

Supplementary Materials

Metabolomics Tools Assisting Classic Screening Methods in Discovering New Antibiotics from Mangrove Actinomycetia in Leizhou Peninsula

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Figure S36. The HSQC spectrum of compound **12**.

Figure S37. The HMBC spectrum of compound **12**.

Table S1. Composition of 12 different media used to isolate actinomycetial strains from 13 mangrove soil samples.

No.	Name	Composition
M1	ISP 2 medium	Glucose 4.0 g, yeast extract 4.0 g, malt extract 10.0 g, water ^a 1000 mL, agar 15.0 g, pH 7.2-7.4
M2	GPT medium	Glucose 10.0 g, peptone 5.0 g, tryptone 3.0 g, NaCl 5.0 g, water ^a 1000 mL, agar 15.0 g, pH 7.2-7.4
M3	R2A medium	R2A agar 18.1 g, water ^a 1000 mL, pH 7.2-7.4
M4	TP medium	Trehalose 5.0 g, proline 1.0 g, peptone 1.0 g, yeast extract 0.5 g, (NH ₄) ₂ SO ₄ 1.0 g, NaCl 1.0 g, CaCl ₂ 2.0 g, K ₂ HPO ₄ 1.0 g, chlorhematin 0.01 mg, MgSO ₄ ·7H ₂ O 1.0 g, vitamin mixture ^d 1.0 mL, water ^a 1000 mL, agar 15.0 g, pH 7.2-7.4
M5	ISP 7 medium	Glycerol 15.0 mL, L-tyrosine 0.5 g, L-asparagine 1.0 g, K ₂ HPO ₄ 0.5 g, MgSO ₄ ·7H ₂ O 0.5 g, NaCl 0.5 g, FeSO ₄ ·7H ₂ O 0.01 g, trace salt solution ^e 1.0 mL, water ^a 1000 mL, agar 15.0 g, pH 7.2-7.4
M6	Arginine-glycerol medium	Arginine 1.0 g, glycerol 6.0 mL, vitamin mixture 1.0 mL, trace salt solution 1.0 mL, water ^a 1000 mL, agar 15.0 g, pH 7.2-7.4
M7	Starch-casein medium	Soluble starch 10.0 g, casein 0.3 g, KNO ₃ 2.0 g, MgSO ₄ ·7H ₂ O 0.05 g, NaCl 30.0 g, K ₂ HPO ₄ 2.0 g, CaCO ₃ 0.02 g, FeSO ₄ ·7H ₂ O 0.01 g, water ^a 1000 mL, agar 15.0 g, pH 7.2-7.4
M8	Asparagine-glycerol medium	L-asparagine 1.0 g, glycerol 10.0 mL, K ₂ HPO ₄ 1.0 g, trace salt solution 1.0 mL, water ^a 1000 mL, agar 15.0 g, pH 7.2-7.4
M9	1/10 ATCC 172 medium	Soluble starch 2.0 g, glucose 1.0 g, yeast extract 0.5 g, CaCO ₃ 1.5 g, N-Z-amine 0.5 g, water ^a 1000 mL, agar 15.0 g, pH 7.2-7.4
M10	RH medium	Raffinose 10.0 g, L-histidine 1.0 g, K ₂ HPO ₄ 1.0 g, MgSO ₄ 1.0 g, FeSO ₄ ·7H ₂ O 0.01 g, water ^a 1000 mL, agar 15.0 g, pH 7.2-7.4
M11	modified ISP 2 medium	Glucose 4.0 g, yeast extract 4.0 g, malt extract 10.0 g, <i>Laminaria japonica</i> juice (kelp juice) 15 mL, water ^b 985 mL, agar 15.0 g, pH 7.2-7.4
M12	modified TP medium	Trehalose 5.0 g, proline 1.0 g, peptone 1.0 g, yeast extract 0.5 g, (NH ₄) ₂ SO ₄ 1.0 g, NaCl 1.0 g, CaCl ₂ 2.0 g, K ₂ HPO ₄ 1.0 g, MgSO ₄ ·7H ₂ O 1.0 g, chlorhematin 0.01 mg, vitamin mixture 1.0 mL, fresh coconut juice 10 mL, water ^c 990 mL, agar 15.0 g, pH 7.2-7.4

^awater: 900.0 mL distilled water and 100.0 mL sea water; ^bwater: 885.0 mL distilled water and 100.0 mL sea water; ^cwater: 890.0 mL distilled water and 100.0 mL sea water

^dvitamin mixture: thiamine (0.10 g); pyridoxine (0.10 g); riboflavin (0.10 g); niacin (0.10 g); biotin (0.10 g); distilled water (100 mL); ^eTrace salt mixture: FeSO₄·7H₂O (0.20 g); MnCl₂·4H₂O (0.01 g); ZnSO₄·7H₂O (0.01 g); distilled water (100 mL).

