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Leaf Photosynthetic Capacity of Sunlit and Shaded Mature Leaves in a Deciduous Forest

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Abstract: A clear understanding of the dynamics of photosynthetic capacity is crucial for accurate modeling of ecosystem carbon uptake. However, such dynamical information is hardly available and has dramatically impeded our understanding of carbon cycles. Although tremendous efforts have been made in coupling the dynamic information of photosynthetic capacity into models, using "proxies" rooted from the close relationships between photosynthetic capacity and other available leaf parameters remains the popular selection. Unfortunately, no consensus has yet been reached on such "proxies", leading them only applicable to limited cases. In this study, we aim to identify if there are close relationships between the photosynthetic capacity (represented by the maximum carboxylation rate, $V_{\rm cmax}$) and leaf traits for mature broadleaves within a cold temperature deciduous forest. This is based on a long-term in situ dataset including leaf chlorophyll content (Chl), leaf nitrogen concentration (Narea, Nmass), leaf carbon concentration (Carea, Cmass), equivalent water thickness (EWT), leaf mass per area (LMA), and leaf gas exchange measurements from which V_{cmax} was derived, for both sunlit and shaded leaves during leaf mature periods from 2014 to 2019. The results show that the $V_{\rm cmax}$ values of sunlit and shaded leaves were relatively stable during these periods, and no statistically significant interannual variations occurred (p > 0.05). However, this is not applicable to specific species. Path analysis revealed that N_{area} was the major contributor to V_{cmax} for sunlit leaves (0.502), while LMA had the greatest direct relationship with $V_{\rm cmax}$ for shaded leaves (0.625). The LMA has further been confirmed as a primary proxy if no leaf type information is available. These findings provide a promising way to better understand photosynthesis and to predict carbon and water cycles in temperate deciduous forests.

Keywords: V_{cmax}; interannual variation; biochemical properties; structural traits

1. Introduction

Photosynthesis is one of the main drivers of the carbon flux and plays a critical role in climate models across various scales from individual to global [1–3]. Currently, carbon uptake is popularly estimated using a well-established biochemical model of photosynthesis developed by Farquhar et al. [4,5], requiring leaf photosynthetic capacity parameters as key inputs. Hence, a clear understanding of leaf photosynthetic capacity is indispensable for predicting responses of carbon flow in ecosystems to climate change [6] and for modeling biosphere-atmosphere interactions at local and regional scales [7,8]. Leaf photosynthetic capacity is often described by the maximum rate of carboxylation (V_{cmax}), determined by the amount, activity, and kinetics of the enzyme ribulose 1,5-bisphosphate carboxylase/oxygenase (Rubisco) [4].

widely across species, plant functionally treated as a constant. However, V_{cmax} in general varies widely across species, plant functional types, and environmental conditions [10–14]. In addition, seasonal variations of it are also apparent [15–17], further complicating the situation and suggesting that the constant-treatment of the parameter in the models is inadequate. Unfortunately, the traditional approach of inferring V_{cmax} by model inversion from photosynthesis measurements [18,19] is so time-consuming and cannot provide highly frequent dynamical information of the parameter required by models.

Tremendous efforts have hence been made to provide spatiotemporal dynamics of the parameter. Searching potential correlations between leaf photosynthetic capacity and other easily measurable quantitative parameters has become a popular practice in recent years, e.g., using nitrogen (N) content on an area basis [20,21] or a mass basis [22], or chlorophyll [23], as "proxies" for providing spatiotemporal dynamics of the parameter, and is getting particularly important for parameterizing terrestrial biosphere models [5,24,25].

However, it is vital to explore the mechanistic relationships between V_{cmax} and leaf parameters, e.g., leaf mass per area (LMA), nitrogen concentration, or chlorophyll content, before this approach can be widely applied. It has been well documented that V_{cmax} is highly correlated with leaf nitrogen concentration either expressed on an area basis [20,21] or a mass basis [22], and it can therefore be estimated using existing V_{cmax} -N relationships. Such associations have widely been used in scaling estimates of leaf photosynthesis to canopy level [26]. A possible reason for the association is that nitrogen is the main component of Rubisco, accounting for a large proportion of the photosynthetic machinery [27,28]. On the other hand, however, the use of the relationship has been criticized because there is a wide range of V_{cmax} for a given value of N_{area} among different studies, and leaf ontogeny or drought may have additional effects on the relationship, which in turn lead to substantial errors [18]. More critically, this relationship is still a subject of debate [28]. No strong relationship could have ever been observed in several studies, e.g., van de Weg et al. [29] and Dusenge et al. [30], whereas more studies claimed that the relationship is relatively stable and robust among different species [31] or under different environmental conditions [27,32,33].

Comparatively, much fewer works have ever focused on using other leaf biochemical properties such as chlorophyll content, or other leaf structural traits to derive V_{cmax} . The leaf chlorophyll content is a key parameter directly linked to photosynthetic potential or primary production and may thus provide valuable information on the physiological status [34,35]. Since the leaf chlorophyll content may be derived from satellite data over different spatial and temporal scales [23], it may therefore provide another promising potential proxy for V_{cmax} . Researches show that leaf chlorophyll distribution is strongly correlated with leaf nitrogen content [36] because a large amount of leaf nitrogen is distributed in chlorophyll pigments [37], suggesting a potential linkage between them. A previous study by Niinemets et al. [38] has reported that the decline in leaf chlorophyll content would have resulted in similar decreases in leaf photosynthetic capacity. Unfortunately, the variation of V_{cmax} in response to the chlorophyll content has been poorly documented up to date, limiting its usefulness as a "proxy" for V_{cmax} .

Previous researches have also shown that most of the variation in V_{cmax} could be explained by the variation in leaf structural traits, usually quantified using LMA [39,40], although the effect of leaf structure on V_{cmax} could be rather complex [41–44]. The LMA determines several leaf-scale processes, including nitrogen and water use efficiency, integrates leaf morphology and composition, and is critical for carbon sequestration and productivity [45]. Furthermore, LMA is a key parameter for plant growth and an important indicator of plant strategies, although it has been used less so in plant physiology [42]. Unfortunately, the controlling function of LMA on photosynthetic capacity remains unclear [25].

Previous studies have further proved that the canopy-level flux is an integrated response in which both sunlit and shaded leaf groups are involved, which respond differently to climate change [46,47]. As a result, most models have optimized the estimation of fluxes in carbon dioxide and water vapor by

separating the canopy structure into sunlit and shaded leaf groups [48–52]. Unfortunately, most models focused much more on sunlit leaves, ignoring the fact that most of the leaves within the canopy are in the shade and that the shaded leaves have their specific photosynthetic characteristics. A recent study by He et al. [47] clearly indicated that the shaded leaf group is particularly important in estimating gross primary production and is not vulnerable to the effects of climate change, suggesting the importance of including the dynamics of leaf photosynthetic capacity for this group in models. Consequently, searching close relationships for providing dynamic information of leaf photosynthetic capacity for shaded leaves is an inevitable step towards improving flux estimations further.

