

Article

Enhanced Degradation of Pharmaceutical Compounds by a Microbubble Ozonation Process: Effects of Temperature, pH, and Humic Acids

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Abstract: This study systematically investigated the feasibility of the microbubble ozonation process to degrade the 17 α -ethinylestradiol, ibuprofen, and atenolol through the comparison with the millibubble ozonation process for elucidating the degradation behavior and mechanisms during the microbubble ozonation processes. The proportions of small microbubbles (diameter 1–25 μ m) were increased with increasing the cavity pump frequency (40 Hz: 51.4%; 50 Hz: 57.5%; 60 Hz: 59.9%). The increased concentrations of O₃ and OH radicals due to the higher specific area of O₃ microbubbles compared to O₃ millibubbles could facilitate their mass transfer at the gas–water interface. Furthermore, the elevated reactivity of O₃ by increasing the temperature might improve the degradation of the pharmaceutical compounds, which was more pronounced for the microbubble ozonated waters than the millibubble ozonated waters. Although the degradation efficiency of the pharmaceutical compounds during the microbubble ozonation processes was significantly influenced by the existence of humic acids compared to the millibubble ozonation process, the increased solubilization rate of O₃ and OH radicals by collapsing O₃ microbubbles enhanced the degradation of the pharmaceutical compounds. Overall, these results clearly showed that the microbubble ozonation process could be an alternative option to conventional ozonation processes for the abatement of the pharmaceutical compounds.

Keywords: microbubbles; millibubbles; OH radicals; ozonation; pharmaceutical compounds

1. Introduction

In recent years, the occurrence and fate of trace organic compounds, including pharmaceutical compounds, personal care products, and endocrine-disrupting chemicals in surface water (i.e., lake water and river water), wastewater, and groundwater have become an emerging concern over the world since a considerable amount of used and/or metabolized organic compounds are discharged from municipal wastewater treatment plants (WWTPs) to the receiving surface water bodies in highly urbanized areas [1–3]. Most of trace organic compounds can be efficiently removed by biological wastewater treatment processes (conventional activated sludge, anoxic-oxic, and anaerobic–anoxic–oxic processes) generally applied in WWTPs, whereas hydrophilic chemical compounds (i.e., pharmaceutical compounds) tend to be recalcitrant to microbial activities, which may increase the concentrations of the used pharmaceutical compounds in lake water and surface water [4,5]. Even though used and/or

metabolized pharmaceutical compounds present at a very low concentration in secondary effluents (i.e., ng L⁻¹ range), the long-term exposure to them can provide adverse and/or harmful effects on human health and aquatic ecosystems [6,7]. Therefore, the application of advanced water treatment techniques and the assessment of their efficiencies have gained considerable attention for reducing the discharge of pharmaceutical compounds from municipal WWTPs to the receiving surface water bodies (e.g., lake water, river water, and groundwater) [8].

A few treatment methods, including coagulation-flocculation, activated carbon adsorption (powdered activated carbon and granular activated carbon), oxidative processes, and membrane processes, are available for the removal of pharmaceutical compounds from surface water and wastewater [9]. Among them, ozonation is considered to be a promising option as a tertiary wastewater treatment step in WWTPs for reducing the discharge of pharmaceutical compounds to river water and lake water, as ozone (O₃) has a high reactivity towards most of trace organic compounds and self-decomposition characteristics to oxygen compared to other oxidizing agents (e.g., chlorine, hydrogen peroxide) [10]. Many researchers found that ozonation is efficient for disinfection, deodorization, and decoloration, and can effectively eliminate pharmaceutical compounds, personal care products, and endocrine-disrupting chemicals in surface water and wastewater [11,12]. However, O₃ gases injected into the aqueous phase generally remains in the gaseous phase because of its low solubility in water (1.0×10^{-6} mol m⁻³·Pa– 1.3×10^{-4} mol m⁻³·Pa), which may allow O₃ gases existing in surface water and wastewater to pass through the reacting zone of the O₃ reactors without the chemical reactions with trace organic compounds [13]. Based on these reasons, it is required to develop a novel method for improving the solubilization rate of O₃ gases injected in surface water and wastewater closely associated with the removal of pharmaceutical compounds [14].

