Marine Drugs ISSN 1660–3397 www.mdpi.com/journal/marinedrugs

**OPEN ACCESS** 

Review

# **Neuroprotective Effects of Marine Algae**

Ratih Pangestuti<sup>1</sup> and Se-Kwon Kim<sup>1,2,\*</sup>

- <sup>1</sup> Marine Biochemistry Laboratory, Department of Chemistry, Pukyong National University, Busan 608–737, Korea; E-Mail: ratihpangestuti@pknu.ac.kr
- <sup>2</sup> Marine Bioprocess Research Center, Pukyong National University, Busan 608–737, Korea
- \* Author to whom correspondence should be addressed; E-Mail: sknkim@pknu.ac.kr; Tel.: +82-51-629-7094; Fax: +82-51-629-7099.

Received: 25 February 2011; in revised form: 12 April 2011 / Accepted: 28 April 2011 / Published: 10 May 2011

**Abstract:** The marine environment is known as a rich source of chemical structures with numerous beneficial health effects. Among marine organisms, marine algae have been identified as an under-exploited plant resource, although they have long been recognized as valuable sources of structurally diverse bioactive compounds. Presently, several lines of studies have provided insight into biological activities and neuroprotective effects of marine algae including antioxidant, anti-neuroinflammatory, cholinesterase inhibitory activity and the inhibition of neuronal death. Hence, marine algae have great potential to be used for neuroprotection as part of pharmaceuticals, nutraceuticals and functional foods. This contribution presents an overview of marine algal neuroprotective effects and their potential application in neuroprotection.

Keywords: marine algae; neuroprotective; neuroprotection

# **1. Introduction**

Ninety percent of the world's living biomass is found in the oceans with marine species comprising approximately half of the total global biodiversity [1,2]. This wide diversity of organisms is being recognized as a reservoir of potent molecules which are elicited by marine organisms to help them survive in the hostile environment [2,3]. Among marine organisms, marine algae have been identified as an under-exploited plant resources [4,5]. The term marine algae, as used herein, generally refer to marine macroalgae or sometimes referred to as seaweeds.

Marine algae can be classified into three classes based on their pigmentation, namely brown, red, and green algae, which are referred to as Phaeophyceae, Rhodophyceae, and Chlorophyceae, respectively [6]. Since the 1940s, production of algal polysaccharides has attained commercial significance through their application as thickening and gelling agents for various industrial applications [7]. Moreover, marine algae are recognized as rich sources of structurally diverse biologically active compounds with great pharmaceutical and biomedical potential. Researchers have revealed that marine algal originated compounds exhibit various biological activities such as anticoagulant [8,9], anti-viral [10,11], antioxidant [12–14], anti-allergic [15], anti-cancer [16], anti-inflammatory [17], anti-obesity [18–20], *etc.* Furthermore, several scientific studies have long been used in food diets as well as traditional remedies in Eastern countries and more recently in Europe and America. Hence, marine algae have great potential to be used in neuroprotection [21].

In recent years, biological activities, nutritional value, and potential health benefits of marine algae have been intensively investigated and reviewed. This review, however, focuses specifically on the neuroprotective effects of marine algae and emphasizes their potential application as future pharmaceutical candidates to prevent neurodegenerative diseases.

# 2. Bioactivities and Neuroprotective Effects of Marine Algae

## 2.1. Antioxidant

Oxidative stress is the result of an imbalance between pro-oxidant and antioxidant homeostasis that leads to the generation of toxic reactive oxygen species (ROS) [22]. Compared to other parts of our body, the central nervous system (CNS) is more sensitive to oxidative stress due to its high oxygen consumption and lipid content. Increased oxidative stress in the CNS will further lead to lipid peroxidation, DNA and protein damage [23]. Oxidative stress in the CNS has been demonstrated to involve excitotoxicity and apoptosis, the two main causes of neuronal death. Furthermore, oxidative stress has also been implicated the progression of Alzheimer's disease (AD), Parkinson's disease (PD), multiple sclerosis (MS) and other neurodegenerative diseases [24,25]. Antioxidants may have a positive effect in the CNS and seem to be a promising approach of neuroprotection therapy, as they can protect the CNS against free radical mediated oxidative damage [26]. However, our endogenous antioxidant defenses are not always completely effective and exposure to damaging environmental factors is increasing, therefore it seems reasonable to propose that exogenous antioxidants could be effective in diminishing the cumulative effects of oxidative damage. Presently, antioxidants constitute a major component of clinical and experimental drugs that are currently considered for prevention of neurodegenerative diseases and therapy [27].

Antioxidant activities of marine algae have been determined by various methods such as 1,1-diphenyl-2-picryl hydrazyl (DPPH) radical scavenging, 2,2'-azinobis-3-ethylbenzo thizoline-6-sulphonate (ABTS) radical scavenging, singlet oxygen quenching activity, lipid peroxide inhibition, superoxide and hydroxyl radical scavenging assays. Lim *et al.* demonstrated that *Neorhodomela aculeate*, which is also known as *Rhodomela confervoides*, was able to scavenge DPPH with an IC<sub>50</sub> = 90  $\mu$ g/mL and at a concentration of 20  $\mu$ g/mL completely suppressed H<sub>2</sub>O<sub>2</sub> induced

lipid peroxidation in rat brain homogenate [28]. Furthermore, Fallarero et al. showed that Halimeda incrassata and Bryothamniom triquetrum are potent ROS scavengers in mouse hypothalamic (GT1-7) cells [29]. Novoa et al. reported that the antioxidant and ROS scavenging activity of B. triquetrum are related to their high phenolic contents [30]. Dieckol, a phenolic compound isolated from brown algae has been shown to scavenge ROS production in murine microglia (BV2) cells [31]. Wijesekara and Kim reported that most phenolic compounds which were purified from marine algae are responsible for marine algal antioxidant activities and protective effects against oxidative stress induced cell damage [32]. Phenolic compounds act as free radical scavengers, reducing agents and metal chelators, and thus effectively inhibit lipid oxidation. In addition, Yan et al. demonstrated that carotenoids have a strong radical scavenging activity and are found as a major antioxidant in marine algae [33,34]. Young and Lowe indicated that structure, physical form, location or site of action, potential interaction with another antioxidant, concentration and partial pressure to oxygen may affect the antioxidant activities of carotenoids in biological systems [35]. Fucoxanthin obtained from *Padina tetrastromatic* has shown higher potential to be used as an antioxidant than  $\beta$ -carotene in modulating antioxidant enzyme in plasma and liver of retinol deficient rats [36,37]. However, the exact mechanisms of action how fucoxanthin exerts antioxidative effect in rat induced by retinol deficiency are not yet completely understood. Moreover, the cytoprotective effect of fucoxanthin against ROS formation induced by H<sub>2</sub>O<sub>2</sub> in monkey kidney fibroblast (Vero) cells has been observed [38]. Two hydroxyl groups present in the ring structure of fucoxanthin may correlate to the inhibition of ROS formation. Indeed, it has been reported that the number of hydroxyl groups on the ring structure is correlated with the effects of ROS suppression. Moreover, it has also been shown that some marine algal sulfated polysaccharides (SPs) can be used as potent antioxidants [39,40]. Antioxidant activity of marine algal SPs depends on their structural features such as degree of sulfating, molecular weight, type of the major sugar and glycosidic branching [41,42]. However, bioactivities of marine algal carotenoids and SPs against oxidative stress in the CNS have not been demonstrated yet.

