

Article

# Removal of Composite Restoration from the Root Surface in the Cervical Region Using Er: YAG Laser and Drill—In Vitro Study

Wojciech Zakrzewski <sup>1</sup>, Maciej Dobrzynski <sup>2</sup>, Piotr Kuropka <sup>3</sup>, Jacek Matys <sup>4</sup>,  
Malgorzata Malecka <sup>5</sup>, Jan Kiryk <sup>6</sup>, Zbigniew Rybak <sup>1</sup>, Marzena Dominiak <sup>6</sup>,  
Kinga Grzech-Lesniak <sup>6</sup>, Katarzyna Wiglusz <sup>7</sup> and Rafal J. Wiglusz <sup>5,\*</sup>

<sup>1</sup> Department of Experimental Surgery and Biomaterial Research, Wrocław Medical University, Bujwida 44, 50-345 Wrocław, Poland; wojciech.zakrzewski@student.umed.wroc.pl (W.Z.); zbigniew.rybak@umed.wroc.pl (Z.R.)

<sup>2</sup> Department of Conservative Dentistry and Pedodontics, Wrocław Medical University, Krakowska 26, 50-425 Wrocław, Poland; maciej.dobrzynski@umed.wroc.pl

<sup>3</sup> Department of Histology and Embriology, Wrocław University of Environmental and Life Sciences, Norwida 31, 50-375 Wrocław, Poland; piotr.kuropka@upwr.edu.pl

<sup>4</sup> Laser Laboratory at Dental Surgery Department, Wrocław Medical University, Krakowska 26, 50-425 Wrocław, Poland; jacek.matys@wp.pl

<sup>5</sup> Institute of Low Temperature and Structure Research, Polish Academy of Sciences, Okolna 2, 50-422 Wrocław, Poland; m.malecka@intibs.pl

<sup>6</sup> Dental Surgery Department, Medical University of Wrocław, 50-425 Wrocław, Poland; jan.kiryk@umed.wroc.pl (J.K.); marzena.dominiak@wp.pl (M.D.); kinga.grzech-lesniak@umed.wroc.pl (K.G.-L.)

<sup>7</sup> Faculty of Pharmacy, Wrocław Medical University, Borowska 211 A, 50-566 Wrocław, Poland; katarzyna.wiglusz@umed.wroc.pl

\* Correspondence: r.wiglusz@intibs.pl; Tel.: +48-71-3954-159; Fax: +48-71-344-10-29

Received: 20 May 2020; Accepted: 3 July 2020; Published: 7 July 2020



**Abstract:** Background: Recently, the defects of the tooth surface in the cervical region are often restored using composite filling materials. It should meet the needs of the patients regarding esthetics and material stability. The aim of the study was to analyze the tooth root surface at the cervical region after the removal of the composite filling material by means of the Erbium-doped Yttrium Aluminium Garnet (Er: YAG) laser or drill using the scanning electron microscopy (SEM) and fluorescence microscopy. Materials and Methods: For the purposes of this study, 14 premolar teeth ( $n = 14$ ) were removed due to orthodontic reasons. The rectangular shape cavities with 3 mm in width and 1.5 mm in height were prepared with a 0.8 mm bur on high-speed contra-angle in the tooth surface just below cemento-enamel junction (CEJ) and filled with the composite material. The composite material was removed with the Er: YAG laser at a power of 3.4 W, energy 170 mJ, frequency 20 Hz, pulse duration 300  $\mu$ s, tip diameter 0.8 mm, air/fluid cooling 3 mL/s, and time of irradiation: 6 sec, at a distance from teeth of 2 mm (G1 group,  $n = 7$ ) or a high-speed contra-angle bur (G2 group,  $n = 7$ ). After the removal of composite material, the surfaces of teeth were examined using the scanning electron microscopy (SEM) and fluorescence microscopy. Results: The Er: YAG irradiation allowed to remove completely the composite material from the tooth cavity. The study confirmed, that the ends of collagen fibers were only partially denatured after the Er: YAG laser application. Conclusion: It has been proved that using the Er: YAG laser is an effective and safe method of composite removal for the dentin surface.

**Keywords:** cervical root surface; Er: YAG laser; laser ablation; smear layer; vaporization

## 1. Introduction

The progress that has been made in the recent years in the field of dentistry enables more proficient and safer work, as well as a predictable treatment outcomes [1–3]. One of the brand-new technologies is a laser that is becoming more and more popular in dental clinics and supports traditional forms of treatment while replacing classic technologies [4–9]. The Erbium-doped Yttrium Aluminium Garnet (Er: YAG) laser presents several advantages over a conventional bur preparation [10]. Conventional bur preparation is a source of bone-conducted noise and vibration that can cause a painful sensation [11]. One of the essential advantages of the laser therapy is a lack of vibrations. Therefore, the local anesthesia is reduced or not needed [12]. When compared to drill during the osteotomy procedure, Pandurić, et al. [13] showed, that the Er: YAG laser produced preparations with regular and sharp edges, without bone fragments and debris, in a shorter preparation time. Laser irradiates rough surfaces, turning them into clean surfaces with opened dentin tubules and without smear layer [14]. This procedure is crucial and enables a complete and exact removal of filling material penetrating dentin tubules, which is impossible to achieve with conventional bur [15].