During the last decades, conventional ozonation (i.e., millibubble ozonation) processes, in which sizes ranged from 2 to 5 mm varies significantly depending on the type of gas spargers, have been used in conventional ozonation processes for drinking water and wastewater treatments [15]. Consequently, a great amount of O₃ gases are necessary for conventional ozonation processes, which can lead to a high level of energy consumption [16]. Some studies recently reported that the use of the microbubble, which is defined as a bubble with a volume equivalent diameter in the range of 1 µm to 100 µm by the International Organization for Standardization (ISO; ISO 20480-1:2017) for ozonation processes is efficient to increase the degradation of trace organic compounds by enhancing the solubilization rate of O₃ in the aqueous phase [15,17,18]. Compared to the millibubble ozonation process, the microbubble ozonation processes can transfer the gaseous phase to the aqueous phase more effectively due to their much lower rising velocity, higher surface area, and greater curvature [19]. Furthermore, the increased concentrations of OH radicals in the aqueous phase by collapsing the O₃ microbubbles may additionally contribute to the decrease of trace organic compounds through the hydrogen abstraction, electron transfer, electrophilic addition, and radical-radical reactions [10,20]. In spite of the several benefits, microbubble ozonation technologies have been rarely applied to enhance sludge solubilization, treat textiles wastewater, and remove pesticides from vegetables in the food industries [18,21,22]. Moreover, a comprehensive study has not been performed yet to assess the feasibility of the microbubble ozonation processes for the reduction of the pharmaceutical compounds. Therefore, the fundamental knowledge of the microbubble ozonation process still remained incomplete, including the degradation behavior and mechanisms of the pharmaceutical compounds.

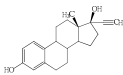
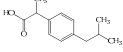
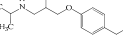
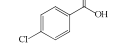
The main purpose of this study was to provide valuable insights into the degradation behavior of the pharmaceutical compounds during the microbubble ozonation processes. Hence, the bubble size effects on the solubilization rate of O₃ and OH radicals were identified and directly correlated to the decrease of the pharmaceutical compounds to evaluate the efficiency of the microbubble ozonation process. Furthermore, the effects on temperature, pH, and humic acids on the degradation efficiency of the pharmaceutical compounds were systematically investigated to elucidate their degradation mechanisms in the microbubble ozonation processes.

2. Materials and Methods

2.1. Pharmaceutical Compounds and Reagents

Three different types of the pharmaceutical compounds, including 17 α -ethinylestradiol (EE2), ibuprofen (IBU), atenolol (ATE), were selected as the representative compounds with various functional groups closely related to the ozone reactivity, and para-chlorobenzoic acid (*p*CBA) was used as a probe chemical for OH radicals (Table 1). Ultrapure water (resistivity > 18 M Ω cm⁻¹) produced by a water purification system (NANOpure Diamond, Barnstead, Dubuque, IA, USA) was utilized to make stock solutions of these trace organic compounds (concentration of each trace organic compound = 1 mM). The buffer solutions were prepared using disodium hydrogen phosphate, potassium dihydrogen phosphate, sodium dihydrogen phosphate, boric acid, sodium hydroxide purchased from Sigma-Aldrich (St. Louis, Missouri, USA) for maintaining the pH of water samples during ozonation experiments (purity > 98%).

Table 1. The second order rate constants for the reactions of the target pharmaceutical compounds with O₃ at pH 7 and OH radicals.

Compounds (Abbreviation, Molar Mass (g mol ⁻¹))	Use	Structure	Electron-Rich Organic Moiety	pK _a	k _{O₃} at pH 7 (M ⁻¹ s ⁻¹)	k _{OH} at pH 7 (M ⁻¹ s ⁻¹)	UVA Detection (nm)	Ref.
17 α -Ethinylestradiol (EE2, 296.40)	Ovulation inhibitor		Phenol	10.4	3.0 × 10 ⁶	9.8 × 10 ⁹	230	[23,24]
Ibuprofen (IBU, 206.28)	Analgesic		Alkyl aromatic	4.9	9.6	7.4 × 10 ⁹	210	[25]
Atenolol (ATE, 266.34)	β -blocker		Secondary amine	9.6	1.7 × 10 ⁴	8.0 × 10 ⁹	251	[26]
Para-Chlorobenzoic acid (<i>p</i> CBA, 156.57)	OH radical probe compound		Chloro-aromatic	4.0	<0.2	5.0 × 10 ⁹	240	[8]