Based on those findings, it can be suggested that among various organisms in the marine environment, marine algae prove to be one of the useful candidates that can protect the CNS against oxidative degradation. Hence, developing novel molecules derived from marine algae which promote antioxidant activity in the CNS may lead to the development of effective neuroprotective agents. Furthermore, it is also important to determine whether antioxidants derived from marine algae can be used as prophylactic neuroprotective agents in order to slow down the progression of neurodegenerative diseases in populations that are at high risk, such as the elderly. Additionally, antioxidant activities of marine algal carotenoids, SPs and other bioactive compounds in the CNS warrant further investigations.

#### 2.2. Anti-Neuroinflammation

Inflammation has been found to be the pathophysiological mechanism underlying many chronic diseases such as cardiovascular disease, diabetes, certain cancers, arthritis, and neurodegenerative diseases [43]. Recent studies demonstrated that resulting production of inflammatory responses and neurotoxic factors in the CNS is sufficient to induce neurodegeneration in a rat model [44]. Several cell types have been demonstrated as contributors in inflammation-mediated neurodegeneration, yet

microglia are implicated as critical components of the immunological insult to neurons [45]. Microglia are the immune cells in the CNS, they enters the system from the blood circulation early in an organism's development and serve a role of immune surveillance [43]. Ramified or resting microglia constitute 5–20% of glial populations in the CNS [46]. Recent study demonstrated that activation of microglia and the resulting production of pro-inflammatory and neurotoxic factors are sufficient to induce neurodegeneration in a rat model. Furthermore, activation of microglia and excessive amounts of pro-inflammatory mediators release by microglia have been observed during the pathogenesis of PD, AD, MS, AIDS dementia complex, as well as post neuronal death in cerebral stroke and traumatic brain injury [44,47]. Therefore, a mechanism to regulate inflammatory response release by microglia may have important therapeutic potential for the treatment of neurodegenerative diseases.

Numerous studies has documented anti-inflammatory activities of marine algae in vitro and in vivo [48]. However, scientific analysis of anti-neuroinflammatory activity of marine algae has been poorly carried out and until now only few studies were reported. Ecklonia cava (Phaeophyceae; Laminareaceae), also known as "sea trumpet", has been reported to possess anti-inflammatory activity [49–51]. E. cava was able to suppress the levels of pro-inflammatory mediators such as nitric oxide (NO), prostaglandine- $E_2$  (PGE<sub>2</sub>) and pro-inflammatory cytokines (tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-6 (IL-6) and interleukin-1 $\beta$ (IL-1 $\beta$ )) in lipopolysaccharides (LPS)-stimulated BV2 cells by blocking nuclear factor- $\kappa$ B (NF- $\kappa$ B) and mitogen-activated protein kinases (MAPKs) activation [31,51]. Furthermore, N. aculeate decreased NO production and inhibiting inducible NO synthase (iNOS) expression in interferon-gamma (IFN- $\gamma$ ) stimulated BV2 cells [28]. A number of bromophenols have been previously isolated from N. aculeate and may be potential anti-neuroinflammatory candidates [52-55]. Another study conducted by Cui et al. [56] provide the first evidence that fucoidan isolated from Laminaria japonica has a potent inhibitory effect against LPS-induced NO production in BV2 cells. In their study, the average molecular weight of fucoidan was 7000 Dalton, consisting of 48% total sugar (including 28% fucose) and 29% sulfate. Fucoidan at a concentration of 125 µg/mL, significantly inhibited NO production to 75% [56]. NO is a cytotoxic, short lived highly diffusible signaling molecule [57]. A number of studies demonstrated that NO generated by iNOS causes injury and cell death of neuron and oligodendrocytes in the CNS, hence NO is implicated in pathogenesis of various neurodegenerative disease [57,58]. Anti-neuroinflammatory activity of another marine algae species, Ulva conglobata has been reported. U. conglobata methanolic extracts were able to suppress the expression of pro-inflammatory enzymes, iNOS and cyclooxygenase-2 (COX-2), which accounted for the large production of NO and PGE<sub>2</sub>, respectively [59,60]. Among other mediators released by microglia, NO and PGE<sub>2</sub> are the main cytotoxic mediators participating in the innate response in the CNS [61,62]. Pro-inflammatory mediators have been found to be elevated in the brain of early AD [63]. For this reasons, agents that inhibit the production of pro-inflammatory mediators have been previously considered as potential candidates for the treatment of neurodegenerative diseases.

Epidemiological studies show that application of non-steroidal anti-inflammatory drugs (NSAIDs) reduces the risk and delays the onset of inflammation in the CNS which further participates in the pathogenesis of some neurodegenerative diseases. NSAIDs mainly act by inhibiting the production of pro-inflammatory mediators. Hence, attenuation of pro-inflammatory mediators in microglia by marine algae demonstrates its potential neuroprotective activity. Furthermore, marine algae as potential anti-neuroinflammatory agents have a great potential application in the pharmaceuticals area as well as

the food industry. There are numerous advantages of marine algae use in pharmaceuticals and functional foods, such as relatively low production costs, low cytotoxicity, safety and wide acceptability. However, further studies are needed with clinical trials for marine algal anti-neuroinflammatory activity.

# 2.3. Cholinesterase Inhibitory Activity

Alzheimer's diseases (AD) is an irreversible, progressive neurodegenerative disease, which resulting in memory loss, behavior disturbances, personality changes and a decline in cognitive abilities [64]. It was stated in the cholinergic hypothesis, that a serious loss of cholinergic function in the CNS contributes significantly to the cognitive symptoms associated with AD [65]. In accordance, neuropathological studies demonstrated that AD was associated with deficiency in the brain neurotransmitter acetylcholine (ACh) [66]. The inhibition of acetylcholinesterase (AChE) enzyme, which catalyzes the breakdown of ACh, may be one of the most realistic approaches to the symptomatic treatment of AD [67]. Presently, a variety of plants has been reported to possess AChE inhibitory activity. *Huperzia serrata*, a Chinese terrestrial herb has been demonstrated to be a potent AChE inhibitor [68]. In addition, Houghton *et al.* reported cholinesterase (ChE) inhibitory activity of *Crinum jagus* and *Crinum glaucum*, two Nigerian *Crinum* species [69]. A number of studies have recently shown AChE inhibitory activity of several marine algae species. A list of marine algae reported to have significant AChE inhibitory activity is presented in Table 1.

