

Supplementary Materials

Isoquinolinequinone derivatives from a marine sponge (Haliclona sp.) regulate intestinal inflammation

Yun Na Kim^{1†}, Yeong Kwang Ji^{2†}, Na-Hyun Kim³, Nguyen Van Tu⁴, Jung-Rae Rho^{2*} and Eun Ju Jeong^{1*}

¹ Department of Agronomy and Medicinal Plant Resources, Gyeongnam National University of Science and Technology, Jinju 52725, Republic of Korea

² Department of Oceanography, Kunsan National University, Gunsan 54150, Republic of Korea

³ Gyeongnam Department of Environment & Toxicology, Korea Institute of Toxicology, 17 Jegok-gil, Munsan-eup 52834, Republic of Korea

⁴ Institute of Tropical Biology, 85 Tran Quoc Toan Street District 3, Ho Chi Minh, Vietnam

* Correspondence: jrrho@kunsan.ac.kr(J.R.); Tel.: +82 63 469 4606(J.R.); ejjeong@gnitech.ac.kr(E.J.J.); Tel.: +82 55 751 3224 (E.J.J.)

Contents

Table S1. Spectral data for compound 8 in CDCl ₃ (500Hz, for ¹ H; 125 Hz for ¹³ C).	13
Table S2. Coordinate for the optimized conformer of compound 1 .	20
Table S3. Experimental and calculated ¹³ C NMR chemical shifts of compound 1 .	21
Figure S1. HRqTOFMS for 1 .	3
Figure S2. ¹ H NMR (500 MHz, CDCl ₃) spectrum of 1 .	4
Figure S3. ¹³ C NMR (125 MHz, CDCl ₃) spectrum of 1 .	4
Figure S4. HSQC NMR (500 MHz, CDCl ₃) spectrum of 1 .	5
Figure S5. HMBC NMR (500 MHz, CDCl ₃) spectrum of 1 .	5
Figure S6. HRqTOFMS for 2 .	6
Figure S7. ¹ H NMR (500 MHz, CDCl ₃) spectrum of 2 .	7
Figure S8. ¹³ C NMR (125 MHz, CDCl ₃) spectrum of 2 .	7
Figure S9. HSQC NMR (500 MHz, CDCl ₃) spectrum of 2 .	8
Figure S10. HMBC NMR (500 MHz, CDCl ₃) spectrum of 2 .	8
Figure S11. HRqTOFMS for 3 .	9
Figure S12. ¹ H NMR (500 MHz, CDCl ₃) spectra of 3(a) and 4(b) .	10
Figure S13. ¹³ C NMR (125 MHz, CDCl ₃) spectra of 3(a) and 4(b) .	11
Figure S14. HRqTOFMS for 8 .	12
Figure S15. ¹ H NMR (500 MHz, CDCl ₃) spectrum of 8 .	14
Figure S16. ¹³ C NMR (125 MHz, CDCl ₃) spectrum of 8 .	14
Figure S17. COSY NMR (500 MHz, CDCl ₃) spectrum of 8 .	15
Figure S18. HSQC NMR (500 MHz, CDCl ₃) spectrum of 8 .	16
Figure S19. HMBC NMR (500 MHz, CDCl ₃) spectrum of 8 .	16
Figure S20. NOESY NMR (500 MHz, CDCl ₃) spectrum of 8 .	17
Figure S21. Cytotoxicity of compounds 1~8 against THP-1 cells.	18
Figure S22. Cytotoxicity of compounds 1~8 against Caco-2 cells.	18
Figure S23. In vitro co-culture system of Caco-2 and THP-1 macrophages	19

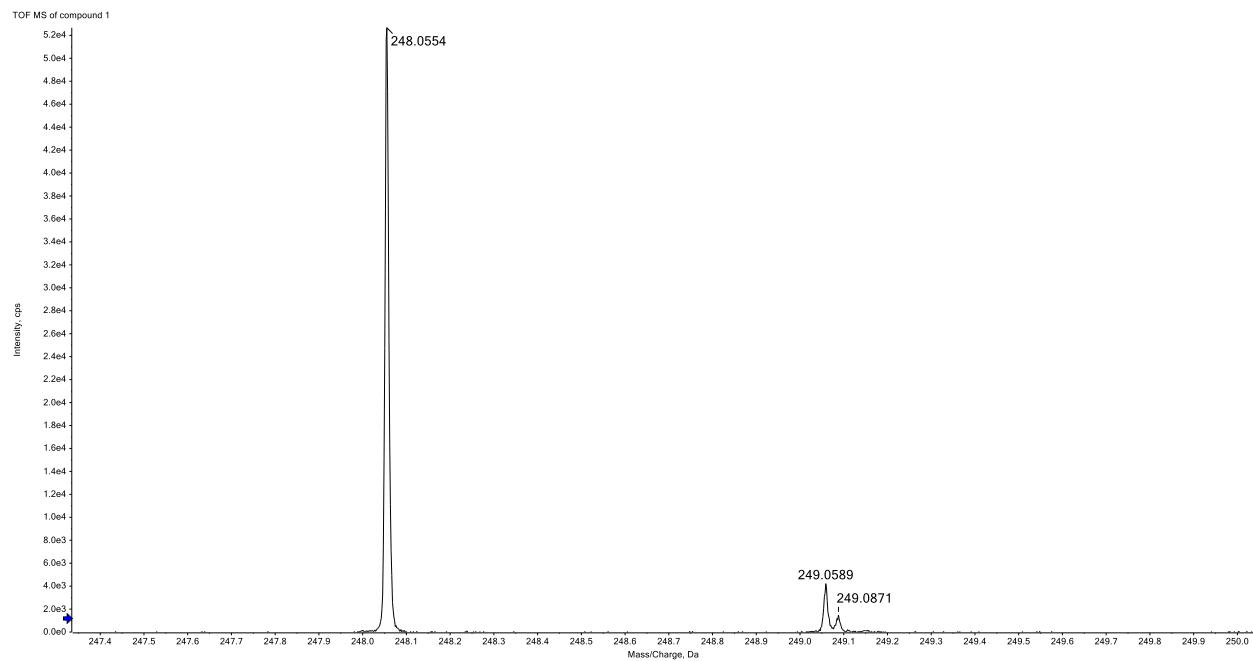


Figure S1. HRqTOFMS for **1**.

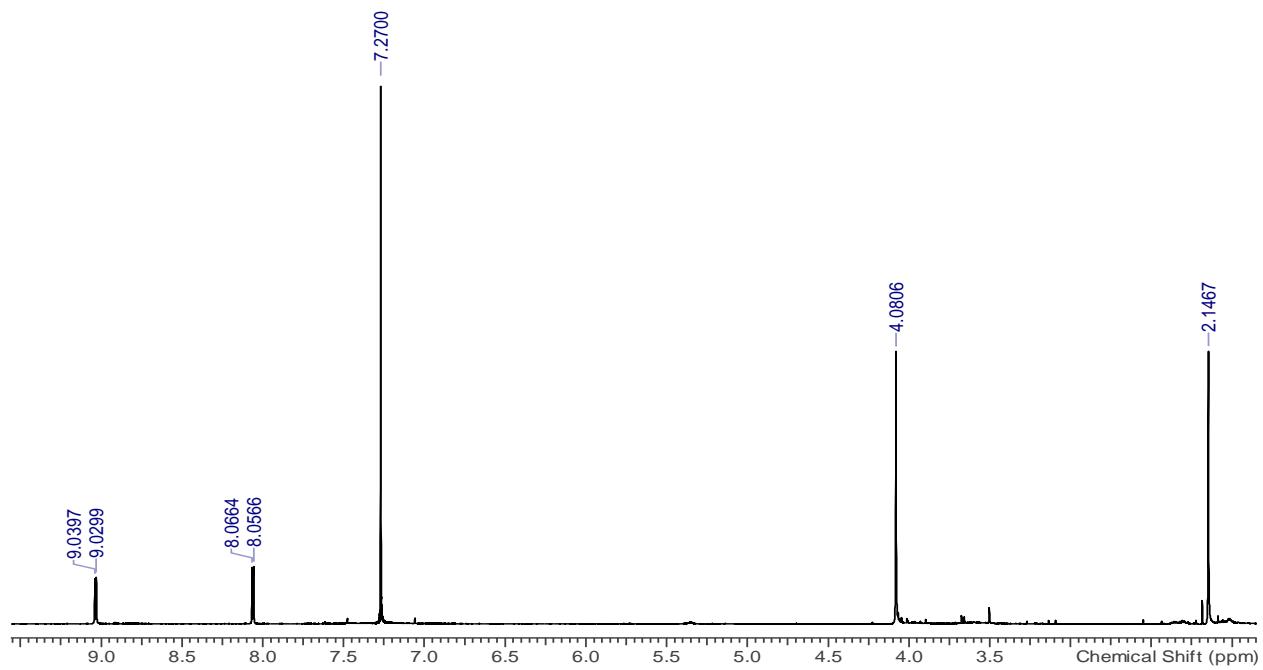


Figure S2. ^1H NMR (500 MHz, CDCl_3) spectrum of **1**.