In this study, we focused on identifying the closest relationships for both sunlit and shaded leaves of a typical temperature deciduous forest by assessing the relationships of photosynthetic capacity with various biochemical and structural parameters. This is based on a long-term field-measured dataset including both leaf photosynthetic capacity and associated biochemical and structural parameters for the two leaf groups. The main objectives are: (1) to provide an overview of long-term (interannual) and short-term (within leaf mature periods) variations in leaf photosynthetic capacity for both leaf groups; (2) to explore potential close relationships between photosynthetic capacity and leaf biochemical and structural parameters in different leaf groups.

2. Materials and Methods

2.1. Study Site

The study site is located at Nakagawane forest (138°06′ E, 35°04′ N) (Figure 1), one of the research forests of Shizuoka University, Japan [53]. This research site has a typical alpine cold-temperate climate with a mean annual air temperature of 16°C and a mean annual precipitation of approximately 2500 mm [54], and an elevation from 390 to 1560 m.



Figure 1. The location of the tower site used in this study: (**a**) inside Japan; (**b**) inside Shizuoka Prefecture, Japan. (Boundary file was obtained from GADM - https://gadm.org/index.html).

2.2. Sample collection

Leaf gas exchange, as well as leaf biochemical and structural properties were measured for all species accessible from the tower at the study site, consisting mainly of *Fagus crenata*, *Betula grossa*, *Carpinus tschonoskii*, *Stewartia monadelpha*, and *Acer shirasawanum*. The tree canopies were separated into the top layer of sunlit leaves and the bottom layer of shaded leaves as well as the middle layer with both sunlit and shaded leaves. However, the samples taken from the middle layer were not included in the analysis in order to avoid possible miscategorization of leaf groups. All samples were taken following

the detached branch method [5] inside the tower of different directions at each layer. Three fully expanded and apparent non-senescing leaves attached to the cut branches from each direction were selected for each round of measurement. They were cut predawn and were immediately recut under water for reestablishing the xylem water column [16,29,40]. After that, the sampled branches were sealed in polyethylene boxes under dark and humid conditions and transported to the laboratory for further determination. Samples were collected from mid-June to mid-September (DOY (Day of Year) 168 to DOY 258) in 2015, 2017, and 2018, while in mid-September (DOY 258) in 2014, and mid-August (DOY 222) for both 2016 and 2019.

2.3. Leaf Gas Exchange Measurements

Leaf gas exchange was measured using the LI-6400 portable photosynthesis system (Li-Cor 6400, Li-Cor, Lincoln, NE, USA) during the maturation periods of 2014–2019 for sunlit leaves and 2014–2018 for shaded leaves. The chamber temperature was set to 25 °C, and the net photosynthetic rate (A)–intercellular CO₂ concentrations (Ci) (A-Ci) curves of light-saturated photosynthesis (at 1000 μ mol·m⁻²·s⁻¹) were obtained. Leaf photosynthetic capacity (maximum carboxylation rate– V_{cmax}) on a leaf area basis was then inferred from the A-Ci curves using the R package "plantecophys" [55].

2.4. Leaf Biochemistry and Structure Measurements

Leaf discs for chlorophyll content were punched on the same leaf samples after gas exchange measurements and then were kept at 4 °C refrigerator prior to extraction with N, N-Dimethylformamide (DMF) for 24 h. A dual-beam scanning ultraviolet-visible spectrophotometer (Ultrospec 3300 pro, Amersham Biosciences, Piscataway, NJ, USA) was used to record absorption at different wavelengths, while the Porra's method was applied to calculate chlorophyll content (Chl) [56]. Leaf areas were measured using digital photographs taken by a high-resolution scanner (EPSON GT-S600; EPSON, Jakarta, Indonesia). Fresh weights of the same leaves were also taken before they were oven-dried for 48 h at 70 °C to constant weight for calculating equivalent water thickness (EWT) and leaf mass per area (LMA). However, there were no EWT and LMA data available for 2015. Finally, the oven-dried leaves were milled to a fine powder, from which leaf nitrogen concentration per unit area (N_{area}) and per unit mass (N_{mass}), as well as carbon concentration per unit area (C_{area}) and per unit mass (C_{mass}) were determined with a gas chromatograph (Sumigraph NC-95A, Shimadzu, Kyoto, Japan) (only for the samples from 2017–2018).

2.5. Statistical Analysis

All photosynthetic, biochemical, and structural parameters were log10-transformed prior to statistical analysis. Repeated measure analysis of variance (ANOVA) was performed to determine the variations among the months and years (p < 0.05). The relationships of photosynthetic capacity with biochemical and leaf structural parameters were analyzed by Pearson's correlation, with the possible direct and indirect effects of each parameter on the photosynthetic capacity being further identified by path analysis [57–59]. Data analysis was conducted using the packages "plantecophys" and "agricolae" in R 3.5.3.

3. Results

3.1. Long-term and Short-term Variations in V_{cmax}

Figure 2 shows the temporal variations of V_{cmax} in the leaf mature periods for both sunlit and shaded leaves (from DOY 168 to DOY 258) for 2015, 2017, and 2018, respectively. For sunlit leaves, V_{cmax} values in these 3 years slightly decreased from mid-June to July but rebound slowly until mid-August (DOY 222) before they slightly declined towards mid-September (Figure 2a). However,

statistical results revealed that there were no significant changes during the months (p > 0.5). Similarly, V_{cmax} also tended to be stable within these months for shaded leaves (p > 0.5, Figure 2b).



Figure 2. Changes in V_{cmax} of sunlit (**a**) and shaded (**b**) leaves from DOY (Day of Year) 168 to DOY 258 in 2015, 2017, and 2018.

The mean $V_{\rm cmax}$ values of sunlit leaves from 2014 to 2019 and of shaded leaves from 2014 to 2018 were used to reveal their interannual variations (Figure 3). As shown in Figure 3, the mean $V_{\rm cmax}$ values varied from 36.80 to 50.79 µmol m⁻² s⁻¹ in the sunlit leaves and from 25.28 to 32.26 µmol m⁻² s⁻¹ in the shaded leaves. The highest mean value of $V_{\rm cmax}$ of sunlit leaves was found for 2019 and of shaded leaves was found for 2017 (50.79 and 32.26 µmol m⁻² s⁻¹, respectively). However, there was no significant yearly variation in both sunlit and shaded leaves (p > 0.05).



Figure 3. Interannual variations in the maximum carboxylation rate (V_{cmax}) at the study site for sunlit leaves from 2014 to 2019 and for shaded leaves from 2014 to 2018. Mean values and standard errors were used to show the trends.

