

## Article

# Thermal Analysis and Kinetic Modeling of Pyrolysis and Oxidation of Hydrochars

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**Abstract:** This study examines the kinetics of pyrolysis and oxidation of hydrochars through thermal analysis. Thermogravimetric analysis (TGA) and differential scanning calorimetry (DSC) techniques were used to investigate the decomposition profiles and develop two distributed activation energy models (DAEM) of hydrochars derived from the hydrothermal carbonization of grape seeds produced at different temperatures (180, 220, and 250 °C). Data were collected at 1, 3, and 10 °C/min between 30 and 700 °C. TGA data highlighted a decomposition profile similar to that of the raw biomass for hydrochars obtained at 180 and 220 °C (with a clear distinction between oil, cellulosic, hemicellulosic, and lignin-like compounds), while presenting a more stable profile for the 250 °C hydrochar. DSC showed a certain exothermic behavior during pyrolysis of hydrochars, an aspect also investigated through thermodynamic simulations in Aspen Plus. Regarding the DAEM, according to a Gaussian model, the severity of the treatment slightly affects kinetic parameters, with average activation energies between 193 and 220 kJ/mol. Meanwhile, the Miura–Maki model highlights the distributions of the activation energy and the pre-exponential factor during the decomposition.



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**Keywords:** hydrothermal carbonization; modeling; kinetics; thermal analysis; Aspen Plus; biomass; hydrochar

## 1. Introduction

The global increase in energy demand, driven mainly by the growing global population, urbanization, and economic growth, is depleting the world's reserves of fossil fuels, already strained by their high consumption rate, heightening environmental concerns, resource depletion, and rising costs. To face the problem, different renewable energy sources have been investigated, like wind, solar, hydropower, geothermal, and biomass. Among the different biomasses, those deriving from waste or agricultural residues, agro-industrial, horticulture, and wood processing, are cheap and convenient [1,2]. Besides, the use and valorizing of waste biomass are part of a circular economy context, where residues become the input for a new process [3]. At present, most of the waste produced in Europe is disposed of without any further treatment resulting in wastefulness of energy and resources and in serious environmental impact. A high-efficient use of waste not only increases its economic value but also results in environmental benefits [4].

In the last decades, biomass waste treatment and management have seen an increasing interest from the research community. Currently, biomass waste has characteristics that limit its use, including a low heating value, low fixed carbon content, high moisture content, low grindability, and non-uniformity. Studies on the conversion of biomass into high-quality biofuels have been performed to avoid the problems caused by direct combustion and find more effective methods of utilization, such as pyrolysis, gasification, torrefaction, and hydrothermal carbonization [5,6].

Despite pyrolysis, gasification, and torrefaction processes reducing moisture content and increasing fixed carbon content and heating value, all of them have a problem concerning high energy consumption and high pollutant emissions [6,7].

Hydrothermal carbonization (HTC) is a promising technique that can turn lignocellulose biomass into carbon-rich, solid, lignite-like fuel.

HTC is a thermochemical treatment able to recover energy from biomass wastes and eliminate organic contaminants and pollutants [8]. Similar to the natural coalification process, HTC lowers both the oxygen and hydrogen content of the feedstock, obtaining a final coal-like product referred to as hydrochar [9]. The process consists of subjecting the feedstock to mild temperatures (180–250 °C) and autogenous pressure in liquid water, ensuring a wide range of applicability for substrates with high moisture, up to 95 wt.% [10]. Hydrochar is endowed with higher calorific value and higher fixed carbon content compared to its precursor biomass [11]. Due to its potential, HTC is a mindful tool to treat biomass waste, recover materials, and provide an energy source. Hydrochar can be used as solid fuel and integrating HTC with other processes (like gasification, pyrolysis, anaerobic digestion [12], and composting [13]) can face shortcomings related to the single-stage, and improve the overall process efficiency. For example, HTC as a pre-treatment can affect the pyrolysis performance [1]. This last factor is important to control because it is the primary stage during the combustion and gasification processes (in view of using hydrochar as a fuel) and because it is a standalone process to produce biofuels (i.e., char, oil, syngas). In this framework, HTC can be used for the pre-conditioning of high moisture content biomass, making it hydrophobic (and thus allowing for a much cheaper dewatering/drying [2]), and then the biomass can undergo the dry pyrolysis treatment [14]. How much HTC could affect the pyrolysis rate depends on the type of feedstock involved [15,16].

In this context, understanding the kinetics of pyrolysis and oxidation of hydrochar could help in designing a thermochemical conversion process.

Even though the application of hydrochar is important in boilers and burners, the kinetics and combustion performance of hydrochar have been poorly studied. Sharma et al. [17] showed that adding hydrochar to pulverized coal helps to decrease pollutant gas emissions. Moreover, the study of process kinetics and the modeling of its control in the industrial field is weak. Different models (e.g., the Kissinger–Akahira–Sunose method and the Coats–Redfern method) have also been adopted to investigate the combustion kinetics of hydrochar [18,19], but the results obtained did not lead to quantitative data that fit well with the experimental values. Due to the importance of the kinetic behavior of hydrochar combustion, additional studies are needed. Actually, the distributed activation energy model (DAEM) is the most comprehensive model to investigate the thermal reaction kinetics of biomass. Bach et al. [20] employed DAEM to study the kinetic performance of combustion of wet-towered forest residues, while Chen et al. [6] applied DAEM to study the kinetic performance of pyrolysis of five lignocellulosic biomasses. Still, Zhang et al. [21] used a multi-Gaussian DAEM to investigate the decomposition characteristics of cellulose, hemicellulose, and lignin in different atmospheres. Therefore, the DAEM offers a higher quality of fit to the experimental data and can provide more information about the kinetics of biomass pyrolysis. DAEM was proposed originally by Vand [22] to model coal pyrolysis and is based on the hypothesis that the decomposition occurs through an infinite number of parallel reactions characterized by a certain distribution of activation energy. The energy distribution is often modeled through a continuous function, like the Gaussian, Weibull, or Boltzmann function [23]. Despite the diffusion of DAEMs to predict kinetics parameters of biomass and coal pyrolysis and oxidation [24–28], studies on the hydrochars are still very rare in the literature [20]. Therefore, this study used thermogravimetric analysis (TGA) to investigate the pyrolytic and oxidative behavior of hydrochars derived from an agro-industrial residue (grape seeds [29]).

In particular, TGA is a technique commonly adopted to analyze and examine biomass degradation in oxidation or pyrolysis patterns, and can be used to deduce kinetics parameters and mechanisms [30,31]. TGA data under inert and oxidative atmosphere were

collected and used to determine kinetic parameters through two DAEMs (Gaussian and Miura–Maki DAEM). To do this, TGA data at different heating rates were collected, and activation energies and pre-exponential factors were computed. Analyses were also performed on the raw biomass and the oil contained inside the seeds. To investigate more in-depth the decomposition, derivative conversion curves were deduced, and the heat involved during the pyrolysis or oxidation was determined through differential scanning calorimetry (DSC). A simple thermodynamic model was also developed in Aspen Plus to compare experimental outputs with thermodynamic predictions. Through the software, pyrolysis and oxidation of the hydrochars were simulated and the outputs were compared with experimental measures from DSC to understand if this modeling approach can be used as a preliminary study of the heat of reactions.

## 2. Materials and Methods

### 2.1. Starting Samples: Grape Seeds and Hydrochars

The starting substrate consists of grape seeds and their corresponding hydrochars produced and characterized in previous work by our group [32]. Grape seeds are a residue of the winemaking industry and were provided by an alcohol producer company in the North of Italy (province of Trento). Hydrochars were obtained at 180, 220, and 250 °C through a solar HTC reactor, at a residence time of two hours, and a dry biomass to water weight ratio (B/W) of 0.3 [32]. Table 1 summarizes the main properties of the samples.

**Table 1.** Grape seeds and hydrochar (HC) properties, from the literature [32] (C: carbon, H: hydrogen, O: oxygen, N: nitrogen, S: sulfur, HHV: higher heating value).

Sample	Ultimate Analysis (wt.%)					Ash (wt.%)	HHV (MJ/kg)
	C	H	O	N	S		
Grape seeds	53.2	6.7	36.2	1.7	0.3	1.9	22.5
HC 180 °C	59.7	6.7	29.4	1.5	0.4	2.3	24.9
HC 220 °C	64.0	6.4	25.3	1.7	0.4	2.2	27.2
HC 250 °C	69.2	6.5	19.4	2.0	0.3	2.5	30.5

### 2.2. Sample Characterization

Grape seeds and hydrochars were characterized through thermogravimetric analysis (TGA), differential scanning calorimetry (DSC), and Fourier-transform infrared spectroscopy (FTIR).

TGA was performed in a thermobalance Mettler TG50 under non-isothermal conditions. 15 mg of whole dried samples (i.e., the entire raw and carbonized grape seeds) were placed into a 70 µL alumina crucible. Runs were performed between 40 and 700 °C at a heating rate of 1, 3, and 10 °C/min, under 100 mL/min of nitrogen or air (20.95% O<sub>2</sub> and 78.08% N<sub>2</sub>). The thermobalance has a sensitivity of  $\pm 1 \times 10^{-3}$  mg and 0.1 °C. The extent of conversion  $\alpha$  at any time  $t$  was computed as:

$$\alpha(t) = \frac{m_0 - m(t)}{m_0 - m_f} = \frac{v(t)}{v_\infty} \quad (1)$$

where  $m_0$ ,  $m(t)$ , and  $m_f$  are the mass at time zero (dry), at a given time  $t$ , and final time, respectively. This corresponds to the ratio between  $v(t)$ , which represents the amount of volatiles evolved up to a certain time  $t$ , and  $v_\infty$ , which represents the amount of volatiles that evolved up to the end of the process.