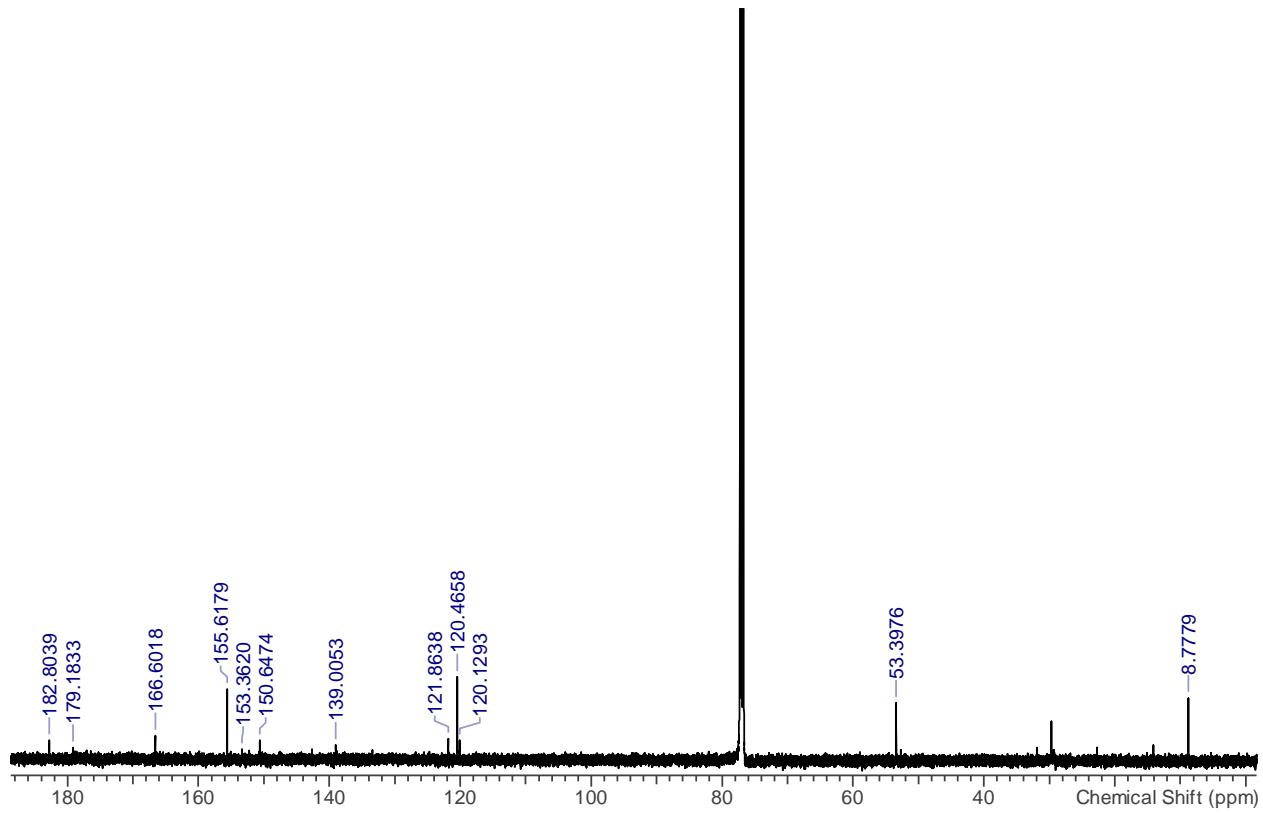


Figure S3. ^{13}C NMR (125 MHz, CDCl_3) spectrum of **1**.

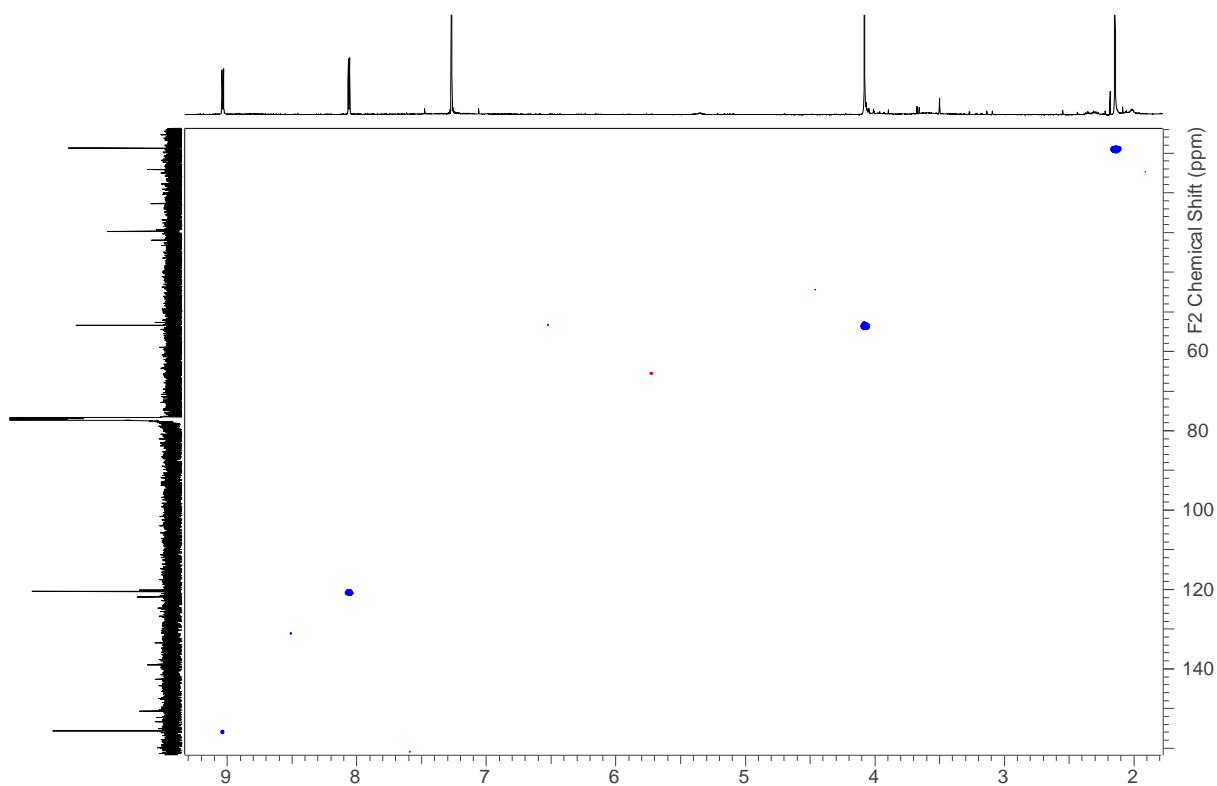


Figure S4. HSQC NMR (500 MHz, CDCl_3) spectrum of **1**.

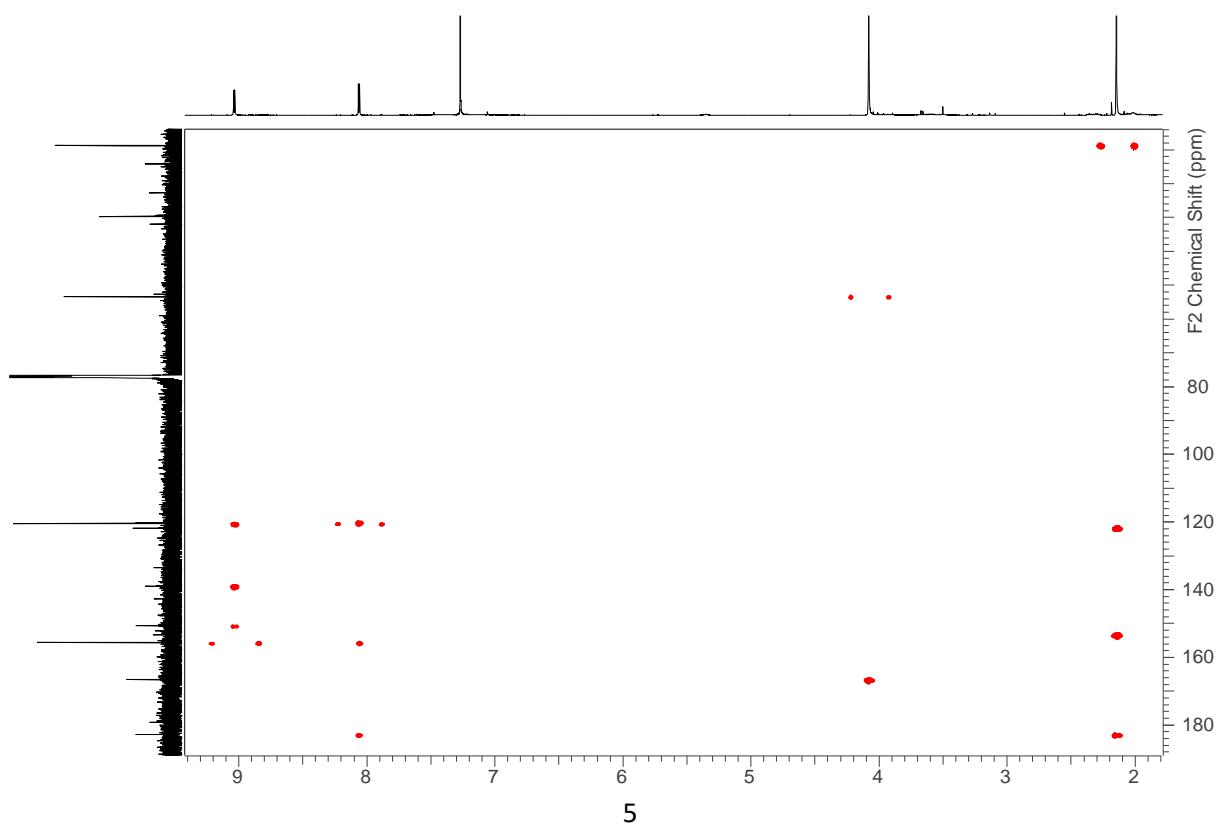


Figure S5. HMBC NMR (500 MHz, CDCl₃) spectrum of **1**.

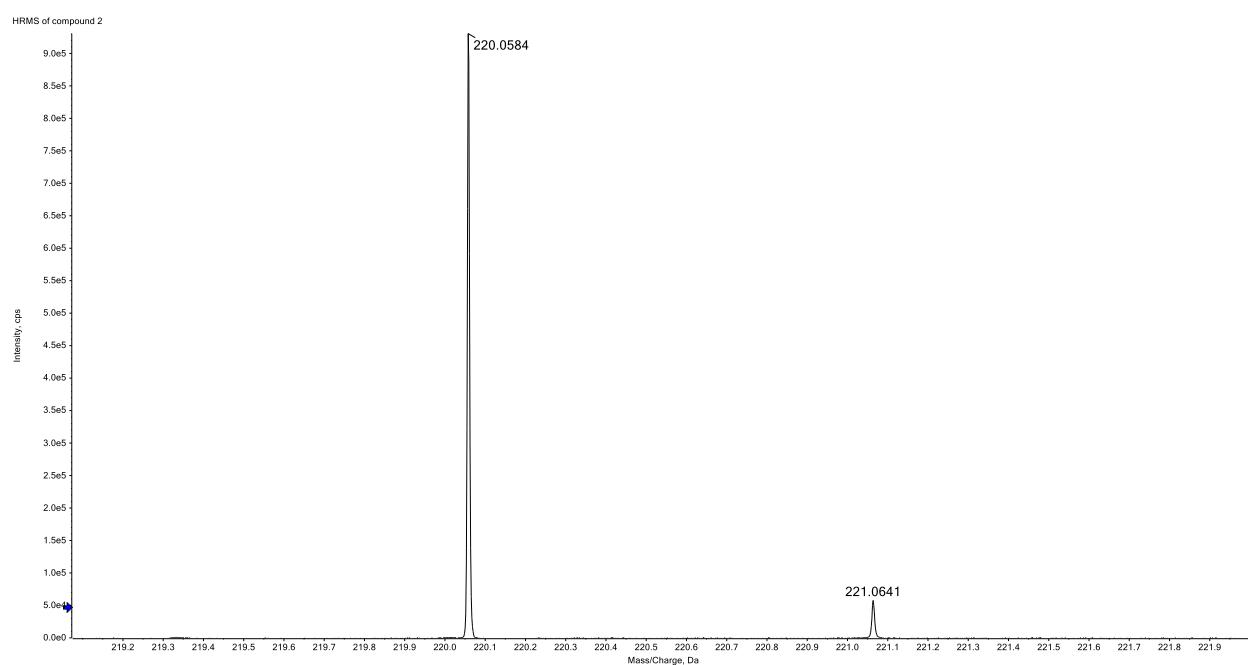


Figure S6. HRqTOFMS for **2**.