2.2. System Description

Figure 1 exhibits the schematic diagram of a lab-scale microbubble ozonation process consisting of a cavity pump (KTM20ND, Nikuni, Kawasaki, Japan), an O₃ generator (the range of 0.17–0.20 mg/s; LAB-01, Ozone Tech, Daejeon, Republic of Korea), an O₃ gas detector (OM 2021, Ozone Tech, Daejeon, Republic of Korea), and three reservoirs made of stainless steel for the feed water (effective volume = 20 L), O₃ dissolution (effective volume = 4 L), and O₃ reaction (effective volume = 10 L). The temperature of feed waters was kept constantly using a temperature controller (Maxircu CR-30, Daihan Scientific, Wonju-si, Gangwon-do, Republic of Korea). The feed water flowed into the cavity pump with O₃ gases produced using pure oxygen when the pressure of the cavity pump was lower than −0.04 MPA by a self-suction effect to produce O₃ microbubbles. The feed water containing O₃ microbubbles was pressurized in the dissolution tank by applying high pressure (Maximum pressure = 0.3 MPa) for increasing the solubilization rate of O₃ gases into the feed water, and then O₃ microbubbles in the feed water were destructed in the reactor under atmospheric pressure conditions.

2.3. Microbubble Ozonation

The feed water samples spiked with trace organic compounds, including *p*CBA, EE2, IBU, and ATE (final concentration of each compound = 2.0 μ M; reaction volume = 20 L) were fed to the microbubble ozonation process. During the microbubble ozonation process, the size distributions of O₃ microbubbles in the aqueous phase were controlled by changing the frequencies of the cavity pump (30–60 Hz). The reduction of the pharmaceutical compounds was investigated under various experimental conditions (temperature: 10–30 °C; pH: 4–10; the concentration of humic acids: 5 mg/L) to compare the efficiencies of the millibubble and microbubble ozonation processes. The feed and

ozonated water samples were collected at 0, 60, 120, 180, 240, and 300 s, and all the samples were stored in the refrigerator at 4 °C prior to analyses.

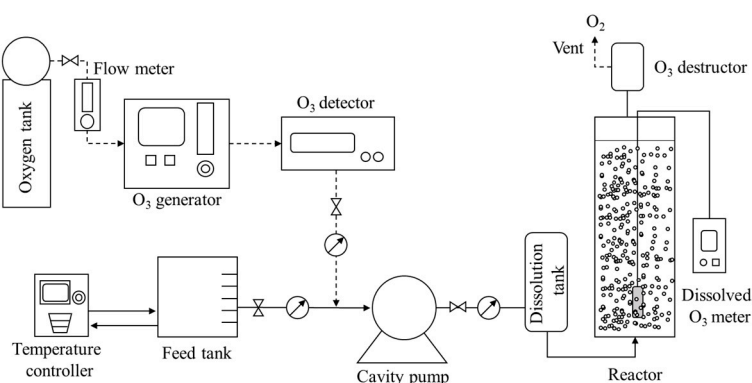


Figure 1. The schematic diagram of the lab-scale microbubble ozonation process.

2.4. Analytical Methods

The size distribution and numbers of microbubbles in the aqueous phase were measured by a liquid particle counter (PC3400, Chemtrac, Norcross, GA, USA). The concentration of O_3 gas and dissolved O_3 in the water samples were determined using the O_3 detector and the indigo method (2518025 Ozone Accuvac® Ampules, Hach, Loveland, CO, USA), respectively [27]. The pH of the solutions was measured by a pH meter (Orion™ star A320, Thermo Fisher Scientific, San Jose, CA, USA). The concentrations of *p*CBA, EE2, IBU, and ATE in the feed and ozonated water samples were analyzed using a UV/Vis spectrophotometer (UV-1280, Shimadzu, Kyoto, Japan) with a 1 cm quartz cuvette (Hellma, Müllheim, Germany) 1 hour after the millibubble and microbubble ozonation processes [8]. To avoid the interferences of the residual O_3 in the aqueous phase on the measurements of the pharmaceutical compounds, sodium thiosulphate pentahydrate ($Na_2S_2O_3 \cdot 5H_2O$; Sigma-Aldrich, St. Louis, Missouri, USA) was used as an O_3 quenching reagent. The corresponding calibration plots of the trace organic compounds were linear over the range of 0.1 Mm–2.0 μ M (R^2 values at pH 7 and 20 °C: *p*CBA = 0.9994, EE2 = 0.9992, IBU = 0.9991; ATE = 0.9993), as shown in Figure 2.

