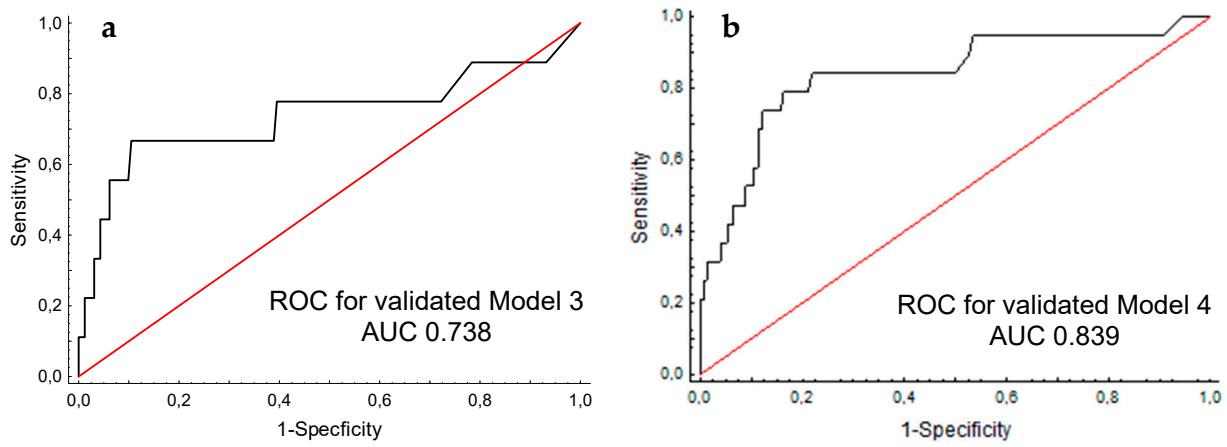


Figure S3. ROC curves for validated logistic regression models for HCC occurrence. Model 3 (**a**) was generated automatically using backward stepwise regression with all the variables significant in the monovariate analysis as an input, and Model 4 (**b**) contained additionally minor allele status in *HDAC3* rs976552/ *CYBRD1* rs884409 for comparison.

Table S5. Polymorphisms associated with biochemical and histopathological data.

