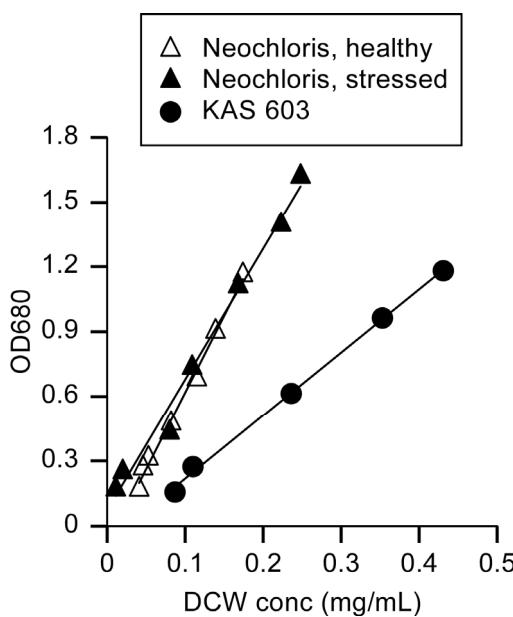


## Supplementary Information

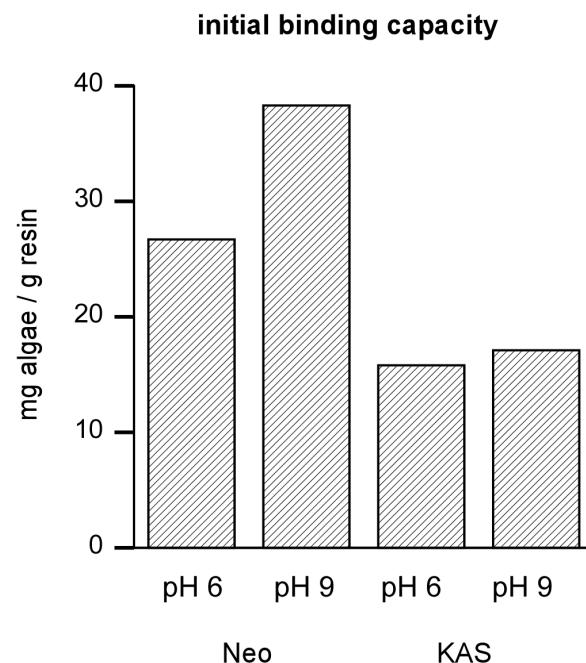
**Table S1.** Distribution of fatty acyl substituents generated from algae. FAME molecular ion assignments and abundance for FAME generated from healthy and stressed *Neochloris*, and KAS 603 are given as mean values ( $\pm$  standard error) relative to dry cell weight.

FA	m/z	healthy	stressed	KAS 603
		<i>Neochloris</i> % ( $\pm$ )	<i>Neochloris</i> % ( $\pm$ )	% ( $\pm$ )
C14:0 (myristate)	243	2.9 (0.5)	2.7 (0.4)	2.6 (0.5)
C16:0 (palmitate)	271	16.3 (3.2)	41.4 (9.9)	50.6 (5.8)
C18:3 (linolenate)	293	22.2 (4.2)	9.7 (3.6)	8.3 (3.9)
C18:2 (linoleate)	295	2.7 (1.4)	2.4 (1.0)	0.6 (0.2)
C18:1 (oleate)	297	2.3 (0.6)	3.9 (0.2)	2.7 (0.6)
C18:0 (stearate)	299	4 (1.2)	9.2 (2.5)	13.3 (1.9)
C20:0 (arachidate)	327	0.9 (0.3)	1.8 (0.6)	9.0 (1.9)
C21:4 (heneicosatetraenate)	333	46.0 (3.5)	26.2 (7.8)	10.4 (5.0)
C23:5 (tricosapentaenate)	359	2.6 (0.6)	2.8 (0.1)	2.4 (0.1)