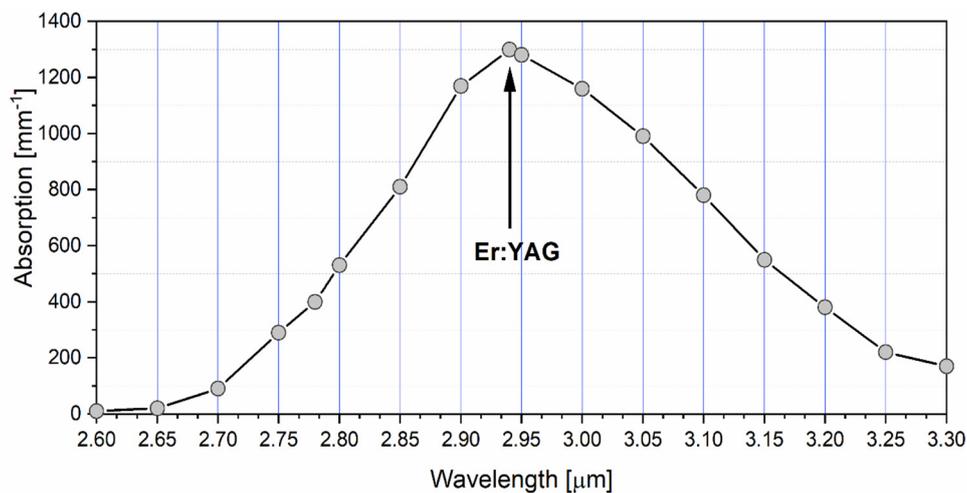
The scientific literature confirmed that both the scope of pulp carbonization and the pulp irritation during irradiation by the Er: YAG laser are lower in comparison with the use of micro-motor and turbine at the same conditions of air/water spray [16–19]. Its wavelength is in the mid-infrared region of the electromagnetic spectrum, so it is readably absorbed by water particles [20]. Special care should be paid to the prevention of thermal damage to the dental pulp with high-power lasers [7,21,22]. High absorption of the Er: YAG laser in water results in its shallow penetration depth [23]. The high absorption of 2940 nm wavelength and its low penetration depth enable the vaporization of the tooth tissues without pulp overheating. For instance, an intrapulpal rise of 5.5 °C caused pulpitis or pulp necrosis in 15% of irradiated teeth [21]. Temperature rise by 10 °C on the external tooth surfaces caused a bone resorption and root ankylosis [22]. Therefore, the application of high-power lasers requires the determination of parameters and a proper treatment protocol in order to limit the thermal damage and to assure the safe composite filling removal expected.

The removal of the composite material using a drill leads to leaving a remnant particles inside the dentinal tubules. It results in the formation of a smear layer which is removed together with the composite when a laser is used during treatment. The smear layer is an amorphous irregular layer formed during biomechanical preparation [24]. According to the scientific literature the Er: YAG laser showed high effectiveness in the removal of composite material from both enamel and dentin surface without the creation of the smear layer [25–27]. Moreover, Almeida et al. obtained shorter time required for the composite resin ablation after brackets debonding in comparison with a conventional drill [25]. Furthermore, another study, in which the Er: YAG laser was used for caries treatment, highlighted that its small spot diameter of 1 mm enabled more detailed and selective ablation while preserving the surrounding tissues [28].

After the composite material removal from the cervical tooth area with a dental bur is crucial to remove the remaining smear layer, which can impede the success of the gingival recession coverage [29,30]. In such a scenario, the laser enables immediate surgical recession coverage thanks to its ability to remove the composite material without the smear layer formation [31]. In comparison, a drill can only remove the composite material, leaving the smear layer intact, which results in a lack of adhesion of gingival tissue over the cavity [32]. It is known that with the Er: YAG laser the dental composite is removed very carefully, especially from the tooth cavity. It could enable better integration of new material, e.g., with the adjacent mucous membrane of exposed cervical areas of the teeth. Frequently, dentists artificially extend the clinical crowns by applying a layer of composites to the root surface instead of providing an appropriate periodontal treatment [33]. The gingiva lacks the ability of attachment to the composite material, which may eventually lead to a gingival recession [34]. Therefore, in order to remove the filling material either a laser or a drill is used [35].

In the case of a hard-tissue in dentistry, a wavelength of a laser plays a key role. The Er: YAG laser wavelength is most suitable for the hard-tissue ablation treatment because it is operated in

the region of the largest absorption peak for water (see Figure 1). Moreover, depending on different water content levels in human dentine and enamel, the absorption coefficients for the Er: YAG lasers are approximately  $150 \text{ mm}^{-1}$  in enamel, and  $200 \text{ mm}^{-1}$  in dentine. Thus, the Er: YAG laser wavelength penetrates approximately 7 micrometers in enamel and 5 micrometers in dentine.



**Figure 1.** The absorption curve of water in the middle infrared region. The position of Erbium-doped Yttrium Aluminium Garnet (Er: YAG) laser ( $2.94 \mu\text{m}$ ) used for hard-tissue ablation has been shown in the plot (based on Handbook of optical materials) [36].

The Er: YAG laser vaporization leads to the thermal rise of the irradiated tissues. Therefore, the aim of the study was to assess the changes in the dentinal surface of the root after the composite removal using  $2940 \text{ nm}$  wavelength by using SEM and fluorescence microscopy. Also, the roughness of the root surface was evaluated with the same methods in comparison with the same procedure performed with the dental drill.

## 2. Materials and Methods

### 2.1. Sample Preparation

For this scientific work, a total of 14 premolar teeth were removed because of orthodontic reasons. They were cleaned from the blood and debris, and rinsed with 0.9% saline solution and frozen in  $-20 \text{ }^{\circ}\text{C}$ . Then they were unfrozen and randomized to one of the study groups. The section material used in the procedures was acquired from the Dental Surgery Department of Wroclaw Medical University.

All the procedures were carried out in accordance with appropriate guidelines and regulations of the Republic of Poland as well as in accordance with the Declaration of Helsinki. The premolar teeth used in the present study were obtained with the consent of the owners and according to all ethical guidelines and requirements applicable in such cases. The experiment in this study was approved by the Ethics Committee of the Wroclaw Medical University (No. KB-132/2019).