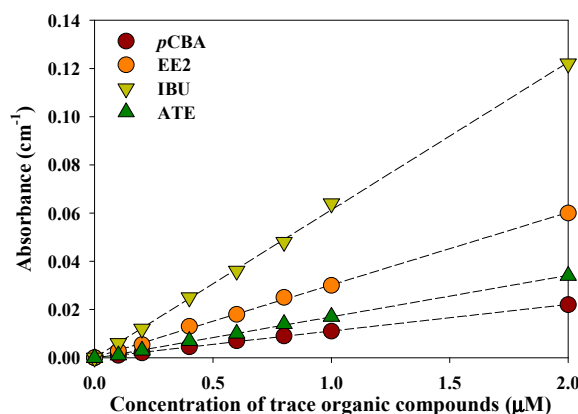


Figure 2. The calibration plots of *p*CBA, EE2, IBU, and ATE in the range of 0.1 μ M to 2.0 μ M (pH 7, temperature = 20 °C).

3. Results and Discussion

3.1. Size Distribution of Microbubbles

The size distribution and the total numbers of microbubbles as a function of the cavity pump frequency (30–60 Hz) are illustrated in Figure 3. In the tested microbubble ozonation process,

the generation of microbubbles was not detected at the cavity pump frequency of 30 Hz, whereas microbubbles seemed to generated stably when the cavity pump frequency was higher than 40 Hz (the total numbers of microbubbles at 40 Hz = 1901; the total numbers of microbubbles at 50 Hz = 2702; the total numbers of microbubbles at 60 Hz = 3368). For the tested frequency range of 40–60 Hz, the size of the microbubbles ranged commonly from 5 to 25 μm . The proportions of microbubbles having diameters of less than 25 μm were significantly increased when the cavity pump frequency was increased: (a) 40 Hz: 1–5 μm = 9.6%, 5–25 μm = 41.8%, 25–50 μm = 23.5%, 50–75 μm = 16.8%, 75–100 μm = 8.3%, (b) 50 Hz: 1–5 μm = 9.8%, 5–25 μm = 47.7%, 25–50 μm = 24.0%, 50–75 μm = 13.5%, 75–100 μm = 5.0%, and (c) 60 Hz: 1–5 μm = 9.9%, 5–25 μm = 50.0%, 25–50 μm = 24.3%, 50–75 μm = 11.8%, 75–100 μm = 4.0%). These results indicated that the size distribution of microbubbles is highly dependent on the swirl speed of the cavity pump [28].

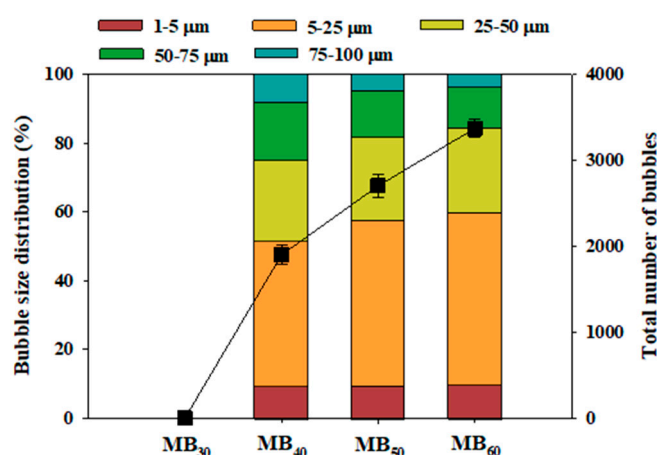


Figure 3. The size distributions and the total number of the microbubbles generated by the cavity pump at the different frequencies (temperature = 20 °C).