Marine algae	Extracts/Compounds	IC <sub>50</sub>	Ref
Caulerpa racemosa	MeOH extracts	$5.5 \text{ mg mL}^{-1}$	[70]
Codium capitatum	MeOH extracts	$7.8 \text{ mg mL}^{-1}$	[70]
Ulva fasciata	MeOH extracts	$4.8 \text{ mg mL}^{-1}$	[70]
Halimeda cuneata	MeOH extracts	5.7 mg mL <sup><math>-1</math></sup>	[70]
Amphiora ephedraea	MeOH extracts	5.1 mg mL <sup><math>-1</math></sup>	[70]
Amphiora bowerbankii	MeOH extracts	5.3 mg mL <sup><math>-1</math></sup>	[70]
Dictyota humifusa	MeOH extracts	$4.8 \text{ mg mL}^{-1}$	[70]
Hypnea valentiae	MeOH extracts	$2.6 \text{ mg mL}^{-1}$	[71]
Padina gymnospora	MeOH extracts	$3.5 \text{ mg mL}^{-1}$	[71]
Ulva reticulate	MeOH extracts	$10 \text{ mg mL}^{-1}$	[71]
Gracilaria edulis	MeOH extracts	$3 \text{ mg mL}^{-1}$	[71]
Ecklonia stolonifera	EtOH extracts	$108.11 \ \mu \text{g mL}^{-1}$	[72]
Ecklonia stolonifera	24-hydroperoxy-24-vinylcholesterol	389.1 μM	[72]
Ecklonia stolonifera	Eckstolonol	42.66 µM	[72]
Ecklonia stolonifera	Eckol	20.56 µM	[72]
Ecklonia stolonifera	Phlorofucofluoroeckol-A	4.89 μM	[72]
Ecklonia stolonifera	Dieckol	17.11 μM	[72]
Ecklonia stolonifera	2-phloroeckol	38.13 µM	[72]
Ecklonia stolonifera	7–phloroeckol	21.11 μM	[72]
Ishige okamurae	MeOH extracts	163.07 μM	[73]
Ishige okamurae	EtOAc extracts	137.25 μM	[73]
Ishige okamurae	6,6'–bieckol	46.42 μM	[73]

**Table 1.** Acetylcholinesterase inhibitory activities of several marine algae.

 $IC_{50}$  values for eserine and galanthamine were 0.004 µg mL<sup>-1</sup> and 0.0007 mg mL<sup>-1</sup>, respectively.

Recently, Myung *et al.* reported that dieckol and phlorofucofluoroeckol possess memory enhancing and AChE inhibitory activity [74]. Furthermore, Yoon et al. screened ethanolic extracts of 27 Korean marine algae, for inhibitory activity on AChE, and found that extracts from Ecklonia stolonifera showed significant inhibitory activity [72]. Two sterols and eight phlorotannins were isolated from E. stolonifera. Eckol, dieckol, 2-phloroeckol and 7-phloroeckol demonstrated selective dose dependent inhibitory activities toward AChE; whereas, eckstolonol and phlorofucofuroeckol-A exhibited inhibitory activities toward both AChE and butyrylcholinesterase (BChE). However, phloroglucinol, which is a monomer, and triphlorethol-A, the opened-chain trimer of phloroglucinol, did not inhibit the cholinesterase (ChE) at the concentrations tested. The exact mechanisms underlying this phenomenon have not yet been identified. However, the possible relation between structure of phlorotannins and AChE inhibitory activity has been reported, it is suggested that phlorotannins as polymers of phloroglucinol have appropriately bulky structures, which is then able to mask the ChE and prevents the binding of the substrates. Moreover, as the phloroglucinol monomer and open-chain trimer of phloroglucinol were not able to inhibit the ChE activity, it may suggest that that degree of polymerization and closed-ring structure of phlorotannins play key roles in the inhibitory potential of phlorotannins toward the ChE [72]. In addition, Hypnea valentiae and Ulva reticulate, two marine algae species from Tamil Nadu, India, also reported to inhibit both AChE and BChE activity [71]. A good balance between AChE and BChE activity has been reported to result in higher efficacy for the treatment of AD [75]. BChE are considered to play a minor role in regulating brain AChE levels. Notably, AChE and BChE mixed inhibition have been found in tacrine and physostigmine, which are licensed drugs used in the treatment of AD.

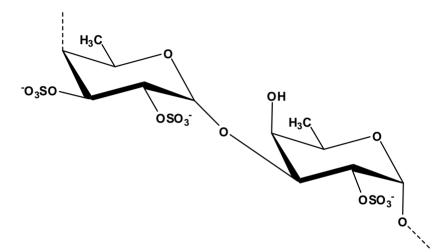
Taken together, marine red, brown and green algae have potential to be use as functional neuroprotective agents due to their effectiveness in inhibiting ChE activity. Furthermore, some compounds derived from marine algae provided mixed type ChE (AChE and BChE) inhibitory activities, which have been considered to be more effective in the treatment of AD. Some AChE synthetic commercial drugs are known to produce side effects. Hence, researchers have a great interest to study natural herbs that can act as AChE inhibitors. Many kinds of marine algae, consumed for centuries in East Asia countries, are well tolerated and lack harmful side effects. Interestingly, several marine algae species have also been demonstrated as potential AChE inhibitors. Hence, AChE inhibitory activity of marine algae should be screened and further studies with clinical trials are also needed.

## 2.4. Inhibition of Neuronal Death

A common pathological hallmark of various neurodegenerative diseases is the loss of particular subsets of neurons [76]. Neurodegeneration of these neural subsets may be a consequence of various forms of neural cell death, including necrosis and apoptosis [77]. A study carried out by Jhamandas *et al.* successfully showed that fucoidan isolated from *Fucus vesiculosus* (Figure 1), was able to protect rat cholinergic neuronal death induced by  $A\beta_{1-42}$  [78]. Fucoidan pretreatment blocked the activation of caspase-9 and caspase-3 have been suggested to mediate the terminal stages of neuronal apoptosis [79]. Caspase-9 and caspase-3 are two of several central components of the machinery responsible for apoptosis. Therefore, the ability of fucoidan to block the activation of

caspase-9 and caspase-3 suggest that inhibition of neuronal death by fucoidan mainly occurs through apoptotic inhibition. In neurodegenerative diseases, apoptosis might be pathogenic, and targeting this process might mitigate neurodegenerative diseases [80]. Furthermore, aqueous extracts of *B*. *triquetrum* has been demonstrated to protect GT1–7 cells death produced by severe (180 min) chemical hypoxia/aglycemia insult, which further reduced the cytotoxicity and early production of free radicals. The protection exerted by *B. triquetrum* extract seems to be linked to its ability to reduce free-radical generation [81]. The authors suggest that the protective effects of *B. triquetrum* extract are partially related to the presence of ferulic acid [81].