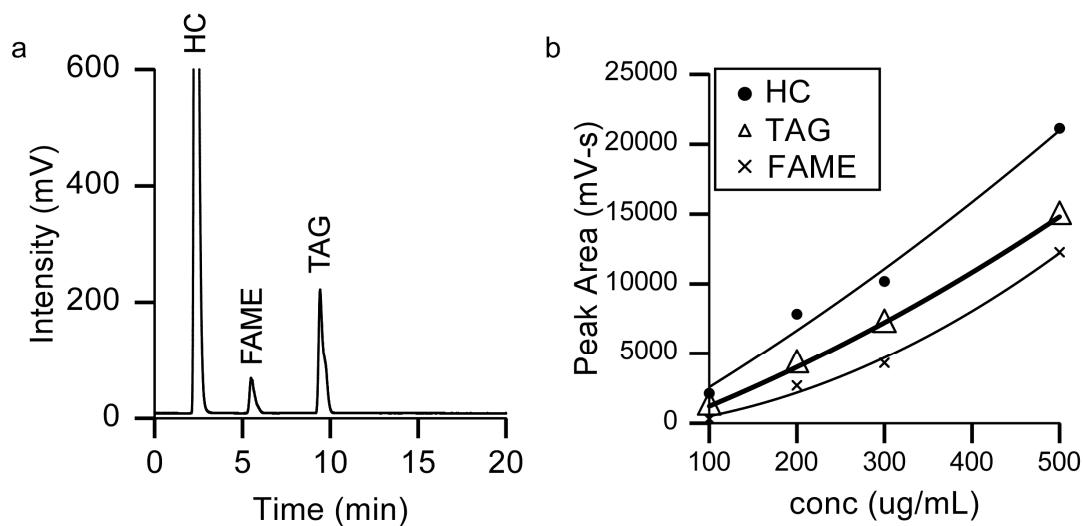
**Figure S1.** Algal biomass quantitation by chlorophyll absorbance detection. Dried cell weight concentration (DCW conc) of healthy *Neochloris*, stressed *Neochloris*, and KAS603 suspensions were fit to spectrophotometric measurement of optical density at 680 nm (OD<sub>680</sub>) ( $R^2 > 0.990$ ).



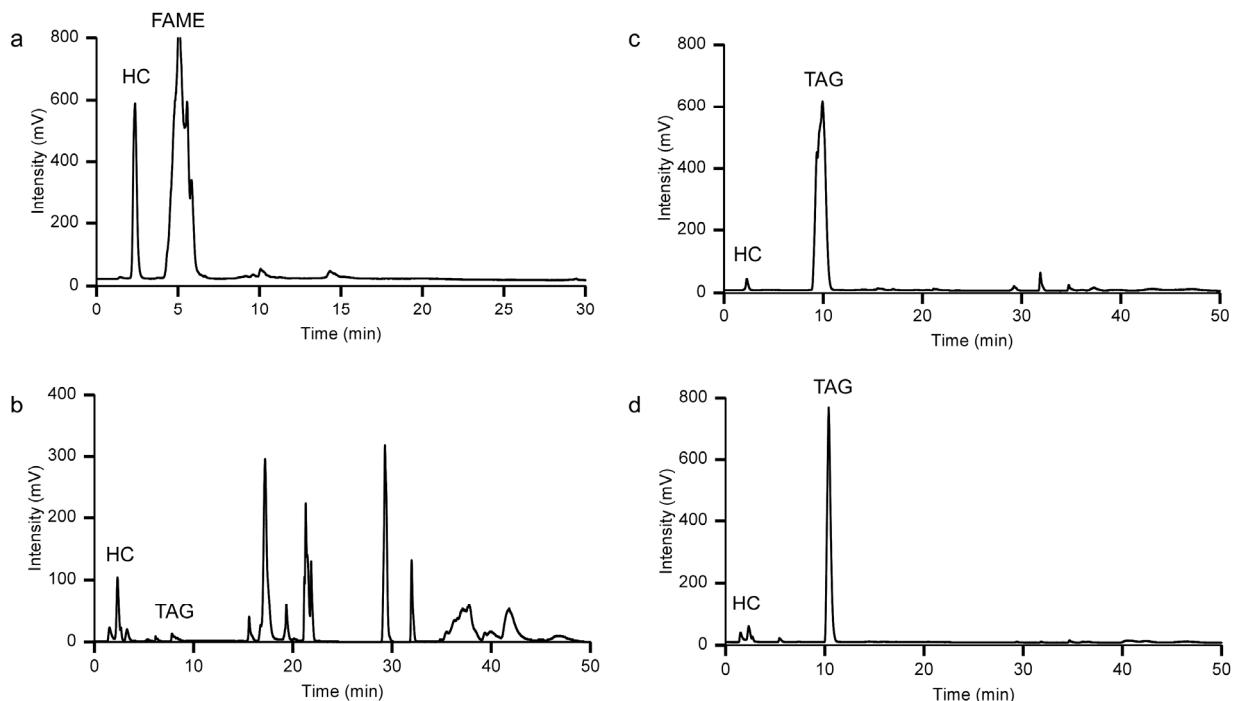
**Figure S2.** Effect of freshwater media pH on algal binding capacity of Amberlite, for algal species *Neochloris oleoabundans* and KAS 603.



