



Article Enhanced Enzymatic Hydrolysis of Rice Straw Pretreated by Oxidants Assisted with Photocatalysis Technology

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Abstract: This work evaluated the effectiveness of rice straw pretreatment using a TiO₂/UV system in the presence of oxidants. The effects of TiO₂ concentrations, pH and photocatalysis time were investigated. Inorganic oxidants including H_2O_2 , $K_2S_2O_8$, and KIO₄ were added to further enhance the effect on enzymatic hydrolysis of rice straw. The TiO₂/UV/ H_2O_2 pretreatment showed a higher amount of released reducing sugar ($8.88 \pm 0.10 \text{ mg/mL}$, compared to $5.47 \pm 0.03 \text{ mg/mL}$ in untreated sample). Composition analyses of rice straw after the TiO₂/UV/ H_2O_2 pretreatment showed partial lignin and hemicellulose removal. Moreover, structural features of untreated and pretreated rice straw were analyzed through FE-SEM, FT-IR, and XRD. This work suggests that H_2O_2 is an efficient addition for photocatalysis pretreatment of rice straw.

Keywords: photocatalysis; oxidants; pretreatment; enzymatic hydrolysis

1. Introduction

Lignocellulosic biomass is an abundant and sustainable energy resource for producing of biofuels and valuable chemicals, such as bioethanol, biobutanol, xylitol and lactic acid [1]. The use of lignocellulosic biomass as an alternative fuel is considered one of the best solutions to the current energy crisis. Furthermore, utilizing lignocellulosic biomass reduces greenhouse gas emissions, promotes rural area economic development, and enhances energy security [2,3]. Despite these advantages, the biomass recalcitrance due to the cuticle and epicuticular waxes, structural heterogeneity, degree of lignification and crystalline cellulose of lignocellulosic biomass is a challenge in the search to obtain high bioethanol production [4,5]. Herein, finding a pretreatment that can break down biomass recalcitrance is the imperative for all forms and types of lignocellulosic biomass [6,7].

A suitable pretreatment technology should be both financially viable and effective [8]. It could also facilitate lignin or hemicellulose removal, disrupt hydrogen bonds in crystalline cellulose and increase enzymatic access to cellulose [7,9]. Among several pretreatments, photocatalytic oxidation has attracted significant attention for the depolymerization of lignin under mild conditions [10]. For a potential photocatalyst, there has been increasing interest in environmental applications of titanium dioxide (TiO₂) due to its commercial availability and chemical stability [11]. It is capable of indirect photolysis or heterogeneous photocatalysis when TiO₂ combines with ultraviolet (UV) light [12]. It is generally accepted that OH radical oxidation and electron–hole reaction occur due to the short wavelength of UV

light to complete the photolysis process [13], and show very fast reaction rates [14,15]. Several groups have used the pretreatment technology of TiO_2/UV to degrade organic contaminants [16,17], and the TiO_2/UV process can successfully pretreat paper mill effluent, black liquor, and olive mill waste water [10,18]. Moreover, some attempts have been made to depolymerize lignin with a simpler structure [11]. However, TiO_2 as a photocatalyst produces hydroxyl radicals that cannot be infinitely increased, and adding an excess amount of titanium dioxide alone will disperse the ultraviolet light in the solution.

To reduce the probability of recombing photogenerated electrons and holes, more holes and increasing hydroxyl radical concentration can be provided for oxidative degradation of organic contaminants. Some researchers have found that the presence of oxidants can promote the performance of TiO₂/UV process [19]. Gözmen [20] observed that increasing periodate ion concentration (1–5 mM) enhances the degradation of basic dyes in the TiO₂/UV system. Inorganic oxidants such as IO_4^- , BrO_3^- , $S_2O_8^{2-}$, and H_2O_2 can quench conduction band electrons and form reactive radical intermediates, so they are used as additives applied in TiO₂/UV [19].

Photocatalysis, are considered to have mild pretreatment conditions and provide an environmentally friendly alternative. The TiO_2/UV process has been successfully applied to pretreat organic species [10]. Related research has focused mainly on using the TiO_2/UV system to degrade commercial lignin powder and lignin solutions [11]. Compared with native biomass, commercial lignin has a less complicated structure. To the best of our knowledge, there have been very few studies based on native biomass, such as rice straw. This study utilized oxidants such as H_2O_2 , $K_2S_2O_8$ and KIO_4 as additives in a TiO_2/UV system. They have high polarizability so that they can capture the electrons ejected from TiO_2 [21] and are more receptive to price. This study: (i) investigated the effects of TiO_2 concentrations, pH, and photocatalysis time; (ii) determined the effectiveness of oxidants such as KIO_4 , $K_2S_2O_8$ and H_2O_2 as additives for rice straw pretreatment with a TiO_2/UV system; (iii) investigated oxidants can improve the pretreatment efficacy and how they function during the process; and (iv) investigated the optimal oxidant to achieve the desirable enzymatic hydrolysis efficiency.