Parameter	Polymorphism												Minor HDAC3 rs976552/ CYBRD1 rs884409		
	HFE C282Y rs1800562 G>A			HFE H63D rs1799945 C>G			HDAC3 rs976552 T>G			CYBRD1 rs884409 T>G			TT in any SNP n=213	GG+GT/GG+GT n=36	p
	GG n=233	GA n=16	p	CC+CG n=240	GG n=9	p	TT+GT n=243	GG n=6	p	TT n=163	GG+GT n=86	p			
Age [yr]	47±1	52±2	0.172	48±1	52±2	0.337	48±1	41±7	0.391	47±1	49±1	0.694	48±1	46±2	0.175
Sex (Male/Female)	138/95	15/1	0.013	90/150	6/3	0.436	150/93	3/3	0.874	99/64	54/32	0.751	83/130	13/23	0.872
HGB [g/dL]	14.7±0.1	15.4±0.3	0.082	14.8±0.1	14.0±0.6	0.291	14.7±0.1	15.1±0.8	0.577	14.8±0.1	14.6±0.2	0.237	14.7±0.1	14.8±0.3	0.858
ALT [IU/L]	111±6.2	167±27	0.019	112±6	170±35	0.050	116±6	57±12	0.064	106±7	131±11	0.036	111±7	139±16	0.079
AST [IU/L]	79±4	91±16	0.382	78±4	123±22	0.016	80±4	39±7	0.035	69±4	98±8	0.002	74±4	111±13	0.002
GGT [IU/L]	101±6	109±19	0.226	103±6	58±12	0.189	102±6	85±116	0.372	87±6	129±13	0.016	97±7	129±17	0.127
Bilirubin [mg/dL]	0.9±0.04	0.8±0.1	0.925	0.9±0.04	0.8±0.1	0.439	0.9±0.04	0.6±0.1	0.142	0.8±0.04	1.0±0.1	0.009	0.8±0.04	0.9±0.1	0.769
Serum iron [µg/dL]	151±5	179±13	0.067	151±4	192±29	0.152	153±4	125±14	0.392	145±5	167±7	0.011	150±5	167±12	0.161
Transferrin saturation [%]	42±1.5	56±6	0.011	43±2	51±9	0.339	43±2	34±3	0.330	40±2	48±3	0.078	42±2	46±5	0.764
Ferritin [ng/mL]	348±27	708±220	0.004	372±30	339±95	0.998	375±30	117±48	0.126	343±35	420±52	0.170	366±33	394±63	0.710
Histopathology n=211	n=197	n=14	p	n=205	n=6	p	n=206	n=5	p	n=140	n=71	p	n=181	n=30	p
Inflammation grade (0-3)	2(1/2)	2(2/3)	0.150	2(2/2)	2(1.5/2)	0.818	2(2/2)	2(2/2)	0.718	2(1/2)	2(2/3)	0.013	2(1/2)	2(2/3)	0.021
Fibrosis grade (0-4)	2(1/3)	2(1/3)	0.842	2(1/3)	1.5(1/2)	0.289	2(1/3)	1(1/2)	0.277	2(1/3)	2(1/3)	0.095	2(1/3)	2(2/3)	0.191
Iron deposits grade (0-3)	1(0/1)	1(0/2)	0.154	0(0/1)	0(0/1)	0.761	0(0/1)	0(0/0)	0.179	0(0/1)	0(0/1)	0.890	0(0/1)	0(0/1)	0.698
Steatosis grade (0-3)	1(0/2)	2(0/2)	0.292	1(0/2)	0.5(0/2)	0.772	1(0/2)	2(0/2)	0.832	1(0/2)	1(0/2)	0.292	1(0/2)	1(0/2)	0.610
Hepatocyte iron deposits present (yes/no)	68/129	8/6	0.161	75/130	1/5	0.871	76/130	0/5	0.217	50/90	26/45	0.926	65/116	11/19	0.883
Hepatocyte steatosis present (yes/no)	117/80	9/5	0.937	123/82	3/3	0.944	123/83	3/2	0.654	76/64	50/21	0.023	107/74	19/11	0.814
Liver fibrosis present (yes/no)	122/75	9/5	0.913	128/77	3/3	0.848	129/77	2/3	0.573	82/58	49/22	0.140	110/71	21/9	0.446

Quantitative biochemical data is shown as mean ± SE; histopathological data is shown as median values with percentiles (25th/ 75th).

Table S6. SNPs associated with hepatic iron-related gene expression normalized to serum iron indices.