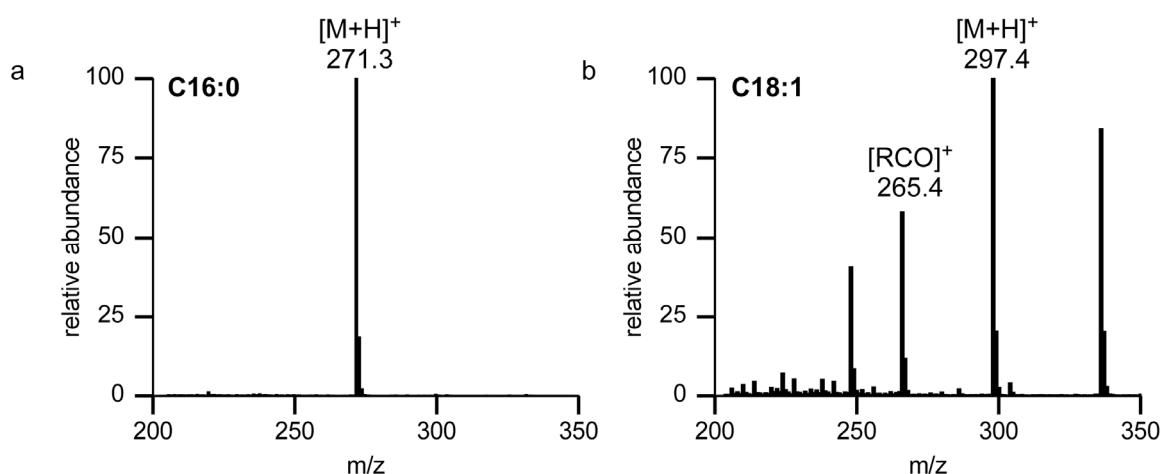
**Figure S3.** Resolution of lipid classes by normal-phase HPLC-ELSD for identification and quantification. Retention standards for hydrocarbon (HC), fatty acid methyl ester (FAME), and triacylglycerol (TAG) (a), calibration of ELSD chromatogram peak areas to lipid standard concentrations (sigmoidal fit,  $R^2 > 0.99$ ) (b).