2. Materials and Methods

2.1. Materials and Chemical Reagents

Rice straw was supplied by the Chinese Academy of Sciences (Xiamen, China). It was cut into small pieces, dried at 60 °C, and then ground and screened to 40 mesh size particles for further use. TiO₂ (99.9%) was purchased from Sigma Aldrich Co., Ltd. (St. Louis, MO, USA) The photocatalytic device was purchased from BiLon Co., Ltd. (Xian, China) The UV lamp (36W, BiLon Co., Ltd., Xian, China) with irradiation wavelength 254 nm. KIO₄, K₂S₂O₈, and H₂O₂ [30% (w/w)] were obtained from Guangzhou Chemical Reagent Factory (Guangzhou, China). All other reagents were analytical grade.

2.2. Photocatalysis Pretreatment

The rice straw powder was first dewaxed in acetone at 20 °C for 16 h [22]. The dewaxed rice straw was cleaned with deionized water and oven dried at 60 °C for 24 h. Sample preparation consisted of soaking 1.0 g rice straw in 40 mL TiO₂ solution. The initial pH (3, 4, 6, 7, 8, and 10) was adjusted by addition of appropriate amounts of HCl (1N) or NaOH (1N). TiO₂ concentrations were 0%, 0.05%, 0.25%, 0.50%, 1.0%, and 2.0% (w/v). The sample preparation was transferred into a 500-mL beaker and exposed to UV irradiation in a device at ambient temperature (30 ± 1 °C), with a magnetic stirrer. The UV lamp was positioned in a quartz glass vessel to maintain a uniform distribution of UV light during pretreatment. The UV lamp was in a fixed position in the center of the bottom of the beaker. UV lamp irradiation times ranged from 0 to 8 h (i.e., 0, 1, 2, 3, 4, 6 and 8 h).

2.3. Enzymatic Hydrolysis of Rice Straw

Untreated and pretreated samples were tested at a solid loading of 2.5% (w/v) in 25-mL conical flasks. Each sample enzymatic hydrolysis reaction contained 0.05 M sodium citrate buffer (pH = 4.8), 0.02% sodium azide, cellulase complex (Novozyme NS220086, 250 FPU mL⁻¹, Copenhagen, Denmark) at a loading of 50 FPU/g and β -glucosidase (Novozyme NS221118, 320 CBU mL⁻¹) at a loading of 40 CBU/g. The reactions were carried out in a horizontal shaker incubator (150 rpm, Jinghong Experimental Equipment Co., Ltd., Shanghai, China) for 48 h at 50 °C. Samples taken after 0 and 48 h of hydrolysis and centrifuged to remove the residue at 7000 rpm were analyzed for released reducing sugar by the dinitrosalicylic acid (DNS) method [23].

2.4. Addition of Oxidants in a TiO₂/UV System

For experiments with $TiO_2/UV/oxidants$, TiO_2 concentrations were 0.50% (w/v). Three types of oxidants were used only for rice straw pretreatment in the TiO_2/UV system: KIO_4 , $K_2S_2O_8$, and H_2O_2 (1–16 mM). The UV irradiation time was 3 h.

2.5. Characterization Analyses

The morphologies of untreated and pretreated rice straw were performed using JSM-7001F Field-Emission Scanning Electron Microscope (JEOL, Tokyo, Japan). The samples were fixed with conductive tape on brass sample stubs and then coated with Au before observation. Images were acquired at a 15-KV acceleration voltage and magnification \times 500.

The untreated and pretreated samples for Fourier Transform Infrared (FTIR) spectroscopic analysis were determined using a Nicolet 6700 spectrometer (Thermo Fisher Scientific, Waltham, MA, USA) over a wavenumber range from 500–4000 cm⁻¹ with 20 scans and a resolution of 4 cm⁻¹. The samples were dispersed of spectroscopic grade potassium bromide (KBr) and pressed into disks to determined.