Figure 1. Chemical structure of fucoidan isolated from Fucus vesiculosus (Adapted from [39]).



# 2.5. Antineurotoxicity

Neurotoxins are a varied groups of compounds, whose highly specific effects on the nervous system of animals, including humans, is by interfering with nerve impulse transmission [82]. They are able to produce neuronal damage or neurodegeration when administered in vivo or in vitro [83]. As an example,  $\beta$ -amyloid (A $\beta$ ) peptides have been demonstrated to possess neurotoxic effect on neuron and glial cells although the precise mechanisms by which this occurs have yet to be elucidated [84]. Excessive accumulation of A $\beta$  in the brain has been characterized as a major pathological hallmark of AD and recently, fucoidan has been reported to block Aβ neurotoxicity in neuronal cell [78]. Fucoidan treatment abolished the inhibitory effect of  $A\beta$  on the phosphorylation of protein kinase C (PKC) which has been demonstrated to stimulate the survival of neurons and prevents AB neurotoxicity. PKC causes GSK-3<sup>β</sup> inactivation and this inactivation in turn leads to the accumulation of cytoplasmic  $\beta$ -catenin and the subsequent translocation of  $\beta$ -catenin to the nucleus, causing TCF/LEF-1-dependent transcriptional activation of growth and differentiation related genes, which is required to stimulate neuronal survival [85]. In addition, Luo et al. showed that fucoidan isolated from L. japonica was able to protect against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced neurotoxicity in animal model of Parkinsonism (C57/BL mice) and dopaminergic (MN9D) cells [86]. The mechanisms of protection provided by fucoidan may partly relate to its antioxidative activity. Furthermore, the results of those studies suggest potential application of fucoidan for PD prevention and or treatment. Moreover, the possible roles of alginates to protects human neuronal (NT2) cells against H<sub>2</sub>O<sub>2</sub>-induced neurotoxicity have previously been demonstrated [87]. *H. incrassata* and *B. triquetrum* at a concentration of 0.2 mg/mL has been shown to protect methyl mercury-induced neurotoxicity in GT1–7 cells [29]. Collectively, marine algae and its bioactive compounds can be used for the development of new generation therapeutic neuroprotective agents against neurotoxins in the CNS.

### 2.6. Other Neuroprotective Activities

Neurite outgrowth is a fundamental neuronal feature and plays an important role in neuronal development during embryogenesis and in the adult brain [88]. *Sargassum macrocarpum* and its two active component, sargaquinoic acid and sargachromechanol, have been shown to promote neurite outgrowth in rat pheochromocytoma (PC12) cells [89–91]. Structure and neurite outgrowth promoting relationship of sargaquinoic acid has been reported by Tsang *et al.* [92]. They reported that quinone is the structural moiety of the sargaquinoic acid molecule which is responsible for the neurite outgrowth-promoting activity. Notably, the hydroxyl group bonded to quinone had a significant effect on neuritogenic activity. In addition, pheophytin a, a chlorophyll-related compound and its analog, vitamin B12 derived from *Sargassum fulvellum* also has potential neurite outgrowth-promoting activity [93,94].

Phlorotannins derived from *Eisenia bicyclis* have been demonstrated to inhibit  $\beta$ -amyloid cleavage enzyme (BACE-1) activity [95]. BACE-1 represents candidate biomarkers of AD, since it initiates the formation of A $\beta$  [96]. When considering that almost all currently available medications for AD are AChE inhibitors, suppression of BACE-1 by phlorotannins will enhance the medications and or therapy for AD patients.

In addition, Lee *et al.* demonstrated that fucoidan treatment resulted in an increase in cell proliferation of human neuroblastoma (SH-SY5Y) cell induced by A $\beta$  [97]. Hence, it may suggest that fucoidan has potential neuroprotective effects.

## 3. Prospects of Marine Algae as Neuroprotective Agents

Neurodegenerative diseases are estimated to surpass cancer as the second most common cause of death among elderly by the 2040s [98,99]. For this reason, a great deal of attention has been expressed by scientists regarding safe and effective neuroprotective agents. Many categories of natural and synthetic neuroprotective agents have been reported. However, synthetic neuroprotective agents are believed to have certain side effects such as dry mouth, tiredness, drowsiness, sleepiness, anxiety or nervousness, difficulty to balance, *etc.* [100]. Hence, nowadays researchers have an interest in studying natural bioactive compounds that can act as neuroprotective agents. Marine algae represent one potential candidate neuroprotective agent. However, development of marine algae as neuroprotective agents still faces several challenges. The rationale for marine algal neuroprotective effects treatment in the CNS is based on established observations and experiments *in vitro* or in animal models only. Up to now, none of the marine algal neuroprotective effects have been examined in human subjects. Therefore, small clinical studies and further large-scale controlled studies are needed. Another important challenge in the development of marine algae as neuroprotective agents is that many drugs failed to provide real neuroprotection in practice. Potential reasons for this failure include inappropriate use of specific neuroprotection/s for a given disease or stage of disease progression or

the use of suboptimal doses [101]. Hence, future studies are needed focusing on the synergistic benefits of consuming different marine algae species, recommended doses and timing of intake, and preparation methods for marine algal bioactive compounds in order to maximize the desired protective effect in the prevention of neurodegenerative diseases.

It has been reported that neurodegenerative diseases in East Asian countries were lower than in Europe (p < 0.0004) [102,103]. Many studies have indicated potential health benefits of marine algae consumption [7,104]. Thus, lower incidence of neurodegenerative diseases in East Asia may correlate to high fish and marine algae consumption by East Asian populations. More recently, there has been growing interest in marine algae and their constituents as functional foods and nutraceuticals with potential health benefit effects as sources of antioxidant to reduce the risk of neurodegenerative diseases. Marine algae are an important source of bioactive ingredients that can be applied to many aspects of processing healthier foods and developing functional neuroprotective foods.

In addition, the wide diversity of marine algae and numerous undiscovered unique metabolites present in marine algae are interesting sources to increase numbers of novel drugs against neurodegenerative diseases. However, large-scale human studies are required to identify the prophylactic and therapeutic neuroprotective effect of marine algae.

# 4. Conclusions

In conclusion, marine algae are a valuable source of neuroprotective agents and could be introduced for the preparation of novel functional ingredients in pharmaceuticals and functional foods as a good approach for the treatment and or prevention of neurodegenerative disease. Marine algae can be suggested as an alternative source to synthetic ingredients that can contribute to neuroprotection by being a part of pharmaceuticals and functional foods. Furthermore, the wide range of biological activities associated with natural compounds derived from marine algae such as phlorotannins, alginates, fucoidan, sargaquinoic acid, SPs and carotenoids increase the potential to expand the neuroprotective effects and health beneficial value of marine algae in the pharmaceutical industry. Until now, most of the biological and neuroprotective activities of marine algae and its natural compounds have been observed *in vitro* or in mouse model systems. Therefore, further research studies are needed in order to investigate marine algae neuroprotective activities in human subjects and further in large-scale controlled studies.