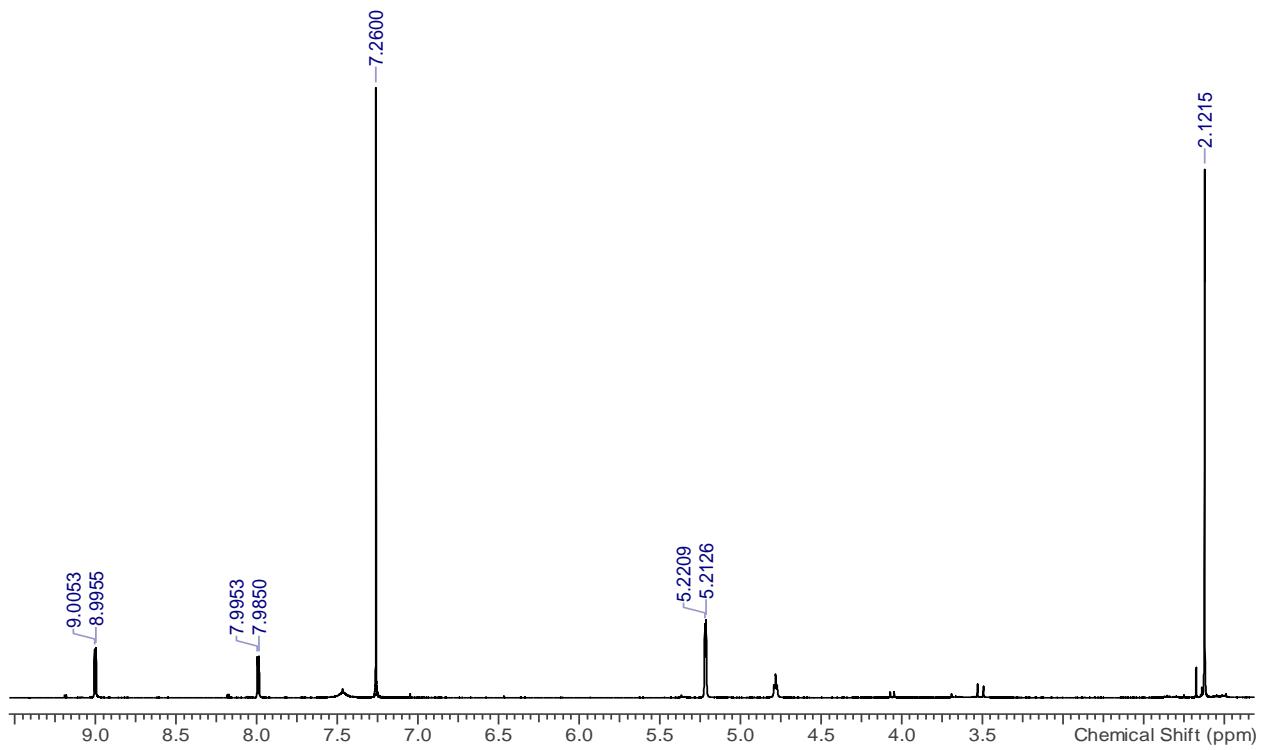


Figure S7. ^1H NMR (500 MHz, CDCl_3) spectrum of **2**.

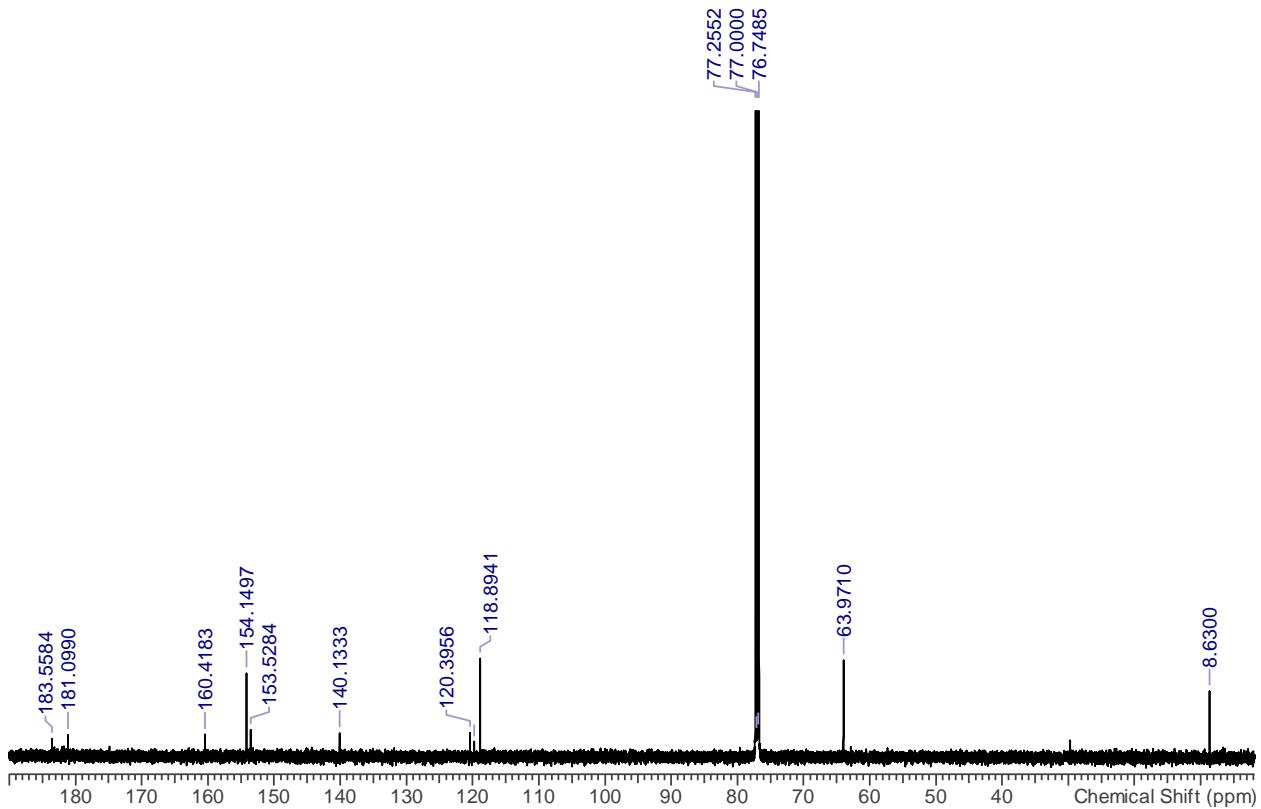


Figure S8. ^{13}C NMR (125 MHz, CDCl_3) spectrum of **2**.

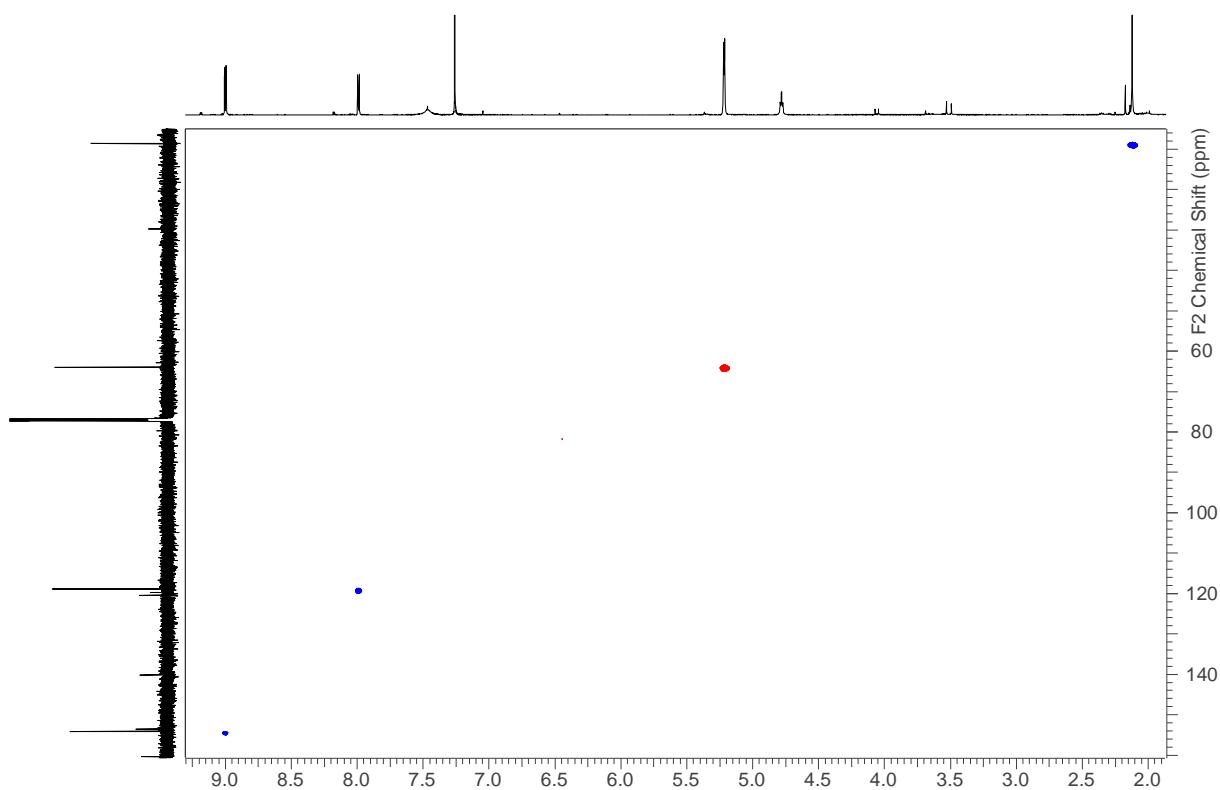


Figure S9. HSQC NMR (500 MHz, CDCl_3) spectrum of **2**.