Species-specific temporal variations in V_{cmax} have also been examined. Two common species, *Fagus crenata* and *Acer shirasawanum* were selected because their relatively large and continuous sample numbers met with the statistical requirement. Contrary to canopy scale behaviors, significant interannual variations in both sunlit and shaded leaves were identified for *Fagus crenata*. Its V_{cmax} in 2014 was significantly higher than in 2017 and 2018 for sunlit leaves (p < 0.001), and was significantly higher than the other three years (2016–2018) for shaded leaves (p < 0.01). In contrast, there were

no statistically significant interannual variations in V_{cmax} for both sunlit and shaded leaves of *Acer shirasawanum* (p > 0.05) from 2014 to 2016, suggesting the behaviors can be very different among different species.

3.2. Relationships of V_{cmax} with Leaf Biochemical and Structural Traits

Significant relationships of V_{cmax} with leaf biochemical and structural traits were detected in all leaf samples (Figure 4). Variations in V_{cmax} were positively correlated with leaf biochemistry (represented by nitrogen concentration and chlorophyll content in this study), showing a relatively stronger relationship with N_{area} (r = 0.54, p < 0.001, Figure 4a) than with Chl (r = 0.46, p < 0.001, Figure 4c). However, additional analysis revealed that there was no significant pairwise relationship between V_{cmax} and N_{mass} (p > 0.05, Figure 4b) in terms of a mass base. On the other hand, besides leaf biochemical properties, V_{cmax} was significantly correlated with leaf structural traits (represented by LMA in this study) across all leaves (r = 0.53, p < 0.001, Figure 4d).



Figure 4. Relationships of V_{cmax} with N_{area} (**a**), N_{mass} (**b**), Chl (**c**), and LMA (**d**) for sunlit leaves (hollow triangles) and shaded leaves (solid triangles). V_{cmax} , maximum carboxylation rate; N_{area} , leaf nitrogen concentration per unit area; N_{mas} , leaf nitrogen concentration per unit mass; Chl, chlorophyll content; LMA, leaf mass per area. Solid lines indicate significant results.

Further analysis based on a mass basis revealed that there were no significant correlations between V_{cmax} and other leaf biochemical and structural traits, including Chl, N_{area} , and LMA (p > 0.05), for both sunlit and shaded leaves.

Depending on the different leaf groups, V_{cmax} exhibited significant relationships with both N_{area} and Chl (r = 0.38 vs. r = 0.37, p < 0.001), while a weak relationship with LMA (r = 0.30, p < 0.05) was found for the sunlit leaves. On the contrary, the correlation between V_{cmax} and LMA proved to be stronger (r = 0.56, p < 0.001) than that with Chl (r = 0.47, p < 0.001) and N_{area} (r = 0.45, p < 0.01) for shaded leaves. Collectively, V_{cmax} was correlated less strongly with leaf biochemical and structural traits in sunlit leaves than in shaded leaves (Figure 4).

3.3. Direct and Indirect Effects of Leaf Biochemical and Structural Traits on V_{cmax}

Path analysis (V_{cmax} as the response variable and leaf biochemical and structural parameters as the independent variables) was carried out to reveal the direct and indirect effects of leaf traits on V_{cmax} . The results based on all samples and on different leaf groups are presented in Table 1.

Table 1. Direct and indirect effects of leaf biochemistry and structure on V_{cmax} in sunlit and shaded leaves.

	EWT	Chl	Narea	Carea	LMA	n
EWT-total	-0.164	0.073	0.221	-0.077	0.267	124
EWT-sun	-0.037	0.182	0.331	-0.274	0.038	63
EWT-sh	-0.197	0.019	0.026	-0.045	0.238	61
Chl-total	-0.072	0.167	0.206	-0.079	0.238	170
Chl-sun	-0.018	0.379	0.226	-0.252	0.035	88
Chl-sh	-0.018	0.208	0.055	-0.138	0.363	82
N _{area} -total	-0.102	0.097	0.356	-0.103	0.293	79
N_{area} -sun	-0.024	0.170	0.502	-0.307	0.039	39
N_{area} -sh	-0.051	0.117	0.099	-0.183	0.469	40
C _{area} -total	-0.110	0.115	0.320	-0.115	0.339	79
Carea-sun	-0.028	0.261	0.422	-0.365	0.050	39
C_{area} -sh	-0.041	0.135	0.085	-0.213	0.544	40
LMA-total	-0.122	0.110	0.288	-0.108	0.361	124
LMA-sun	-0.026	0.250	0.366	-0.343	0.053	63
LMA-sh	-0.075	0.121	0.074	-0.185	0.625	61

EWT, equivalent water thickness; Chl, chlorophyll content; N_{area} , leaf nitrogen concentration per unit area; C_{area} , leaf carbon concentration per unit area; LMA, leaf mass per area; V_{cmax} , maximum carboxylation rate. Bold diagonal values indicate direct effects; n is the number of leaf samples.

Among the examined variables, Chl, N_{area} , and LMA had positive effects on V_{cmax} in all leaves. The variable that influenced V_{cmax} most significantly was LMA (0.361), followed by N_{area} (0.356).

For the sunlit leaves, path analysis showed that the maximum positive direct effect was from N_{area} (0.502), followed by Chl (0.379) and LMA (0.053). Although LMA was strongly correlated with V_{cmax} (r = 0.3, p < 0.05), its direct effect was surprisingly low (0.053, less than 0.100). However, LMA exhibited an impressive positive indirect effect via N_{area} (0.366).

For the shaded leaves, LMA was the only variable that strongly correlated with V_{cmax} if considering the direct effect. It exhibited a significant positive correlation (r = 0.56, p < 0.001) and an ever-higher direct path coefficient value (0.625) with V_{cmax} . Although correlation analysis suggested that Chl, N_{area} , and C_{area} also had high and significant relationships (r = 0.47, 0.45, and 0.51, respectively) with V_{cmax} , their direct effects on V_{cmax} , however, were relatively low (path coefficient = 0.208, 0.099, and -0.213, respectively). Nonetheless, high indirect effects on V_{cmax} were detected for each of them through LMA, following the order C_{area} (0.544), N_{area} (0.469), and Chl (0.363).

4. Discussion

4.1. Intra-annual and Interannual Variations in V_{cmax}

Similar to Niinemets et al. [38], the result obtained in this study confirmed that no statistically significant variation can be distinguished for V_{cmax} during the leaf maturity period, irrespective of the leaf group. The V_{cmax} stabilized from the middle of June (around DOY 168) to the middle of September (around DOY 258), suggesting there is a platform of V_{cmax} values surrounding the peak values, termed as "leaf maturity period". However, such values should be used with caution, since they were obtained from canopy-layer-scale statistics rather than from specific species. There are apparent seasonal variations of leaf photosynthetic capacity beyond this period [16,32,60,61]. Up to now, most eco-physiological studies mainly focus on mature leaves and show that the photosynthetic apparatus is essentially constant in the mature period (e.g., Niinemets et al. [38]). Such adaptationconstancy

of photosynthetic mechanism on one hand greatly simplifies the estimation of the carbon balance, while on the other hand, unfortunately, it dramatically increases simulation deviation, as previous researches proved that only emphasizing on measuring mature leaves ignores the demographics of the leaves [17], leading to an overestimation of the photosynthetic productivity [62,63]. Furthermore, V_{cmax} also varies across species and locations [3]. Consequently, the traditional treatment of constant V_{cmax} in gas exchange models might be applicable only to the leaf maturity period.