Table S2. Information on genera distribution of actinomycetial strains isolated from 13 different mangrove soil samples.

Genus	S1	S2	S3	S4	S5	S6	S7	S8	S9	S10	S11	S12	S13	Isolates
<i>Micromonospora</i>	9	-	4	4	23	11	10	8	20	13	15	3	1	121
<i>Streptomyces</i>	34	8	2	-	15	-	2	22	6	8	8	4	7	116
<i>Microbacterium</i>	2	5	4	2	1	-	-	7	4	4	-	-	7	36
<i>Rhodococcus</i>	7	6	-	-	11	-	-	1	-	-	4	4	2	35
<i>Brachybacterium</i>	-	-	-	-	-	-	1	1	8	1	-	-	17	28
<i>Isoptericola</i>	-	1	-	-	-	-	-	-	13	8	-	-	3	25
<i>Cellulosimicrobium</i>	3	9	4	1	2	-	-	1	-	-	-	-	1	21
<i>Brevibacterium</i>	2	2	2	2	2	-	-	-	2	3	-	-	2	17
<i>Serinibacter</i>	-	-	-	1	-	-	-	3	8	-	-	-	1	13
<i>Agromyces</i>	-	2	2	-	-	-	1	3	-	1	-	1	-	10
<i>Micrococcus</i>	-	-	-	-	-	1	-	6	-	1	1	1	-	10
<i>Mycolicibacterium</i>	2	1	2	-	-	-	-	1	1	2	-	-	1	10
<i>Kocuria</i>	-	1	1	1	-	-	-	3	1	1	1	-	-	9
<i>Gordonia</i>	2	3	-	-	-	-	-	1	-	-	-	-	-	6
<i>Mycobacterium</i>	-	-	-	-	-	-	-	-	-	6	-	-	-	6
<i>Aeromicrobium</i>	1	-	1	-	1	1	-	-	1	-	-	-	-	5
<i>Arthrobacter</i>	-	1	-	-	-	-	1	1	-	-	1	1	-	5
<i>Citricoccus</i>	-	-	-	1	1	-	-	3	-	-	-	-	-	5
<i>Janibacter</i>	2	1	2	-	-	-	-	-	-	-	-	-	-	5
<i>Nocardia</i>	2	-	1	-	-	-	-	-	-	-	2	-	-	5
<i>Corynebacterium</i>	2	-	2	-	-	-	-	-	-	-	-	-	-	4
<i>Glutamicibacter</i>	-	1	1	-	-	-	-	-	-	-	-	-	2	4
<i>Agrococcus</i>	1	-	-	-	-	-	-	2	-	-	-	-	-	3
<i>Intrasporangium</i>	2	-	-	-	-	-	-	-	-	-	-	-	-	2
<i>Kineococcus</i>	-	-	-	-	-	-	-	-	2	-	-	-	-	2
<i>Phycoccus</i>	-	-	-	-	-	-	-	-	-	-	-	-	2	2
<i>Serinicoccus</i>	-	-	-	-	-	-	-	-	-	-	-	-	2	2
<i>Sinomonas</i>	1	-	1	-	-	-	-	-	-	-	-	-	-	2
<i>Actinomadura</i>	1	-	-	-	-	-	-	-	-	-	-	-	-	1
<i>Actinopolymorpha</i>	-	-	-	-	-	-	-	-	1	-	-	-	-	1
<i>Blastococcus</i>	-	-	-	-	-	-	-	-	-	-	-	1	-	1
<i>Demequina</i>	-	-	-	-	-	-	-	-	-	-	-	-	1	1
<i>Georgenia</i>	-	-	-	1	-	-	-	-	-	-	-	-	-	1
<i>Gulosibacter</i>	1	-	-	-	-	-	-	-	-	-	-	-	-	1
<i>Jonesia</i>	-	-	1	-	-	-	-	-	-	-	-	-	-	1
<i>Leucobacter</i>	-	-	-	-	-	-	-	-	1	-	-	-	-	1
<i>Motilibacter</i>	-	-	-	-	-	-	-	-	1	-	-	-	-	1

<i>Mumia</i>	-	-	-	-	-	-	-	-	1	-	-	-	-	1
<i>Salinibacterium</i>	-	-	-	-	-	1	-	-	-	-	-	-	-	1
<i>Streptacidiphilus</i>	-	-	-	-	-	-	-	-	-	-	1	-	-	1
Isolates	74	41	30	13	56	14	15	63	70	48	33	15	49	521
Genera	17	13	15	8	8	4	5	15	15	11	8	7	14	40

:- No isolate

Table S3. Information on genera distribution of actinomycetial strains recovered from 12 different cultural media.