### 2.2. Composite Restoration Procedure

The rectangular shape cavities with 3 mm in width and 1.5 mm in height were prepared with a 0.8 mm high-speed, contra-angle bur on the buccal surface of teeth just below the cemento-enamel junction (CEJ). Etching of the cavity's surface was performed within 15 s. OptiBond Solo Plus (Kerr, Italy) was used as a bonding agent, and curing time was 40 s. Then, cavity was filled with the composite material (Filtek Ultimate A3D, 3M, St. Paul, MN, USA) (see Figure 2). The depth of the cavities was up to 0.4 mm (half of the bur diameter).



**Figure 2.** The image of a model preparation of the root surface in the cervical region.

### 2.3. Composite Removal Techniques

#### 2.3.1. G1 Group (Er: YAG laser)

The study group ( $n = 7$ ) of composite filling materials was irradiated using the Er: YAG laser (LightWalker, Fotona, Ljubljana, Slovenia) with a wavelength of 2940 nm at a power of 3.4 W, energy 170 mJ, frequency 20 Hz, pulse duration 300  $\mu$ s, tip diameter 0.8 mm, air/fluid cooling 3 mL/s, and time of irradiation: 6 sec, at a distance of 2 mm from the tooth surface with an “S” shape movement (motion technique). Removal of the composites was performed accordingly to our previously described technique for orthodontics ceramic brackets debonding [37]. Additionally, filling material from the surrounding walls of the cavity was removed.

#### 2.3.2. G2 Group (Dental Bur)

The control group ( $n = 7$ ) of composite filling materials was treated with the use of a dental bur. The composite material was removed with a high-speed coarse diamond bur (MLX 534, no. 801, ISO 023, 150  $\mu$ m, Poldent, Warsaw, Poland) using a dental turbine and a water cooling. The remaining composite material was removed with low-speed fine diamond bur (F 514, no. 801, ISO 023, 45  $\mu$ m, Poldent, Warsaw, Poland) using micromotor and water cooling. The whole composite material was removed during the procedure in both groups. Moreover, it was carried out by the same, experienced operator.

### 2.4. Fluorescence Microscope Analysis

The test material after fixation was analyzed directly in a Nikon Eclipse 80i fluorescence microscope (Nikon, Tokyo, Japan) using a UV-2A filter (EX-330–380 nm, DM-400 nm, BF-420 nm) (Nikon, Tokyo, Japan). The magnification during an examination was 40 $\times$ .

### 2.5. Scanning Electron Microscopy

The collection of the teeth from both groups (laser and drill) were fixed in 2.5% glutaraldehyde using 7.4 phosphate buffer. Subsequently, the samples were rinsed in a phosphate buffer and then dehydrated in an acetone series (from 50–100%). The teeth were dried, mounted on the stubs and sputter-coated with graphite. Material testing was analyzed in a SEM Evo LS 15 (Zeiss, Oberkochen,

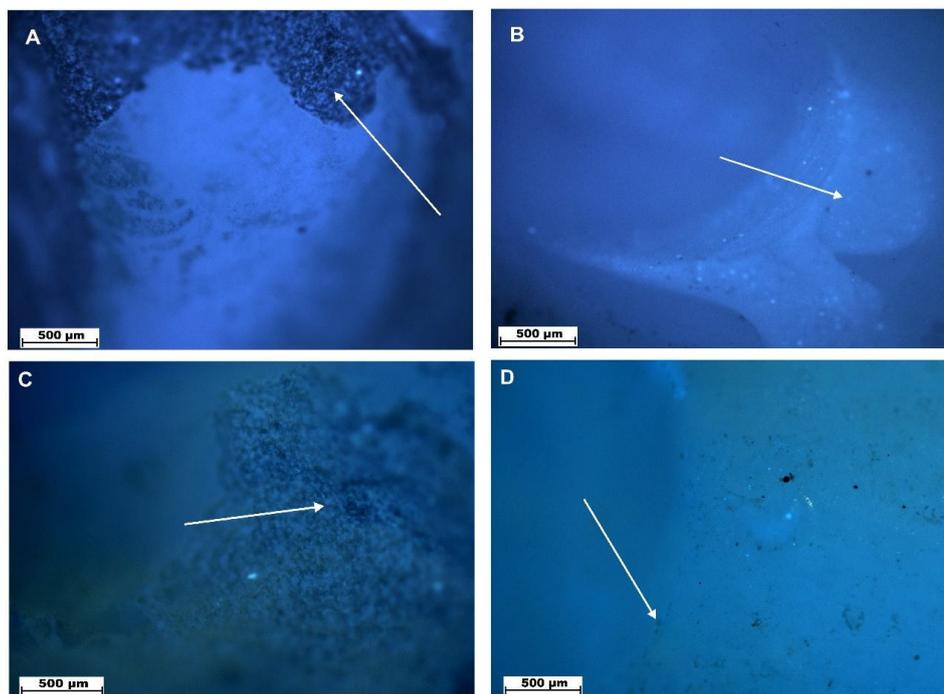
Germany). SEM settings during the examination of a surface modified by drill were as follows: WD 5.3 or 5.4 mm, 5.00 kV, spot 4.0, magnification 500× or 5000×, and 5.00 keV.

### 3. Results

The completion of the restoration removal in the both groups was verified by means of the SEM and fluorescence microscope inspection. The control group examination with the use of the fluorescence microscope and the scanning electron microscopy (SEM) showed an unstable arrangement of the fibers with a smear layer and residual composite material of the bottom of the cavity, while the study group analyzed with the use of the Er: YAG laser showed regular bottom surface of the cavity without composite residues and smear layer. The fluorescence microscope and SEM images showed the overall results related to the whole cavity surface. Similar dentin surface characteristics were also found in other samples of the studied groups (laser—G1 and drill—G2).