DSC was performed in a Mettler DSC20 calorimeter between 40 and 600 °C, at a heating rate of 10 °C/min, under nitrogen or air at a flow rate of 100 mL/min. About 15 mg of material was placed into a 40 µL aluminum crucible. The heat released/absorbed was computed by integrating the measured heat flow over time in the 40–600 °C range, while the sample mass variation was recorded after every run.

FTIR spectra of samples were acquired using PerkinElmer FT-IR spectrometer Spectrum One. Spectra were obtained over a wavenumber range of 4000–650  $\text{cm}^{-1}$ . Dried samples were ground to be compacted into a thin film and a flat top-plate Zinc Selenide crystal was used for the detection.

### 2.3. Distributed Activation Energy Models (DAEMs)

Two types of DAEMs (the Gaussian and the Miura–Maki [33]) were developed in software codes to model the decomposition of hydrochars, grape seeds, and grape seed oil.

In particular, when a first-order reaction is assumed to model biomass degradation, the variation of  $\alpha$  vs. time can be expressed as the product between the reaction rate  $k$  and the conversion function  $f$ :

$$\frac{d\alpha}{dt} = k(T)f(\alpha) \quad (2)$$

By adopting the Arrhenius theory,  $k(T)$  can be expressed by Equation (3):

$$k(T) = k_0 \exp\left(-\frac{E}{RT}\right) \quad (3)$$

where  $k_0$  represents the pre-exponential factor,  $E$  is the activation energy, and  $R$  is the universal gas constant.

To deduce the kinetic parameters,  $f(\alpha)$  is usually assumed in advance (in the so called model-fitting methods) and then forced to fit experimental data [23]. In particular, non-isothermal models usually adopt a constant heating rate  $\beta$  equal to  $dT/dt$ , allowing the re-arrangement of Equation (3) into (4), which can be solved using thermogravimetric data [23].

$$\frac{d\alpha}{dT} \cong \frac{d\alpha}{dt} \frac{dt}{dT} = \frac{d\alpha}{dt} \frac{1}{\beta} = \frac{k(T)}{\beta} f(\alpha) = \frac{k_0}{\beta} \exp\left(-\frac{E}{RT}\right) f(\alpha) \quad (4)$$

Among model-fitting methods, DAEM assumes that pyrolysis can be described through an infinite number of irreversible first-order parallel reactions, with different rate parameters, which occur simultaneously [23,34]. In particular, the DAEM for a reaction order equal to 1 can be obtained by re-arranging Equations (2)–(4) as:

$$1 - \alpha = \int_0^\infty \exp\left(-\int_{T_0}^T \frac{k_0}{\beta} \exp\left(-\frac{E}{RT}\right) dT\right) f(E) dE \quad (5)$$

Equation (5) does not have an analytical solution and there are two families of methods to solve it. One is referred to as distribution-fitting method and the other as isoconversional methods. The distribution-fitting method assumes  $f(E)$  (like a Gaussian, Weibull, or Boltzmann distribution) and forces to fit the TGA data by applying a certain numerical method [23]. Meanwhile, isoconversional methods do not assume  $f(\alpha)$ , but adopt a series of thermogravimetric data at different heating rates to directly compute the activation energy. The reaction rate at a certain  $\alpha$  is a function of temperature. Among these distribution-free methods, the Miura–Maki integral method is a common one [35].

#### 2.3.1. Gaussian Model

In the Gaussian DAEM, the distribution function  $f(E)$  is assumed to be a Gaussian distribution with mean activation energy  $E_0$  and standard deviation  $\sigma$ , which is:

$$f(E) = \frac{1}{\sigma\sqrt{2\pi}} \exp\left(-\frac{(E - E_0)^2}{2\sigma^2}\right) \quad (6)$$

Applying the Coats–Redfern [36] and Fisher et al. [37] approximations, Equation (7) is obtained.

$$1 - \alpha = \int_0^{\infty} \left( \exp \left( -\frac{k_0 RT^2}{\beta E} \exp \left( -\frac{E}{RT} \right) \right) \right) \frac{1}{\sigma \sqrt{2\pi}} \exp \left( -\frac{(E - E_0)^2}{2\sigma^2} \right) dE \quad (7)$$

To solve Equation (7), the authors applied the simplification reported by Anthony and Howard [38], which consists in keeping  $k_0$  constant and equal to  $1.67 \times 10^{13} \text{ s}^{-1}$ , and the minimization procedure proposed by Güneş and Güneş [39] that solves the integral using the Simpson's 1/3 rule. To avoid oscillations in the results,  $dE$  was kept constant at 50 kJ/mol, while the extremes of integration were iteratively adjusted according to  $E_0$ . Then, the best solution was calculated by iterating the solution of the integral for several values of  $E_0$  and  $\sigma$  and minimizing  $h$ , i.e., the root mean quadratic error between the model output and the experimental data:

$$h = \sqrt{\frac{\sum_{j=1}^n (\alpha_{TGA,j} - \alpha_{DAEM,j})^2}{n}} \quad (8)$$

where  $\alpha_{TGA,j}$  and  $\alpha_{DAEM,j}$  are the experimental and calculated extent of conversion, respectively, and  $n$  is the number of available data points.

The code was developed in MATLAB and a direct search technique was adopted [39]. The code is reported in Supplementary Materials. Firstly, a larger grid procedure was used for both  $E_0$  and  $\sigma$ , with a step size equal to 5. The first three values minimizing  $h$  were obtained. Then, for these last three values, an iteration process using  $E_0 \pm 5$  and  $\sigma \pm 5$  with a step equal to 1 was set. Finally, final values were used to solve the integral of Equation (7) and to calculate  $\alpha_{DAEM}$ .

### 2.3.2. Miura–Maki Model

The Miura–Maki model adopts some assumptions to determine  $f(E)$  and  $k_0$  without any a priori assumption on the energy distribution [33]. In particular, Miura and Maki approximated [33,35] the so called  $\Phi$  function (Equation (9)) to a step-function  $E = E_s$  for a selected temperature  $T$ .

$$\Phi(E, T) = \exp \left( -k_0 \int_0^t \exp \left( -\frac{E}{RT} \right) dt \right) \cong \exp \left( -\frac{k_0}{\beta} \int_0^T \exp \left( -\frac{E}{RT} \right) dT \right) \quad (9)$$

Thus, Equation (5) can be written as:

$$\alpha \cong 1 - \int_{E_s}^{\infty} f(E) dE = \int_0^{E_s} f(E) dE \quad (10)$$

with  $E_s$  so that  $\Phi(E_s, T) \cong 0.58$ .

By the Miura and Maki approach, only a reaction having an activation energy  $E_s$  occurs at a given  $T$  and  $\beta$ —the model simplifies an actual reaction system by a set of  $N$  reactions, characterized by their own activation energy and pre-exponential factor.

Approximating the function  $\Phi(E, T)$  as:

$$\Phi(E, T) \cong \exp \left( -\frac{k_0 RT^2}{\beta E} \exp \left( -\frac{E}{RT} \right) \right) \quad (11)$$

the extent of conversion for the  $j$ -th reaction is expressed by Equation (12):

$$1 - \frac{\Delta v}{\Delta v_{\infty}} \cong \exp \left( -\frac{k_0 RT^2}{\beta E} \exp \left( -\frac{E}{RT} \right) \right) \quad (12)$$

where  $\Delta v$  and  $\Delta v_\infty$  represent the amount of volatiles evolved and the effective volatile content for the  $j$ -th reaction.

After some mathematical steps [35], and after imposing  $1 - \Delta v / (\Delta v_\infty) = \Phi(E, T) \cong 0.58$ , the representative equation of the Miura–Maki model is given by Equation (13).

$$\ln\left(\frac{\beta}{T^2}\right) = \ln\left(\frac{k_0 R}{E}\right) + 0.6075 - \frac{E}{RT} \quad (13)$$

Compared to the Gaussian DAEM, the approach of Miura [35] and Miura and Maki [33] relies on the dependence of  $k_0$  on  $E$ . In particular, they proposed a simple procedure to estimate  $f(E)$  and  $k_0$ , consisting of the following steps:

- (1) measurement of  $\alpha$  vs.  $T$  relationship for at least three heating rates;
- (2) calculation of  $\beta/T^2$  at selected  $\alpha$  values from the  $\alpha$  vs.  $T$  relationship obtained in (1) for each heating rate;
- (3) plotting of  $\ln(\beta/T^2)$  vs.  $-1/(RT)$  at the selected  $\alpha$  values, and determination of  $E$  and  $k_0$  from the Arrhenius plot at different  $\alpha$  values using the relationship represented by Equation (13):  $E$  can be estimated by the slope, and  $k_0$  by the intercept of each line with the  $y$  axis; and
- (4) plotting the  $\alpha$  value against the activation energy  $E$ , as calculated in (3), and differentiating the  $\alpha$  vs.  $E$  relationship to obtain  $f(E)$ .

#### 2.4. Aspen Plus Model

The energy aspects of pyrolysis were thermodynamically investigated by performing some Aspen Plus simulations reproducing the pyrolysis of hydrochars. The model is based on thermodynamic equilibrium and the minimization of the Gibbs energy, without considering any kinetic phenomena. In particular, the thermodynamic properties of the conventional components were calculated by adopting the Peng–Robinson method.