Genus	M1	M2	M3	M4	M5	M6	M7	M8	M9	M10	M11	M12	Isolates
<i>Micromonospora</i>	17	23	12	-	15	7	1	10	12	19	5	-	121
<i>Streptomyces</i>	7	6	12	5	21	10	10	18	12	9	5	1	116
<i>Microbacterium</i>	9	1	4	-	1	1	-	1	5	3	10	1	36
<i>Rhodococcus</i>	3	4	7	-	1	5	2	1	3	7	2	-	35
<i>Brachybacterium</i>	3	1	1	3	3	1	4	1	5	-	6	-	28
<i>Isoptericola</i>	7	4	2	-	4	-	-	-	1	1	6	-	25
<i>Cellulosimicrobium</i>	3	2	3	1	3	-	1	1	2	1	3	1	21
<i>Brevibacterium</i>	-	2	3	2	2	2	-	-	4	-	-	2	17
<i>Serinibacter</i>	2	1	1	1	-	1	-	2	3	1	1	-	13
<i>Mycolicibacterium</i>	3	2	-	-	-	-	-	3	-	-	2	-	10
<i>Micrococcus</i>	-	2	-	-	-	3	-	1	-	-	3	1	10
<i>Agromyces</i>	1	-	2	-	-	1	4	-	-	-	1	1	10
<i>Kocuria</i>	-	-	-	-	1	-	-	-	-	1	4	3	9
<i>Mycobacterium</i>	2	-	1	-	-	-	-	-	-	-	3	-	6
<i>Gordonia</i>	-	2	1	-	-	-	-	-	1	1	1	-	6
<i>Aeromicrobium</i>	-	1	2	-	1	-	-	-	-	-	1	-	5
<i>Arthrobacter</i>	1	1	-	-	-	2	-	-	-	-	-	1	5
<i>Citricoccus</i>	1	-	-	-	-	2	-	-	-	-	-	2	5
<i>Janibacter</i>	-	-	1	1	1	-	1	-	-	-	1	-	5
<i>Nocardia</i>	2	-	1	-	-	-	-	1	-	-	1	-	5
<i>Corynebacterium</i>	1	-	1	-	-	-	1	-	-	-	1	-	4
<i>Glutamicibacter</i>	-	-	1	-	-	-	-	-	-	-	-	3	4
<i>Agrococcus</i>	-	1	-	-	-	1	1	-	-	-	-	-	3
<i>Intrasporangium</i>	-	-	-	-	1	1	-	-	-	-	-	-	2
<i>Kineococcus</i>	1	1	-	-	-	-	-	-	-	-	-	-	2
<i>Phycococcus</i>	-	-	-	-	-	-	-	-	-	-	2	-	2
<i>Sinomonas</i>	1	-	-	-	1	-	-	-	-	-	-	-	2
<i>Serinicoccus</i>	-	-	-	-	-	-	-	-	-	-	2	-	2
<i>Actinomadura</i>	-	-	1	-	-	-	-	-	-	-	-	-	1
<i>Actinopolymorpha</i>	-	-	-	-	1	-	-	-	-	-	-	-	1
<i>Blastococcus</i>	-	-	-	-	-	-	-	-	-	-	1	-	1
<i>Demequina</i>	-	-	-	-	-	-	-	-	-	-	1	-	1
<i>Georgenia</i>	-	-	-	-	-	-	-	-	-	-	-	1	1
<i>Gulosibacter</i>	-	-	-	-	-	-	-	-	1	-	-	-	1

<i>Jonesia</i>	-	-	-	-	-	-	-	-	-	-	-	1	1
<i>Leucobacter</i>	-	-	-	-	-	-	-	-	1	-	-	-	1
<i>Motilibacter</i>	-	1	-	-	-	-	-	-	-	-	-	-	1
<i>Mumia</i>	1	-	-	-	-	-	-	-	-	-	-	-	1
<i>Salinibacterium</i>	-	-	-	-	-	-	-	-	-	-	1	-	1
<i>Streptacidiphilus</i>	-	-	-	-	1	-	-	-	-	-	-	-	1
Isolates	65	55	56	13	57	37	25	39	50	43	63	18	521
Genera	18	17	18	6	15	13	9	10	12	9	23	12	40

-: No isolate

Table S4. Antibacterial activities of 179 actinomycetal strains by the paper disc diffusion method.