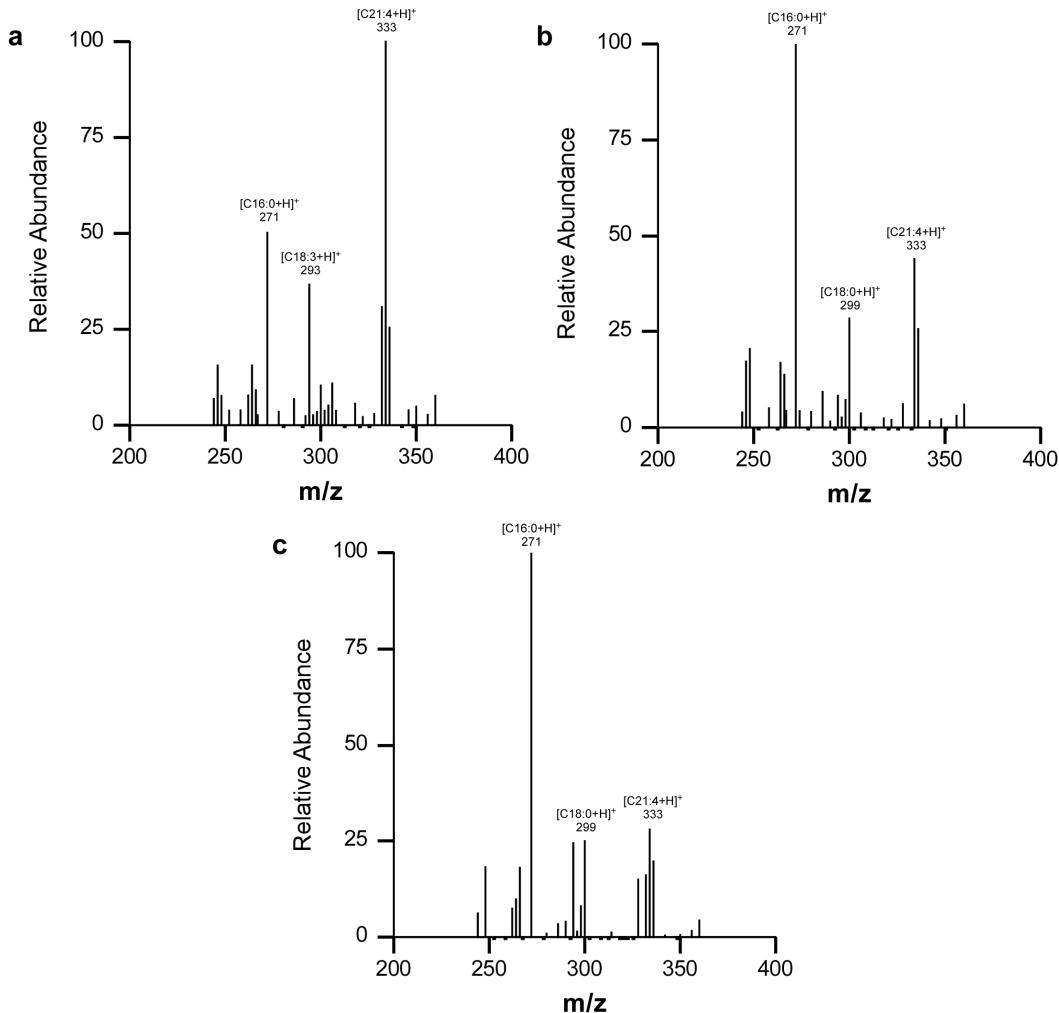
**Figure S4.** Normal-phase HPLC-ELSD chromatograms from a typical hexane extract from resin column elution shows algal HC and FAME recovered after transesterification of KAS 603 (**a**); this is compared to chromatograms from the total algal lipid extracts for healthy *Neochloris* (**b**); stressed *Neochloris* (**c**); and KAS 603 (**d**). Note differences in time axis intervals between (a) and (b–d).



**Figure S5.** HPLC-APCI-MS positive mode spectra for FAME standards methyl palmitate (C16:0) (**a**) and methyl oleate (C18:1) (**b**) show characteristic molecular ion  $[M+H]^+$  and occasionally fragment  $[RCO]^+$  species.



**Figure S6.** Typical APCI-MS positive mode spectra for FAME generated from healthy *Neochloris* (**a**), stressed *Neochloris* (**b**), and *Chlorella* (**c**) are dominated by C16:0, C18:0 or C18:3, and C21:4 species.



**Figure S7.** Use of ethanol as an alternate transesterification alcohol to methanol. Comparison of fatty acid ester yield relative to dry cell weight (DCW) for methanol and ethanol with 5% sulfuric acid catalyst for resin-bound *Neochloris* and KAS 603.