#### 3.1. Teeth Surface in a Fluorescence Microscope

Figure 3 has shown the visible differences in the surface of teeth after the laser (Figure 3A,C) and a drill treatment in the wall of prepared cavity (Figure 3B,D). A—The visible surface does not contain any residues of the filling material, while the dentin collagen fibers undergo a slight material melting, which makes the surface rough (arrow). B—residues of the filling material visible (arrow). C and D—bottom of the cavity. C—visible rough surface of the cavity bottom (arrow) that does not contain any residues after the treatment. D—visible small residues after the mechanical action (arrow), magnitude 40×. The similar image can be observed on all surfaces related to the prepared cavity.

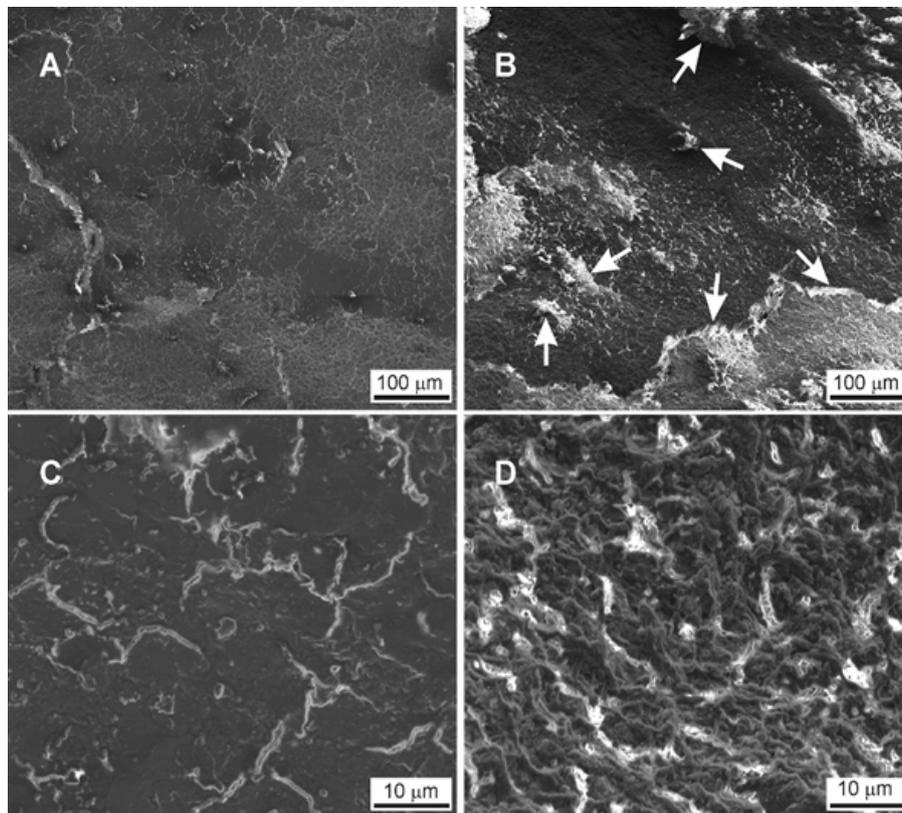


**Figure 3.** Surfaces of teeth after laser (A,C) and drill treatment (B,D) examined by fluorescence microscope, magnification 40×.

#### 3.2. Teeth Surface Analysis in a SEM Microscope

In Figure 4, the bottom of prepared cavity has been shown. In the pictures on the left are presented the results of the preparation with the use of the drill (Figure 4A,C) and on the right with the use of the laser (Figure 4B,D). The effect of mechanical action on dentin is visible. The dentine collagen fibres are partially detached from the matrix and form numerous loose remains lying on the surface

of the bottom of the prepared cavity. The arrangement of the fibres is unstable, which results in the foundation for new collagen fibres growing in the cavity. In addition, there are numerous small pieces of a composite material in the cavity (white arrow). In the case of a laser, the ends of collagen fibres are partially melted, creating a homogeneous, rough surface, a similar image can be observed on all the surfaces of the cavity prepared.



**Figure 4.** Analysis of tooth surface after laser (A,C) and drill (B,D) preparation.

#### 4. Discussion

The success of the composite removal from the root surface in the cervical region of the tooth could be a complex problem, especially in case of the patients with the gingival recessions. The main aim of the study was to assess the changes in the dentinal surface of the root after the composite removal using the Er: YAG laser and/or a dental bur with the help of SEM and fluorescence microscopies. Both analyses showed an unstable arrangement of the fibers with a smear layer and residual composite material on the bottom of the cavity in the control group (dental bur). Moreover, the study group with the Er: YAG laser showed the regular bottom surface of the cavity without a composite debris and a smear layer. However, it has been found that denatured/melted collagen fiber ends the partially closed dentinal tubules. Surfaces treated by the lasers appear rough and free from harmful residues, lipopolysaccharides that may interfere with the adhesion of connective tissue cells that might be useful for root conditioning in periodontal therapy [15,38].

The evaluation of root dentin surface after the composite material removal by a dental bur on a high-speed contra-angle handpiece showed a residual composite resin with the formation of a thick hybrid layer. The appearance and the dimension of the resin tags observed in our present study are similar to other findings [39,40]. The use of a diamond dental bur evoked a presence of a smear layer that penetrated the dentinal tubules up to around 5 micrometers, which formed smear plugs. The crucial procedure before the tooth restoration or recession coverage procedure is to remove the smear layer in order to open dentinal tubules and establish a highly adhesive structure of the tooth surface [41]. This

procedure forces the clinicians to use a phosphoric acid EDTA (EthyleneDiamineTetraacetic Acid) to open the dentinal tubules that can irritate the pulp [42,43].

The Erbium-doped yttrium aluminum garnet (Er: YAG) laser is an effective tool that can be used not only for the cavity preparation [44,45], but also for composite filling removal due to its selectivity and effectivity [46,47]—the effect of the Er: YAG laser.