Simulations were performed on grape seeds, hydrochars, and a commercial coal. Table 2 reports a summary of all input data, while Figure 1 shows the general Aspen scheme. The stream REACTANT represents the input feedstock entering the process, defined through a non-conventional stream characterized by its ultimate analysis and HHV (grape seeds and hydrochar in Table 1, coal Illinois No. 6 from literature [40]). Then, the RYIELD reactor, which is fictitious, splits the non-conventional stream into a conventional one (ELEMENTS) based on its elemental distribution. This stream enters the RGIBBS reactor, which computes the product distribution as a function of the operating conditions (i.e., temperature and pressure) by minimizing the Gibbs energy of the product mixture. The final products considered in the model are carbon graphite, carbon dioxide ( $\text{CO}_2$ ), carbon monoxide ( $\text{CO}$ ), water ( $\text{H}_2\text{O}$ ), hydrogen ( $\text{H}_2$ ), oxygen ( $\text{O}_2$ ), methane ( $\text{CH}_4$ ), acetylene ( $\text{C}_2\text{H}_2$ ), ethylene ( $\text{C}_2\text{H}_4$ ), and ethane ( $\text{C}_2\text{H}_6$ ), to ensure a good model performance. Then, the net heat required/produced by the process is given by stream QOUT, while QIN represents the heat required to sustain RYIELD. Runs were performed between 300 and 900 °C (every block and stream were kept isotherm to avoid any sensible heat contribution) to deduce the amount of heat involved during the reaction. The product distribution at different temperatures was also investigated. Data were referred per gram of dry feedstock entering the process.

**Table 2.** Summary of parameters used in the Aspen Plus simulations.

Feedstock	Grape seeds, HC 180 °C, HC 220 °C, HC 250 °C, Coal
Products (at equilibrium)	C (graphite), $\text{CO}_2$ , $\text{CO}$ , $\text{H}_2$ , $\text{O}_2$ , $\text{H}_2\text{O}$ , $\text{CH}_4$ , $\text{C}_2\text{H}_2$ , $\text{C}_2\text{H}_4$ , $\text{C}_2\text{H}_6$
Equation of state	Peng–Robinson
Pyrolysis temperature	300–900 °C

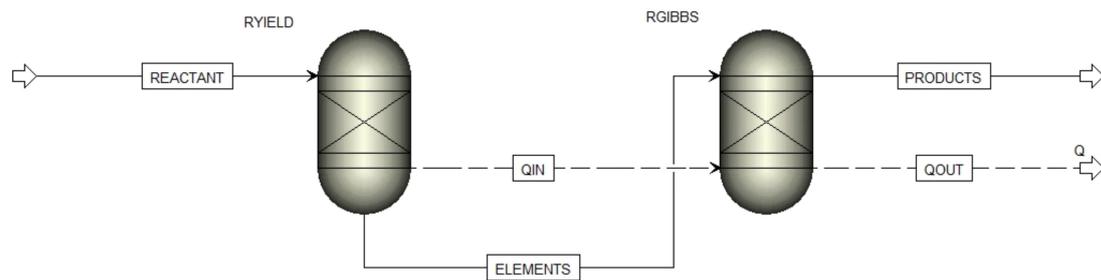


Figure 1. Aspen Plus scheme used to model pyrolysis.

### 3. Results and Discussion

#### 3.1. Derivative Thermogravimetric Curves

Figure 2 shows the derivative thermogravimetric curves (DTGA) of grape seeds and hydrochars under pyrolytic and oxidative conditions, at different heating rates. Table 3 shows a sum of the position of local peaks, with their temperatures and intensities.

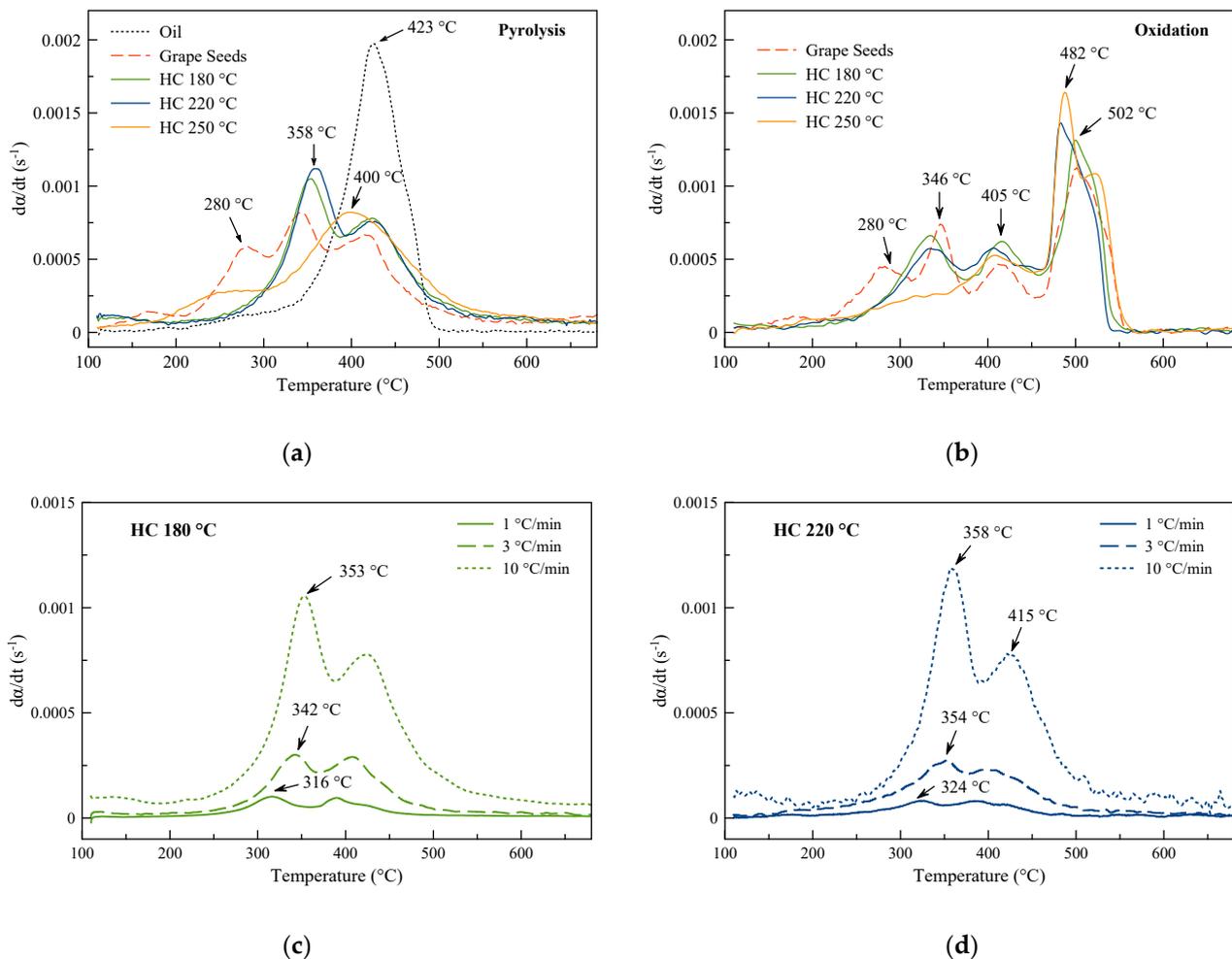
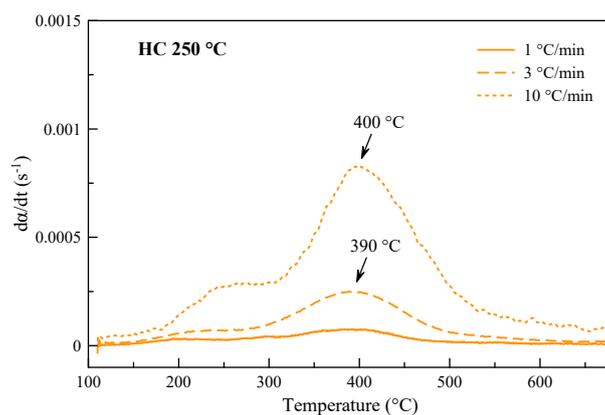


Figure 2. Cont.



(e)

**Figure 2.** Derivative thermogravimetric (DTGA) curves: (a) during pyrolysis at 10 °C/min; (b) during oxidation at 10 °C/min; (c) effect of the heating rate during pyrolysis of hydrochar 180 °C; (d) effect of the heating rate during pyrolysis of hydrochar 220 °C; (e) effect of the heating rate during pyrolysis of hydrochar 250 °C. HC: hydrochars,  $d\alpha/dt$ : derivative conversion curve of the extent of conversion  $\alpha$  with respect to time  $t$ .