Our results indicate that the mean values of V_{cmax} in the time of leaf maturation did not vary significantly among years, leading to no interannual variations in the context of statistics. This finding is rather inconsistent with those observed in previous studies [16,18,64,65]. Again, our canopy-layer approach of statistics has explained the discrepancy, since a further species-specific examination revealed that there existed statistically significant interannual variations in different species within the canopy. Furthermore, according to [18], the interannual variations of V_{cmax} are primarily caused by water stress. Whereas no water stress has been confirmed in our research site and may have further explained the discrepancy.

Taken together, the results obtained in this study should be interpreted with caution since they were rooted in canopy scale statistics. Even so, our results indicated that there were relatively constant V_{cmax} within leaf mature periods across years and may thus provide a feasible modeling approach from leaf functional types, rather than from distinctive species, for understanding canopy scale performance.

4.2. Relationships of V_{cmax} with Leaf Biochemical and Structural Traits for Both Leaf Groups

In general, V_{cmax} is proportional to leaf nitrogen concentration when expressed on an area basis (N_{area}) and has been confirmed for a wide range of species or plant functional types around the world [8,39,66,67]. The results obtained in this study again indicate that there are stronger relationships for the area-based than the mass-based approaches, but all relationships were strongly affected by LMA. This confirms that the functional interpretation of the V_{cmax} - N_{area} relationship requires the analysis of the parallel variation of LMA [68]. More directly, a significant correlation between V_{cmax} and LMA was found during leaf maturation for all leaves, similar to those reported by other studies [16,18,39,40]. However, our results suggest that LMA is positively correlated with V_{cmax} , inconsistent with previous claims that thick leaves should have low photosynthetic rates [66]. Such discrepancy reminds us that the relationship between photosynthesis and LMA could be rather complex and requires a detailed understanding of how this physiological process relates to plant strategy or environmental conditions case by case. Even so, our results at least suggest that both LMA and N_{area} are useful parameters for estimating photosynthetic capacity, as claimed by Han et al. [69].

Since the relationship determined only by simple correlation analysis may limit the prediction of selection responses, it is therefore of necessity to partition direct and indirect cause-and-effect [70] in order to identify more accurate and robust relationships.

Path analysis demonstrated the importance of N_{area} as the primary indicator of V_{cmax} in the sunlit leaves, with Chl and LMA being less important. A stronger direct effect of N_{area} than of Chl with V_{cmax} in this leaf group suggests that leaf N_{area} is of greater importance in controlling V_{cmax} than Chl, which is in disagreement with Croft et al. [35], in which chlorophyll exerted a strong influence over V_{cmax} and the use of chlorophyll as a proxy to model V_{cmax} was recommended. Our finding, nevertheless, was in accordance with the results of Wright et al. [66] and Han et al. [69], where indicated that leaf nitrogen concentration was most suitable to predict photosynthetic capacity. Clearly, the strong correlation between V_{cmax} and N_{area} has a physiological basis, which can be explained by the high proportion of nitrogen in leaves stored in Rubisco [71], and the proportion is largely constant [72]. The results again suggest that N_{area} , rather than Chl, is more closely related to V_{cmax} , at least for sunlit leaves. This finding, thus, should have important implications for canopy integration schemes when used to calculate photosynthesis in gas exchange models.

On the other hand, path analysis clearly indicated that LMA was the single factor that directly affected V_{cmax} most in the shaded leaves. Even though significant positive correlations of V_{cmax} with Chl

and N_{area} were also obtained, they were attributed preliminarily to the indirect effects of the parameters through LMA. Previous studies have shown that the growth of shaded leaves is mainly restricted by carbohydrate supply [73], leading to a significant positive relationship between photosynthetic capacity and LMA, which is commonly reported for shaded leaves (e.g., Poorter et al. [42]). It is clear that LMA is associated with resource investment in structural components [74], while leaf photosynthetic capacity is related to biomass investment, making LMA the main driver of the photosynthetic capacity in the shaded leaves. Further, variations in LMA can be attributed to light acclimation, which in turn drives leaf photosynthetic capacity in response to the light environment [75]. The importance of exploring the relationship between V_{cmax} and LMA has also been pointed out by Grassi and Bagnaresi [68] by providing a functional interpretation of acclimation and elucidating the role of physiological plasticity. The results obtained in this study indicate the importance of leaf structure in determining photosynthetic capacity during leaf maturation, which is critical information that needs to be included in gas exchange models.

Furthermore, significant positive relationships of LMA with Chl (Figure 5a) and N_{area} (Figure 5b) were identified, all suggesting that the structural traits of fully mature leaves are important in determining photosynthetic capacity.



Figure 5. Relationships of LMA with Chl (**a**) and N_{area} (**b**) for sunlit and shaded leaves. LMA, leaf mass per area; Chl, chlorophyll content; N_{area} , leaf nitrogen concentration per unit area. Solid lines indicate significant results.

4.3. Implications for Understanding Gas Exchange and for Climate Models

Our results demonstrate that V_{cmax} is closely correlated with N_{area} , at least for sunlit leaves, challenging the results reported by Croft et al. [35], in which reported chlorophyll instead of N_{area} should be used as a reliable proxy to model photosynthetic capacity. Theoretically, the strong correlation between photosynthetic capacity and nitrogen concentration in the leaves can integrate the influence of differences in the light environment during the growth process [76] and has been implemented in physiologically-based gas exchange or carbon cycle models. We confirm the robustness of the relationship and suggest that N_{area} can be used for estimating V_{cmax} . Since the reliable ways to estimate leaf nitrogen concentration from remote sensing data are supported by the relationships between vegetative indices and nitrogen concentration [77–80] even at a large scale, we predict that V_{cmax} may be retrieved from remote sensing information via the bridge of N_{area} .

On the other hand, an increasing number of studies have indicated that the shaded leaves account for half the total increase in gross primary production and hence play a critical role in global carbon and water cycling [47], calling the attention to the contributions from shaded leaves. Previous studies also concluded that differently shaded portions might have altered the relationships between remote sensing indices (e.g., photochemical reflectance index and light use efficiency) [81]. As a result, future gas exchange models should have the ability to encompass both sunlit and shaded leaves, with the later much less been studied yet. Our results clearly indicate that the photosynthesis of shaded leaves

is mainly determined by structural properties rather than by biochemical constraints. Accordingly, LMA, rather than N_{area} or Chl, should be used for retrieving V_{cmax} for this group of leaves. Hence, leaf structure therefore not only plays a major role in optical properties but also may serve as an easily measured accurate index for photosynthetic capacity and productivity. For this reason, it should be incorporated into models to better understand carbon dynamics in terrestrial ecosystems.