3.2. Bubble Size Effects on Solubilization Rate of O_3 and OH Radicals

Figure 4 represents the changes in the solubilization rate of O_3 and OH radicals in the aqueous phase depending on the size distribution of O_3 microbubbles. As shown in Figure 4a, dissolved O_3 was not detected for only the microbubble treated water at the cavity pump frequency of 60 Hz (MB₆₀) and the concentration of dissolved O_3 in the ozonated waters was gradually increased with increasing the the cavity pump frequency (millibubble ozonation at 30 Hz (O_3) = 0.50 mg L⁻¹; microbubble ozonation at 40 Hz (O_3 -MB₄₀) = 0.64 mg L⁻¹; microbubble ozonation at 50 Hz (O_3 -MB₅₀) = 0.88 mg L⁻¹; microbubble ozonation at 60 Hz (O_3 -MB₆₀) = 1.67 mg L⁻¹). The possible explanation for this observation is that the higher specific area (surface area per volume) of O_3 microbubbles with smaller diameters (1–25 μm) compared to O_3 millibubbles can enhance the dissolution of gaseous O_3 molecules into the aqueous phase [29]. The size effects of O_3 microbubbles on the solubilization rate of OH radicals were examined using pCBA (Figure 4b). Similarly to the trends in the solubilization rate of O_3 , the relative residual concentrations (C/C_0) of pCBA were rapidly decreased with increasing the cavity pump frequency (MB₆₀ = 1.00; O_3 = 0.93; O_3 -MB₄₀ = 0.89; O_3 -MB₅₀ = 0.85; O_3 -MB₆₀ = 0.60). Based on these observations, it can be concluded that the increased specific area of O_3 microbubbles plays a critical role in the mass transfer of OH radicals at the gas–water interface [30,31]. As there were no noticeable changes in the relative residual concentration (C/C_0) of pCBA for the microbubble treated water, it is evident that OH radicals cannot be produced by collapsing microbubbles without O_3 at the neutral condition (pH 7). In contrast to the current study, it has been previously reported that OH radicals can be generated during the destruction of microbubbles without O_3 at the acidic condition [30].

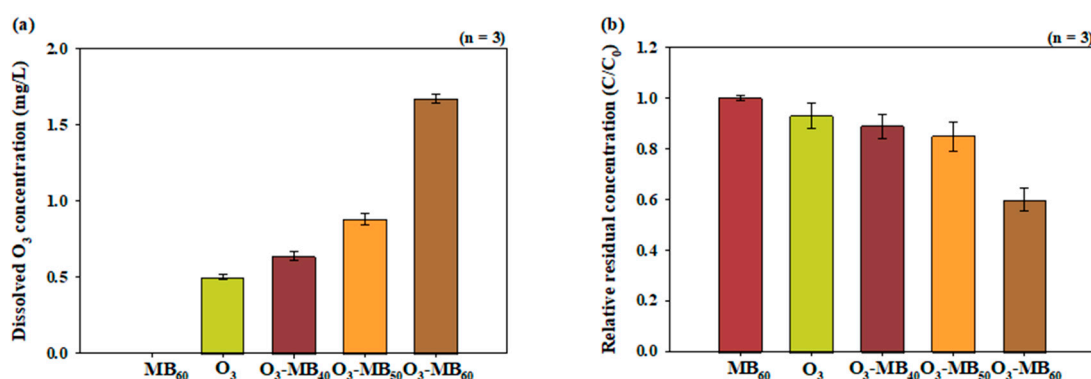


Figure 4. The changes in the concentrations of dissolved O₃ and the degradation of *p*CBA in the aqueous phase as a function of the cavity pump frequency (the initial concentration of *p*CBA = 2.0 μM; pH = 7.0; temperature = 20 °C; reaction time = 300 seconds; MB₆₀ = microbubble at 60 Hz without O₃; O₃ = millibubble ozonation at 30 Hz; O₃-MB₄₀ = microbubble ozonation at 40 Hz; O₃-MB₅₀ = microbubble ozonation at 50 Hz; O₃-MB₆₀ = microbubble ozonation at 60 Hz): (a) the concentration of dissolved O₃ and (b) the relative residual concentration (C/C₀) of *p*CBA.