X-ray diffraction analysis was tested using an Ultima X-ray diffractometer of Rigaku Corporation (40 KV, 20 mA, Tokyo, Japan) with Cu K α radiation. The 2 θ range was from 5° to 40° in steps of 0.02° at a rate of 8° min⁻¹ [24]. The crystallinity index (CrI) of untreated and pretreated samples was calculated according to Equation (1) [25].

$$\mathrm{CrI\%} = \frac{I_{002} - I_{am}}{I_{002}} \times 100\% \tag{1}$$

where I_{002} is the intensity of the crystalline regions ($2\theta = 22.5^{\circ}$) and I_{am} is the intensity of amorphous portion ($2\theta = 18.7^{\circ}$).

3. Results

3.1. Optimization of Conditions for Photocatalysis Pretreatment

The variables used in the pretreatment were TiO₂ concentrations, pH solution, and photocatalysis irradiation time. To optimize pretreatment conditions, the selected TiO₂ concentrations were 0%, 0.05%, 0.25%, 0.50%, 1.0%, and 2.0% (w/v). The effect of TiO₂ concentration on enzymatic hydrolysis of rice straw is illustrated in Figure 1a, in which the released amount of reducing sugar increased with the TiO₂ concentration from 0.05% to 1.0% (w/v). The highest released reducing sugar was 8.04 ± 0.15 mg/mL, observed at TiO₂ concentration of 1.0% (w/v). Higher TiO₂ concentration absorbed more energy during UV irradiation process. However, adding an excess amount of TiO₂ alone will disperse the ultraviolet light in the solution, markedly decreasing the enzymatic hydrolysis efficiency. Consequently, optimal TiO₂ concentration was 1.0% (w/v).



(c)

Figure 1. (a) Effect of TiO₂ concentration on enzymatic hydrolysis of rice straw; (b) effect of pH on enzymatic hydrolysis of rice straw; and (c) effect of photocatalysis time on enzymatic hydrolysis of rice straw. The different letters above the bars in figures indicate significantly difference ($p \le 0.05$, n = 3).

The solution pH has a significant effect on photocatalysis reactions taking place on the TiO₂ surface. Thus, the pH of solution was studied for enzymatic hydrolysis of rice straw in the pH range of 3–10 at a fixed concentration of TiO₂ (1.0% (w/v)). Mechanisms of the TiO₂/UV process on the effect of pH are complex and the solution pH dictates the surface charge properties of the photocatalyst [26]. TiO₂ is an amphoteric metal oxide, and, at more acidic values, the surface will be positively charged. Conversely, the surface is negatively charged. A pH that is too high or too low is not suitable for TiO₂/UV on enzymatic hydrolysis of rice straw, as shown in Figure 1b. The pH value of 8 was selected as the most suitable photocatalysis pretreatment.

Photocatalysis times and their effects on enzymatic hydrolysis of rice straw were investigated. Figure 1c shows that the released reducing sugar increased when irradiation time ranged from 0 to 8 h. The TiO_2/UV pretreatment time was prolonged from 1 to 3 h. As the irradiation time increased to 8 h, enzymatic hydrolysis of rice straw was reduced. Therefore, optimal conditions for the TiO_2/UV pretreatment were 3 h, pH 8, and TiO_2 concentration 1.0% (w/v).

3.2. Effect of Added Oxidants in TiO₂/UV Pretreatment on Enzymatic Hydrolysis of Rice Straw

In subsequent experiments, the reducing dosage of TiO_2 was chosen as a concentration of 0.50% (w/v). The added oxidants in a TiO₂/UV system function as electronic scavengers, as well as the oxidant itself. The released reducing sugar was 5.47 ± 0.03 mg/mL of TiO₂/UV pretreatment, for which TiO₂ concentration was 0.50% (w/v) without oxidant added. As indicated in Figure 2, adding inorganic oxidants increased the released reducing sugar of rice straw in the TiO₂/UV system. The optimal released reducing sugar for TiO₂/UV/K₂S₂O₈, TiO₂/UV/KIO₄ and TiO₂/UV/H₂O₂ pretreatments were 6.80 ± 0.10 mg/mL, 6.90 ± 0.20 mg/mL and 8.88 ± 0.10 mg/mL, respectively. These results confirm previous reports that TiO_2/UV /oxidant provided better photocatalysis than TiO_2/UV alone. For increasing hydrolysis, H_2O_2 was more effective than $K_2S_2O_8$ or KIO_4 , and excess $K_2S_2O_8$ and KIO_4 reduced the hydrolysis efficiency of rice straw in a TO_2/UV system. The excess of $K_2S_2O_8$ inhibited photocatalysis process, due to the increase of sulfate radical anion in solution, was adsorbed on the TiO_2 surface. The adsorbed sulfate radical anion also reacted with hydroxyl radicals, thus reducing the photocatalytic activity [27]. Further increase of KIO_4 decreased the photocatalytic efficiency, as the excess periodate ion in solution may scavenge hydroxyl radicals [19]. Released reducing sugar increased with increasing H_2O_2 concentration up to 13 mM, but exceeding 13 mM of H_2O_2 concentration decreased the hydrolysis efficiency. In particular, the addition of 13 mM H₂O₂ had a greater impact than the TiO₂ concentration of 0.50% in the TiO₂/UV system. The released reducing sugar from $TiO_2/UV/H_2O_2$ pretreatment was greater than $TiO_2/UV/K_2S_2O_8$ and $TiO_2/UV/KIO_4$ pretreatment. The addition of oxidants in a TiO₂/UV system are generally applied for treating reactive dyes and organic contaminants in industrial wastewater [19,28]. Prior studies have proposed that the role of oxidants in TiO_2/UV system increased the formation of hydroxyl radicals and impeded electron-hole recombination. Hence, a proper concentration of H_2O_2 was important for enzymatic hydrolysis of rice straw.