YAG laser on composite filling and the dentin lying underneath is crucial. The pattern of a dentin surface after the laser application is strictly related to the amount of water component in different parts of the root dentin [37,48,49]. The 2940 nm wavelength is characterized by the highest absorption coefficient (absorption peak) in water [50]. This feature influences a variable speed of dental root vaporization in a specific part of the dentin. The intertubular dentin has a more considerable amount of water than the peritubular zone [51]. This causes a faster ablation of the intertubular dentin and increases the roughness of its surface after the Er: YAG laser application [48,49].

A disadvantage of the Er: YAG laser ablation is an increase in the temperature which can melt or carbonize the dentin. In our present study, the melting of the dentin was insignificant, however, we found a slight fusion of the ends of collagen fibers. That process caused a partial blockage of dentin canals that can disturb the adhesion of filling materials or interrupt the formation of new clinical attachment after the periodontal recession coverage [52,53]. In order to solve this problem as well as open dentinal tubules as well as to increase its adhesion, the application of sodium subchloride was recommended by many authors [53–55]. Furthermore, any residues left in the cavity will result in inefficient adhesion [29] of gingival tissue to the tooth's surface. In that case, the filling removal and the cavity preparation with the use of drill is not enough to successfully finish the treatment of patient with such recessions. Our study showed that the application of the Er: YAG laser resulted in the regular bottom surface of the cavity without the composite residues after the filling material removal. Our results were similar to the study of Almeida et al. [25] who confirmed significantly better effectiveness of the Er: YAG laser than the conventional technique for removing the composite remnants. Nevertheless, contrary to our findings, the study of Correa-Afonso et al. [46] showed that the Er: YAG laser at 250 mJ, 2–10 Hz presented a higher amounts of the remaining restorative material. The Er: YAG laser application with a scanning technique was utilized in our present research at a lower energy (170 mJ) and higher frequency (20 Hz) at a distance of 2 mm, enabling the removal of the composite restoration completely in a way which is safer for the pulp.

The safeness of the dental pulp is of supreme value during high power laser irradiation [37]. We irradiated the composite filling without contact with a water spray cooling. However, others recommended laser-assisted composite vaporization by placing the laser tip perpendicular to the material [56]. To avoid the injury of the laser tip or mirror, which may occur during perpendicular laser irradiation due to the reflection of the beam, it can be recommended to use the Er: YAG laser from a slight distance of at least 1–2 mm with minimal angulation of the laser tip [37]. Within the restrictions of this ex vivo study, the results suggest that it is safe to use the Er: YAG laser during the composite vaporization employing our present method described. Considering the limitations of the in vitro experiment additional studies to confirm the results of the research in a human model for a different tooth cavities dimension are needed.

## 5. Conclusions

This research proves the superiority of the Er: YAG laser over the drill use in case of the composite filling removal. The Er: YAG laser application showed the regular bottom surface of the cavity without composite debris and smear layer. In contrast, the use of dental bur for composite material removal showed an unstable arrangement of the fibers with a smear layer and residual composite material on the bottom of the cavity. Moreover, the preparation with the use of the dental bur resulted in the formation of an uneven cavity surface with the composite residues.

**Author Contributions:** Conceptualization, methodology, software, formal analysis, investigation, writing-original draft preparation, review and editing, supervision, project administration, funding acquisition, R.J.W. and M.D.

(Maciej Dobrzynski); data curation, investigation, resources, writing-original draft preparation, W.Z., K.W. and Z.R.; software; validation; formal analysis; investigation; resources; writing-original draft preparation, M.D. (Marzena Dominiak), P.K., J.M., J.K., K.G.-L., and M.M. All authors have read and agreed to the published version of the manuscript.

**Funding:** Financial support of the National Science Centre in the course of realization of the Projects “Preparation and characterization of biocomposites based on nanoapatites for theranostics” (no. UMO-2015/19/B/ST5/01330) and “Preparation and modulation of spectroscopic properties of YXZO<sub>4</sub>, where X and Z - P<sup>5+</sup>, V<sup>5+</sup>, As<sup>5+</sup>, doped with “s<sup>2</sup>-like” ions and co-doped with rare earth ions” (no. UMO-2019/33/B/ST5/02247) are gratefully acknowledge.

**Acknowledgments:** The authors would like to thank K. Rola for SEM measurements.

**Conflicts of Interest:** The authors declare that there is no conflict of interests regarding the publication of this paper.