3.3. Effects of Temperature on Degradation of the Pharmaceutical Compounds

The relative residual concentrations (C/C₀) of the selected pharmaceutical compounds, including EE2 (phenol), IBU (alkyl aromatic), and ATE (secondary amine), in the millibubble and microbubble ozonated waters under different temperature conditions (10–30 °C) are illustrated in Figure 5. Although the solubilization rate of O₃ and OH radicals in the aqueous phase was decreased with increasing the temperature of the solutions (the concentration of dissolved O₃ during millibubble ozonation: 0.65 mg/L at 10 °C, 0.50 mg/L at 20 °C, 0.40 mg/L at 30 °C; the concentration of dissolved O₃ during microbubble ozonation: 2.16 mg/L at 10 °C, 1.67 mg/L at 20 °C, 1.32 mg/L at 30 °C), the relative residual concentrations (C/C₀) of EE2, IBU, and ATE were gradually reduced during both millibubble (C/C₀ at 10 °C: EE2 = 0.80, IBU = 0.90; ATE = 0.90; C/C₀ at 20 °C: EE2 = 0.79, IBU = 0.88; ATE = 0.87; C/C₀ at 30 °C: EE2 = 0.78, IBU = 0.86; ATE = 0.86) and microbubble ozonation processes (C/C₀ at 10 °C: EE2 = 0.66, IBU = 0.77; ATE = 0.79; C/C₀ at 20 °C: EE2 = 0.61, IBU = 0.75, ATE = 0.77; C/C₀ at 30 °C: EE2 = 0.60, IBU = 0.73, ATE = 0.71) as the increase of the temperature could elevate the reactivity of O₃ with the pharmaceutical compounds [32]. This phenomenon was much more significant for the microbubble ozonated waters than the millibubble ozonated waters, which was probably due to the fact that the increased concentrations of O₃ and OH radicals in the microbubble ozonated waters might strongly enhance the ozonation efficiency of the pharmaceutical compounds [29–31]. In the tested temperature range of 10–30 °C, EE2 was found to be most reactive with O₃ because it included a phenolic moiety. Furthermore, an intermediate reactivity with O₃ was found for ATE containing a secondary amine moiety, and IBU showed the highest relative residual concentration because of its alkyl aromatic moiety, which is less reactive with O₃ [8]. These observations implied that the degradation of the pharmaceutical compounds during the millibubble and microbubble ozonation processes were strongly influenced by both the solubilization rate and the reactivity of O₃ and OH radicals with the pharmaceuticals.

3.4. Effects of pH on the Degradation of Pharmaceutical Compounds

The variations in the relative residual concentrations (C/C₀) of the pharmaceutical compounds during the millibubble and microbubble ozonation processes as a function of the pH of the solutions are depicted in Figure 6. In general, the reactivity of the pharmaceutical compounds with O₃ strongly depended on their pK_a values (EE2 = 10.4; IBU = 4.9; ATE = 9.6) [23–26]. Therefore, the relative residual concentrations of EE2 and ATE were proportionally reduced with increasing the pH of the solutions during the millibubble (C/C₀ at pH 4: EE2 = 0.80; ATE = 0.89; C/C₀ at pH 7: EE2 = 0.79;

ATE = 0.87; C/C_0 at pH 10: EE2 = 0.73; ATE = 0.79) and microbubble ozonation processes (C/C_0 at pH 4: EE2 = 0.64, ATE = 0.79; C/C_0 at pH 7: EE2 = 0.61, ATE = 0.77; C/C_0 at pH 10: EE2 = 0.55, ATE = 0.63). These observations could be explained by the higher reactivity of the deprotonated phenolic and secondary amine moieties with O_3 compared to the non-deprotonated phenolic and secondary amine moieties [33]. In the case of IBU, its relative residual concentration in the millibubble ozonated and microbubble ozonated waters was gradually increased with increasing the pH of the solutions (C/C_0 of IBU in the millibubble ozonated water: pH 4 = 0.85, pH 7 = 0.88, pH 10 = 0.92; C/C_0 of IBU in the microbubble ozonated water: pH 4 = 0.72, pH 7 = 0.74, pH 10 = 0.80). These results might be attributed to the accelerated decomposition of O_3 in the aqueous phase at a higher pH value, which was intimately associated with the reduction of IBU [33]. The increased degradation of IBU containing an alkyl aromatic moiety, which is not reactive with O_3 over the tested pH range of 4–10 in the microbubble ozonated waters (C/C_0 of IBU = 0.72–0.80) compared to other millibubble ozonated waters (C/C_0 of IBU = 0.85–0.92), supported the assumption that the increased concentrations of O_3 and OH radicals by collapsing O_3 microbubbles greatly facilitated the abatement of the pharmaceutical compounds [15,29,31].

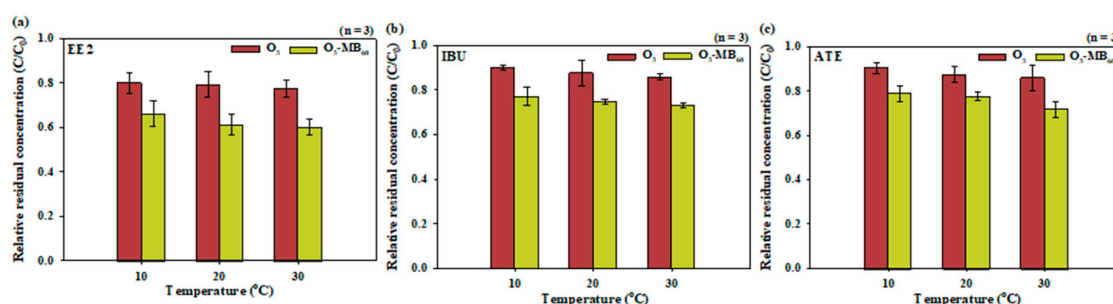


Figure 5. The effects of the temperature on the degradation of the pharmaceutical compounds during the millibubble and microbubble ozonation processes (the initial concentration of each pharmaceutical compound = 2.0 μ M; cavity pump frequency = 60 Hz; pH = 7; O_3 = millibubble ozonation at 30 Hz; O_3 -MB₆₀ = microbubble ozonation at 60 Hz): (a) EE2, (b) IBU, and (c) ATE.