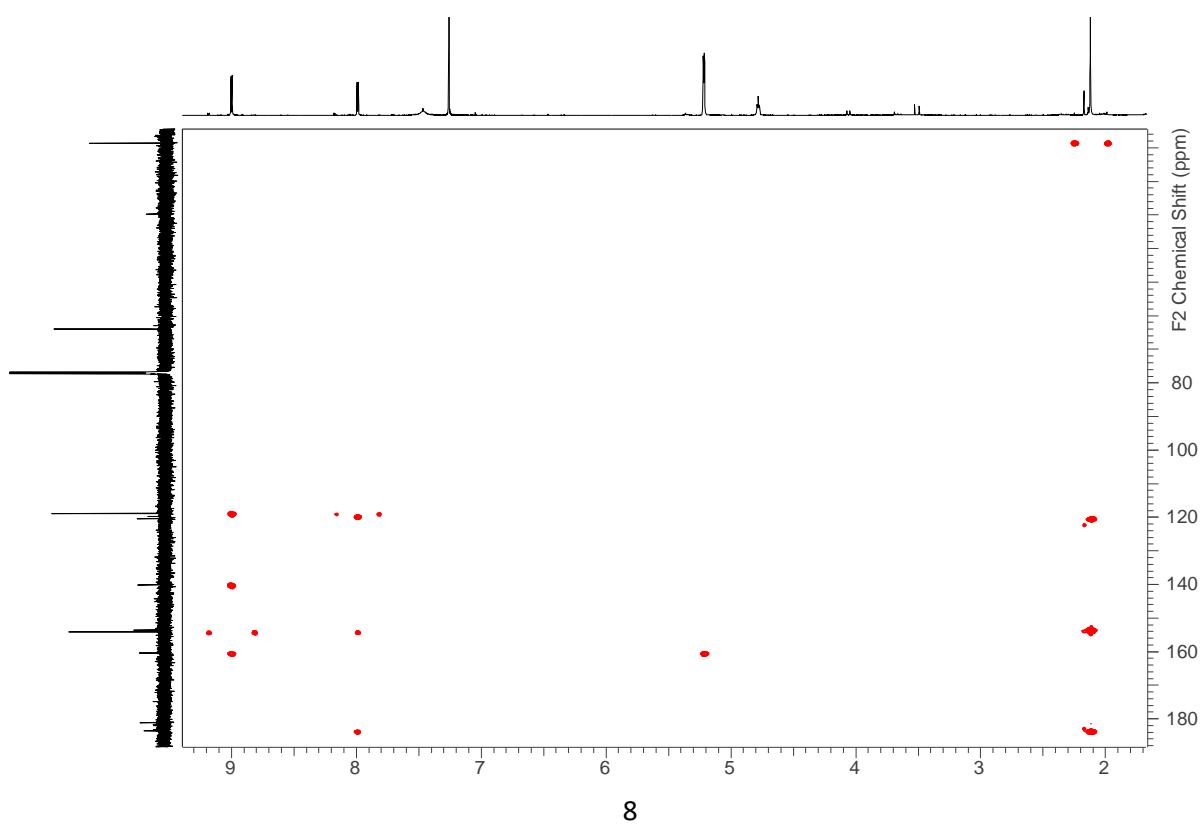


Figure S10. HMBC NMR (500 MHz, CDCl₃) spectrum of **2**.

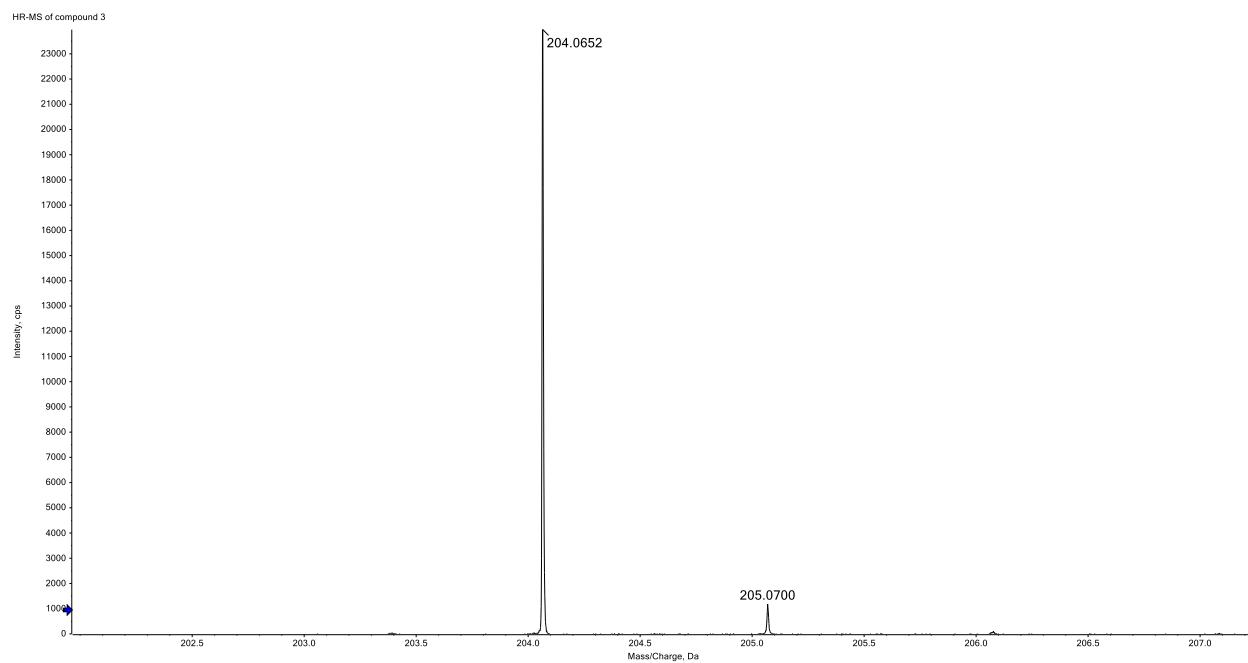
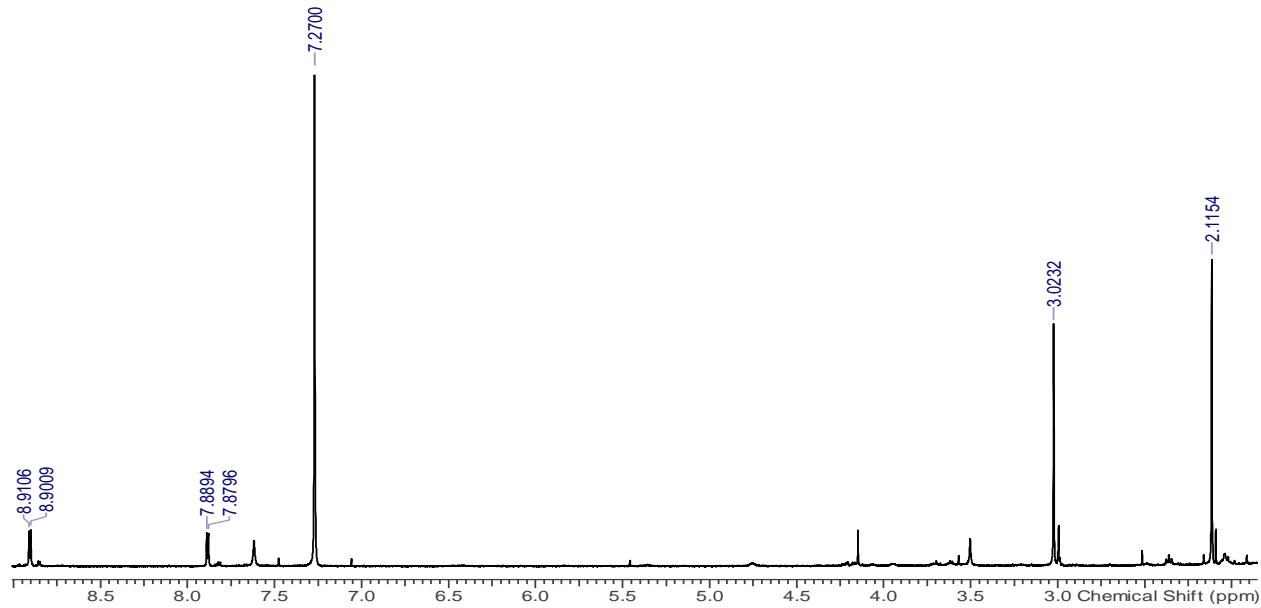


Figure S11. HRqTOFMS for **3**.

