## Aldehydes-aided lignin-first deconstruction strategy for facilitating lignin monomers and fermentable glucose production from poplar wood

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### **Appendix A. Supplementary material**

#### Chemicals

All the reagents were purchased from Sigma Chemical Co. (Beijing, China), except for cellulase (Cellic@ CTec2, 100 FPU/ml), which was kindly provided from Novozymes (Beijing, China). 5% Ru on carbon catalyst (Evonik Noblyst® P3060 5% Ru), (2-methoxy-4-methylphenol, >98%). methylguaiacol ethylguaiacol (4-ethyl-2-methoxyphenol, >98%), propylguaiacol (2-methoxy-4-propylphenol, >99%), (4-ethoxy-3-methoxybenzaldehyde, >98%), guaiac-aldehyde methylvanillate (4-Hydroxy-3- methoxy-benzoic acid methyl >97%), propylsyringol ester, (2,6-dimethoxy-4-propylphenol, 95%), propionaldehydsyringol [3-(4-Hydroxy-3,5-dimethoxyphenyl)-propionaldehyde, 95%], allylsyringol (4-allyl-2,6-dimethoxyphenol, 95%), ethanonesyringol [1-(4-Hydroxy-3,5dimethoxy-phenyl)-ethanone, 95%], 1,4-dioxane (99%), formaldehyde solution (36.5 wt % in H<sub>2</sub>O), acetaldehyde solution (40 wt% in H<sub>2</sub>O), sodium acetate, acetic acid, sulfuric acid and fuming hydrochloric acid (37%), all were analytical reagents. Methanol (>99%) and tetrahydrofuran (THF, >99%) were chromatographic grade reagent. Dimethylsulfoxide was deuterium reagent. All the reagents were used without further purification.

#### Solvents

Sodium acetate buffer: 2.1 g sodium acetate was added into 500 mL deionized water, and stirred until dissolved. The buffer was prepared by adjusting the pH to 4.8 with acetic acid.

#### The analysis of sugar

The filtrate of enzymatic hydrolysis was diluted with ultrapure water, and filtered through a 0.45 µm water phase needle filter (Jinteng, Tianjin; Diameter:13 mm,

Aperture pore, 0.45  $\mu$ m; Texture, PES). The final filtrate was directly added into injected bottle for HPAEC detection.

HPAEC system (Dionex ICS5000) with pulsed amperometric detector and an ion exchange Carbopac PA-1 column (4×250 mm). The neutral sugars were separated in 18 mM NaOH (carbonate free and purged with nitrogen) with post column addition of 0.3 M NaOH at a rate of 0.5 mL/min. Run time was 45 min, followed by a 10 min elution with 0.2 M NaOH to wash the column and then a 15 min elution with 18 mM NaOH to re-equilibrate the column. Calibration was performed with a standard solution of L-arabinose, L-glucose, L-galactose, D-mannose, D-xylose, glucuronic acid, and galacturonic acids. Measurements were conducted with two parallels, and reproducibility of the values was found within the range of 5%. The content of sugar was calculated as follow,

# $Sugar\% = \frac{C \times Dilute \text{ fold} \times Volume \times Conversion \text{ fraction}}{m \times Sugar\% \text{ in substrate}}$

Where, C, the concentration of sugar calculated by HPAEC system, mg/L;

Dilute fold, dilute with ultrapure water to ensure the result of HPAEC system within 25 mg/L;

Volume, the total volume of filtrate, mL;

Conversion fraction, 0.9 for hexose, 0.88 for pentose;

m, the mass of substrate, mg;

Sugar% in substrate, obtained from component analysis of substrate;

#### **Component analysis of substrate**

3 mL 72%H<sub>2</sub>SO<sub>4</sub> was added into a hydrolysis of bottle with 300 mg substrate and hydrolyzed in a water bath at 30 °C for 1 h, stirred every 10 minutes to make it hydrolyzed as completely as possible. After the strong acid hydrolysis, 84 mL of deionized water was added to reduce the concentration of H<sub>2</sub>SO<sub>4</sub> to 4%, and the bottle was placed in a autoclave at 121 °C for 1 h. After the reaction, the supernatant was filtered and used for the determination of sugar content with HPAEC.

#### Correction

The yield of lignin fractions was corrected to exclude the impact of the attached aldehydes according to the results of 2D-HSQC NMR.

	No incorporation	FA	AA
S	226	240	254
G	196	210	224

Mass of the monolignols with solvent incorporation.