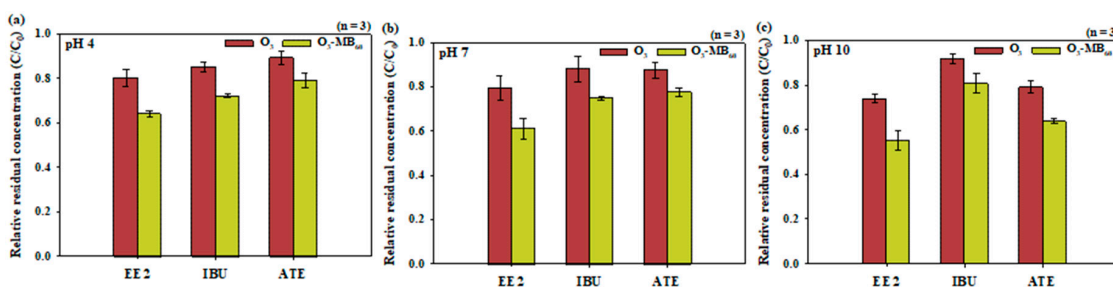


Figure 6. The pH effects on the degradation of the pharmaceutical compounds during the millibubble and microbubble ozonation processes (the initial concentration of each pharmaceutical compound = 2.0 μ M; cavity pump frequency = 60 Hz; temperature = 20 °C, reaction time = 300 seconds; O_3 = millibubble ozonation at 30 Hz; O_3 -MB₆₀ = microbubble ozonation at 60 Hz): (a) pH 4, (b) pH 7, and (c) pH 10.

3.5. Effects of Humic Acids on the Degradation of Pharmaceutical Compounds

The inferences on humic acids on the abatement of the pharmaceutical compounds during the millibubble and microbubble ozonation processes are compared in Figure 7. The relative residual concentrations of (C/C_0) of EE2, IBU, and ATE in the millibubble and microbubble ozonated waters were gradually decreased with increasing the exposure time to O_3 neither without nor with humic acids since the reduction of organic compounds was directly proportional to the oxidant exposure (the

time-dependent oxidant concentrations integrated over the reaction time) [34]. In the presence of humic acids, a considerable decrease in the degradation of the pharmaceutical compounds was found for either the millibubble (C/C_0 without humic acids after 300 seconds: EE2 = 0.80, IBU = 0.89, ATE = 0.88; C/C_0 with humic acids after 300 seconds: EE2 = 0.84, IBU = 0.95, ATE = 0.93) or microbubble ozonated waters (C/C_0 without humic acids after 300 seconds: EE2 = 0.60, IBU = 0.78, ATE = 0.74; C/C_0 with humic acids after 300 seconds: EE2 = 0.71, IBU = 0.86, ATE = 0.84). Despite of the higher degradation efficiency of EE2 and ATE during the microbubble ozonation process, the disturbance of humic acids on the decrease of the pharmaceutical compounds was more pronounced for the microbubble ozonated waters (the difference between the relative residual concentrations ($\Delta C/C_0$) of EE2 without and with humic acids after 300 seconds = 0.11; $\Delta C/C_0$ of ATE without and with humic acids after 300 seconds = 0.10) than the millibubble ozonated waters ($\Delta C/C_0$ of EE2 without and with humic acids after 300 seconds = 0.04; $\Delta C/C_0$ of ATE without and with humic acids after 300 seconds = 0.05) because of the phenolic moieties ubiquitous in humic acids, which might react readily with dissolved O_3 and OH radicals during the microbubble ozonation processes [35,36]. A similar behavior has been previously found for the oxidative treatment of dissolved organic matter with Cl_2 , and ClO_2 [27,37]. Compared to EE2 and ATE, the interference effects of humic acids on the degradation of IBU during the microbubble ozonation process ($\Delta C/C_0$ of IBU without and with humic acids after 300 seconds = 0.07) were not significantly different from those on the abatement of IBU during the millibubble ozonation process ($\Delta C/C_0$ of IBU without and with humic acids after 300 seconds = 0.05) since the reaction of the phenolic moiety in humic acids with O_3 could produce O_3 , which was readily decomposed to OH radicals in association with the decrease of IBU [33,38]. The enhanced degradation efficiency of the pharmaceutical compounds in the microbubble ozonated waters is evidence that the microbubble ozonation process is considered to be a promising option as an alternative to the conventional ozonation processes for the reduction of the pharmaceutical compounds.