(a)



(b)

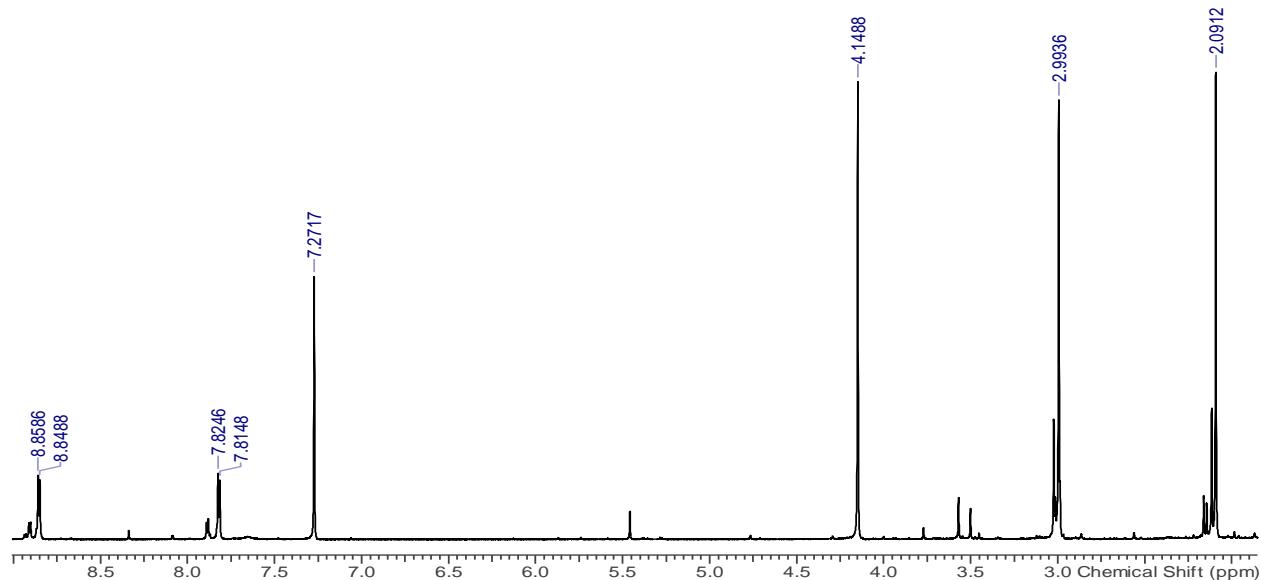
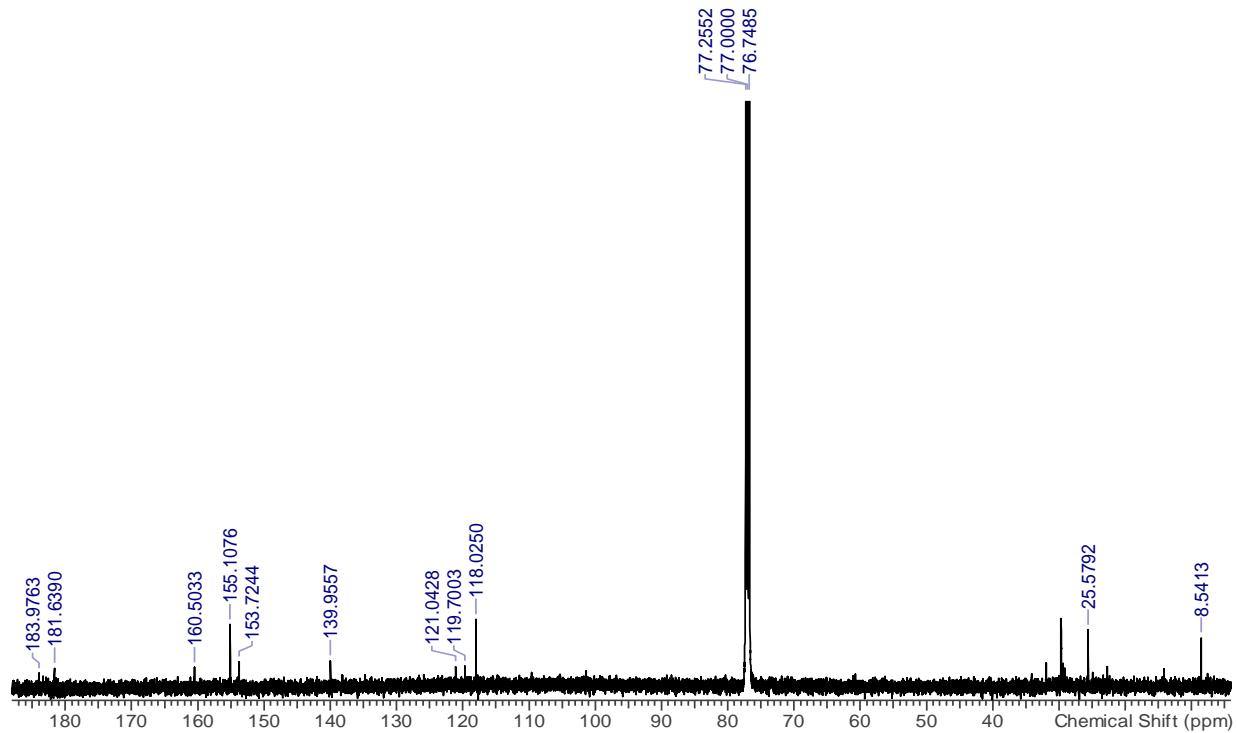


Figure S12. ¹H NMR (500 MHz, CDCl₃) spectra of **3(a)** and **4(b)**.

(a)



(b)

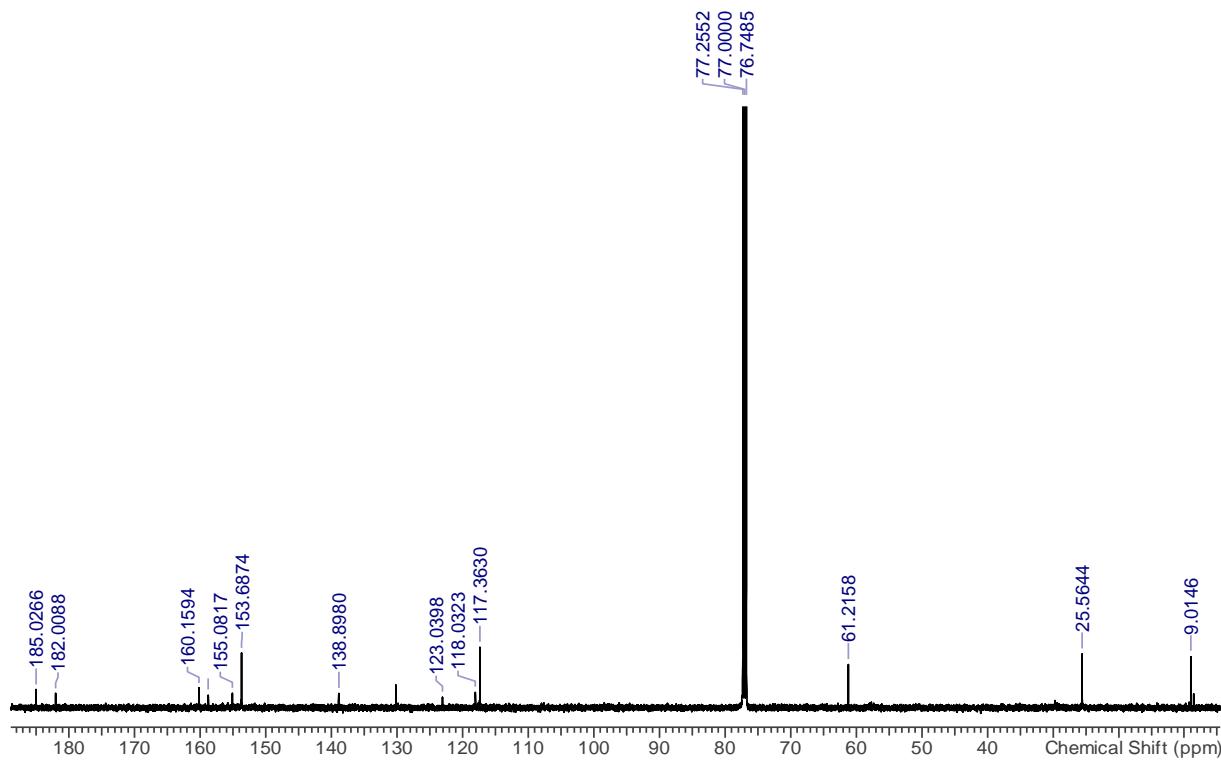


Figure S13. ^{13}C NMR (125 MHz, CDCl_3) spectra of **3(a)** and **4(b)**.

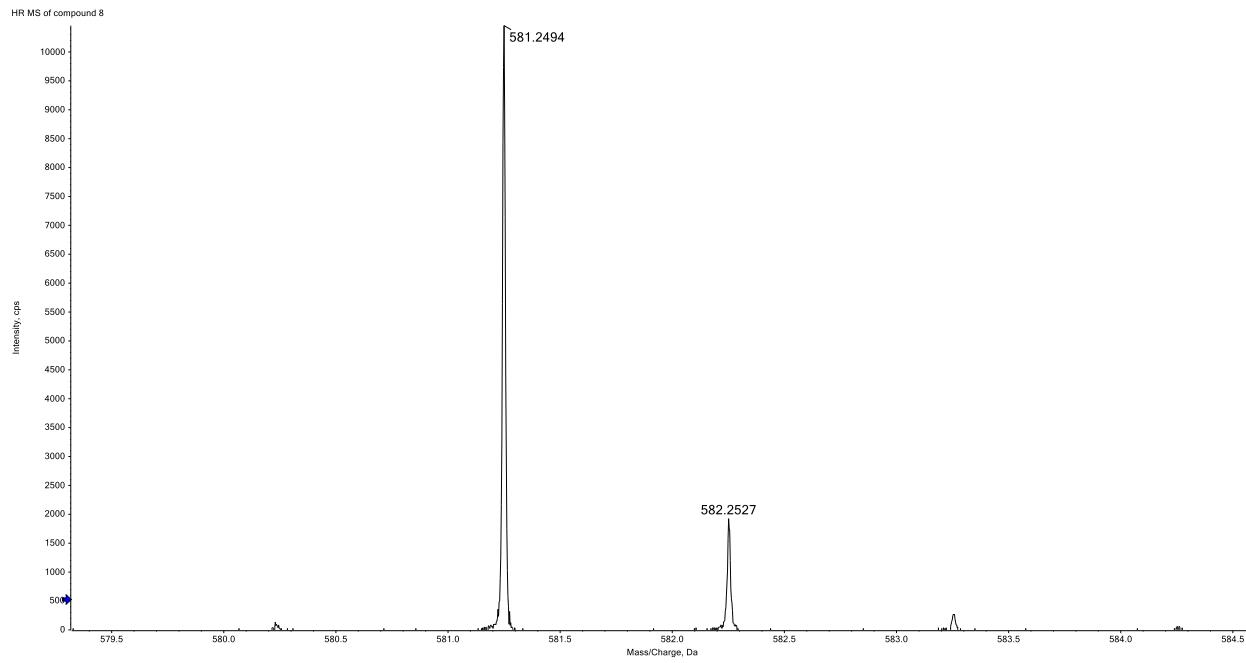


Figure S14. HRqTOFMS for **8**.