Correction factors of the monolignols with different aldehydes

Correction factor	FA	AA
S	0.0619	0.1239
G	0.0714	0.1429

The correction yield of lignin during the lignin-first strategy (%)

	LFA	$L_{AA}$	L <sub>BM-FA</sub>	L <sub>BM-AA</sub>
Correction yield	68.0	85.1	82.5	86.5

Based on the total lignin in the biomass

	conditions			
Labels	$\delta_C/\delta_H \left( L_{Control} \right)$	$\delta_C/\delta_H (L_{FA})$	$\delta_C\!/\delta_H\left(L_{AA}\right)$	Assignments
$\mathbf{B}_{\beta}$	53.5/3.02	53.5/3.02	53.5/3.02	$C_{\beta}$ -H <sub><math>\beta</math></sub> in resinol substructures (B)
-OCH <sub>3</sub>	55.5/3.69	55.5/3.69	55.5/3.69	C-H in methoxyls
$A_{\gamma}$	59.5/3.70 and 3.56	59.5/3.70 and 3.56	59.5/3.70 and 3.56	$C_{\gamma}$ - $H_{\gamma}$ in $\beta$ - $O$ -4 substructures (A)
$A'_{\gamma}$	64.1/4.47	64.1/4.47		$C_{\gamma}$ -H <sub><math>\gamma</math></sub> in $\gamma$ -acylated $\beta$ -O-4 substructures (A)
$A''_{\gamma}$		68.2/3.99 and 3.67	68.2/3.99 and 3.67	$C_{\gamma}$ -H <sub><math>\gamma</math></sub> in shifted $\beta$ -O-4 substructures (A)
$\mathbf{B}_{\gamma}$	71.4/4.17 and 3.86	71.4/4.17 and 3.86	71.4/4.17 and 3.86	$C_{\gamma}$ -H <sub><math>\gamma</math></sub> in resinol substructures (B)
$A_{\alpha}$	71.7/4.85	71.7/4.85	71.7/4.85	$C_{\alpha}$ -H <sub><math>\alpha</math></sub> in $\beta$ -O-4 substructures (A)
$A''_{\alpha}$		73.4/4.24	73.4/4.24	$C_{\alpha}$ -H <sub><math>\alpha</math></sub> in shifted $\beta$ -O-4 substructures (A)
$A_{\beta}(G)$	83.5/4.41		83.5/4.41	$C_{\beta}$ -H <sub><math>\beta</math></sub> in $\beta$ -O-4 linked to a G/H unit (A)
$A_{\beta}''(G)$		81.7/4.48	81.7/4.48	$C_\beta\text{-}H_\beta$ in shifted $\beta\text{-}O\text{-}4$ linked to a G unit (A)
$\mathbf{B}_{\alpha}$	84.9/4.62	84.9/4.62	84.9/4.62	$C_{\alpha}$ -H <sub><math>\alpha</math></sub> in resinol substructures (B)
$A_{\beta}(S)$	86.0/4.12			$C_{\beta}$ -H <sub><math>\beta</math></sub> in $\beta$ -O-4 linked to a S unit (A)
S <sub>2,6</sub>	103.5/6.62	103.5/6.62	103.5/6.62	C <sub>2,6</sub> -H <sub>2,6</sub> in syringyl units (S)
S' <sub>2,6</sub>	106.2/7.27			C <sub>2,6</sub> -H <sub>2,6</sub> in oxidized(C=O) phenolic syringyl units (S)
G <sub>2</sub>	110.6/6.91	110.6/6.91		C <sub>2</sub> -H <sub>2</sub> in guaiacyl units (G)
G <sub>5</sub>	114.9/6.76	114.9/6.76	114.9/6.76	C <sub>5</sub> -H <sub>5</sub> in guaiacyl units (G)
G <sub>6</sub>	118.8/6.77	118.8/6.77	118.8/6.77	C <sub>6</sub> -H <sub>6</sub> in guaiacyl units (G)
PB <sub>2,6</sub>	131.2/7.66	131.2/7.66	131.2/7.66	C <sub>2,6</sub> -H <sub>2,6</sub> in <i>p</i> -hydroxybenzoate units (S)

Table S1 Assignments of  ${}^{13}C$ - ${}^{1}H$  cross-signals in the HSQC spectra of lignin obtained from different