**Table 3.** DTGA peak temperature and decomposition rate of pyrolysis and oxidation (10 °C/min).

DTGA Peak		Pyrolysis					Oxidation			
		Oil	Grape Seeds	HC 180 °C	HC 220 °C	HC 250 °C	Grape Seeds	HC 180 °C	HC 220 °C	HC 250 °C
1	T (°C)	-	280	-	-	-	280	-	-	-
	$d\alpha/dt$ (s <sup>-1</sup> )	-	$5.9 \times 10^{-4}$	-	-	-	$4.4 \times 10^{-4}$	-	-	-
2	T (°C)	-	343	353	358	-	346	334	336	-
	$d\alpha/dt$ (s <sup>-1</sup> )	-	$8.2 \times 10^{-4}$	$1.0 \times 10^{-3}$	$1.1 \times 10^{-3}$	-	$7.4 \times 10^{-4}$	$6.6 \times 10^{-4}$	$5.7 \times 10^{-4}$	-
3	T (°C)	423	421	424	428	400	415	415	406	409
	$d\alpha/dt$ (s <sup>-1</sup> )	$2.0 \times 10^{-3}$	$6.6 \times 10^{-4}$	$7.8 \times 10^{-4}$	$7.7 \times 10^{-4}$	$8.2 \times 10^{-4}$	$4.6 \times 10^{-4}$	$6.2 \times 10^{-4}$	$5.8 \times 10^{-4}$	$5.3 \times 10^{-4}$
4	T (°C)	-	-	-	-	-	502	502	482	480–523
	$d\alpha/dt$ (s <sup>-1</sup> )	-	-	-	-	-	$1.1 \times 10^{-3}$	$1.3 \times 10^{-3}$	$1.4 \times 10^{-3}$	$1.6\text{--}1.1 \times 10^{-3}$

Overall, both grape seeds and hydrochars mostly decompose between 200 and 500 °C, indicating that most of the volatile matter is removed in this range. Grape seeds present three main decomposition peaks, occurring at 280, 343, and 421 °C under a pyrolytic atmosphere. These peaks can be attributed to the overlapped decomposition of hemicellulose, lignin plus cellulose, and oil, which are the main constituents of grape seeds (around 7% cellulose, 31% hemicellulose, 44% lignin, and 10–15% oil [41,42]). Indeed, it is well known that under pyrolytic conditions, due to its amorphous and little polymerized structure, hemicellulose is very reactive at low temperatures (in the range of 250–300 °C [21,43]). In the DTGA curves, this reactivity translates into the first decomposition peak. Meanwhile, the second peak can be attributed to the overlapped decomposition of lignin and cellulose, which both generally decompose in the 300–350 °C range [43]. Considering the low content of cellulose (around 7% [41,42]), lignin clearly dominates at this temperature. Beyond this decomposition, lignin also contributes by forming a decomposition “baseline” to the profile, overlapping the other components. Indeed, due to its high polymerized structure and higher heterogeneity, it decomposes also (less intensively than at 300–350 °C) in a broad range of temperatures, from around 230° up to 600 °C [43]. Therefore, during the pyrolysis of grape seeds, its decomposition could overlap to that of the other constituents, forming a “baseline” to the overall profile. Meanwhile, comparing the measured curve with that of

grape seed oil from previous work by our group [28], it is clear that the peak occurring at 423 °C can be attributed to the decomposition of the oil. Regarding the hydrochars, through HTC, grape seeds undergo carbonization, decreasing their atomic O/C and H/C ratios with a natural reduction of the volatile matter as the harshness of the process increases [32]. In DTGA curves, this translates in flattening the first decomposition peak at around 280 °C, which is indeed absent for all the hydrochars. Therefore, the authors can affirm that hemicellulosic compounds are mostly degraded during HTC. Interestingly, the 250 °C hydrochar exhibits a broad and not intense decomposition between 200 and 300 °C. Since hemicellulose degrades during HTC, this reactivity can be explained by the presence of re-polymerized compounds from the aqueous phase. Indeed, during HTC, sugar-derived compounds dissolved in the aqueous phase (like 5-HMF) can undergo condensation and re-polymerization, forming a solid phase called “secondary char” [44]. Beyond this, fatty acids formed during the hydrolysis of the oil could also be embedded in the solid phase, conferring to the sample an extra-reactivity at low temperatures. Both grape seeds and 180 and 220 °C hydrochars show their highest reactivity at around 350 °C, attributable to the degradation of lignin and cellulosic derived compounds. Differences can be due to both the heterogeneity of the feedstock and the synergistic behavior among the constituents, which are present in different ratios in the various samples. Conversely, the 250 °C hydrochar, after the initial slow decomposition at 200–300 °C, shows only one main decomposition peak at 400 °C. Therefore, while the 180 and 220 °C hydrochars tend to follow the feedstock profile (2nd and 3rd peaks), the 250 °C hydrochar highly deviates from that, highlighting the impact of the HTC operating temperature on the devolatilization profile. Overall, it is important to highlight to positive effect of the HTC treatment: after HTC, curves tend to shift towards the right region of the graph, demonstrating a higher thermal stability during the degradation process.

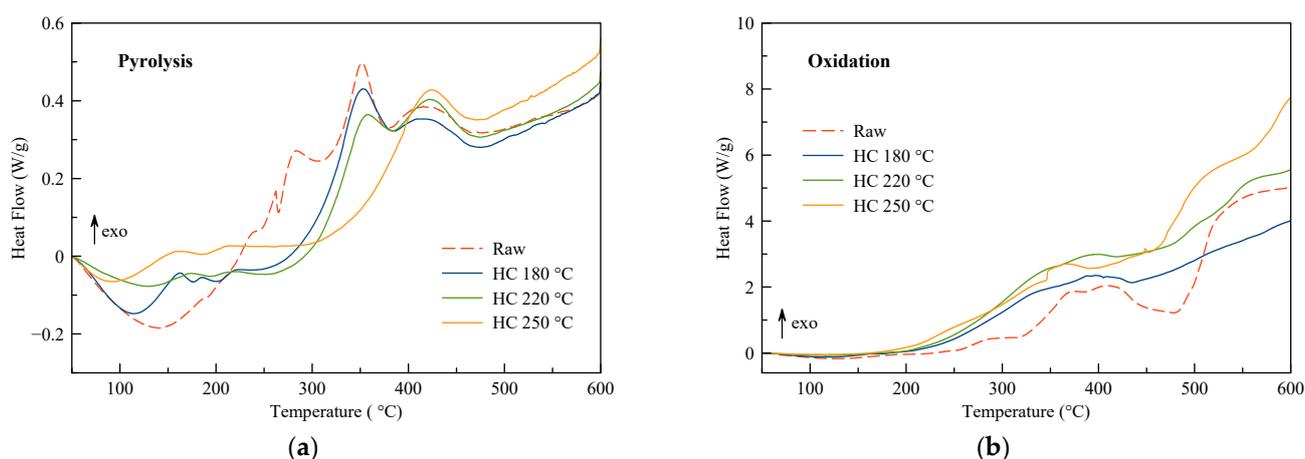
Regarding oxidative conditions (Figure 2b), as expected by typical combustion profiles of biomass fuels [20], all the profiles can be divided into two regions: a first phase during which the feedstock devolatilizes to char, and a second phase where char oxidation occurs. The first phase, between 250 and 450 °C, resembles the pyrolytic behavior: hemicellulose, lignin plus cellulose, and oil volatilize in the same temperature ranges as during pyrolysis. Then, the second phase, 480–520 °C, corresponds to the oxidation of the char formed during the first phase. This phase is predominant with respect to the previous phase. As for pyrolysis, the 180 and 220 °C hydrochars resemble the behavior of grape seeds, exhibiting three main DTGA peaks. Meanwhile, the 250 °C hydrochar presents only two DTGA peaks, demonstrating the harshness of severe HTC conditions on the feedstock structure and composition. Oxidation and pyrolysis TGA curves where  $(1-\alpha)$  is plotted vs. T are reported in Supplementary Materials Figure S1.

Figure 2c–e show the effect of the heating rate on DTGA curves. The heating rate does not affect the shape of the decomposition profile. Meanwhile, the DTGA peak moves towards higher temperatures at increasing heating rates, which can be due to heat and mass transfer phenomena and intrinsic devolatilization kinetics. Indeed, at higher rates, the mismatch between the temperature in the furnace and the particle is higher, and therefore there is a delay between the temperature measured and the decomposition stage [45]. In addition, the  $d\alpha/dt$  is higher at higher rates because, at a fixed  $\alpha$ ,  $dt$  is smaller. In addition, higher rates cause a bigger gradient between the surface and core temperature of the sample [45].

### 3.2. DSC Curves

Figure 3 shows DSC curves under nitrogen and oxidative atmosphere. Under an inert atmosphere, both grape seeds and hydrochars show an initial endothermic phase followed by an exothermic one at higher temperatures. The transition from an endo- to an exothermic behavior occurs in a temperature range between 200 and 290 °C. The heat request comprises both the heat necessary to evaporate water and that to trigger endothermic reactions. Water derives from both some residual moisture and some produced

during condensation reactions occurring at the beginning of pyrolysis. Meanwhile, after the transition temperature, heat is released during pyrolysis. This behavior was already observed in literature with hydrochars derived from other feedstock [40,46] and can be attributed to the presence of non-oxidized oxygen on the hydrochar, like in the form of carboxylic groups. Indeed, this oxygen is enough to oxidize the elemental carbon, leading to a heat release. As shown in Table 4, the net energy balance along the temperature spectrum 40–600 °C is positive for all the samples, highlighting that overall pyrolysis releases thermal energy. Not surprisingly and consistently with elemental analysis (Table 1), a higher heat release is associated with a higher volatile matter loss (Table 4). Details on the progressive integration of DSC data are given later in the paper (namely, in Section 3.5).



**Figure 3.** Differential scanning calorimetry (DSC) curves of grape seeds and hydrochars (HC) at 10 °C/min: (a) under inert and (b) oxidative atmosphere (> 0 exothermic, < 0 endothermic); per grams of dry feedstock.