Table S1. Spectral data for compound **8** in CDCl₃ (500Hz, for ¹H; 125 Hz for ¹³C)

no	$\delta^{13}\text{C}$, mult	δH , mult(<i>J</i> Hz)
1	58.4, CH	3.62, m
3	56.0, CH	2.63, dt(11.4, 2.6)
4	25.8, CH ₂	1.22, ddd(11.4, 11.4, 3.7); 2.74, dd(11.4, 2.6)
5	185.7, C	
6	129.7, C	
7	155.1, C	
8	181.5, C	
9	136.9, C	
10	141.62, C	
11	55.1, CH	4.04, br d(2.6)
13	57.7, CH	3.23, m
14	71.8, CH	3.89, br s
15	185.7, C	
16	128.0, C	
17	156.1, C	
18	183.2, C	
19	136.2, C	
20	141.58, C	
21	56.1, CH ₂	2.77, dd(11.0, 3.2); 3.11, dd(11.0, 2.2)
22	62.8, CH ₂	4.26, dd(11.4, 2.5); 4.32, dd(11.4, 2.9)

24	167.0, C	
25	126.7, C	
26	139.4, CH	5.93, qq(7.2,1.4)
27	15.6, CH ₃	1.79, dq(7.2, 1.4)
6-CH ₃	8.67, CH ₃	1.91, s
16-CH ₃	8.65, CH ₃	1.94, s
25-CH ₃	20.3, CH ₃	1.57, br s
7-OCH ₃	60.9, CH ₃	3.96, s
14-OCH ₃	59.2, CH ₃	3.54, s
17-OCH ₃	60.8, CH ₃	4.00, s
12-NCH ₃	42.1, CH ₃	2.46, s

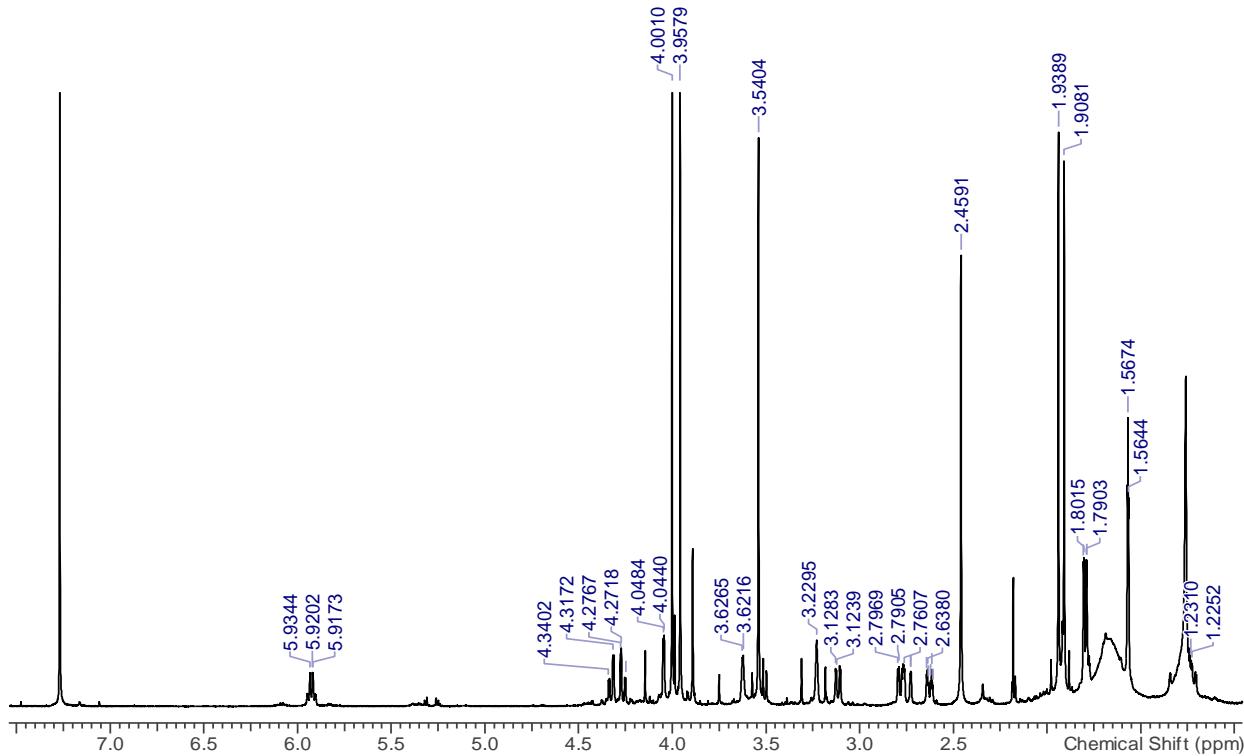


Figure S15. ^1H NMR (500 MHz, CDCl_3) spectrum of **8**.

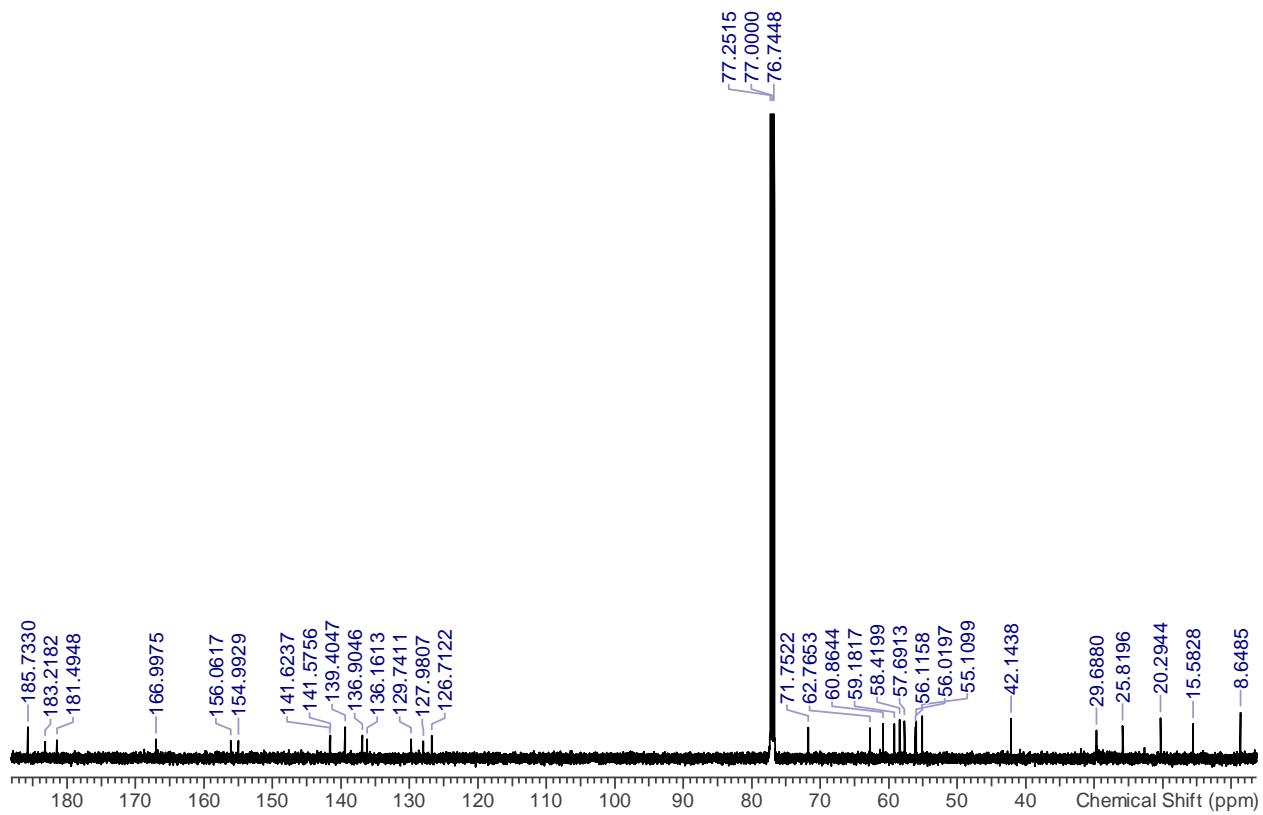


Figure S16. ^{13}C NMR (125 MHz, CDCl_3) spectrum of **8**.