Parameter	HFE C282Y rs1800562 G>A			HFE S65C rs1800730 A>T			HFE H63D rs1799945 C>G			HDAC2 rs3778216 C>T			TMPRSSR6 rs855791 C>T		
	GG n=233	GA n=16	p	AA n=241	AT n=8	p	CC n=172	GC+GG n=77	p	CC n=132	CT+TT n=117	p	CC+CT n=220	TT n=29	p
sFerritin	347±27	707±221	0.004	367±30	459±150	ns	388±39	331±35	ns	155±6	149±7	ns	381±32	289±60	ns
sFe	150±5	178±13	ns	152±4	150±38	ns	149±5	161±8	ns	410±45	323±34	ns	154±5	141±12	ns
Gene expression	n=116	n=8		n=118	n=6		n=85	n=39		n=71	n=53		n=111	n=13	
HAMP	1.6±0.1	1.9±0.3	ns	1.6±0.1	2.9±0.6	0.029	1.9±0.3	1.5±0.1	ns	1.8±0.1	1.6±0.2	ns	1.7±0.1	1.6±0.3	ns
FPN1	14±1	10±2	ns	14±0.8	13±3.4	ns	13±1	14±2	ns	13±1.0	15±1.3	ns	14±1	11±2	ns
Ratio*															
Tfr2/ sFerritin	13±2	3±1	0.008	5±1	12±2	ns	11±4	13±2	ns	10±2	16±4	ns	12±2	11±3	ns
HAMP/ sFerritin	9±2	4±1	ns	8±2	9±2	ns	6±1	10±2	ns	9±2	8±2	ns	9±2	11±3	ns
HJV/ sFerritin	10±2	2±1	0.007	4±1	10±2	ns	8±2	10±2	ns	7±1	13±3	0.046	9±3	11±4	ns
Bmp6/ sFerritin	10±1	2±1	0.010	4±1	10±1	ns	7±1	11±2	ns	7±1	12±2	0.049	9±1	11±4	ns
Id1/ sFerritin	10±1	6±3	ns	6±1	10±1	ns	8±1	10±1	ns	9±1	10±1	ns	9±1	13±4	ns
HO1/ sFerritin	17±2	4±1	0.017	6±1	16±2	ns	14±1	17±3	ns	12±2	21±4	0.030	16±2	16±4	ns
FPN1/sFerritin	2±0.4	0.3±0.1	0.008	2±0.3	0.6±0.1	ns	1.8±0.4	1.7±0.7	ns	1.3±0.2	2.4±0.7	ns	1.8±0.4	1.7±0.6	ns
Tfr2/ sFe	9±1	5±1	0.009	14±5	8±1	ns	8±1	9±1	ns	8±1	9±1	ns	9±1	9±1	ns
HAMP/ sFe	9±2	7±1	ns	23±7	8±1	0.001	9±1	9±1	ns	9±1	8±1	ns	9±1	10±2	ns
HJV/ sFe	7±0.5	4±0.3	0.006	10±3	6±0.4	ns	6±1	7±1	ns	6±1	6±1	ns	6±0.5	8±1	0.022
Bmp6/ sFe	7±1	4±1	0.016	14±6	7±1	ns	6±0.4	8±1	ns	6±0.5	5±1	ns	7±1	8±1	0.031
Id1/ sFe	10±1	9±2	ns	18±4	9±1	0.021	9±1	10±1	ns	10±1	9±1	ns	10±1	10±2	ns
HO1/ sFe	13±1	8±1	0.034	17±4	12±1	ns	11±1	13±1	ns	11±1	15±2	ns	12±1	13±2	ns
FPN1/sFe	1.4±0.1	0.7±0.2	0.041	1.3±0.1	1.4±0.3	ns	1.3±0.1	1.4±0.2	ns	1.2±0.2	1.5±0.2	ns	1.3±0.1	1.3±0.3	ns

*ratio of relative gene expression in liver biopsy normalized to serum ferritin (sFerritin) or serum iron (sFe)

Table S7. Spearman rank correlation coefficients for associations between hepatic gene expression and baseline biochemical parameters for CHC patients.

Relative gene expression	Parameter								
	Age	ALT	AST	GGT	Bilirubin	sFe	TS	sFerritin	Liver inflammation
<i>Tfr2</i>	ns	ns	ns	ns	ns	ns	ns	ns	ns
<i>HAMP</i>	0.196 ^d	ns	ns	0.221 ^d	ns	0.394 ^a	0.467 ^a	0.641 ^a	ns
<i>HJV</i>	-0.201 ^d	ns	-0.223 ^d	ns	ns	ns	ns	ns	-0.293 ^c
<i>Bmp6</i>	-0.214 ^d	ns	ns	ns	ns	ns	ns	ns	ns
<i>Id1</i>	0.282 ^d	0.245 ^d	0.244 ^d	0.319 ^c	0.240 ^d	0.492 ^a	0.548 ^a	0.595 ^a	ns
<i>HO1</i>	ns	0.308 ^c	0.319 ^c	0.262 ^d	0.239 ^d	0.207 ^d	ns	0.292 ^d	ns
<i>FPN1</i>	ns	ns	ns	ns	ns	ns	ns	ns	ns
<i>Tim3</i>	ns	ns	ns	ns	ns	ns	ns	ns	ns
<i>PD-1</i>	ns	0.462 ^b	0.422 ^c	ns	0.290 ^d	0.247 ^d	ns	ns	0.267 ^d
<i>CTLA4</i>	ns	0.557 ^a	0.526 ^a	ns	0.279 ^d	0.304 ^d	0.246 ^d	0.230 ^d	0.301 ^d

^a, p<0.00001; ^b, p<0.0001; ^c, p<0.001; ^d, p<0.05; ns, not significant; sFe, serum iron; sFerritin, serum ferritin.