[illegible]

[illegible]

M76 (MW724585)	<i>Janibacter indicus</i> CGMCC 1.12511 ^T (99.88)	-	-	-	-	-	-	-	-	-	-	-	-
M81 (MW724586)	<i>Citricoccus alkalitolerans</i> YIM 70010 ^T (99.63)	-	-	-	-	-	-	-	-	-	-	-	-
M84 (MW724587)	<i>Micromonospora auratinigra</i> DSM 44815 ^T (99.07)	-	-	-	-	-	-	-	-	-	-	-	-
M86 (MW724588)	<i>Aeromicrobium tamense</i> SSW1-57 ^T (99.87)	-	-	-	-	-	-	-	-	-	-	-	-
M90 (MW724589)	<i>Kocuria dechangensis</i> NEAU-ST5-33 ^T (99.09)	-	-	-	-	-	-	-	-	-	-	-	-
M91 (MW724590)	<i>Brevibacterium permense</i> VKM AC-2280 ^T (99.87)	-	-	-	-	-	-	-	-	-	-	-	-
M102 (MW724591)	<i>Arthrobacter gandavensis</i> R812 ^T (99.60)	-	-	-	-	-	-	-	-	-	-	-	-
M104 (MW724592)	<i>Streptomyces griseoflavus</i> LMG 19344 ^T (99.75)	-	-	-	-	-	-	-	-	-	-	-	-
M106 (MW724593)	<i>Streptomyces geysiriensis</i> NBRC 15413 ^T (99.75)	14	9	7	-	16	-	10	8	14	20	11	13
M107 (MW724594)	<i>Streptomyces andamanensis</i> KC-112 ^T (99.75)	-	-	-	-	-	-	-	-	7	7	-	-
M108 (MW724595)	<i>Phycococcus endophyticus</i> IP6SC6 ^T (98.61)	-	-	-	-	-	-	-	-	-	-	-	-
M110 (MW724596)	<i>Agromyces tropicus</i> CM9-9 ^T (99.88)	-	-	-	-	-	-	-	-	-	-	-	-
M111 (MW724597)	<i>Streptomyces smyrnaeus</i> SM3501 ^T (99.74)	-	-	-	-	9	-	-	-	13	16	17	15
M112 (MW724598)	<i>Micromonospora globispora</i> S2901 ^T (99.63)	-	-	-	-	-	-	-	-	-	-	-	-
M118 (MW724599)	<i>Microbacterium hominis</i> NBRC 15708 ^T (98.91)	-	-	-	-	-	-	-	-	-	-	-	-
M129 (MW724600)	<i>Streptomyces nanshensis</i> SCSIO 01066 ^T (99.63)	-	-	-	-	-	-	-	-	-	-	-	-
M131 (MW724601)	<i>Cellulosimicrobium marinum</i> RS-7-4 ^T (100.00)	-	-	-	-	-	-	-	-	-	-	-	-
M133 (MW724602)	<i>Blastococcus aggregatus</i> DSM 4725 ^T (99.61)	-	-	-	-	-	-	-	-	-	-	-	-
Y2 (MW724603)	<i>Streptomyces hygroscopicus</i> subsp. <i>hygroscopicus</i> NBRC 13472 ^T (99.63)	-	-	-	-	-	-	-	-	16	21	18	23
Y3 (MW724604)	<i>Streptomyces aurantiogriseus</i> NBRC 12842 ^T (99.40)	-	-	-	-	-	-	-	-	7	8	-	-
Y4 (MW724605)	<i>Streptomyces albogriseolus</i> NRRL B-1305 ^T (100.00)	12	13	-	-	17	10	10	9	17	26	-	16
Y8 (MW724606)	<i>Streptomyces pseudogriseolus</i> NRRL B-3288 ^T (100.00)	-	-	-	-	-	-	-	-	7	-	11	-
Y9 (MW724607)	<i>Streptomyces pluripotens</i> MUSC 135 ^T (99.87)	14	17	7	8	16	-	17	-	16	23	-	-
Y13 (MW724608)	<i>Streptomyces qinglanensis</i> 172205 ^T (100.00)	-	-	-	-	-	-	-	-	8	8	14	-
Y14 (MW724609)	<i>Streptomyces coelicoflavus</i> NBRC 15399 ^T (100.00)	-	-	-	-	-	-	-	-	-	-	10	-
Y15 (MW724610)	<i>Streptomyces cellulosae</i> NBRC 13027 ^T (100.00)	-	-	-	-	-	-	-	-	8	11	-	13

[illegible]

[illegible]

[illegible]

[illegible]

M116 (MW724713)	<i>Serinicoccus profundus</i> MCCC 1A05965 ^T (98.69)		-	-	-	-	-	-	-	-	-	-		
Methanol			-	-	-	-	-	-	-	-	-	-		
Levofloxacin (10 µg)			36	39	26	19	40	24	28	23	24	15	19	-

Paper disk diameter, 6 mm; -, no Inhibitory zone.