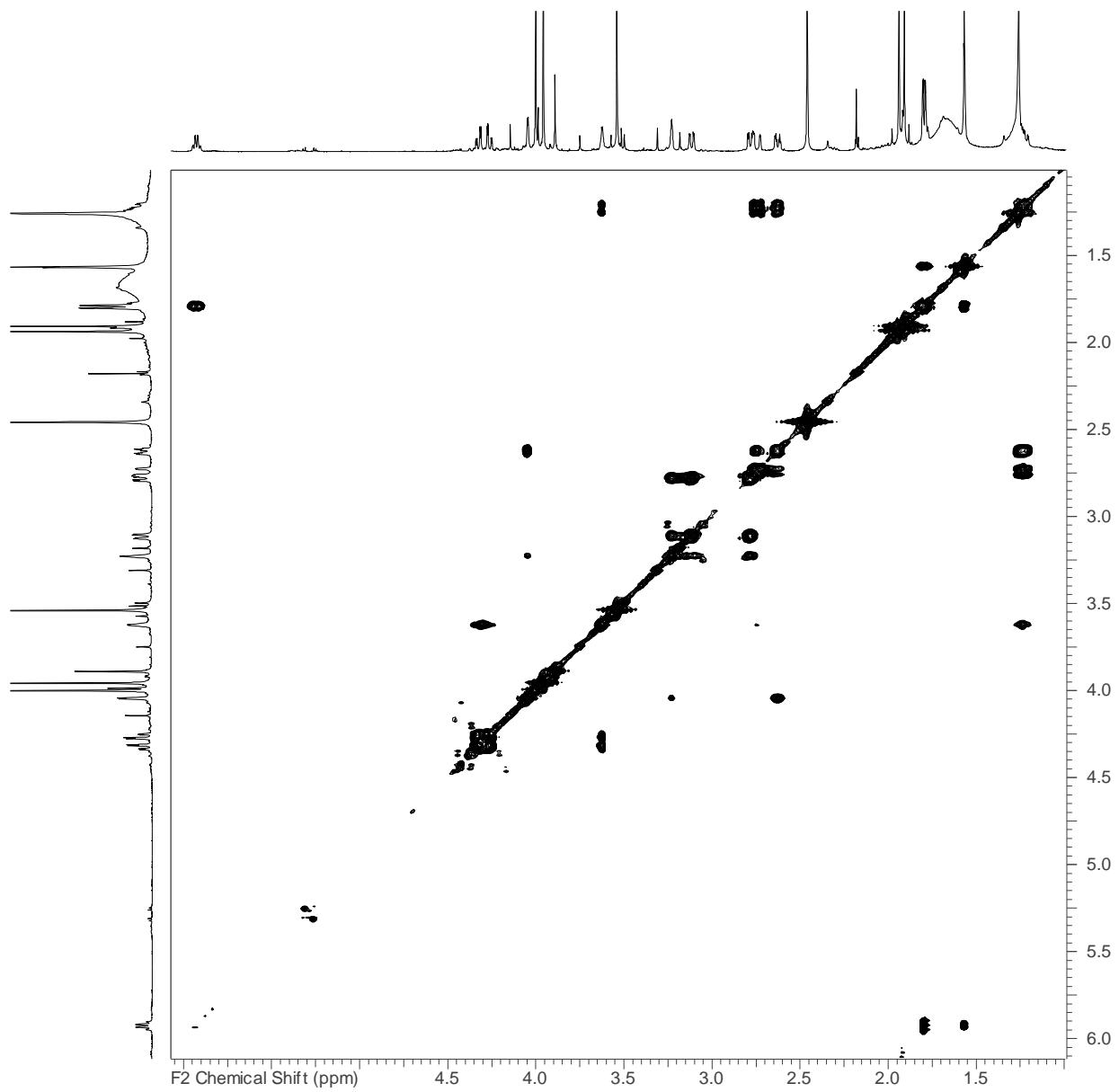


Figure S17. COSY NMR (500 MHz, CDCl_3) spectrum of **8**.

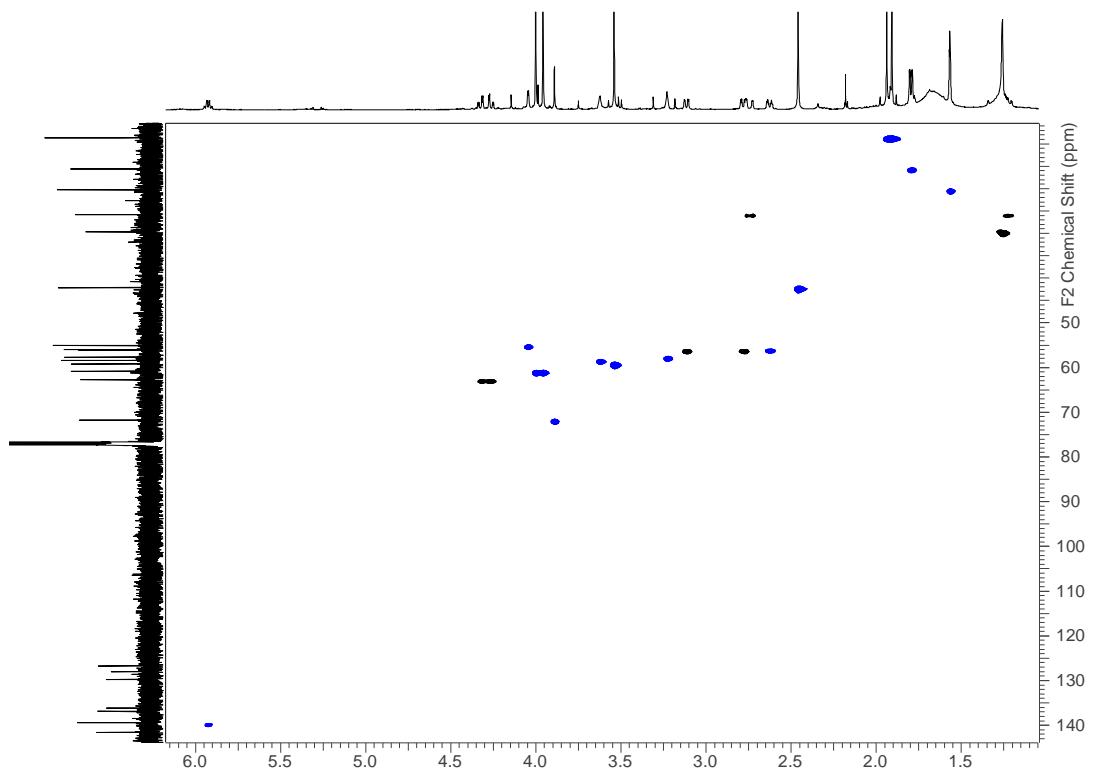


Figure S18. HSQC NMR (500 MHz, CDCl_3) spectrum of **8**.

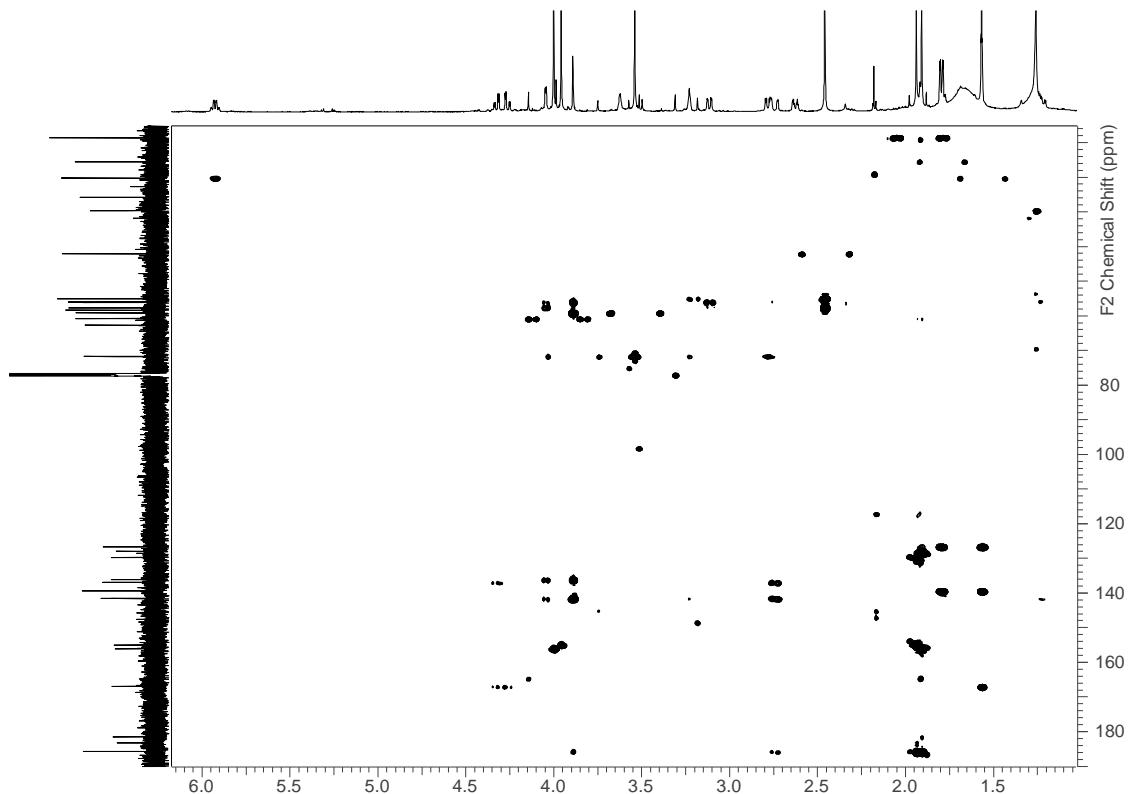


Figure S19. HMBC NMR (500 MHz, CDCl_3) spectrum of **8**.

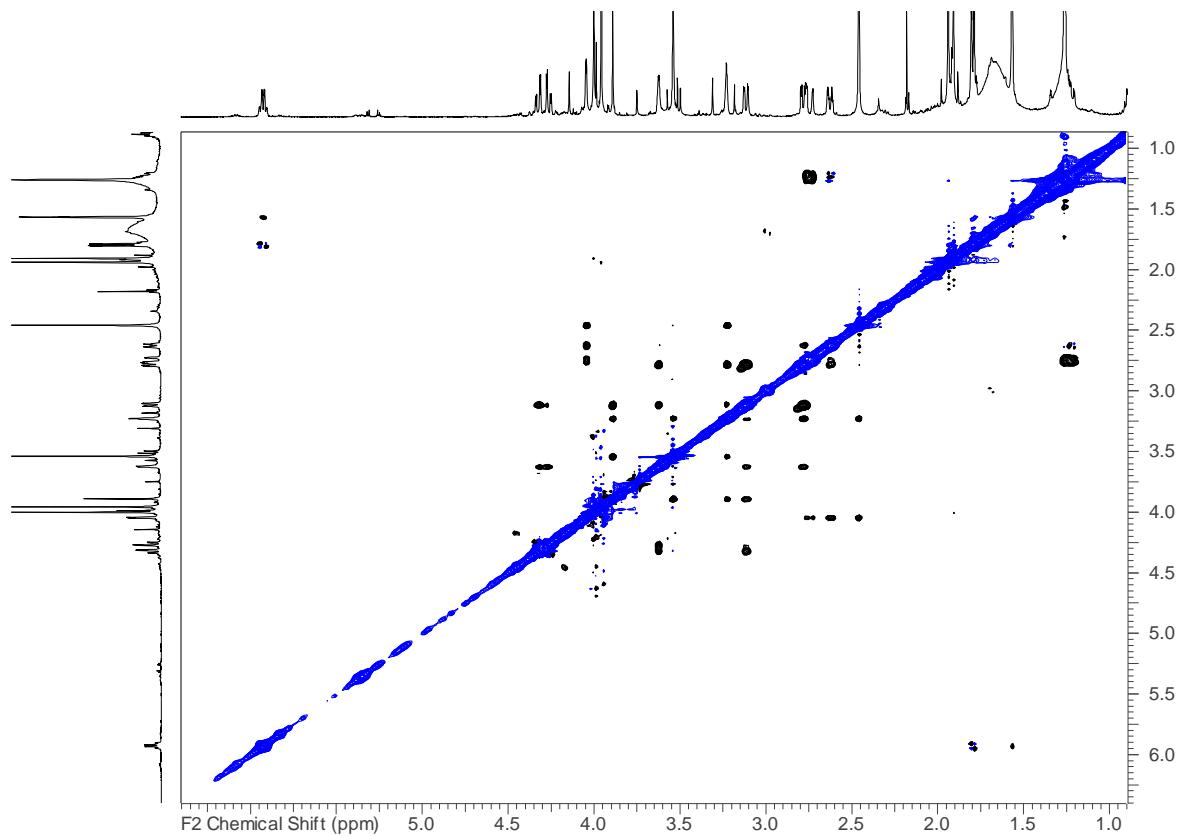


Figure S20. NOESY NMR (500 MHz, CDCl_3) spectrum of **8**.