Although statistically significant, moderate correlations of LMA with V_{cmax} nevertheless prevent the recommendation of using it as a sole proxy, suggesting further studies on this topic are required. There is increasing evidence that leaf phosphorus content plays a role in determining photosynthetic capacity, as significant positive relationships were observed [5,40,44,82]. Even so, identifying potential close relationships between leaf traits and photosynthetic capacity should be a promising step for including dynamic information of it to gas exchange models.

5. Conclusions

Our results reveal that leaf nitrogen concentration and LMA are more closely related to photosynthetic capacity in the sunlit leaves and shaded leaves, respectively, based on long-term field-measured data in a typical cold temperature deciduous forest. The results are important for understanding carbon uptake across a wide range of environmental conditions, carbon, or climate models, and deserve close attention. In the future, gas exchange models should be coupled with the dynamic information of V_{cmax} for different leaf groups, and their spatiotemporal variations should be provided by the respective proxies.

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References

- Sitch, S.; Smith, B.; Prentice, I.C.; Arneth, A.; Bondeau, A.; Cramer, W.; Kaplan, J.O.; Levis, S.; Lucht, W.; Sykes, M.T.; et al. Evaluation of ecosystem dynamics, plant geography and terrestrial carbon cycling in the LPJ dynamic global vegetation model. *Glob. Chang. Biol.* 2003, *9*, 161–185. [CrossRef]
- Canadell, J.G.; Le Quéré, C.; Raupach, M.R.; Field, C.B.; Buitenhuis, E.T.; Ciais, P.; Conway, T.J.; Gillett, N.P.; Houghton, R.A.; Marland, G. Contributions to accelerating atmospheric CO2 growth from economic activity, carbon intensity, and efficiency of natural sinks. *Proc. Natl. Acad. Sci. USA* 2007, *104*, 18866–18870. [CrossRef]
- Bonan, G.B.; Lawrence, P.J.; Oleson, K.W.; Levis, S.; Jung, M.; Reichstein, M.; Lawrence, D.M.; Swenson, S.C. Improving canopy processes in the Community Land Model version 4 (CLM4) using global flux fields empirically inferred from FLUXNET data. J. Geophys. Res. 2011, 116, 1–22. [CrossRef]
- 4. Farquhar, G.D.; von Caemmerer, S.; Berry, J.A. A biochemical model of photosynthetic CO2 assimilation in leaves of C3 species. *Planta* **1980**, *149*, 78–90. [CrossRef]
- Domingues, T.F.; Meir, P.; Feldpausch, T.R.; Saiz, G.; Veenendaal, E.M.; Schrodt, F.; Bird, M.; Djagbletey, G.; Hien, F.; Compaore, H.; et al. Co-limitation of photosynthetic capacity by nitrogen and phosphorus in West Africa woodlands. *Plant Cell Environ.* 2010, *33*, 959–980. [CrossRef]
- Hikosaka, K.; Nabeshima, E.; Hiura, T. Seasonal changes in the temperature response of photosynthesis in canopy leaves of Quercus crispula in a cool-temperate forest. *Tree Physiol.* 2007, 27, 1035–1041. [CrossRef] [PubMed]
- 7. Santiago, L.S.; Mulkey, S.S. A test of gas exchange measurements on excised canopy branches of ten tropical tree species. *Photosynthetica* **2003**, *41*, 343–347. [CrossRef]
- 8. Kattge, J.; Knorr, W.; Raddatz, T.; Wirth, C. Quantifying photosynthetic capacity and its relationship to leaf nitrogen content for global-scale terrestrial biosphere models. *Glob. Chang. Biol.* **2009**, *15*, 976–991. [CrossRef]