## Acknowledgements

This study was supported by a grant from the Marine Bioprocess Research Center of the Marine Bio 21 Project funded by the Ministry of Land, Transport and Maritime, Republic of Korea.

# References

- 1. Kim, S.; Wijesekara, I. Development and biological activities of marine-derived bioactive peptides: A review. *J. Funct. Foods* **2010**, *2*, 1–9.
- 2. Swing, J. What future for the oceans? *Foreign Aff.* **2003**, *82*, 139–152.

- 3. Alonso, D.; Castro, A.; Martinez, A. Marine compounds for the therapeutic treatment of neurological disorders. *Expert Opin. Ther. Patents* **2005**, *15*, 1377–1386.
- 4. Heo, S.J.; Hwang, J.Y.; Choi, J.I.; Han, J.S.; Kim, H.J.; Jeon, Y.J. Diphlorethohydroxycarmalol isolated from Ishige okamurae, a brown algae, a potent [alpha]-glucosidase and [alpha]-amylase inhibitor, alleviates postprandial hyperglycemia in diabetic mice. *Eur. J. Pharmacol.* **2009**, *615*, 252–256.
- 5. Pangestuti, R.; dan Limantara, L. Rumput Laut, Zamrud Tak Tergali Dari Laut. *BioS* 2010, *2*, 2–10.
- 6. Khan, S.; Kong, C.; Kim, J.; Kim, S. Protective effect of *Amphiroa dilatata* on ROS induced oxidative damage and MMP expressions in HT1080 cells. *Biotech. Bioproc. Eng.* **2010**, *15*, 191–198.
- 7. Burtin, P. Nutritional value of seaweeds. *EJEAFChe* **2003**, *2*, 498–503.
- 8. Matsubara, K.; Matsuura, Y.; Hori, K.; Miyazawa, K. An anticoagulant proteoglycan from the marine green alga, *Codium pugniformis. J. Appl. Phycol.* **2000**, *12*, 9–14.
- 9. Athukorala, Y.; Lee, K.; Kim, S.; Jeon, Y. Anticoagulant activity of marine green and brown algae collected from Jeju Island in Korea. *Bioresour. Technol.* **2007**, *98*, 1711–1716.
- 10. Artan, M.; Li, Y.; Karadeniz, F.; Lee, S.; Kim, M.; Kim, S. Anti-HIV-1 activity of phloroglucinol derivative, 6, 6'-bieckol, from Ecklonia cava. *Bioorgan. Med. Chem.* **2008**, *16*, 7921–7926.
- 11. Huheihel, M.; Ishanu, V.; Tal, J.; Arad, S. Activity of *Porphyridium* sp. *polysaccharide* against herpes simplex viruses *in vitro* and *in vivo*. *J. Biochem. Biophys. Meth.* **2002**, *50*, 189–200.
- 12. Heo, S.J.; Park, E.J.; Lee, K.W.; Jeon, Y.J. Antioxidant activities of enzymatic extracts from brown seaweeds. *Bioresour. Technol.* **2005**, *96*, 1613–1623.
- 13. Park, P.; Heo, S.; Park, E.; Kim, S.; Byun, H.; Jeon, B.; Jeon, Y. Reactive oxygen scavenging effect of enzymatic extracts from *Sargassum thunbergii*. J. Agr. Food Chem. 2005, 53, 6666–6672.
- 14. Zou, Y.; Qian, Z.; Li, Y.; Kim, M.; Lee, S.; Kim, S. Antioxidant effects of phlorotannins isolated from *Ishige okamurae* in free radical mediated oxidative systems. *J. Agr. Food Chem.* **2008**, *56*, 7001–7009.
- 15. Li, Y.; Lee, S.; Le, Q.; Kim, M.; Kim, S. Anti-allergic effects of phlorotannins on histamine release via binding inhibition between IgE and Fc RI. J. Agr. Food Chem. 2008, 56, 12073–12080.
- Kong, C.S.; Kim, J.A.; Yoon, N.Y.; Kim, S.K. Induction of apoptosis by phloroglucinol derivative from *Ecklonia cava* in MCF-7 human breast cancer cells. *Food Chem. Toxicol.* 2009, 47, 1653–1658.
- Kim, M.; Rajapakse, N.; Kim, S. Anti inflammatory effect of *Ishige okamurae* ethanolic extract via inhibition of NF B transcription factor in RAW 264.7 cells. *Phytother. Res.* 2009, 23, 628–634.
- 18. Maeda, H.; Hosokawa, M.; Sashima, T.; Miyashita, K. Dietary combination of fucoxanthin and fish oil attenuates the weight gain of white adipose tissue and decreases blood glucose in obese/diabetic KK-Ay Mice. J. Agr. Food Chem. 2007, 55, 7701–7706.