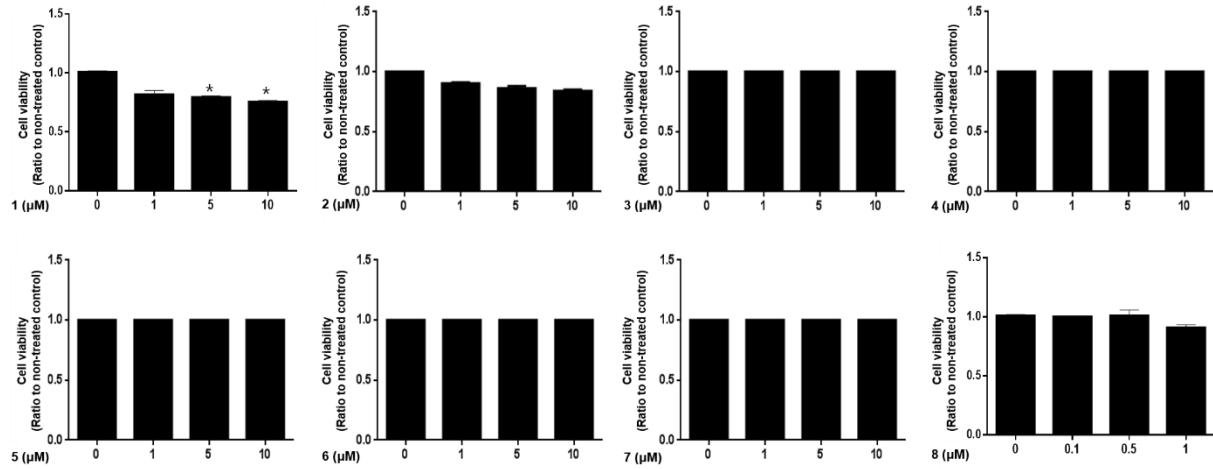


Figure S21: Cytotoxicity of compounds 1~8 against THP-1 cells. THP-1 macrophages were treated with each compound (1-8) at three concentrations indicated. After 24 h of incubation, cell viability was measured using a CCK-8 assay as described in materials and methods. Results are presented as the means \pm SDs of triplicate experiments; * $p < 0.05$ compared to non-treated control cells.

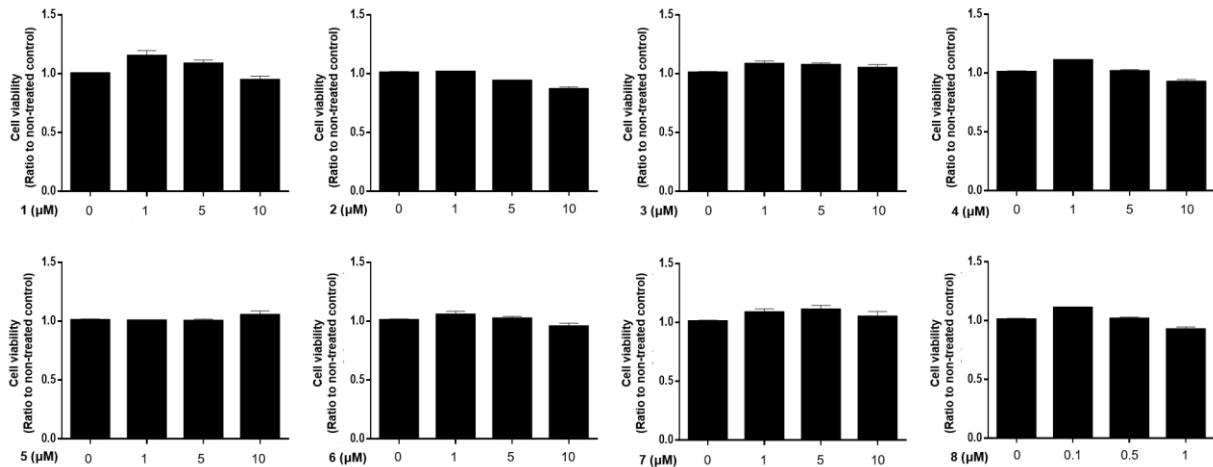


Figure S22: Cytotoxicity of compounds 1~8 against Caco-2 cells. Caco-2 epithelial cells were treated with each compound (1-8) at three concentrations indicated. After 24 h of incubation, cell viability was measured using a CCK-8 assay as described in materials and methods. Results are presented as the means \pm SDs of triplicate experiments.

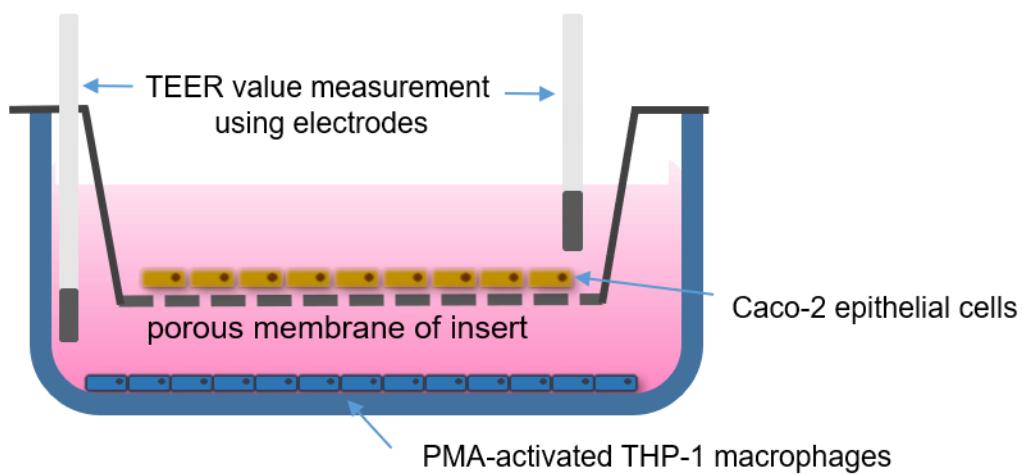


Figure S23: In vitro co-culture system of Caco-2 and THP-1 macrophages

Table S2. Coordinate for the optimized conformer of compound **1**.

Center Number	Atoms	Atomic type	X	Y	Z
1	C	0	-0.67446	2.634854	-0.07909
2	C	0	1.39187	0.84104	0.126022
3	C	0	-0.9777	1.277806	-0.03478
4	C	0	0.668978	3.013855	-0.0239
5	N	0	1.683583	2.145527	0.07913
6	C	0	0.074713	0.351406	0.064985
7	C	0	-0.24241	-1.087272	0.094751
8	C	0	-1.67882	-1.497746	0.045607
9	C	0	-2.71373	-0.618451	-0.04327
10	C	0	-2.41532	0.822146	-0.09279
11	O	0	-3.31192	1.657879	-0.17941
12	C	0	-4.15117	-1.043825	-0.09725
13	O	0	-1.86253	-2.826589	0.090559
14	O	0	0.603697	-1.979409	0.154744
15	C	0	2.594372	-0.058939	0.322516
16	O	0	3.03689	-0.333707	1.415289
17	O	0	3.118615	-0.437844	-0.84919
18	C	0	4.279801	-1.293805	-0.76906
19	H	0	-1.47213	3.365122	-0.15612
20	H	0	0.947774	4.064041	-0.06236
21	H	0	-4.24031	-2.129762	-0.05697
22	H	0	-4.62406	-0.676746	-1.01445
23	H	0	-4.71065	-0.605298	0.736071
24	H	0	-0.97109	-3.227938	0.149854
25	H	0	4.02259	-2.219952	-0.25053

26	H	0	5.08601	-0.785596	-0.23576
27	H	0	4.562039	-1.491612	-1.80214

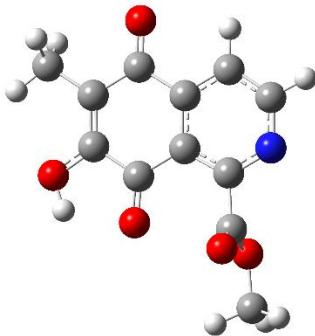


Table S3. Experimental and calculated ^{13}C NMR chemical shifts of compound 1.

	Experimental (Exp.) data Chemical shifts (ppm)	Calculated (Cald.) Data Shielding shifts (ppm)	Calculated (Cald.) Data Chemical shifts (ppm)	Difference between Exp. and Cald.
1	150.6	27.8983	152.1	-1.5
3	155.6	22.6133	157.1	-1.5
4	120.5	61.8512	119.5	1.0
5	182.8	-5.9330	184.5	-1.7
6	121.9	58.2206	123.5	-1.1
7	153.4	26.0895	153.8	-0.4
8	179.2	-1.8495	180.6	-1.4
9	120.1	62.3536	119.0	1.1
10	139.0	42.4741	138.1	0.9
11	166.6	10.4171	168.8	-2.2
6-CH ₃	8.8	175.9468	10.0	-1.2
OCH ₃	53.4	131.1725	53.0	0.4

Scaling factor for MPW1PW91/6-311+G(2d,p)//B3LYP/6-31+G(d,p)

- slope : -1.0420
- intercept : 186.3567

(<http://cheshirenmr.info/ScalingFactors.htm>)