Table S5. The trioxacarcin-type antibiotics isolated from actinomycetial strains.

Name	UVmax (nm)	Exact mass	Molecular Formula	Ref.
LL-D49194 α 1	230, 270, 399	992	C ₄₈ H ₆₄ O ₂₂	[1,2]
LL-D49194 β 1	230, 270, 399	1010	C ₄₈ H ₆₆ O ₂₃	[1,2]
LL-D49194 β 2	230, 270, 399	950	C ₄₆ H ₆₂ O ₂₁	[1,2]
LL-D49194 β 3	230, 270, 399	806	C ₃₉ H ₅₀ O ₁₈	[1]
LL-D49194 γ	225, 270, 399(sh)	-	-	[1]
LL-D49194 δ	230, 270, 399	-	-	[1]
LL-D49194 ϵ	230, 270, 399	1052	C ₅₀ H ₆₈ O ₂₄	[1]
LL-D49194 ζ	230, 270, 399	-	-	[1]
LL-D49194 η	230, 270, 399	848	C ₄₁ H ₅₂ O ₁₉	[1]
LL-D49194 ω 1	230, 270, 399	968	C ₄₆ H ₆₄ O ₂₂	[1]
LL-D49194 ω 2	230, 270, 399	-	-	[1]
LL-D49194 ω 3	230, 270, 399	866	C ₄₁ H ₅₄ O ₂₀	[1]
Parimycin	260, 423, 447	396	C ₂₂ H ₂₀ O ₇	[3]
trioxacarcin A	233, 271, 399	876	C ₄₂ H ₅₂ O ₂₀	[4,5]
trioxacarcin B	233, 271, 399	894	C ₄₂ H ₅₄ O ₂₁	[4,5]
trioxacarcin C	233, 271, 399	878	C ₄₂ H ₅₄ O ₂₀	[4,5]
trioxacarcin D	270, 396	834	C ₄₀ H ₅₀ O ₁₉	[5]
trioxacarcin E	-	740	C ₃₄ H ₄₄ O ₁₈	[5]
trioxacarcin F	-	912	C ₄₂ H ₅₆ O ₂₂	[5]
Gutingimycin	269, 399	1027	C ₄₇ H ₅₇ N ₅ O ₂₁	[5,6]
DC-45-A1	-	704	C ₃₄ H ₄₀ O ₁₆	[7-9]
DC-45-A2	-	518	C ₂₅ H ₂₆ O ₁₂	[7-9]

-: no data

Table S6. The information of mangrove-derived soil samples in different sites of Leizhou Peninsula, China.

Samples	Sampling sites	The characteristic of soil	Longitude (E)	Latitude (N)	Sampling depth
Sample 1	He'an Town, Xuwen County	Rhizosphere soil of <i>Sonneratia apetala</i>	110.3708830	20.6395330	5-10 cm under surface
Sample 2	He'an Town, Xuwen County	Rhizosphere soil of <i>Sonneratia apetala</i>	110.3708830	20.6396330	5-10 cm under surface
Sample 3	He'an Town, Xuwen County	Rhizosphere soil of <i>Aegiceras corniculatum</i>	110.3709170	20.6395830	5-10 cm under surface
Sample 4	Dongsong Island, He'an Town, Xuwen County	Rhizosphere soil of <i>Avicennia marina</i>	110.3666000	20.6785170	5-10 cm under surface
Sample 5	Dongsong Island, He'an Town, Xuwen County	Rhizosphere soil of <i>Rhizophora stylosa</i>	110.3665170	20.6787170	5-10 cm under surface
Sample 6	Dongsong Island, He'an Town, Xuwen County	Rhizosphere soil of <i>Kandelia candel</i>	110.3665330	20.6787170	5-10 cm under surface
Sample 7	Dongsong Island, He'an Town, Xuwen County	Muddy soil of intertidal zone in mangrove	110.3674500	20.6799330	5-10 cm under surface
Sample 8	Dongsong Island, He'an Town, Xuwen County	Rhizosphere soil of <i>Sonneratia apetala</i>	110.3686330	20.6665330	5-10 cm under surface
Sample 9	Maichen Town, Xuwen County	Rhizosphere soil of <i>Rhizophora stylosa</i>	110.0107830	20.4205000	5-10 cm under surface
Sample 10	Maichen Town, Xuwen County	Rhizosphere soil of <i>Avicennia marina</i>	110.0107670	20.4205670	5-10 cm under surface
Sample 11	Techeng Island, Xiashan District	Rhizosphere soil of <i>Avicennia marina</i>	110.4404500	21.1553670	5-10 cm under surface
Sample 12	Techeng Island, Xiashan District	Rhizosphere soil of <i>Avicennia marina</i>	110.4403830	21.1554000	5-10 cm under surface
Sample 13	Techeng Island, Xiashan District	Rhizosphere soil of <i>Avicennia marina</i>	110.4413500	21.1574500	5-10 cm under surface