- 19. Tsukui, T.; Konno, K.; Hosokawa, M.; Maeda, H.; Sashima, T.; Miyashita, K. Fucoxanthin and fucoxanthinol enhance the amount of docosahexaenoic acid in the liver of KKAy obese/diabetic mice. *J. Agr. Food Chem.* **2007**, *55*, 5025–5029.
- 20. Kong, C.; Kim, J.; Ahn, B.; Vo, T.; Yoon, N.; Kim, S. 1-(3,5-Dihydroxyphenoxy)-7-(2,4,6-trihydroxyphenoxy)-2,4,9-trihydroxydibenzo-1,4-dioxin inhibits adipocyte differentiation of 3T3-L1 fibroblasts. *Mar. Biotechnol.* **2010**, *12*, 299–307.
- 21. Zarros, A. In which cases is neuroprotection useful. Adv. Altern. Think. Neurosci. 2009, 1, 3–5.
- 22. Barnham, K.J.; Masters, C.L.; Bush, A.I. Neurodegenerative diseases and oxidative stress. *Nat. Rev. Drug. Discov.* **2004**, *3*, 205–214.
- Akyol, Ö.; Herken, H.; Uz, E.; FadIllIolu, E.; Ünal, S.; Söüt, S.; Özyurt, H.; Sava, H. The indices of endogenous oxidative and antioxidative processes in plasma from schizophrenic patients\* 1:: The possible role of oxidant/antioxidant imbalance. *Progr. Neuro-Psychopharmacol. Biol. Psychiatr.* 2002, 26, 995–1005.
- 24. Migliore, L.; Copped è, F. Environmental-induced oxidative stress in neurodegenerative disorders and aging. *Mutat. Res-Gen. Tox. En.* **2009**, *674*, 73–84.
- 25. Behl, C.; Moosmann, B. Antioxidant neuroprotection in Alzheimer's disease as preventive and therapeutic approach. *Free Rad. Biol. Med.* **2002**, *33*, 182–191.
- Andersen, J. Oxidative stress in neurodegeneration: Cause or consequence? *Nat. Rev. Neurosci.* 2004, 5, S18–S25.
- 27. Moosmann, B.; Behl, C. Antioxidants as treatment for neurodegenerative disorders. *Expert Opin. Investig. Drugs* **2002**, *11*, 1407–1435.
- 28. Lim, C.; Jin, D.; Sung, J.; Lee, J.; Choi, H.; Ha, I.; Han, J. Antioxidant and anti-inflammatory activities of the methanolic extract of *Neorhodomela aculeate* in hippocampal and microglial cells. *Biol. Pharm. Bull.* **2006**, *29*, 1212–1216.
- Fallarero, A.; Loikkanen, J.J.; Männistö, P.T.; Casta ñeda, O.; Vidal, A. Effects of aqueous extracts of *Halimeda incrassata* (Ellis) Lamouroux and *Bryothamnion triquetrum* (S.G.Gmelim) Howe on hydrogen peroxide and methyl mercury-induced oxidative stress in GT1–7 mouse hypothalamic immortalized cells. *Phytomedicine* 2003, *10*, 39–47.
- Vidal Novoa, A.; Motidome, M.; Mancini Filho, J.; Fallarero Linares, A.; Tanae, M.; Torres, L.; Lapa, A. Actividad antioxidante y ácidos fen dicos del alga marina *Bryothamnion triquetrum* (SG Gmelim) Howe; Antioxidant activity related to phenolic acids in the aqueous extract of the marine seaweed *Bryothamnin triquetrum* (SG Gmelim) Howe. *Rev. Bras. Ciânc. Farm.(Impr.)* 2001, *37*, 373–382.
- 31. Jung, W.; Heo, S.; Jeon, Y.; Lee, C.; Park, Y.; Byun, H.; Choi, Y.; Park, S.; Choi, I. Inhibitory effects and molecular mechanism of dieckol isolated from marine brown alga on COX-2 and iNOS in microglial cells. *J. Agr. Food Chem.* **2009**, *57*, 4439–4446.
- 32. Wijesekara, I.; Yoon, N.; Kim, S. Phlorotannins from *Ecklonia cava* (Phaeophyceae): Biological activities and potential health benefits. *BioFactors* **2010**, *306*, 408–414.
- 33. Yan, X.; Chuda, Y.; Suzuki, M.; Nagata, T. Fucoxanthin as the major antioxidant in *Hijikia fusiformis*, a common edible seaweed. *Biosci. Biotech. Biochem.* **1999**, *63*, 605–607.
- 34. Nomura, T.; Kikuchi, M.; Kubodera, A.; Kawakami, Y. Proton-donative antioxidant activity of fucoxanthin with 1, 1-diphenyl-2-picrylhydrazyl (DPPH). *IUBMB Life* **1997**, *42*, 361–370.

- 36. Sangeetha, R.; Bhaskar, N.; Baskaran, V. Comparative effects of β-carotene and fucoxanthin on retinol deficiency induced oxidative stress in rats. *Mol. Cell. Biochem.* **2009**, *331*, 59–67.
- 37. Ravi Kumar, S.; Narayan, B.; Vallikannan, B. Fucoxanthin restrains oxidative stress induced by retinol deficiency through modulation of Na+ Ka+-ATPase and antioxidant enzyme activities in rats. *Eur. J. Nutr.* **2008**, *47*, 432–441.
- Heo, S.; Ko, S.; Kang, S.; Kang, H.; Kim, J.; Kim, S.; Lee, K.; Cho, M.; Jeon, Y. Cytoprotective effect of fucoxanthin isolated from brown algae *Sargassum siliquastrum* against H<sub>2</sub>O<sub>2</sub>-induced cell damage. *Eur. Food Res. Tech. A* 2008, 228, 145–151.
- 39. Wijesekara, I.; Pangestuti, R.; Kim, S. Biological activities and potential health benefits of sulfated polysaccharides derived from marine algae. *Carbohyd. Polym.* **2010**, *84*, 14–21.
- 40. Jiao, G.; Yu, G.; Zhang, J.; Ewart, H. Chemical structures and bioactivities of sulfated polysaccharides from marine algae. *Mar. Drugs* **2011**, *9*, 196–223.
- 41. Qi, H.; Zhang, Q.; Zhao, T.; Chen, R.; Zhang, H.; Niu, X.; Li, Z. Antioxidant activity of different sulfate content derivatives of polysaccharide extracted from *Ulva pertusa* (Chlorophyta) *in vitro*. *Int. J. Biol. Macromol.* **2005**, *37*, 195–199.
- 42. Zhang, Q.; Li, N.; Zhou, G.; Lu, X.; Xu, Z.; Li, Z. *In vivo* antioxidant activity of polysaccharide fraction from *Porphyra haitanesis* (Rhodephyta) in aging mice. *Pharmacol. Res.* **2003**, *48*, 151–155.
- 43. Allen, N.; Barres, B. Neuroscience: Glia—More than just brain glue. *Nature* **2009**, *457*, 675–677.
- 44. Liu, B.I.N.; Gao, H.M.; Wang, J.Y.; Jeohn, G.H.; Cooper, C.L.; Hong, J.S. Role of nitric oxide in inflammation-mediated neurodegeneration. *Ann. N. Y. Acad. Sci.* **2002**, *962*, 318–331.
- 45. Block, M.; Zecca, L.; Hong, J. Microglia-mediated neurotoxicity: Uncovering the molecular mechanisms. *Nat. Rev. Neurosci.* **2007**, *8*, 57–69.
- 46. Kim, S.; de Vellis, J. Microglia in health and disease. J. Neurosci. Res. 2005, 81, 302–313.
- 47. Lull, M.E.; Block, M.L. Microglial activation and chronic neurodegeneration. *Neurotherapeutics* **2010**, *7*, 354–365.
- 48. Abad, M.; Bedoya, L.; Bermejo, P. Natural marine anti-inflammatory products. *Mini Rev. Med. Chem.* **2008**, *8*, 740–754.
- 49. Maegawa, M.; Yokohama, Y.; Aruga, Y. Critical light conditions for young *Ecklonia cava* and *Eisenia bicyclis* with reference to photosynthesis. *Hydrobiologia* **1987**, *151*, 447–455.
- 50. Serisawa, Y.; Yokohama, Y.; Aruga, Y.; Tanaka, J. Photosynthesis and respiration in bladelets of *Ecklonia cava* Kjellman (Laminariales, Phaeophyta) in two localities with different temperature conditions. *Phycol. Res.* **2001**, *49*, 1–11.
- 51. Jung, W.K.; Ahn, Y.W.; Lee, S.H.; Choi, Y.H.; Kim, S.K.; Yea, S.S.; Choi, I.; Park, S.G.; Seo, S.K.; Lee, S.W.; Choi, I.W. Ecklonia cava ethanolic extracts inhibit lipopolysaccharide-induced cyclooxygenase-2 and inducible nitric oxide synthase expression in BV2 microglia via the MAP kinase and NF-[kappa]B pathways. *Food Chem. Toxicol.* 2009, 47, 410–417.
- 52. Zhao, J.; Fan, X.; Wang, S.; Li, S.; Shang, S.; Yang, Y.; Xu, N.; Lü, Y.; Shi, J. Bromophenol derivatives from the red alga *Rhodomela confervoides*. *J. Nat. Prod.* **2004**, *67*, 1032–1035.