**Table 4.** Details of DSC curves: integrals, mass loss, and transition point from endothermic to exothermic behavior; per kilograms of dry feedstock. The positive values of “Net energy” correspond to exothermal behavior during thermal treatments.

	Pyrolysis				Oxidation			
	Raw	HC 180 °C	HC 220 °C	HC 250 °C	Raw	HC 180 °C	HC 220 °C	HC 250 °C
Endothermic <sup>1</sup> (J/g)	118	90	67	23	98	54	31	19
Exothermic <sup>1</sup> (J/g)	705	587	585	598	4406	4928	6678	7533
Net energy <sup>1</sup> (J/g)	587	497	518	575	4309	4874	6647	7514
Mass loss <sup>1</sup> (%)	61.2	56.2	55.4	43.7	73.7	69.0	73.3	69.0

<sup>1</sup> Integral and mass losses computed/measured in the range 40–600 °C.

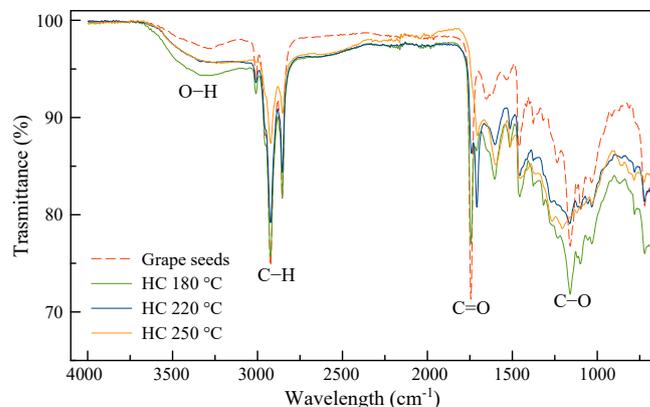
As for DTGA curves, samples show heat profiles with local peaks. Indeed, grape seeds show three main peaks, at 280, 352, and 410–425 °C, in correspondence with the degradation of hemicellulose, lignin plus cellulose, and oil. The 180 and 220 °C hydrochars show two peaks, while the 250 °C shows only one at 420 °C. A comparison between DTGA and DSC curves under pyrolytic atmosphere is reported in Supplementary Materials Figure S2.

As expected from the nature of the process, under oxidative conditions (Figure 3b), the amount of heat released increases as the carbonization degree of the sample increases. All the samples show a slight endothermic behavior at low temperature, justified by the absence or low presence of oxidative reactions at this condition. Overall, the heat release profile is broad and rises to the final observed temperature. Indeed, at 600 °C, a certain quantity of matter that undergoes oxidation is still present. Even if less sharp than under an inert atmosphere, all the samples show a peak at 350–450 °C that can be associated with

the oxidation of lignin-like compounds. Grape seeds also show a small inflection at 280 °C, probably due to the oxidation of hemicellulosic compounds.

### 3.3. FTIR

Figure 4 reports the FTIR spectra of the samples. The deflection at 3300  $\text{cm}^{-1}$  indicates the stretching of the hydroxyl group, occurring generally in the range 3700–3200  $\text{cm}^{-1}$  [47]. Increasing HTC temperature results in a decreasing intensity in O-H stretching, because dehydration phenomena occur during HTC treatment. The peak at 3010  $\text{cm}^{-1}$  represents =C-H group stretch, which tends to gradually disappear in the hydrochars. This fact can be related to double bonds present in the oil, which degrades, and it is even not present in HC 250 °C. Purnomo et al. [48] identified hydrogen bonded to the unsaturated carbon chain (C-H), whereas peaks at lower wavelength represent attachment to the saturated carbon chain. The peak at 1744  $\text{cm}^{-1}$  indicates a typical bending of carbonyl group  $>\text{C}=\text{O}$ , possibly in the form of aldehyde [5], and/or the ester group in glyceride compounds. It tends to attenuate by increasing HTC treatment severity. Whereas grape seeds and HC 180 °C still present a peak, the HC 220 °C spectrum moves toward a lower wavelength. In the HC 250 °C spectrum the peak disappeared, confirming degradation and formation of new compounds during the hydrothermal treatment. Furthermore, Diaz et al. [41] state that peaks around 1030  $\text{cm}^{-1}$  indicate C-O and C-O-C stretching, confirming that both decarboxylation and dehydration occurred. The grape seed spectrum exhibits a shoulder at a wavelength equal to 1655  $\text{cm}^{-1}$ , attributed to stretching of double bond  $>\text{C}=\text{C}<$  [47], that tends to flatten out in hydrochars. Unsaturated double-bond  $>\text{C}=\text{C}<$  could be related to the presence of oil [6]—grape seed oil is mainly composed of linoleic acid [48,49], having two double bonds C=C in position 9 and 12. In general, functional groups act a stretch toward the region at a shorter wavelength.



**Figure 4.** Fourier transform infrared spectroscopy (FTIR) curves of grape seeds and hydrochars (HC).

### 3.4. Kinetic Models

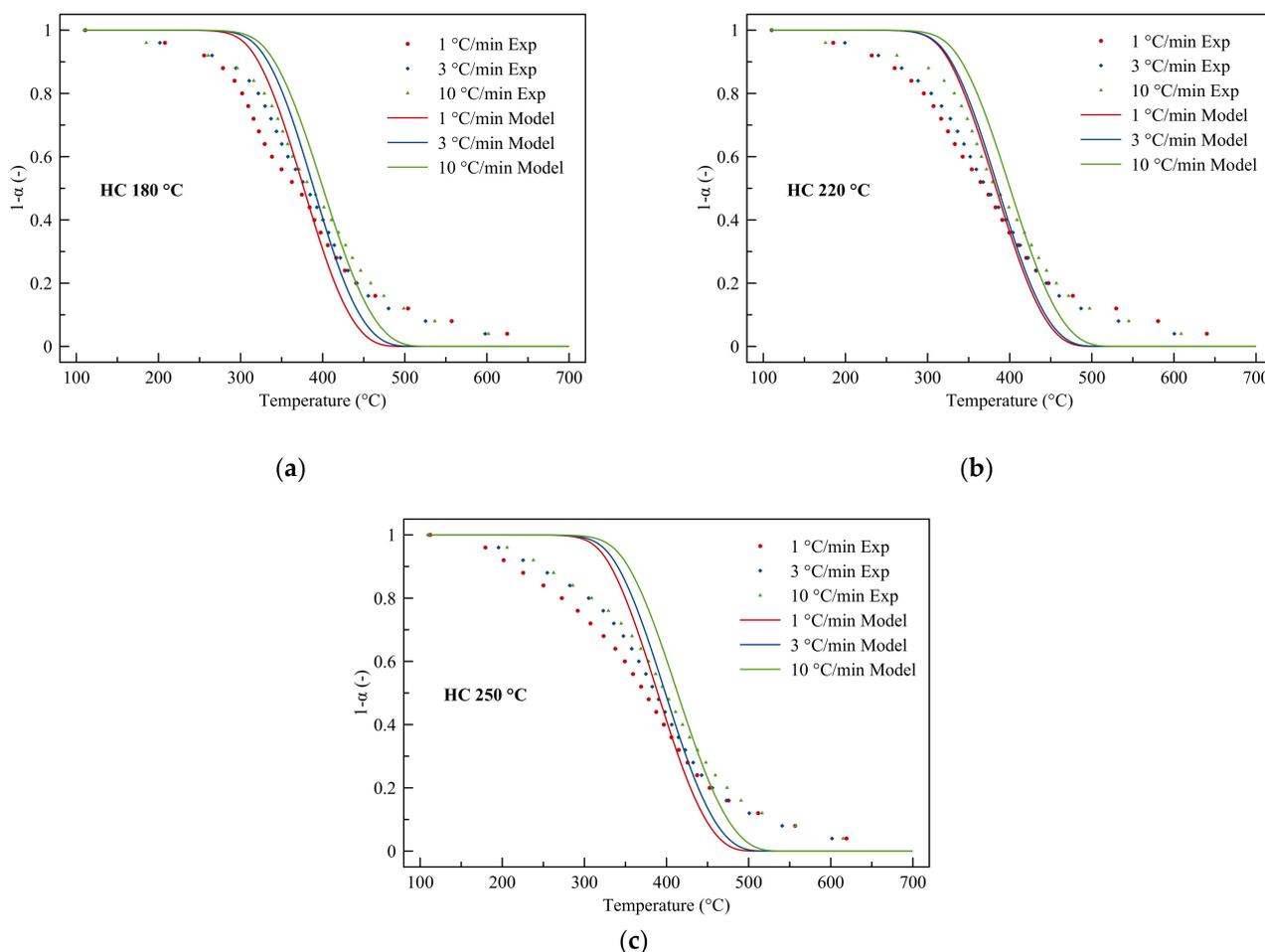
#### 3.4.1. Gaussian Model

Table 5 shows  $E_0$ ,  $\sigma$ , and  $h$  computed through the Gaussian model at different heating rates for the various substrates, while Figure 5 shows the experimental data compared with the modeling curves for pyrolysis (data and curves relevant to oxidation are available in Supplementary Materials Figure S3). Experimental data are the starting point to compute DTGA curves reported in Figure 2.

**Table 5.** Values of  $E_0$  and  $\sigma$  for the Gaussian model for the different substrates and heating rates; per moles of dry feedstock.  $k_0$  is let constant and equal to  $1.67 \times 10^{13} \text{ s}^{-1}$ .