## References

1. Verma, S.; Chaudhari, P.; Maheshwari, S.; Singh, R. Laser in dentistry: An innovative tool in modern dental practice. *Natl. J. Maxillofac Surg.* **2012**, *3*, 124. [[CrossRef](#)] [[PubMed](#)]
2. Grzech-Leśniak, K.; Nowicka, J.; Pajczkowska, M.; Matys, J.; Szymonowicz, M.; Kuropka, P.; Rybak, Z.; Dobrzyński, M.; Dominiak, M. Effects of Nd:YAG Laser Irradiation on the Growth of *Candida Albicans* and *Streptococcus Mutans*: In Vitro Study. *Lasers Med. Sci.* **2019**, *34*, 129–137. [[CrossRef](#)]
3. Coluzzi, D.J. Fundamentals of dental lasers: Science and instruments. *Dent. Clin. N. Am.* **2004**, *48*, 751–770. [[CrossRef](#)]
4. Matys, J.; Świder, K.; Flieger, R. Laser instant implant impression method: A case presentation. *Dent. Med. Probl.* **2017**, *54*, 110–116. [[CrossRef](#)]
5. Perveen, A.; Molardi, C.; Fornaini, C. Applications of laser welding in dentistry: A state-of-the-art review. *Micromachines* **2018**, *9*, 209. [[CrossRef](#)]
6. Luke, A.M.; Mathew, S.; Altawash, M.M.; Madan, B.M. Lasers: A review with their applications in oral medicine. *J. Lasers Med. Sci.* **2019**, *10*, 324–329. [[CrossRef](#)] [[PubMed](#)]
7. Matys, J.; Grzech-Leśniak, K.; Flieger, R.; Dominiak, M. Assessment of an impact of a diode laser mode with wavelength of 980 nm on a temperature rise measured by means of k-02 thermocouple: Preliminary results. *Dent. Med. Probl.* **2016**, *53*. [[CrossRef](#)]
8. Walsh, L.J. The current status of laser applications in dentistry. *Aust. Dent. J.* **2003**, *48*, 146–155. [[CrossRef](#)] [[PubMed](#)]
9. Martens, L.C. Laser physics and a review of laser applications in dentistry for children. *Eur. Arch. Paediatr. Dent.* **2011**, *12*, 61–67. [[CrossRef](#)]
10. Matys, J.; Hadzik, J.; Dominiak, M. Schneiderian Membrane Perforation Rate and Increase in Bone Temperature During Maxillary Sinus Floor Elevation by Means of Er. *Implant Dent.* **2017**, *26*, 238–244. [[CrossRef](#)]
11. Ozaki, M.; Baba, A.; Ishii, K.; Takagi, H.; Motokawa, W. Measurement of bone conduction characteristics for transmitted vibration sounds of tooth drilling. *Pediatr. Dent. J.* **2007**, *17*, 148–155. [[CrossRef](#)]
12. Takamori, K.; Furukawa, H.; Morikawa, Y.; Katayama, T.; Watanabe, S. Basic study on vibrations during tooth preparations caused by high-speed drilling and Er:YAG laser irradiation. *Lasers Surg. Med.* **2003**, *32*, 25–31. [[CrossRef](#)] [[PubMed](#)]
13. Pandurić, D.G.; Bago, I.; Katanec, D.; Žabkar, J.; Miletić, I.; Anić, I. Comparison of Er:YAG Laser and Surgical Drill for Osteotomy in Oral Surgery: An Experimental Study. *J. Oral Maxillofac. Surg.* **2012**, *70*, 2515–2521. [[CrossRef](#)] [[PubMed](#)]
14. Korkut, E.; Torlak, E.; Gezgin, O.; Özer, H.; Şener, Y. Antibacterial and Smear Layer Removal Efficacy of Er:YAG Laser Irradiation by Photon-Induced Photoacoustic Streaming in Primary Molar Root Canals: A Preliminary Study. *Photomed. Laser Surg.* **2018**, *36*, 480–486. [[CrossRef](#)]
15. Yazici, A.R.; Baseren, M.; Gorucu, J. Clinical Comparison of Bur- and Laser-prepared Minimally Invasive Occlusal Resin Composite Restorations: Two-year Follow-up. *Oper. Dent.* **2010**, *35*, 500–507. [[CrossRef](#)]
16. Fornaini, C. Er:YAG and adhesion in conservative dentistry: Clinical overview. *Laser Ther.* **2013**, *22*, 31–35. [[CrossRef](#)]