- 53. Xu, N.; Fan, X.; Yan, X.; Li, X.; Niu, R.; Tseng, C.K. Antibacterial bromophenols from the marine red alga *Rhodomela confervoides*. *Phytochemistry* **2003**, *62*, 1221–1224.
- 54. Fan, X.; Xu, N.J.; Shi, J.G. Bromophenols from the red alga *Rhodomela confervoides*. J. Nat. Prod. 2003, 66, 455–458.
- 55. Ma, M.; Zhao, J.; Wang, S.; Li, S.; Yang, Y.; Shi, J.; Fan, X.; He, L. Bromophenols coupled with methyl γ-ureidobutyrate and bromophenol sulfates from the red alga *Rhodomela confervoides*. *J. Nat. Prod.* **2006**, *69*, 206–210.
- Cui, Y.; Zhang, L.; Zhang, T.; Luo, D.; Jia, Y.; Guo, Z.; Zhang, Q.; Wang, X.; Wang, X.M. Inhibitory effect of fucoidan on nitric oxide production in lipopolysaccharide activated primary microglia. *Clin. Exp. Pharmacol. Physiol.* **2010**, *37*, 422–428.
- 57. Heales, S.; Bolaños, J.; Stewart, V.; Brookes, P.; Land, J.; Clark, J. Nitric oxide, mitochondria and neurological disease. *Biochim. Biophys. Acta* **1999**, *1410*, 215–228.
- 58. Lee, J.; Grabb, M.; Zipfel, G.; Choi, D. Brain tissue responses to ischemia. J. Clin. Invest. 2000, 106, 723–731.
- 59. Jin, D.; Lim, C.; Sung, J.; Choi, H.; Ha, I.; Han, J. *Ulva conglobata*, a marine algae, has neuroprotective and anti-inflammatory effects in murine hippocampal and microglial cells. *Neurosci. Lett.* **2006**, *402*, 154–158.
- Salvemini, D.; Manning, P.; Zweifel, B.; Seibert, K.; Connor, J.; Currie, M.; Needleman, P.; Masferrer, J. Dual inhibition of nitric oxide and prostaglandin production contributes to the antiinflammatory properties of nitric oxide synthase inhibitors. J. Clin. Invest. 1995, 96, 301–308.
- 61. Vane, J.; Botting, R. New insights into the mode of action of anti-inflammatory drugs. *Inflamm. Res.* **1995**, *44*, 1–10.
- 62. Bosc á L.; Zeini, M.; Trav és, P.; Hortelano, S. Nitric oxide and cell viability in inflammatory cells: A role for NO in macrophage function and fate. *Toxicology* **2005**, *208*, 249–258.
- Blasko, I.; Stampfer-Kountchev, M.; Robatscher, P.; Veerhuis, R.; Eikelenboom, P.; Grubeck-Loebenstein, B. How chronic inflammation can affect the brain and support the development of Alzheimer's disease in old age: The role of microglia and astrocytes. *Aging Cell* 2004, *3*, 169–176.
- 64. Pietrini, P.; Alexander, G.; Furey, M.; Hampel, H.; Guazzelli, M. The neurometabolic landscape of cognitive decline: *In vivo* studies with positron emission tomography in Alzheimer's disease. *Int. J. Psychophysiol.* **2000**, *37*, 87–98.
- 65. Bartus, R.T. On neurodegenerative diseases, models, and treatment strategies: Lessons learned and lessons forgotten a generation following the cholinergic hypothesis. *Exp. Neurol.* **2000**, *163*, 495–529.
- 66. Tabet, N. Acetylcholinesterase inhibitors for Alzheimer's disease: Anti-inflammatories in acetylcholine clothing! *Age Ageing* **2006**, *35*, 336–338.
- 67. Pangestuti, R.; Kim, S.K. Neuroprotective properties of chitosan and its derivatives. *Mar. Drugs* **2010**, *8*, 2117–2128.
- 68. Cheng, D.H.; Ren, H.; Tang, X.C. Huperzine A, a novel promising acetylcholinesterase inhibitor. *Neuroreport* **1996**, *8*, 97–101.

- 69. Houghton, P.J.; Agbedahunsi, J.M.; Adegbulugbe, A. Choline esterase inhibitory properties of alkaloids from two *Nigerian Crinum* species. *Phytochemistry* **2004**, *65*, 2893–2896.
- 70. Stirk, W.; Reinecke, D.; van Staden, J. Seasonal variation in antifungal, antibacterial and acetylcholinesterase activity in seven South African seaweeds. *J. Appl. Phycol.* **2007**, *19*, 271–276.
- 71. Yoon, N.; Chung, H.; Kim, H.; Choi, J. Acetyl and butyrylcholinesterase inhibitory activities of sterols and phlorotannins from *Ecklonia stolonifera*. *Fish. Sci.* **2008**, *74*, 200–207.
- 72. Yoon, N.Y.; Lee, S.H.; Yong, L.; Kim, S.K. Phlorotannins from Ishige okamurae and their acetyl- and butyrylcholinesterase inhibitory effects. *J. Funct. Foods* **2009**, *1*, 331–335.
- Suganthy, N.; Karutha Pandian, S.; Pandima Devi, K. Neuroprotective effect of seaweeds inhabiting South Indian coastal area (Hare Island, Gulf of Mannar marine biosphere reserve): Cholinesterase inhibitory effect of *Hypnea valentiae* and *Ulva reticulata*. *Neurosci. Lett.* 2010, 468, 216–219.
- 74. Myung, C.; Shin, H.; Bao, H.; Yeo, S.; Lee, B.; Kang, J. Improvement of memory by dieckol and phlorofucofuroeckol in ethanol-treated mice: Possible involvement of the inhibition of acetylcholinesterase. *Arch. Pharm. Res.* **2005**, *28*, 691–698.
- 75. Greig, N.; Lahiri, D.; Sambamurti, K. Butyrylcholinesterase: An important new target in Alzheimer's disease therapy. *Int. Psychogeriatr.* **2002**, *14*, 77–91.
- 76. Mattson, M.P. Apoptosis in neurodegenerative disorders. *Nat. Rev. Mol. Cell Biol.* 2000, *1*, 120–130.
- 77. Bains, J.S.; Shaw, C.A. Neurodegenerative disorders in humans: The role of glutathione in oxidative stress-mediated neuronal death. *Brain Res. Rev.* **1997**, *25*, 335–358.
- Jhamandas, J.H.; Wie, M.B.; Harris, K.; MacTavish, D.; Kar, S. Fucoidan inhibits cellular and neurotoxic effects of β-amyloid (Aβ) in rat cholinergic basal forebrain neurons. *Eur. J. Neurosci.* 2005, *21*, 2649–2659.
- 79. Cowan, C.M.; Thai, J.; Krajewski, S.; Reed, J.C.; Nicholson, D.W.; Kaufmann, S.H.; Roskams, A.J. Caspases 3 and 9 send a pro-apoptotic signal from synapse to cell body in olfactory receptor neurons. *J. Neurosci.* **2001**, *21*, 7099–7109.
- 80. Vila, M.; Przedborski, S. Targeting programmed cell death in neurodegenerative diseases. *Nat. Rev. Neurosci.* **2003**, *4*, 365–375.
- 81. Fallarero, A.; Peltoketo, A.; Loikkanen, J.; Tammela, P.; Vidal, A.; Vuorela, P. Effects of the aqueous extract of Bryothamnion triquetrum on chemical hypoxia and aglycemia-induced damage in GT1–7 mouse hypothalamic immortalized cells. *Phytomedicine* **2006**, *13*, 240–245.
- Patockaa, J.; Stredab, L. Brief review of natural nonprotein neurotoxins. ASA Newslett. 2002, 89, 16–24.
- 83. Segura-Aguilar, J.; Kostrzewa, R. Neurotoxins and neurotoxic species implicated in neurodegeneration. *Neurotox. Res.* **2004**, *6*, 615–630.
- 84. Butterfield, D.A. Amyloid  $\beta$ -peptide (1–42)-induced oxidative stress and neurotoxicity: Implications for neurodegeneration in Alzheimer's disease brain. A review. *Free Rad. Res.* **2002**, *36*, 1307–1313.