3.3. Composition Analysis of Untreated and Pretreated Rice Straw

Table 1 summarizes the compositions of untreated and pretreated rice straw. Optimum conditions were chosen for the three oxidants to analyze chemical compositions and selected the corresponding control group of oxidants for comparison. The contents of untreated rice straw were determined as lignin (18.68 \pm 0.30%), hemicellulose (32.98 \pm 1.10%), and cellulose (37.47 \pm 0.57%), similar to previous studies [29,30]. The H₂O₂ alone, H₂O₂/UV pretreatment and TiO₂/UV/H₂O₂ pretreatment of lignin contents were 17.04 \pm 0.74%, 17.45 \pm 0.99% and 16.10 \pm 0.67%, respectively. Lignin levels of TiO₂/UV/K₂S₂O₈ and TiO₂/UV/KIO₄ pretreatments were 18.41 \pm 0.33% and 17.69 \pm 0.54%, respectively. This demonstrated that the TiO₂/UV/H₂O₂ pretreatment improved enzymatic hydrolysis of rice straw. Some research has proposed lignin removal as an essential factor for enzymatic saccharification of lignocellulosic biomasses [31]. Hemicellulose contents by TiO₂/UV/K₂S₂O₈ and

 $TiO_2/UV/KIO_4$ pretreatment were 31.46 \pm 0.28% and 32.63 \pm 0.31% compared with 32.98 \pm 1.10% for the untreated. Öhgren [32] reported that hemicellulose hinders enzyme access to cellulose, reducing the cellulase accessibility of lignocellulose biomass. As noted above, the components of rice straw after $TiO_2/UV/K_2S_2O_8$ and $TiO_2/UV/KIO_4$ pretreatment showed no significant difference in the content of lignin, cellulose and hemicellulose. Composition analysis demonstrated that $TiO_2/UV/H_2O_2$ pretreatment improved enzymatic efficiency more than pretreatment with other oxidants to support TiO_2/UV .



Figure 2. Effect of added different oxidants concentration in TiO₂/UV pretreatment on enzymatic hydrolysis of rice straw. The different letters above the bars indicate significant difference ($p \le 0.05$, n = 3).

Pretreatment	Cellulose (%)	Hemicellulose (%)	Lignin (%)
Untreated	37.47 ± 0.57	32.98 ± 1.10	18.68 ± 0.30
H_2O_2	39.68 ± 0.64	32.49 ± 0.62	17.04 ± 0.74
H_2O_2/UV	40.59 ± 1.27	32.16 ± 1.14	17.45 ± 0.99
$TiO_2/UV/H_2O_2$	42.00 ± 0.70	30.14 ± 1.12	16.10 ± 0.67
$K_2S_2O_8$	40.21 ± 0.15	31.04 ± 0.81	18.12 ± 0.74
$UV/K_2S_2O_8$	40.58 ± 0.39	31.05 ± 1.40	18.70 ± 0.10
$TiO_2/UV/K_2S_2O_8$	40.77 ± 0.66	31.46 ± 0.28	18.41 ± 0.33
KIO ₄	40.21 ± 0.81	31.90 ± 0.60	17.75 ± 0.23
UV/KIO_4	41.81 ± 0.10	31.76 ± 0.25	18.05 ± 0.59
TiO ₂ /UV/KIO ₄	39.72 ± 0.69	32.63 ± 0.31	17.69 ± 0.54

 Table 1. Chemical composition of untreated and pretreated rice straw.