17. Zeitouni, J.; Clough, B.; Zeitouni, S.; Saleem, M.; Al Aisami, K.; Gregory, C. The effects of the Er:YAG laser on trabecular bone micro-architecture: Comparison with conventional dental drilling by micro-computed tomographic and histological techniques. *F1000Research* **2017**, *6*, 1133. [[CrossRef](#)]
18. DenBesten, P.K.; White, J.M.; Pelino, J.E.P.; Furnish, G.; Silveira, A.; Parkins, F.M. The safety and effectiveness of an Er:YAG laser for caries removal and cavity preparation in children. *Med. Laser Appl.* **2001**, *16*, 215–222. [[CrossRef](#)]
19. Sonntag, K.D.; Klitzman, B.; Burkes, E.J.; Hoke, J.; Moshonov, J. Pulpal response to cavity preparation with the Er:YAG and Mark III free electron lasers. *Oral Surg. Oral Med. Oral Pathol. Oral Radiol. Endod.* **1996**, *81*, 695–702. [[CrossRef](#)]
20. Featherstone, J.D.B.; Fried, D. Fundamental Interactions of Lasers with Dental Hard Tissues. *Med. Laser Appl.* **2001**, *16*, 181–194. [[CrossRef](#)]
21. Zach, L.; Cohen, G. Pulp response to externally applied heat. *Oral Surg. Oral Med. Oral Pathol.* **1965**, *19*, 515–530. [[CrossRef](#)]
22. Eriksson, A.R.; Albrektsson, T. Temperature threshold levels for heat-induced bone tissue injury: A vital-microscopic study in the rabbit. *J. Prosthet. Dent.* **1983**, *50*, 101–107. [[CrossRef](#)]
23. Apel, C.; Meister, J.; Ioana, R.S.; Franzen, R.; Hering, P.; Gutknecht, N. The Ablation Threshold of Er:YAG and Er:YSGG Laser Radiation in Dental Enamel. *Lasers Med. Sci.* **2002**, *17*, 246–252. [[CrossRef](#)] [[PubMed](#)]
24. Mirseifinejad, R.; Tabrizzade, M.; Davari, A.; Mehravar, F. Efficacy of Different Root Canal Irrigants on Smear Layer Removal after Post Space Preparation: A Scanning Electron Microscopy Evaluation. *Iran. Endod. J.* **2017**, *12*, 185–190. [[PubMed](#)]
25. Almeida, H.C.; Vedovello Filho, M.; Vedovello, S.A.S.; Young, A.A.A.; Ramirez-Yañez, G.O. Er:YAG laser for composite removal after bracket debonding: A qualitative SEM analysis. *Int. J. Orthod. Milwaukee.* **2009**, *20*, 9–13.
26. Dumore, T.; Fried, D. Selective ablation of orthodontic composite by using sub-microsecond IR laser pulses with optical feedback. *Lasers Surg. Med.* **2000**, *27*, 103–110. [[CrossRef](#)]
27. Lizarelli, R.D.F.Z.; Moriyama, L.T.; Bagnato, V.S. Ablation of composite resins using Er:YAG laser—Comparison with enamel and dentin. *Lasers Surg. Med.* **2003**, *33*, 132–139. [[CrossRef](#)]
28. Kornblit, R.; Trapani, D.; Bossù, M.; Muller-Bolla, M.; Rocca, J.P.; Polimeni, A. The use of Erbium:YAG laser for caries removal in paediatric patients following Minimally Invasive Dentistry concepts. *Eur. J. Paediatr. Dent.* **2008**, *9*, 81–87.
29. Blomlöf, J.P.; Blomlöf, L.B.; Lindskog, S.F. Smear removal and collagen exposure after non-surgical root planing followed by etching with an EDTA gel preparation. *J. Periodontol.* **1996**, *67*, 841–845. [[CrossRef](#)]
30. Dilsiz, A.; Aydin, T.; Yavuz, M.S. Root surface biomodification with an Er:YAG laser for the treatment of gingival recession with subepithelial connective tissue grafts. *Photomed. Laser Surg.* **2010**, *28*, 511–517. [[CrossRef](#)]
31. Wang, X.; Cheng, X.; Liu, B.; Liu, X.; Yu, Q.; He, W. Effect of Laser-Activated Irrigations on Smear Layer Removal from the Root Canal Wall. *Photomed. Laser Surg.* **2017**, *35*, 688–694. [[CrossRef](#)] [[PubMed](#)]
32. Tabrizzadeh, M.; Shareghi, A. The Effect of Preparation Size on Efficacy of Smear Layer Removal; A Scanning Electron Microscopic Study. *Iran. Endod. J.* **2015**, *10*, 169–173. [[PubMed](#)]
33. Wahbi, M.A.; Al Sharief, H.S.; Tayeb, H.; Bokhari, A. Minimally invasive use of coloured composite resin in aesthetic restoration of periodontally involved teeth: Case report. *Saudi Dent. J.* **2013**, *25*, 83–89. [[CrossRef](#)] [[PubMed](#)]
34. Khier, S.; Hassan, K. Efficacy of composite restorative techniques in marginal sealing of extended class v cavities. *ISRN Dent.* **2011**, *2011*, 180197. [[CrossRef](#)]
35. Chiniforush, N.; Morshedi, E.; Torabi, S.; Arami, S.; Shahabi, S.; Tabatabaie, M. Assessing microleakage of composite restorations in class V cavities prepared by Er:YAG laser irradiation or diamond bur. *J. Conserv. Dent.* **2014**, *17*, 216. [[CrossRef](#)]
36. Weber, M. *Handbook of Optical Materials*; CRC Press: Boca Raton, FL, USA, 2002.
37. Grzech-Leśniak, K.; Matys, J.; Zmuda-Stawowiak, D.; Mroczka, K.; Dominiak, M.; Brugnera, A.; Gruber, R.; Romanos, G.E.; Sculean, A. Er:YAG Laser for Metal and Ceramic Bracket Debonding: An In Vitro Study on Intrapulpal Temperature, SEM, and EDS Analysis. *Photomed. Laser Surg.* **2018**, *36*, 595–600. [[CrossRef](#)]