		Grape Seeds	Oil	HC 180 °C	HC 220 °C	HC 250 °C	
Pyrolysis	$E_0$ (kJ/mol)	1 °C/min	-	-	205	207	209
		3 °C/min	-	-	203	202	206
		10 °C/min	193	218	200	200	204
	$\sigma$ (kJ/mol)	1 °C/min	-	-	15	16	17
		3 °C/min	-	-	14	15	16
		10 °C/min	16	11	15	15	16
	h (-)	1 °C/min	-	-	0.083	0.090	0.089
		3 °C/min	-	-	0.061	0.072	0.073
		10 °C/min	0.084	0.022	0.064	0.062	0.076
Oxidation	$E_0$ (kJ/mol)	1 °C/min	-	-	211	214	219
		3 °C/min	-	-	215	215	220
		10 °C/min	213	-	214	213	221
	$\sigma$ (kJ/mol)	1 °C/min	-	-	14	15	15
		3 °C/min	-	-	15	14	15
		10 °C/min	17	-	16	15	15
	h (-)	1 °C/min	-	-	0.057	0.065	0.065
		3 °C/min	-	-	0.071	0.066	0.066
		10 °C/min	0.102	-	0.084	0.077	0.077

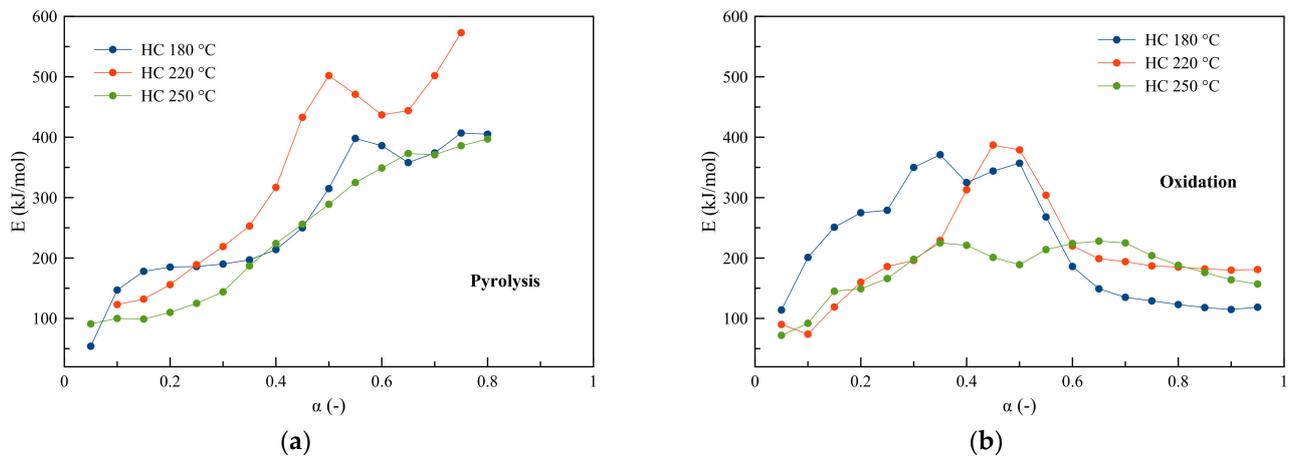
Data demonstrate that for both pyrolysis and oxidation, the effect of the heating rate on  $E_0$  and  $\sigma$  is negligible. Therefore, every substrate can be associated with one single pair of  $E_0$  and  $\sigma$ , regardless of the heating rate, which agrees with previous studies performed on other biomass substrates [28]. Under both inert and oxidative atmospheres, the Gaussian DAEM adopted does not highlight the effects of both HTC and its severity on the starting feedstock (their maximum relative difference is 3.7%). This is not surprising: the model basis on the assumption of approximating the entire decomposition profile, which includes more stages, through a one-step profile characterized by a Gaussian distribution. Therefore, the final values of  $E_0$  and  $\sigma$  can be considered as a sort of weighted average among the values of the single components (biomass constituents), i.e., hemicellulosic, lignin-like plus cellulosic compounds, and oil. Probably, the difference in the relative composition of the hydrochars is not enough to affect the final values. In particular, under an inert atmosphere,  $E_0$  passes from 193 kJ/mol of grape seeds to 200–209 kJ/mol of hydrochars, while  $\sigma$  ranges between 14 and 17 kJ/mol. Values agree with those reported in the literature on the single components. For example, during pyrolysis, the activation energies usually range in 80–116 kJ/mol for hemicellulose, 195–286 kJ/mol for cellulose, and 176–300 kJ/mol for lignin [35,42]. The pyrolysis of oil alone has an  $E_0$  of 218 kJ/mol. With values between 211 and 215 kJ/mol, the activation energy of oxidation is the same for grape seeds and hydrochars obtained at 180 and 220 °C. Meanwhile, it slightly increases at 220 kJ/mol for the 250 °C hydrochar.



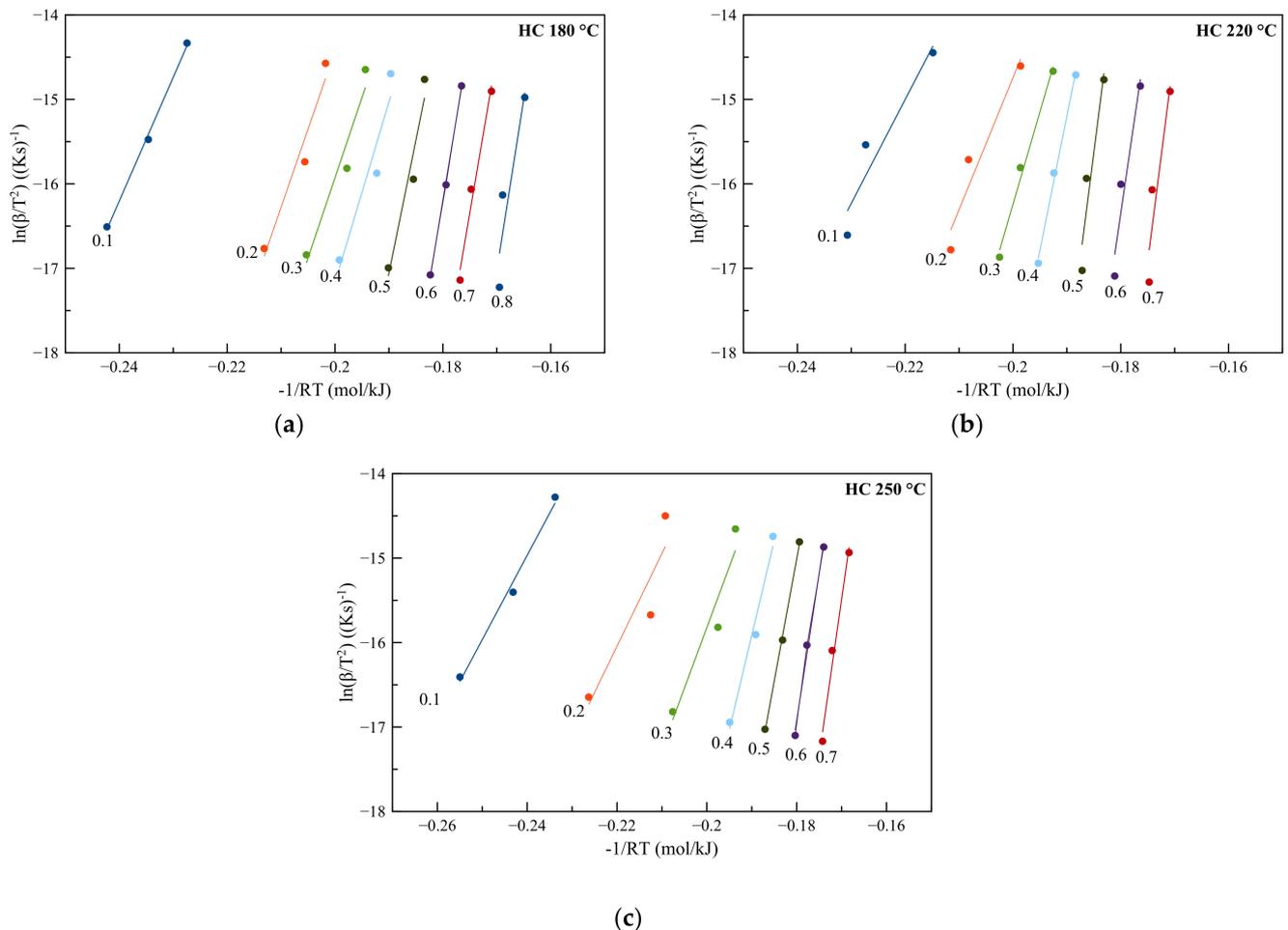
**Figure 5.**  $1-\alpha$  vs. temperature: comparison between experimental data (dots) and predicted data (lines) computed through the Gaussian model, during pyrolysis of different samples. (a) hydrochar 180  $^{\circ}\text{C}$ ; (b) hydrochar 220  $^{\circ}\text{C}$ ; (c) hydrochar 250  $^{\circ}\text{C}$ .