3.4. FE-SEM, XRD and FTIR Profile of Untreated and Pretreated Rice Straw

FE-SEM was performed to observe the morphological structures of untreated and pretreated rice straw and determine structural changes of the sample surfaces after photocatalysis pretreatment. Untreated rice straw (Figure 3a) presented flat and well-ordered surface structure. FE-SEM images show that the $TiO_2/UV/H_2O_2$ pretreated sample (Figure 3b) has a different morphology, disrupting the extensive, well-ordered surface of rice straw. Moreover, the surface has loose and irregular structure, with some cracks on the surface (marked with red circle). Changes of pretreated rice straw were beneficial to the enzymatic hydrolysis process due to the enhancement of cellulose accessibility to the enzyme. FTIR spectroscopy identified the changes of organic groups of untreated and pretreated rice straw, as shown in Figure 4 (marked with red circle). An untreated sample of rice straw was

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observed and compared with a pretreated sample obtained at the conditions of $TiO_2/UV/H_2O_2$. The band at 3415 cm⁻¹ was subject to O–H stretching of the hydroxyl group. Among the peaks, 2918 cm⁻¹ (C–H stretching of methyl and methyene groups), 1059 cm⁻¹ (C–O stretching vibration) and 898 cm⁻¹ (C–H deformation vibration) were assigned as the characteristic peaks of cellulose and hemicellulose. A decrease was observed in the absorbance of the 1641 cm⁻¹ of conjugated carbonyl stretch in lignin [33]. Moreover, lignin-related peaks had some changes in the vibration at the absorbance bands of 1514 cm⁻¹ (aromatic skeletal vibrations) and 1319 cm⁻¹ (C–O vibration in the syringyl ring). With $TiO_2/UV/H_2O_2$, the rice straw was weaker than the untreated sample, indicating greater lignin removal.



Figure 3. (a) FE-SEM images of untreated rice straw; and (b) FE-SEM images of $TiO_2/UV/H_2O_2$ pretreatment of 13 mM H_2O_2 at the condition of pH = 4, irradiation time of 3 h and 0.50% (w/v) concentration of TiO_2 .

Figure 5 shows XRD diffraction of untreated and $TiO_2/UV/H_2O_2$ pretreated rice straw. The CrI of untreated and $TiO_2/UV/H_2O_2$ pretreatment rice straw was calculated according to Equation (1). The CrI of untreated sample was 43.90%, and the CrI of $TiO_2/UV/H_2O_2$ pretreatment was 46.63%. $TiO_2/UV/H_2O_2$ pretreatment increased the intensity of the removal of amorphous components such as hemicellulose and lignin. Additionally, this pretreatment influenced enzymatic hydrolysis that released reducing sugar. There were similar observations of increased crystallinity index for biomass pretreatments such as previous surfactant-assisted ultrasound-ionic liquid-based pretreatment and

concentrated phosphoric acid plus hydrogen peroxide pretreatment, where the crystallinity index increased by deconstruction of the crystalline cellulose [34,35].



Figure 4. FT-IR images of untreated rice straw (**a**) and $\text{TiO}_2/\text{UV}/\text{H}_2\text{O}_2$ pretreatment (**b**) at the condition of pH = 4, irradiation time of 3 h and 0.50% (w/v) concentration of TiO₂.



Figure 5. XRD analysis of untreated rice straw (**a**) and $\text{TiO}_2/\text{UV}/\text{H}_2\text{O}_2$ pretreatment (**b**) at the condition of pH = 4, irradiation time 3 h and 0.50% (w/v) concentration of TiO₂.

4. Conclusions

This work suggests adding oxidants can boost lignin removal and improve the release of reducing sugar of rice straw. Among the oxidants, H_2O_2 addition in the TiO₂/UV system for pretreated rice straw was most effective to reduce the dosage of TiO₂ under the same pretreatment conditions. Optimal TiO₂/UV/H₂O₂ pretreatment conditions for rice straw were adding 13 mM H₂O₂ at pH = 4, irradiation time of 3 h and 0.50% (w/v) concentration of TiO₂. Results of the composition and

characterization analyses indicated that $TiO_2/UV/H_2O_2$ pretreatment efficiently provided lignin removal and significant changes in morphology.

Author Contributions: K.-L.C. and X.-Q.W. conceived and designed the experiments. K.-L.C. and X.-Q.W. wrote the paper. X.-Q.W. performed the experiments. K.-L.C. and X.-Q.W. analyzed the data. Y.-J.H., H.D. and J.-y.L. analyzed results and provided comments to the paper. Y.-C.L. corrected and approved the manuscript. All authors contributed to this work by collaboration.

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