38. Yamaguchi, H.; Kobayashi, K.; Osada, R.; Sakuraba, E.I.; Nomura, T.; Arai, T.; Nakamura, J. Effects of irradiation of an Erbium: YAG laser on root surfaces. *J. Periodontol.* **1997**, *68*, 1151–1155. [[CrossRef](#)]
39. Prati, C.; Chersoni, S.; Mongiorgi, R.; Pashley, D.H. Resin-infiltrated dentin layer formation of new bonding systems. *Oper. Dent.* **1998**, *23*, 185–194.
40. Perdigão, J.; Ramos, J.C.; Lambrechts, P. In vitro interfacial relationship between human dentin and one-bottle dental adhesives. *Dent. Mater.* **1997**, *13*, 218–227. [[CrossRef](#)]
41. Perdigão, J. Dentin bonding-Variables related to the clinical situation and the substrate treatment. *Dent. Mater.* **2010**, *26*. [[CrossRef](#)]
42. Gilpatrick, R.O.; Johnson, W.; Moore, D.; Turner, J. Pulpal response to dentin etched with 10% phosphoric acid. *Am. J. Dent.* **1996**, *9*, 125–129. [[PubMed](#)]
43. Gonçalves, L.F.; Fernandes, A.P.; Cosme-Silva, L.; Colombo, F.A.; Martins, N.S.; Oliveira, T.M.; Araujo, T.H.; Sakai, V.T. Effect of EDTA on TGF- $\beta$ 1 released from the dentin matrix and its influence on dental pulp stem cell migration. *Braz. Oral Res.* **2016**, *30*, e131. [[CrossRef](#)] [[PubMed](#)]
44. Buyukhatipoglu, I.; Secilmis, A. The use of Erbium: Yttrium-aluminum-garnet laser in cavity preparation and surface treatment: 3-year follow-up. *Eur. J. Dent.* **2015**, *9*, 284–287. [[CrossRef](#)] [[PubMed](#)]
45. Al-Batayneh, O.B.; Seow, W.K.; Walsh, L.J. Assessment of Er:YAG laser for cavity preparation in primary and permanent teeth: A scanning electron microscopy and thermographic study. *Pediatr. Dent.* **2014**, *36*, 90–94.
46. Correa-Afonso, A.M.; Palma-Dibb, R.G.; Pécora, J.D. Composite filling removal with erbium:yttrium-aluminium-garnet laser: Morphological analyses. *Lasers Med. Sci.* **2010**, *25*, 1–7. [[CrossRef](#)]
47. Fried, W.A.; Chan, K.H.; Darling, C.L.; Fried, D. Use of a DPSS Er:YAG laser for the selective removal of composite from tooth surfaces. *Biomed. Opt. Express* **2018**, *9*, 5026. [[CrossRef](#)]
48. Bertrand, M.F.; Hessleyer, D.; Muller-Bolla, M.; Nammour, S.; Rocca, J.P. Scanning electron microscopic evaluation of resin-dentin interface after Er:YAG laser preparation. *Lasers Surg. Med.* **2004**, *35*, 51–57. [[CrossRef](#)]
49. Sasaki, L.H.; Lobo, P.D.C.; Moriyama, Y.; Watanabe, I.-S.; Villaverde, A.B.; Tanaka, C.S.-I.; Moriyama, E.H.; Brugnera, A., Jr. Tensile bond strength and SEM analysis of enamel etched with Er:YAG laser and phosphoric acid: A comparative study In vitro. *Braz. Dent. J.* **2008**, *19*, 57–61. [[CrossRef](#)]
50. Matys, J.; Flieger, R.; Dominiak, M. Assessment of Temperature Rise and Time of Alveolar Ridge Splitting by Means of Er:YAG Laser, Piezosurgery, and Surgical Saw: An Ex Vivo Study. *Biomed. Res. Int.* **2016**, *2016*, 9654975. [[CrossRef](#)]
51. Berkovitz, B.; Moxham, B.; Linden, R.; Sloan, A. Master Dentistry Volume 3 Oral Biology E-Book: Oral Anatomy, Histology. Available online: [https://books.google.pl/books?id=Slcpvee98VAC&pg=PA163&lpg=PA163&dq=The+intertubular+dentin+has+a+more+considerable+amount+of+water+than+the+peritubular+zone&source=bl&ots=MAImWtbxrN&sig=ACfU3U1AtWbWszv5wqcFGPXqBOMJi8y\\_Q&hl=en&sa=X&ved=2ahUKewiSkeW7mq7oAhUFpIsKHYDBDZsQ6AEwCXoECAYQAQ#v=onepage&q=Theintertubulardentinhasa~moreconsiderableamountofwaterthanthe~peritubularzone&f=false](https://books.google.pl/books?id=Slcpvee98VAC&pg=PA163&lpg=PA163&dq=The+intertubular+dentin+has+a+more+considerable+amount+of+water+than+the+peritubular+zone&source=bl&ots=MAImWtbxrN&sig=ACfU3U1AtWbWszv5wqcFGPXqBOMJi8y_Q&hl=en&sa=X&ved=2ahUKewiSkeW7mq7oAhUFpIsKHYDBDZsQ6AEwCXoECAYQAQ#v=onepage&q=Theintertubulardentinhasa~moreconsiderableamountofwaterthanthe~peritubularzone&f=false) (accessed on 22 March 2020).
52. Dilsiz, A.; Aydin, T.; Canakci, V.; Cicek, Y. Root surface biomodification with Nd:YAG laser for the treatment of gingival recession with subepithelial connective tissue grafts. *Photomed. Laser Surg.* **2010**, *28*, 337–347. [[CrossRef](#)]
53. Bahrololoomi, Z.; Dadkhah, A.; Alemrajabi, M. The Effect of Er:YAG laser irradiation and different concentrations of sodium hypochlorite on shear bond strength of composite to primary teeth's dentin. *J. Lasers Med. Sci.* **2017**, *8*, 29–35. [[CrossRef](#)]
54. Lahmouzi, J.; Farache, M.; Umana, M.; Compere, P.; Nyssen-Behets, C.; Samir, N. Influence of sodium hypochlorite on Er:YAG Laser-irradiated dentin and its effect on the quality of adaptation of the composite restoration margins. *Photomed. Laser Surg.* **2012**, *30*, 655–662. [[CrossRef](#)]

55. Olivi, G.; Olivi, M. Lasers in Restorative Dentistry: A Practical Guide. Available online: [https://books.google.pl/books?id=h2B1CgAAQBAJ&pg=PA79&lpg=PA79&dq=sodium+hypochlorite+removal+of+melted+collagen+fiber+after+laser&source=bl&ots=wJgBeyLoDg&sig=ACfU3U1awK3pdxSzm8QMzAUqrCGbWv9t6A&hl=en&sa=X&ved=2ahUKEwiGsd3jm67oAhUj\\_SoKHXVqDisQ6AEwAHoECAkQAQ#v=onepage&q=sodiumhypochloriteremovalofmeltedcollagenfiberafterlaser&f=false](https://books.google.pl/books?id=h2B1CgAAQBAJ&pg=PA79&lpg=PA79&dq=sodium+hypochlorite+removal+of+melted+collagen+fiber+after+laser&source=bl&ots=wJgBeyLoDg&sig=ACfU3U1awK3pdxSzm8QMzAUqrCGbWv9t6A&hl=en&sa=X&ved=2ahUKEwiGsd3jm67oAhUj_SoKHXVqDisQ6AEwAHoECAkQAQ#v=onepage&q=sodiumhypochloriteremovalofmeltedcollagenfiberafterlaser&f=false) (accessed on 22 March 2020).
56. Oskoe, P.A.; Oskoe, S.S.; Rikhtegaran, S.; Pournaghi-Azar, F.; Gholizadeh, S.; Aleyasin, Y.; Kasraei, S. Effect of various laser surface treatments on repair shear bond strength of aged silorane-based composite. *J. Lasers Med. Sci.* **2017**, *8*, 186–190. [CrossRef]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).