- 9. Verheijen, L.M.; Brovkin, V.; Aerts, R.; Bönisch, G.; Cornelissen, J.H.C.; Kattge, J.; Reich, P.B.; Wright, I.J.; Van Bodegom, P.M. Impacts of trait variation through observed trait-climate relationships on performance of an Earth system model: A conceptual analysis. *Biogeosciences* **2013**, *10*, 5497–5515. [CrossRef]
- 10. Kattge, J.; Knorr, W. Temperature acclimation in a biochemical model of photosynthesis: A reanalysis of data from 36 species. *Plant Cell Environ.* **2007**, *30*, 1176–1190. [CrossRef] [PubMed]
- 11. Rogers, A. The use and misuse of V c,max in Earth System Models. *Photosynth. Res.* **2014**, *119*, 15–29. [CrossRef] [PubMed]
- De Kauwe, M.G.; Lin, Y.S.; Wright, I.J.; Medlyn, B.E.; Crous, K.Y.; Ellsworth, D.S.; Maire, V.; Prentice, I.C.; Atkin, O.K.; Rogers, A.; et al. A test of the "one-point method" for estimating maximum carboxylation capacity from field-measured, light-saturated photosynthesis. *New Phytol.* 2016, 210, 1130–1144. [CrossRef] [PubMed]
- Wang, H.; Prentice, I.C.; Cornwell, W.K.; Keenan, T.F.; Davis, T.W.; Ian, J.; Evans, B.J.; Peng, C.; Ryde, N.; Impacts, C.; et al. A universal model for carbon dioxide uptake by plants 2. *Nat. Plants* 2017. [CrossRef] [PubMed]
- Bloomfield, K.J.; Prentice, I.C.; Cernusak, L.A.; Eamus, D.; Medlyn, B.E.; Rumman, R.; Wright, I.J.; Boer, M.M.; Cale, P.; Cleverly, J.; et al. The validity of optimal leaf traits modelled on environmental conditions. *New Phytol.* 2019, 221, 1409–1423. [CrossRef]
- 15. Wilson, K.B.; Baldocchi, D.D.; Hanson, P.J. Leaf age affects the seasonal pattern of photosynthetic capacity and net ecosystem exchange of carbon in a deciduous forest. *Plant Cell Environ.* **2001**, *24*, 571–583. [CrossRef]
- Wang, Q.; Ho, A.; Tenhunen, J.; Kakubari, Y. Annual and seasonal variations in photosynthetic capacity of Fagus crenata along an elevation gradient in the Naeba Mountains, Japan. *Tree Physiol.* 2008, 28, 277–285. [CrossRef]
- 17. Albert, L.P.; Wu, J.; Prohaska, N.; de Camargo, P.B.; Huxman, T.E.; Tribuzy, E.S.; Ivanov, V.Y.; Oliveira, R.S.; Garcia, S.; Smith, M.N.; et al. Age-dependent leaf physiology and consequences for crown-scale carbon uptake during the dry season in an Amazon evergreen forest. *New Phytol.* **2018**, *219*, 870–884. [CrossRef]
- Grassi, G.; Vicinelli, E.; Ponti, F.; Cantoni, L.; Magnani, F. Seasonal and interannual variability of photosynthetic capacity in relation to leaf nitrogen in a deciduous forest plantation in northern Italy. *Tree Physiol.* 2005, 25, 349–360. [CrossRef]
- 19. Zheng, T.; Chen, J.; He, L.; Arain, M.A.; Thomas, S.C.; Murphy, J.G.; Geddes, J.A.; Black, T.A. Inverting the maximum carboxylation rate (Vcmax) from the sunlit leaf photosynthesis rate derived from measured light response curves at tower flux sites. *Agric. For. Meteorol.* **2017**, *236*, 48–66. [CrossRef]
- Walcroft, A.; Le Roux, X.; Diaz-Espejo, A.; Dones, N.; Sinoquet, H. Effects of crown development on leaf irradiance, leaf morphology and photosynthetic capacity in a peach tree. *Tree Physiol.* 2002, 22, 929–938. [CrossRef]
- 21. Meir, P.; Levy, P.E.; Grace, J.; Jarvis, P.G. Photosynthetic parameters from two contrasting woody vegetation types in West Africa. *Plant Ecol.* **2007**, *192*, 277–287. [CrossRef]
- 22. Vårhammar, A.; Wallin, G.; Mclean, C.M.; Dusenge, M.E.; Medlyn, B.E.; Hasper, T.B.; Nsabimana, D.; Uddling, J. Photosynthetic temperature responses of tree species in Rwanda: Evidence of pronounced negative effects of high temperature in montane rainforest climax species. *New Phytol.* 2015, 206, 1000–1012. [CrossRef] [PubMed]
- Croft, H.; Chen, J.M.; Zhang, Y.; Simic, A. Modelling leaf chlorophyll content in broadleaf and needle leaf canopies from ground, CASI, Landsat TM 5 and MERIS reflectance data. *Remote Sens. Environ.* 2013, 133, 128–140. [CrossRef]
- 24. Reich, P.B.; Hobbie, S.E.; Lee, T.; Ellsworth, D.S.; West, J.B.; Tilman, D.; Knops, J.M.H.; Naeem, S.; Trost, J. Nitrogen limitation constrains sustainability of ecosystem response to CO2. *Nature* **2006**, *440*, 922–925. [CrossRef] [PubMed]
- 25. Cernusak, L.A.; Hutley, L.B.; Beringer, J.; Holtum, J.A.M.; Turner, B.L. Photosynthetic physiology of eucalypts along a sub-continental rainfall gradient in northern Australia. *Agric. For. Meteorol.* **2011**, *151*, 1462–1470. [CrossRef]
- 26. Baldocchi, D.D.; Harley, P.C. Scaling carbon dioxide and water vapour exchange from leaf to canopy in a deciduous forest. II. Model testing and application. *Plant Cell Environ.* **1995**, *18*, 1157–1173. [CrossRef]

- 27. Meir, P.; Kruijt, B.; Broadmeadow, M.; Barbosa, E.; Kull, O.; Carswell, F.; Nobre, A.; Jarvis, P.G. Acclimation of photosynthetic capacity to irradiance in tree canopies in relation to leaf nitrogen concentration and leaf mass per unit area. *Plant Cell Environ.* **2002**, *25*, 343–357. [CrossRef]
- 28. Domingues, T.F.; Berry, J.A.; Martinelli, L.A.; Ometto, J.P.H.B.; Ehleringer, J.R. Parameterization of canopy structure and leaf-level gas exchange for an Eastern Amazonian tropical rain forest (Tapajós national forest, Pará, Brazil). *Earth Interact.* 2005, *9*. [CrossRef]
- 29. van de Weg, M.J.; Meir, P.; Grace, J.; Ramos, G.D. Photosynthetic parameters, dark respiration and leaf traits in the canopy of a Peruvian tropical montane cloud forest. *Oecologia* **2012**, *168*, 23–34. [CrossRef]
- Dusenge, M.E.; Wallin, G.; Gårdesten, J.; Niyonzima, F.; Adolfsson, L.; Nsabimana, D.; Uddling, J. Photosynthetic capacity of tropical montane tree species in relation to leaf nutrients, successional strategy and growth temperature. *Oecologia* 2015, 177, 1183–1194. [CrossRef]
- 31. Ellsworth, D.S.; Reich, P.B.; Naumburg, E.S.; Koch, G.W.; Kubiske, M.E.; Smith, S.D. Photosynthesis, carboxylation and leaf nitrogen responses of 16 species to elevated pCO2 across four free-air CO2 enrichment experiments in forest, grassland and desert. *Glob. Chang. Biol.* **2004**, *10*, 2121–2138. [CrossRef]
- Misson, L.; Tu, K.; Boniello, R.A.; Goldstein, A.H. Seasonally of photosynthetic parameters in a multi-specific and vertically complex forest ecosystem in the Sierra Nevada of California. *Tree Physiol.* 2006, 26, 729–741. [CrossRef] [PubMed]
- 33. Braune, H.; Müller, J.; Diepenbrock, W. Integrating effects of leaf nitrogen, age, rank, and growth temperature into the photosynthesis-stomatal conductance model LEAFC3-N parameterised for barley (Hordeum vulgare L.). *Ecol. Model.* **2009**, 220, 1599–1612. [CrossRef]
- Riccardi, M.; Mele, G.; Pulvento, C.; Lavini, A.; D'Andria, R.; Jacobsen, S.E. Non-destructive evaluation of chlorophyll content in quinoa and amaranth leaves by simple and multiple regression analysis of RGB image components. *Photosynth. Res.* 2014, 120, 263–272. [CrossRef] [PubMed]
- 35. Croft, H.; Chen, J.M.; Luo, X.; Bartlett, P.; Chen, B.; Staebler, R.M. Leaf chlorophyll content as a proxy for leaf photosynthetic capacity. *Glob. Chang. Biol.* **2017**, *23*, 3513–3524. [CrossRef] [PubMed]
- Daughtry, C.S.T.; Walthall, C.L.; Kim, M.S.; De Colstoun, E.B.; McMurtrey Iii, J.E. Estimating Corn Leaf Chlorophyll Concentration from Leaf and Canopy Reflectance. *Remote Sens. Environ.* 2000, 74, 229–239. [CrossRef]
- 37. Filella, I.; Serrano, L.; Serra, J.; Peñuelas, J. Evaluating Wheat Nitrogen Status with Canopy Reflectance Indices and Discriminant Analysis. *Crop Sci.* **1995**, *35*, 1400–1450. [CrossRef]
- Niinemets, Ü.; Kull, O.; Tenhunen, J.D. Within-canopy variation in the rate of development of photosynthetic capacity is proportional to integrated quantum flux density in temperate deciduous trees. *Plant Cell Environ.* 2004, 27, 293–313. [CrossRef]
- 39. Díaz-Espejo, A.; Walcroft, A.S.; Fernández, J.E.; Hafidi, B.; Palomo, M.J.; Girón, I.F. Modeling photosynthesis in olive leaves under drought conditions. *Tree Physiol.* **2006**, *26*, 1445–1456. [CrossRef]
- Bloomfield, K.J.; Domingues, T.F.; Saiz, G.; Bird, M.I.; Crayn, D.M.; Ford, A.; Metcalfe, D.J.; Farquhar, G.D.; Lloyd, J. Contrasting photosynthetic characteristics of forest vs. savanna species (Far North Queensland, Australia). *Biogeosciences* 2014, *11*, 7331–7347. [CrossRef]
- 41. Aranda, X.; Agustí, C.; Joffre, R.; Fleck, I. Photosynthesis, growth and structural characteristics of holm oak resprouts originated from plants grown under elevated CO2. *Physiol. Plant.* **2006**, *128*, 302–312. [CrossRef]
- 42. Poorter, H.; Niinemets, Ü.; Poorter, L.; Wright, I.J.; Villar, R. Causes and consequences of variation in leaf mass per area (LMA): A meta-analysis. *New Phytol.* **2009**, *182*, 565–588. [CrossRef] [PubMed]
- 43. Walker, A.P.; Beckerman, A.P.; Gu, L.; Kattge, J.; Cernusak, L.A.; Domingues, T.F.; Scales, J.C.; Wohlfahrt, G.; Wullschleger, S.D.; Woodward, F.I. The relationship of leaf photosynthetic traits—Vcmax and Jmax—to leaf nitrogen, leaf phosphorus, and specific leaf area: A meta-analysis and modeling study. *Ecol. Evol.* 2014, 4, 3218–3235. [CrossRef] [PubMed]
- 44. Bahar, N.H.A.; Ishida, F.Y.; Weerasinghe, L.K.; Guerrieri, R.; O'Sullivan, O.S.; Bloomfield, K.J.; Asner, G.P.; Martin, R.E.; Lloyd, J.; Malhi, Y.; et al. Leaf-level photosynthetic capacity in lowland Amazonian and high-elevation Andean tropical moist forests of Peru. *New Phytol.* **2017**, *214*, 1002–1018. [CrossRef] [PubMed]
- 45. Duursma, R.A.; Marshall, J.D.; Nippert, J.B.; Chambers, C.C.; Robinson, A.P. Estimating leaf-level parameters for ecosystem process models: A study in mixed conifer canopies on complex terrain. *Tree Physiol.* **2005**, *25*, 1347–1359. [CrossRef] [PubMed]