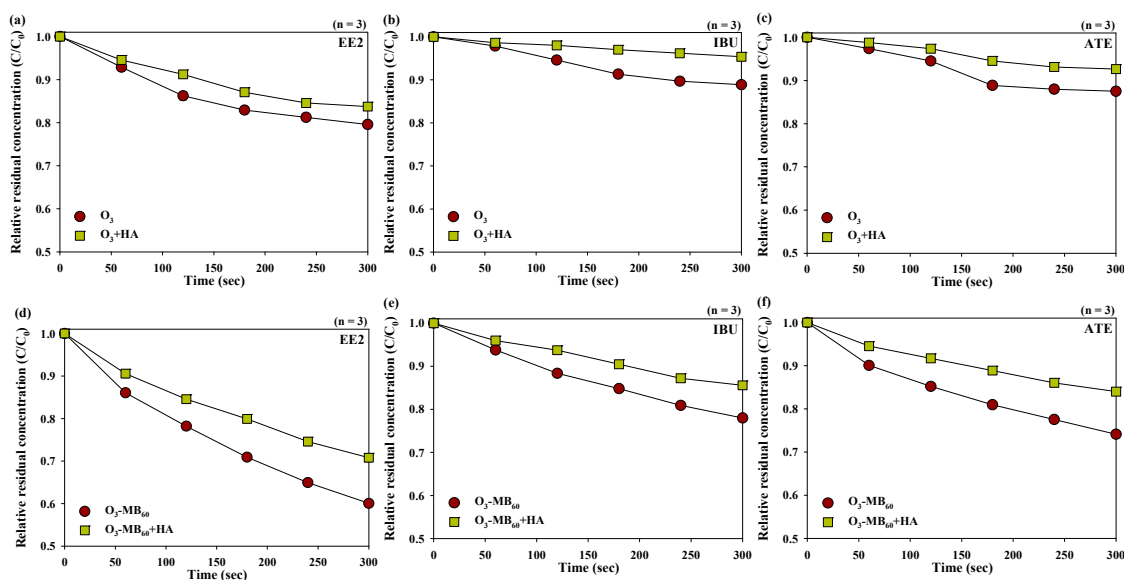


Figure 7. The effects of humic acids on the degradation of the pharmaceutical compounds during the (a–c) millibubble ozonation and (d–f) microbubble ozonation processes (the initial concentration of each pharmaceutical compound = 2.0 μ M; the concentration of humic acid = 5 mg/L; cavity pump frequency = 60 Hz; pH = 7.0; temperature = 20 $^{\circ}$ C).

4. Conclusions

In this study, the potential of the microbubble ozonation process for the abatement of the pharmaceutical compounds was evaluated and compared to the conventional ozonation process (i.e.,

millibubble ozonation) to provide deeper insights into the degradation behavior of the pharmaceutical compounds during the microbubble ozonation processes. The major outcomes are:

- The fractions of microbubbles whose diameters ranged from 1–25 μm gradually increased with increasing the cavity pump frequency (40 Hz: 51.4%; 50 Hz: 57.5%; 60 Hz: 59.9%).
- The increase of the solubilization rate of O_3 and OH radicals induced by their elevated mass transfer at the gas–water interface through the destruction of O_3 microbubbles with the higher specific area than O_3 millibubbles might substantially improve the degradation of the pharmaceutical compounds.
- Both the solubilization rate and the reactivity of O_3 and OH radicals provided strong effects on the degradation of the pharmaceutical compounds during the millibubble and microbubble ozonation processes over the temperature range of 10–30 $^\circ\text{C}$.
- The changes in the degradation of pharmaceutical compounds depending on the pH of the solution and the presence of humic acids were much more pronounced for the microbubble ozonated waters than the millibubble ozonated waters.
- The facilitatory and inhibitory effects of humic acids on the degradation of the pharmaceutical compounds were more significant in the microbubble ozonated waters compared to the millibubble ozonated waters.

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