### 3.4.2. Miura–Maki Model

Figure 6 shows the values of  $E$ , while Tables S1 and S2 (Supplementary Materials) report the values of both  $E$  and  $k_0$  with the correlation coefficient  $R^2$ . Figure 7 shows the plot of  $\ln(\beta/T^2)$  vs.  $-1/RT$  at different values of  $\alpha$ . Each line corresponds to a certain  $\alpha$ , while the three points on each line correspond to different heating rates. Each  $\alpha$  corresponds to a certain temperature and the sequence  $T_{\beta=1\text{ }^{\circ}\text{C}/\text{min}} < T_{\beta=3\text{ }^{\circ}\text{C}/\text{min}} < T_{\beta=10\text{ }^{\circ}\text{C}/\text{min}}$  has to be respected. The maximum value of conversion for which the correct sequence of temperature was respected is defined with  $\alpha_{\text{max}} < 1$ . This was found in the range of  $\alpha$  0.1–0.8 for pyrolysis and 0.1–0.9 for oxidation. Then, for the valid range of  $\alpha$ ,  $E$  and  $k_0$  were determined graphically using the schemes in Figure 7;  $E$  is given by the slope of each line for each  $\alpha$ , while  $k_0$  is obtained from the intercept and by applying Equation (13).



**Figure 6.** Distribution of  $E$  at different  $\alpha$  of hydrochars, computed through the Miura–Maki model during (a) pyrolysis and (b) oxidation; per mol of dry feedstock.



**Figure 7.** Miura–Maki diagrams of pyrolysis of: (a) hydrochar 180 °C; (b) hydrochar 220 °C; (c) hydrochar 250 °C.

Under pyrolysis, the fitting is satisfactory, with a  $R^2$  always higher than 0.95, except for one outlier at  $\alpha = 0.7$  for the 220 °C hydrochar. Results show that  $\alpha$  hugely affects the kinetic parameters. All the hydrochars present an  $E$  that increases with the degree of conversion: it rapidly passes from 100–147 kJ/mol (at  $\alpha = 0.1$ ) to 371–386 kJ/mol in a

range of  $\alpha$  0.6–0.7. This phenomenon can be explained by the progressive increase of the degree of carbonization with temperature. As pyrolysis proceeds, the biomass undergoes volatilization and is converted into a high-carbon content matrix. This char is more difficult to degrade than the volatile matter and therefore causes a higher  $E$ . Generally, values are considerably higher than those commonly found in literature for biomass substrates (for example, for lignin it is 237–267 kJ/mol, and for pine wood it is 186–271 kJ/mol [27]), which can be explained by the nature of the substrate. Meanwhile,  $k_0$  reaches its maximum at an  $\alpha$  of 0.5–0.6, with maximum values of  $10^{24-27}$  for the 180 and 250 °C hydrochars and  $10^{38}$  for the 220 °C one.

Figure 6b shows how  $E$  of different hydrochars vary during oxidation. For the 180 and 220 °C hydrochars,  $E$  increases up to  $\alpha = 0.3-0.5$  to values of 357–379 kJ/mol. Meanwhile, the 250 °C hydrochar shows a very similar trend to the 220 °C one up to an  $\alpha$  of 0.35, and then stabilizes at much lower values of 221–225 kJ/mol. The 180 °C hydrochar shows higher values of  $E$  at the beginning of the conversion. Therefore, the HTC severity decreases the kinetic parameters of both volatile matter and char produced during the process. Similar trends were observed by Bach et al. [20] on hydrochars produced from wood residues. Meanwhile, pre-exponential factors  $k_0$  show a very similar trend to  $E$  (they reach their maxima in correspondence of the maximum of  $\alpha$ , arriving up to  $10^{24-27}$  for the 180 and 220 °C hydrochars, and  $10^{14}$  for the 250 °C one).

#### 3.4.3. Comparison and Suggestions for Future Work

Table 6 reports a comparison table with the average values of  $E$ ,  $\sigma$ , and  $k_0$ , computed through the Gaussian and Miura–Maki models.

**Table 6.** Average values of  $E$ ,  $\sigma$ , and  $k_0$  computed through the Gaussian and Miura–Maki models ( $k_0$  constant and equal to  $1.67 \times 10^{13} \text{ s}^{-1}$  for the Gaussian model).

Model	Atmosphere	Parameter	Grape Seeds	Oil	Hydrochar		
					180 °C	220 °C	250 °C
Gaussian	Pyrolysis	$E$ (kJ/mol)	193	218	203	203	206
		$\sigma$ (kJ/mol)	16	11	15	15	16
	Oxidation	$E$ (kJ/mol)	213	-	213	214	220
		$\sigma$ (kJ/mol)	17	-	15	15	15
Miura-Maki	Pyrolysis	$E$ (kJ/mol)	-	-	265	339	239
		$k_0$ ( $\text{s}^{-1}$ )	-	-	$7.6 \times 10^{27}$	$6.0 \times 10^{38}$	$9.8 \times 10^{24}$
	Oxidation	$E$ (kJ/mol)	-	-	222	215	181
		$k_0$ ( $\text{s}^{-1}$ )	-	-	$3.4 \times 10^{26}$	$1.0 \times 10^{26}$	$2.9 \times 10^{13}$

Due to its starting hypothesis, i.e., approximating the profile with a single Gaussian curve, the Gaussian model does not highlight the effect of the severity of the HTC process on  $E$ , which ranges between 193 and 206 kJ/mol for pyrolysis. This does not occur for the Miura–Maki model, which assumes a distribution of  $E$  that varies on the entire range leading to more diverse average values. Indeed, the Gaussian approach models a decomposition involving several stages through a single decomposition stage characterized by a Gaussian profile of the activation energy. Therefore, this assumption “averages” the profiles, canceling the differences among the samples obtained at different severities. Meanwhile, the Miura–Maki model avoids this averaging since it does not impose a similar strong assumption on the distribution.

Since the literature lacks the modeling of the kinetics of pyrolysis of hydrochar derived from grape seeds, the comparison of the results obtained was possible considering typical values of activation energy for decomposition of the main constituents of lignocellulosic materials. In particular, cellulose decomposition activation energy ranges between 175 and 279 kJ/mol [21,27,50] hemicellulose is in the range 132–186 kJ/mol [27,51,52], whereas

lignin ranges from 62 to 271 kJ/mol [27,50–52], with the broadest range. These values seem to prove the consistency of the results obtained in the present work.

In general, the Gaussian model has the advantage of being valid over the entire range of conversion, while the Miura–Maki model requires always that the temperature sequence must be satisfied, an aspect that is not obvious (in this study this occurs only for  $\alpha$  in the range of 0.1–0.8). Overall, both the models suffer from a strong starting hypothesis:  $k_0$  fixed for the Gaussian model and a strong integral simplification in the Miura–Maki model that can lead to an overestimation of the average values of the kinetics parameters [53].

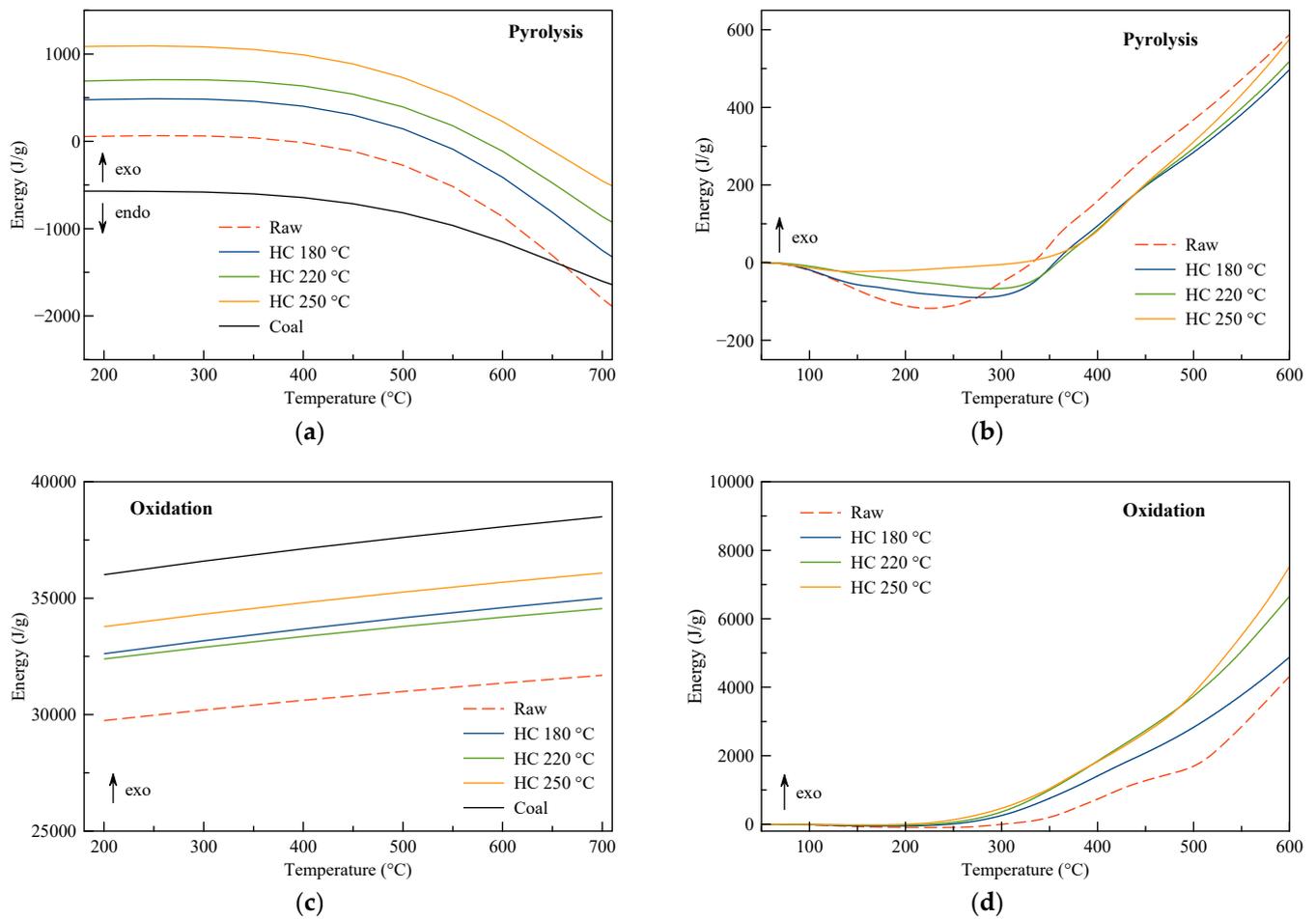
To improve the prediction, the authors suggest extending the single Gaussian model to a Multi Gaussian, in which the entire decomposition profile is divided into single decomposition peaks, each one approximated by a Gaussian curve [21]. Indeed, the single approach suits well homogeneous substrates (like the oil) characterized by a single decomposition peak, but seems too simple for complex substrates like biomasses. As a result, future works should include a comparison with other kinetics models.