- Keenan, T.F.; Niinemets, Ü. Global leaf trait estimates biased due to plasticity in the shade. *Nat. Plants* 2016, 3, 1–6. [CrossRef]
- He, L.; Chen, J.M.; Gonsamo, A.; Luo, X.; Wang, R.; Liu, Y.; Liu, R. Changes in the Shadow: The Shifting Role of Shaded Leaves in Global Carbon and Water Cycles Under Climate Change. *Geophys. Res. Lett.* 2018, 45, 5052–5061. [CrossRef]
- 48. Chen, J.; Liu, J.; Cihlar, J.; Goulden, M. Daily canopy photosynthesis model through temporal and spatial scaling for remote sensing applications. *Ecol. Model.* **1999**, *124*, 99–119. [CrossRef]
- 49. Chen, J.M.; Mo, G.; Pisek, J.; Liu, J.; Deng, F.; Ishizawa, M.; Chan, D. Effects of foliage clumping on the estimation of global terrestrial gross primary productivity. *Global Biogeochem. Cycles* **2012**, *26*, 1–18. [CrossRef]
- 50. Chen, J.M.; Croft, H.; Zheng, T. Exploring the feasibility of global mapping of the leaf carboxylation rate. *Int. Geosci. Remote Sens. Symp.* **2016**, 2016, 1703–1706.
- 51. Dai, Y.; Dickinson, R.E.; Wang, Y.P. A two-big-leaf model for canopy temperature, photosynthesis, and stomatal conductance. *J. Clim.* **2004**, *17*, 2281–2299. [CrossRef]
- 52. Luo, X.; Chen, J.M.; Liu, J.; Black, T.A.; Croft, H.; Staebler, R.; He, L.; Arain, M.A.; Chen, B.; Mo, G.; et al. Comparison of Big-Leaf, Two-Big-Leaf, and Two-Leaf Upscaling Schemes for Evapotranspiration Estimation Using Coupled Carbon-Water Modeling. *J. Geophys. Res. Biogeosci.* **2018**, *123*, 207–225. [CrossRef]
- 53. Sonobe, R.; Wang, Q. Assessing hyperspectral indices for tracing chlorophyll fluorescence parameters in deciduous forests. *J. Environ. Manag.* 2018, 227, 172–180. [CrossRef] [PubMed]
- 54. Sonobe, R.; Wang, Q. Towards a universal hyperspectral index to assess chlorophyll content in deciduous forests. *Remote Sens.* **2017**, *9*, 191. [CrossRef]
- 55. Duursma, R.A. Plantecophys—An R package for analysing and modelling leaf gas exchange data. *PLoS ONE* **2015**, *10*, e0143346. [CrossRef]
- 56. Porra, R.J. Recent progress in porphyrin and chlorophyll biosynthesis. *Photochem. Photobiol.* **1997**, *65*, 492–516. [CrossRef]
- 57. Tadesse, T.; Singh, H.; Weyessa, B. Correlation and path coefficient analysis among seed yield traits and oil content in Ethiopian linseed germplasm. *Int. J. Sustain. Crop Prod.* **2009**, *4*, 8–16.
- 58. Akohoue, F.; Achigan-Dako, E.G.; Coulibaly, M.; Sibiya, J. Correlations, path coefficient analysis and phenotypic diversity of a West African germplasm of Kersting's groundnut [Macrotyloma geocarpum (Harms) Maréchal & Baudet]. *Genet. Resour. Crop Evol.* **2019**, *66*, 1825–1842.
- 59. Ramos, J.C.; Bertol, I.; Bandeira, D.H.; Barbosa, F.T.; Zangiski, F. Path coefficient analysis, a different approach to identify soil quality indicators. *Rev. Bras. Eng. Agric. e Ambient.* **2019**, *23*, 545–551. [CrossRef]
- 60. Wilson, K.B.; Baldocchi, D.D.; Hanson, P.J. Spatial and seasonal variability of photosynthetic parameters and their relationship to leaf nitrogen in a deciduous forest. *Tree Physiol.* **2000**, *20*, 565–578. [CrossRef]
- Wang, Q.; Tenhunen, J.; Falge, E.; Bernhofer, C.H.; Granier, A.; Vesala, T. Simulation and scaling of temporal variation in gross primary production for coniferous and deciduous temperate forests. *Glob. Chang. Biol.* 2004, *10*, 37–51. [CrossRef]
- Xu, L.; Baldocchi, D.D. Seasonal trends in photosynthetic parameters and stomatal conductance of blue oak (Quercus douglasii) under prolonged summer drought and high temperature. *Tree Physiol.* 2003, 23, 865–877. [CrossRef] [PubMed]
- Muraoka, H.; Koizumi, H. Photosynthetic and structural characteristics of canopy and shrub trees in a cool-temperate deciduous broadleaved forest: Implication to the ecosystem carbon gain. *Agric. For. Meteorol.* 2005, 134, 39–59. [CrossRef]
- 64. Noda, H.M.; Muraoka, H.; Nasahara, K.N.; Saigusa, N.; Murayama, S.; Koizumi, H. Phenology of leaf morphological, photosynthetic, and nitrogen use characteristics of canopy trees in a cool-temperate deciduous broadleaf forest at Takayama, central Japan. *Ecol. Res.* **2015**, *30*, 247–266. [CrossRef]
- Coste, S.; Roggy, J.C.; Imbert, P.; Born, C.; Bonal, D.; Dreyer, E. Leaf photosynthetic traits of 14 tropical rain forest species in relation to leaf nitrogen concentration and shade tolerance. *Tree Physiol.* 2005, 25, 1127–1137. [CrossRef]
- 66. Wright, I.J.; Westoby, M.; Baruch, Z.; Bongers, F.; Cavender-Bares, J.; Chapin, T.; Diemer, M.; Reich, P.B.; Ackerly, D.D.; Flexas, J.; et al. The worldwide leaf economics spectrum. *Nature* **2004**, *428*, 821–827. [CrossRef]
- 67. Hikosaka, K. Interspecific difference in the photosynthesis–nitrogen relationship: Patterns, physiological causes, and ecological importance. *J. Plant Res.* **2004**, *117*, 481–494. [CrossRef]