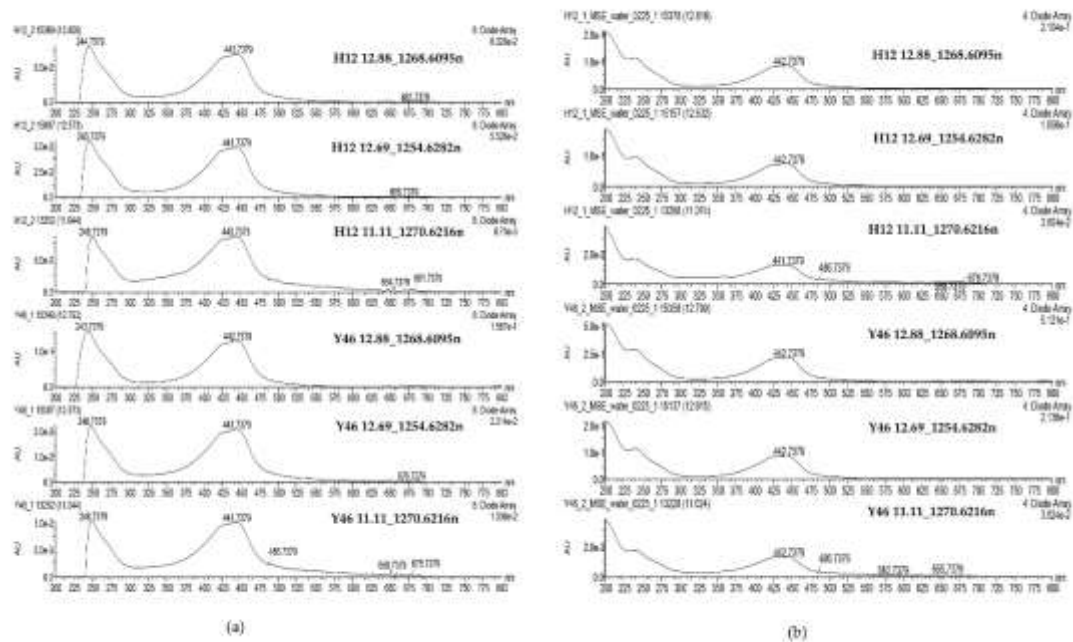


Figure S1. The UV spectra of three outliers (1-3) of samples Y46 and H12. (a) LC condition: ACN and water containing 0.1% HCOOH; (b) LC condition: ACN and water