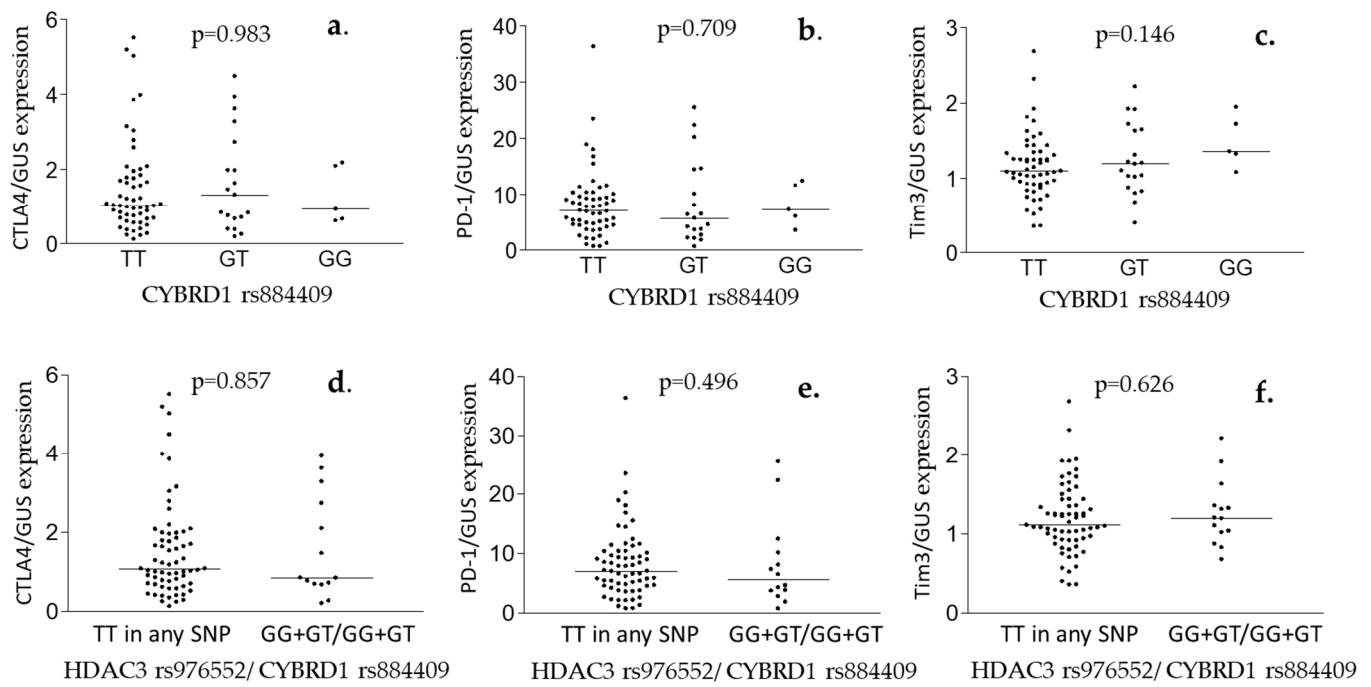


Figure S4. Hepatic expression of co-inhibitory receptors in relation to selected genotypes. Hepatic expression of *CTLA4* (a, d), *PD-1* (b, e), and *Tim3* (c, f) in samples from CHC patients with different *CYBRD1* rs884409 (a-c) and *HDAC3* rs976552/*CYBRD1* rs884409 (d-f) genotypes. Shown are the p values from a Kruskal-Wallis test (a-c) or Mann-Whitney U test (d-f).