Then, applying other DAEMs (like Kissinger–Akahira–Sunose method and the Coats–Redfern method) on the same feedstock could help to validate the results. Apart from contributing to understanding the mechanisms behind the decomposition, kinetic parameters can help in technological design and optimization. Moving to a larger scale, integration with other modeling techniques is necessary. Among these, it is worth mentioning models that consider the effects of particle distribution and geometry on heat transfer phenomena (like fractal models [54,55]), statistical models [56], and comprehensive computational models [57].

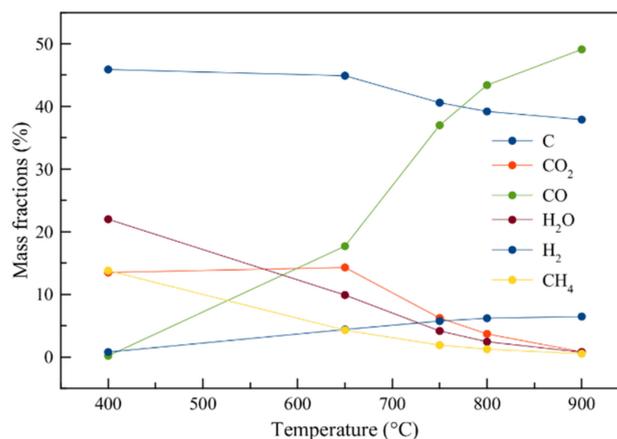
### 3.5. Aspen Plus Model

Figure 8a,c show the trend of heat produced/required as computed by Aspen Plus under pyrolytic and oxidative atmosphere.

Regarding pyrolysis, all samples follow a similar trend, characterized by an exothermic behavior at low temperatures and an endothermic one at high temperatures. Figure 9 reports the trends of the products at different pyrolysis temperatures resulting from the 180 °C hydrochar (the other samples present the same qualitative trends) and can help to interpret the heat trends. At low temperatures, thermodynamics favors exothermic reactions like char oxidation to CO<sub>2</sub> and CO, oxidation of hydrogen, and methanation. Conversely, higher temperatures enhance endothermic reactions, like the Boudouard reaction (and therefore the conversion of CO<sub>2</sub> into CO), and methane steam reforming (thus the conversion of CH<sub>4</sub> and H<sub>2</sub>O to CO and H<sub>2</sub>). At low temperatures, more carbonized substrates show a higher exothermic behavior due to a higher carbon partial oxidation and methane production (they present a higher C/H ratio than less carbonized substrates). Conversely, at higher temperatures, less carbonized samples show a higher endothermic behavior, probably due to the higher production of CO. As expected from its high C/O ratio and low volatile matter content, the profile of coal is flatter than hydrochar, and it is always endothermic. The different starting points can be explained by the higher ash content of coal than hydrochars (13.7% vs. 1.9–2.5%): ash, being not reactive, lowers the energy input/output per unit of mass involved. The slight difference in the coal profile with the previous work of our group [40] is due to the different inlet temperatures (isotherm to the reactors here vs. at ambient temperature in [31]).



**Figure 8.** Energy released (> 0) or absorbed (< 0) by the samples computed from: (a) Aspen Plus during pyrolysis; (b) DSC data during pyrolysis at 10 °C/min; (c) Aspen Plus during oxidation; (d) DSC data during oxidation at 10 °C/min; per grams of dry feedstock.



**Figure 9.** Aspen Plus results: main products from pyrolysis of hydrochar 180 °C.

Besides, Figure 8b shows the progressive integration of pyrolytic DSC data; computing the energy integral is necessary because DSC measures the instantaneous heat flow absorbed or released (which depends on the “history” of the sample due to the thermal program), while thermodynamics considers the energy absorbed or released when converting all the reactants into products at that determined temperature (and therefore the values obtained from thermodynamics do not depend on the thermal history of the sample). Over-

all, DSC data show a trend towards a cumulative exothermic behavior, whose final value corresponds to the integral reported in Table 4. The different behavior concerning Aspen data can be due to the nature of the thermodynamic approach. Indeed, real systems do not behave as predicted by thermodynamics at low temperatures because of reaction kinetics constraints. Moreover, as previously mentioned, thermodynamics does not consider the “history” of what happened before a certain condition. Indeed, inside the DSC, the starting material progressively undergoes pyrolysis, and the material at every time is the result of what happened before (i.e., the thermal program). In addition, thermodynamics does not consider typical “out of equilibrium” products, like tarry compounds, always present in real systems. Therefore, the thermodynamic approach requires particular attention and criticism before being used.

Regarding oxidation, both thermodynamic and DSC curves are clearly exothermic, with values that progressively increase with temperature. Under both atmospheres, absolute values differ between the Aspen and DSC approaches. This is due to the mass involved: DSC curves measure the energy absorbed/released by an amount of material that progressively decreases with time (the process is occurring), while Aspen predicts the energy absorbed/released by the same mass at every temperature. Since the energy is normalized by the amount of starting feedstock (and not the mass measured at every temperature), the amount of heat computed through the DSC approach is smaller than the Aspen one. Finally, it is interesting to note that in an oxidative environment, and for the whole range of temperature investigated, the thermodynamic approach foresees the complete oxidation of the biomass with the production essentially of CO<sub>2</sub>, H<sub>2</sub>O, and residual ash (equal to about 2% of the biomass fed to the reactor, Table 1). Conversely, in an oxidative environment in DSC, at the maximum temperature reached of 600 °C, there is still a substantial mass of non-oxidized carbon: the residual mass at 600 °C is of the order of 30% (Table 4). Through this critical comparison, the two approaches, thermodynamic and DSC, turn out to actually be very distant from each other, which is an aspect worthy of interest.

#### 4. Conclusions

This work investigated the kinetic behavior during pyrolysis and oxidation of hydrochars derived from an agro-industrial residue (grape seeds). The topic was approached through experimental techniques (TGA and DSC) that were used to develop two different DAEMs for biomass decomposition. Indeed, there is a certain lack in the literature regarding hydrochar decomposition, especially for the computation of the kinetic parameters. Therefore, this work covers both the experimental aspects and the comparison of two DAEMs using a critical approach. Thermogravimetric analysis and differential scanning calorimetry highlighted the importance of the HTC severity on the decomposition profiles. Indeed, hydrochars obtained at 180 and 220 °C still decompose with two/three stages (pyrolysis/oxidation) due to the presence of lignocellulosic constituents and oil that only partially degraded during HTC. Meanwhile, the 250 °C hydrochar shows a more stable profile, with one/two decomposition peaks, highlighting that HTC temperature hugely affected the structure of the biomass. Interestingly, all hydrochars show a certain exothermic behavior during pyrolysis.

DAEMs predicted the kinetic parameters involved during the pyrolysis and oxidation processes, i.e., the activation energy and the pre-exponential factor. The single-stage Gaussian model does not highlight differences among the various hydrochars obtained at different HTC temperatures, with average values of activation energy in the range 203–206 kJ/mol for pyrolysis and 213–220 kJ/mol for oxidation. Actually, the Gaussian DAEM turned out to be unsatisfactory to model complex feedstock such as hydrochars characterized by multi-decomposition peaks. Meanwhile, the Miura–Maki model, even if not applicable over the entire decomposition region, enabled the determination of the distribution of activation energies: the more severe the HTC process, the lower the activation energy values.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/en15030950/s1>, Figure S1: (1- $\alpha$ ) at different temperatures of feedstock during: (a) pyrolysis; (b) oxidation. Heating rate: 10 °C/min.; Figure S2: Comparison between DTGA and DSC curves during pyrolysis at 10 °C/min of: (a) grape seeds and hydrochar 180 °C; (b) hydrochars 220 and 250 °C; Figure S3: Comparison between experimental and predicted data (1- $\alpha$  vs temperature) computed through the Gaussian model, during oxidation of different samples: (a) hydrochar 180 °C; (b) hydrochar 220 °C; (c) hydrochar 250 °C; Table S1: Details of Miura-Maki model, pyrolysis (on a dry basis); Table S2: Details of Miura-Maki model, oxidation (on a dry basis); Gaussian DAEM – MATLAB code.

**Author Contributions:** G.G.: methodology, software, data curation, writing—original draft preparation; G.I.: data curation, visualization, writing—original draft preparation; L.F. (Luca Fambri): experimental planning, writing—review and editing, supervision, project administration; L.F. (Luca Fiori): writing—review and editing, supervision, project administration. All authors have read and agreed to the published version of the manuscript.

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