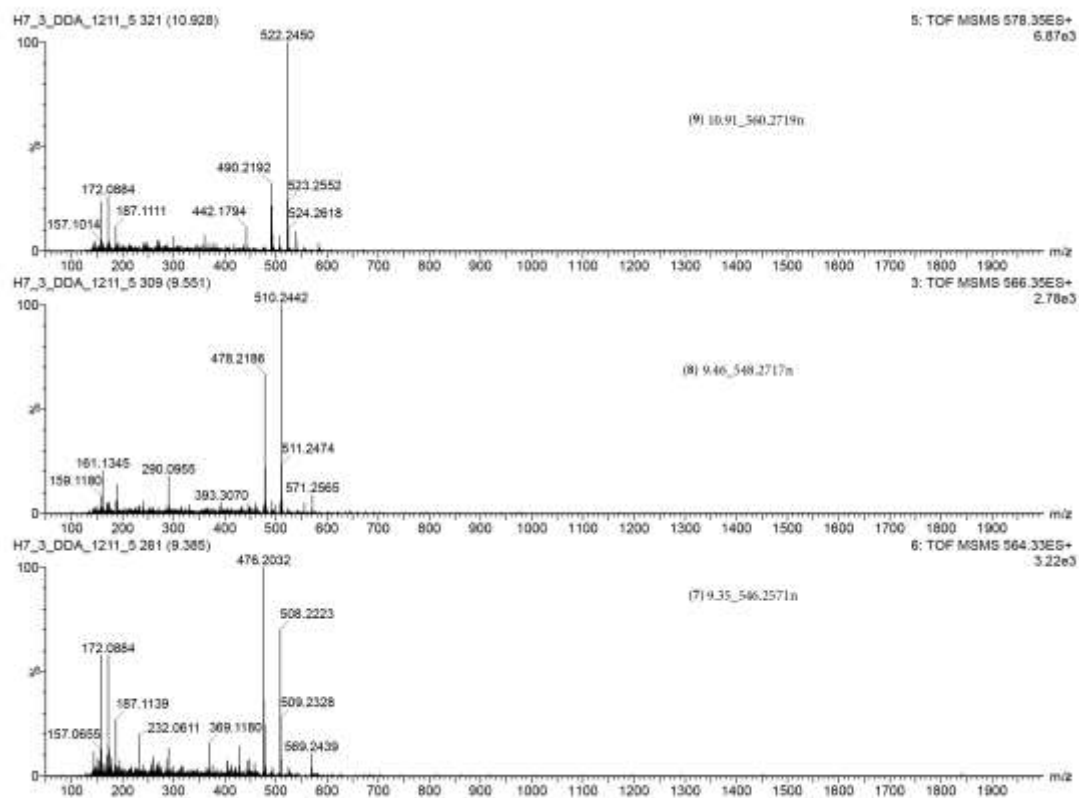


Figure S5. The MS/MS spectra of three revised compounds (7-9) in H7 acquired by DDA method.