- 85. Garrido, J.; Godoy, J.; Alvarez, A.; Bronfman, M.; Inestrosa, N. Protein kinase C inhibits amyloid {beta} peptide neurotoxicity by acting on members of the Wnt pathway. *FASEB J.* **2002**, *16*, 1982–1984.
- Luo, D.; Zhang, Q.; Wang, H.; Cui, Y.; Sun, Z.; Yang, J.; Zheng, Y.; Jia, J.; Yu, F.; Wang, X. Fucoidan protects against dopaminergic neuron death *in vivo* and *in vitro*. *Eur. J. Pharmacol.* 2009, 617, 33–40.
- 87. Eftekharzadeh, B.; Khodagholi, F.; Abdi, A.; Maghsoudi, N. Alginate protects NT2 neurons against H<sub>2</sub>O<sub>2</sub>-induced neurotoxicity. *Carbohyd. Polym.* **2010**, *79*, 1063–1072.
- 88. Khodosevich, K.; Monyer, H. Signaling involved in neurite outgrowth of postnatally born subventricular zone neurons *in vitro*. *BMC Neurosci.* **2010**, *11*, 18:1–18:11.
- 89. Tsang, C.; Ina, A.; Goto, T.; Kamei, Y. Sargachromenol, a novel nerve growth factor-potentiating substance isolated from Sargassum macrocarpum, promotes neurite outgrowth and survival via distinct signaling pathways in PC12D cells. *Neuroscience* **2005**, *132*, 633–643.
- 90. Tsang, C.; Kamei, Y. Sargaquinoic acid supports the survival of neuronal PC12D cells in a nerve growth factor-independent manner. *Eur. J. Pharmacol.* **2004**, *488*, 11–18.
- 91. Kamei, Y.; Sagara, A. Neurite outgrowth promoting activity of marine algae from Japan against rat adrenal medulla pheochromocytoma cell line, PC12D. *Cytotechnology* **2002**, *40*, 99–106.
- 92. Tsang, C.; Sagara, A.; Kamei, Y. Structure-activity relationship of a neurite outgrowth-promoting substance purified from the brown alga, *Sargassum macrocarpum*, and its analogues on PC12D cells. *J. Appl. Phycol.* **2001**, *13*, 349–357.
- 93. Ina, A.; Hayashi, K.; Nozaki, H.; Kamei, Y. Pheophytin a, a low molecular weight compound found in the marine brown alga *Sargassum fulvellum*, promotes the differentiation of PC12 cells. *Int. J. Dev. Neurosci.* **2007**, *25*, 63–68.
- Ina, A.; Kamei, Y. Vitamin B 12, a chlorophyll-related analog to pheophytin a from marine brown algae, promotes neurite outgrowth and stimulates differentiation in PC12 cells. *Cytotechnology* 2006, 52, 181–187.
- 95. Jung, H.; Oh, S.; Choi, J. Molecular docking studies of phlorotannins from *Eisenia bicyclis* with BACE1 inhibitory activity. *Bioorgan. Med. Chem. Lett.* **2010**, *20*, 3211–3215.
- 96. Tang, K.; Hynan, L.; Baskin, F.; Rosenberg, R. Platelet amyloid precursor protein processing: A bio-marker for Alzheimer's disease. *J. Neurol. Sci.* **2006**, *240*, 53–58.
- Lee, H.R.; Do, H.; Lee, S.R.; Sohn, E.S.; Pyo, S.; Son, E. Effects of fucoidan on neuronal cell proliferation-association with NO production through the iNOS pathway. *J. Food Sci. Nutr.* 2007, *12*, 74–78.
- 98. Bjarkam, C.R.; Sørensen, J.C.; Sunde, N.Å.; Geneser, F.A.; Østergaard, K. New strategies for the treatment of Parkinson's disease hold considerable promise for the future management of neurodegenerative disorders. *Biogerontology* **2001**, *2*, 193–207.
- 99. Ansari, J.; Siraj, A.; Inamdar, N. Pharmacotherapeutic approaches of Parkinson's disease. *Int. J. Pharmacol.* **2010**, *6*, 584–590.
- 100. Narang, S.; Gibson, D.; Wasan, A.D.; Ross, E.L.; Michna, E.; Nedeljkovic, S.S.; Jamison, R.N. Efficacy of dronabinol as an adjuvant treatment for chronic pain patients on opioid therapy. *J. Pain* **2008**, *9*, 254–264.

- Gilgun-Sherki, Y.; Melamed, E.; Offen, D. Oxidative stress induced-neurodegenerative diseases: The need for antioxidants that penetrate the blood brain barrier. *Neuropharmacology* 2001, 40, 959–975.
- 102. Mishra, S.; Palanivelu, K. The effect of curcumin (turmeric) on Alzheimer's disease: An overview. *Ann. Indian Acad. Neurol.* **2008**, *11*, 13–19.
- 103. Jorm, A.F.; Jolley, D. The incidence of dementia: A meta-analysis. *Neurology* **1998**, *51*, 728–733.
- 104. Smit, A.J. Medicinal and pharmaceutical uses of seaweed natural products: A review. J. Appl. Phycol. 2004, 16, 245–262.

Samples Availability: Available from the authors.

© 2011 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).