Entry <sup>a</sup>	Α	В	С	D	Ε
Lignin	L <sub>Control</sub>	L <sub>FA</sub>	LAA	L <sub>FA</sub>	LFA
Solvent	THF	THF <sup>b</sup>	THF	MeOH	Dioxane
Con/w% <sup>c</sup>	16.55	42.57	33.00	33.07	26.58
1	1.16 <sup>c</sup> (7.01 <sup>d</sup> )	ND	1.30 (3.94)	ND <sup>e</sup>	ND
2	1.23 (7.43)	1.15 (2.70)	1.33 (4.03)	1.15 (3.48)	1.22 (4.59)
3	1.41 (8.52)	3.51 (8.25)	2.56 (7.76)	2.68 (8.10)	2.33 (8.77)
4	2.47 (14.93)	2.77 (6.51)	2.73 (8.27)	3.03 (9.16)	2.73 (10.27)
5	1.50 (9.06)	1.15 (2.70)	2.63 (7.97)	1.03 (3.12)	1.08 (4.06)
6	4.20 (25.38)	17.00 (39.93)	15.38 (46.61)	10.81 (32.69)	7.30 (27.47)
7	3.07 (18.55)	13.44 (31.57)	3.42 (10.36)	13.08 (39.56)	8.83 (33.22)
8	ND	1.92 (4.51)	1.96 (5.94)	1.29 (3.90)	1.55 (5.83)
9	1.51 (9.12)	1.63 (3.83)	1.69 (5.12)	ND	1.54 (5.79)

**Table S2** Detailed yields of the main aromatic products from the lignin depolymerization reaction over different conditions (based on the weight of starting lignin)

<sup>a</sup> (A)  $L_{Control}$  degraded in THF as solvent system. (B)  $L_{FA}$  degraded in THF as solvent system. (C)  $L_{AA}$  degraded in THF as solvent system. (D)  $L_{FA}$  degraded in MeOH as solvent system. (E)  $L_{FA}$  degraded in dioxane as solvent system.

<sup>b</sup> THF, tetrahydrofuran

° The conversion ratio of lignin is based on the weight of starting lignin

<sup>d</sup> The selectivity of monomer is based on the total monomer yield

<sup>c</sup> Not detected

F (	Retention time		Character and	
Entry	(min)	Component	Structure	
1	18.455	2-Methoxy-4-methyl-phenol	0 H	
2	22.295	4-Ethyl-2-methoxy-phenol	OH O	
3	26.216	2-Methoxy-4-propyl-phenol	OH O	
4	29.493	4-Ethoxy-3-methoxy-benzaldehyde		
5	32.809	4-Hydroxy-3-methoxy-benzoic acid methyl ester	OH OH	
6	34.385 (36.026)	2,6-Dimethoxy-4-propyl-phenol		
7	37.406	3-(4-Hydroxy-3,5-dimethoxy- phenyl)-propionaldehyde		
8	39.701	4-Allyl-2,6-dimethoxy-phenol		
9	42.16	1-(4-Hydroxy-3,5-dimethoxy-phen yl)-ethanone		

Table S3 The main components of the degraded products

Sample	Cellulose	Hemicelluloses	Klason lignin	Acid-soluble lignin
Raw	45.82	21.06	21.80	0.98
R <sub>Control</sub>	78.62	6.40	3.19	3.34
$\mathbf{R}_{\mathrm{FA}}$	74.53	3.00	7.59	3.76
R <sub>AA</sub>	89.19	0.68	3.09	2.63
$R_{H\text{-}FA}$	80.25	0.36	13.79	0.54
R <sub>H-AA</sub>	84.08	0.15	3.71	0.61

 Table S4
 The composition analysis of the control and delignified substrates under different conditions

	24 h	18 h
	24 H	48 11
R <sub>Control</sub>	40.10	61.40
$\mathbf{R}_{\mathrm{FA}}$	13.28	15.40
R <sub>AA</sub>	50.38	75.12
R <sub>H-FA</sub>	28.17	30.38
R <sub>H-AA</sub>	58.19	85.14

Table S5 The glucose yield of the substrates after lignin extraction.

#### **Figure Caption**

Fig. S1. The chemical reaction occurs between aldehydes (formaldehyde and acetaldehyde) and  $\alpha$ -and  $\gamma$ -OH in the side-chain of lignin.

Fig. S2. GC-MS Chromatographic of the products obtaining from the degradation of different lignin samples and solvent system. (A)  $L_{Control}$  degraded in THF as solvent system. (B)  $L_{FA}$  degraded in THF as solvent system. (C)  $L_{AA}$  degraded in THF as solvent system. (D)  $L_{FA}$  degraded in MeOH as solvent system. (E)  $L_{FA}$  degraded in dioxane as solvent system.

Fig. S3. XRD spectra of the raw material and treated substrates





Fig. S1



Fig. S2



Fig. S3