- 68. Grassi, G.; Bagnaresi, U. Foliar morphological and physiological plasticity in Picea abies and Abies alba saplings along a natural light gradient. *Tree Physiol.* **2001**, *21*, 959–967. [CrossRef]
- 69. Han, Q.; Kawasaki, T.; Nakano, T.; Chiba, Y. Leaf-age effects on seasonal variability in photosynthetic parameters and its relationships with leaf mass per area and leaf nitrogen concentration within a Pinus densiflora crown. *Tree Physiol.* **2008**, *28*, 551–558. [CrossRef]
- 70. Mashilo, J.; Shimelis, H.; Odindo, A. Correlation and path coefficient analyses of qualitative and quantitative traits in selected bottle gourd landraces. *Acta Agric. Scand. Sect. B Soil Plant Sci.* **2016**, *66*, 558–569. [CrossRef]
- Whitehead, D.; Walcroft, A.S.; Scott, N.A.; Townsend, J.A.; Trotter, C.M.; Rogers, G.N.D. Characteristics of photosynthesis and stomatal conductance in the shrubland species mānuka (Leptospermum scoparium) and kānuka (Kunzea ericoides) for the estimation of annual canopy carbon uptake. *Tree Physiol.* 2004, 24, 795–804. [CrossRef] [PubMed]
- 72. Evans, J.R.; Seemann, J.R. The allocation of protein nitrogen in the photosynthetic apparatus: Costs, consequences, and control. *Photosynthesis* **1989**, *8*, 183–205.
- 73. Anteb, N.P.R.; Hernandez, R.; Medina, E.M. The Photosynthetic Capacity and Leaf Nitrogen Concentration as Related to Light Regime in Shade Leaves of a Montane Tropical Forest Tree, Tetrochidium rubrivenium. *Funct. Ecol.* **1996**, *10*, 491. [CrossRef]
- 74. Novriyanti, E.; Watanabe, M.; Makoto, K.; Takeda, T.; Hashidoko, Y.; Koike, T. Photosynthetic nitrogen and water use efficiency of acacia and eucalypt seedlings as afforestation species. *Photosynthetica* **2012**, *50*, 273–281. [CrossRef]
- 75. Prieto, J.A.; Louarn, G.; Perez Peña, J.; Ojeda, H.; Simonneau, T.; Lebon, E. A leaf gas exchange model that accounts for intra-canopy variability by considering leaf nitrogen content and local acclimation to radiation in grapevine (Vitis vinifera L.). *Plant Cell Environ.* **2012**, *35*, 1313–1328. [CrossRef]
- 76. Schultz, H.R. Extension of a Farquhar model for limitations of leaf photosynthesis induced by light environment, phenology and leaf age in grapevines (Vitis vinifera L. cvv. White Riesling and Zinfandel). *Funct. Plant Biol.* **2003**, *30*, 673–687. [CrossRef]
- 77. Ollinger, S.V.; Reich, P.B.; Frolking, S.; Lepine, L.C.; Hollinger, D.Y.; Richardson, A.D. Nitrogen cycling, forest canopy reflectance, and emergent properties of ecosystems. *Proc. Natl. Acad. Sci. USA* 2013, 110, E2437. [CrossRef]
- 78. Knyazikhin, Y.; Schull, M.A.; Stenberg, P.; Mõttus, M.; Rautiainen, M.; Yang, Y.; Marshak, A.; Carmona, P.L.; Kaufmann, R.K.; Lewis, P.; et al. Hyperspectral remote sensing of foliar nitrogen content. *Proc. Natl. Acad. Sci. USA* 2013, *110*, E185–E192. [CrossRef]
- 79. Townsend, P.A.; Serbin, S.P.; Kruger, E.L.; Gamon, J.A. Disentangling the contribution of biological and physical properties of leaves and canopies in imaging spectroscopy data. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, E1074. [CrossRef]
- 80. Yang, H.; Yang, X.; Heskel, M.; Sun, S.; Tang, J. Seasonal variations of leaf and canopy properties tracked by ground-based NDVI imagery in a temperate forest. *Sci. Rep.* **2017**, *7*, 1–10. [CrossRef]
- 81. Mõttus, M.; Takala, T.L.H.; Stenberg, P.; Knyazikhin, Y.; Yang, B.; Nilson, T. Diffuse sky radiation influences the relationship between canopy PRI and shadow fraction. *ISPRS J. Photogramm. Remote Sens.* **2015**, *105*, 54–60. [CrossRef]
- 82. Domingues, T.F.; Martinelli, L.A.; Ehleringer, J.R. Ecophysiological traits of plant functional groups in forest and pasture ecosystems from eastern Amazônia, Brazil. *Plant Ecol.* **2007**, *193*, 101–112. [CrossRef]



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