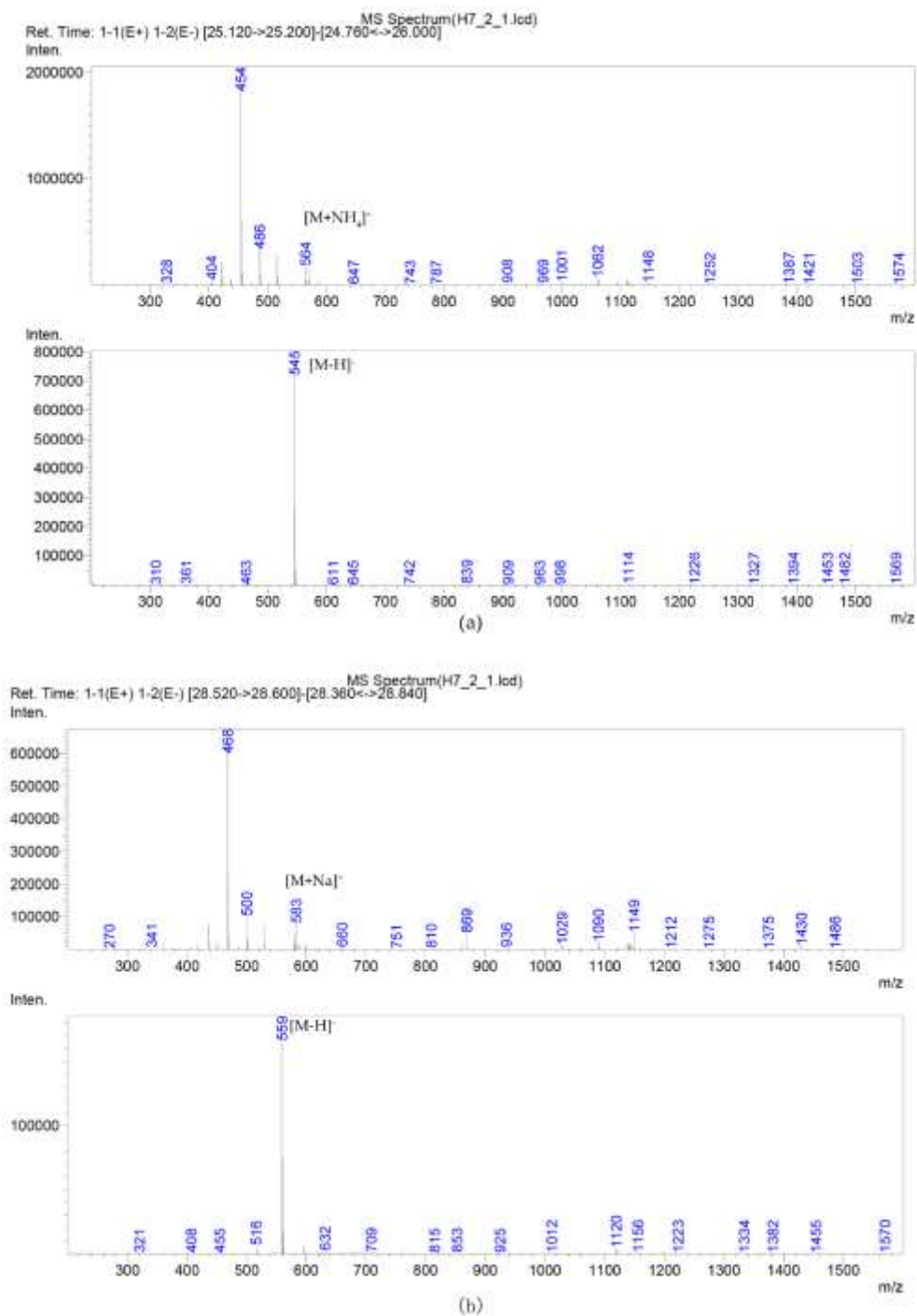


Figure S6. The positive and negative MS spectra of two revised compounds in the LC-MS of H7 (a: 9.35_546.2571n (7); b: 10.91_560.2719n (9)).

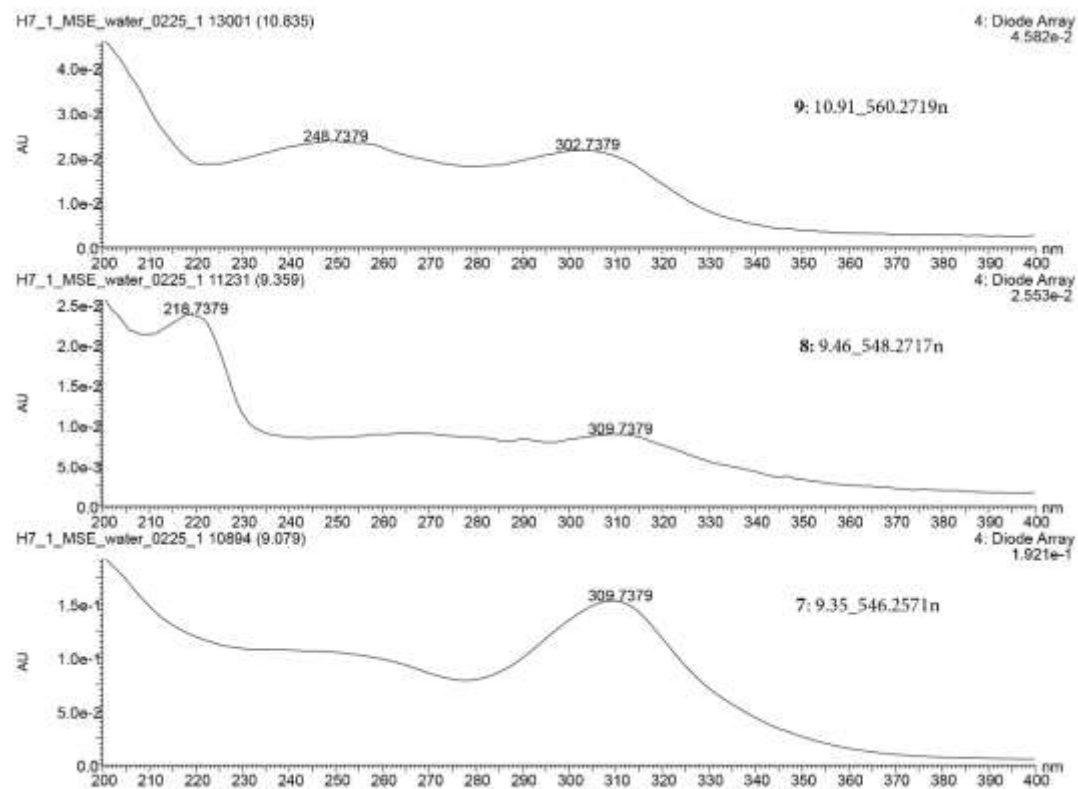


Figure S7. The UV spectra of three compounds 7-9 of the H7 eluting with ACN-H₂O (7: 9.35_546.2571n; 8: 9.46_548.2717n; 9: 10.91_560.2719n).

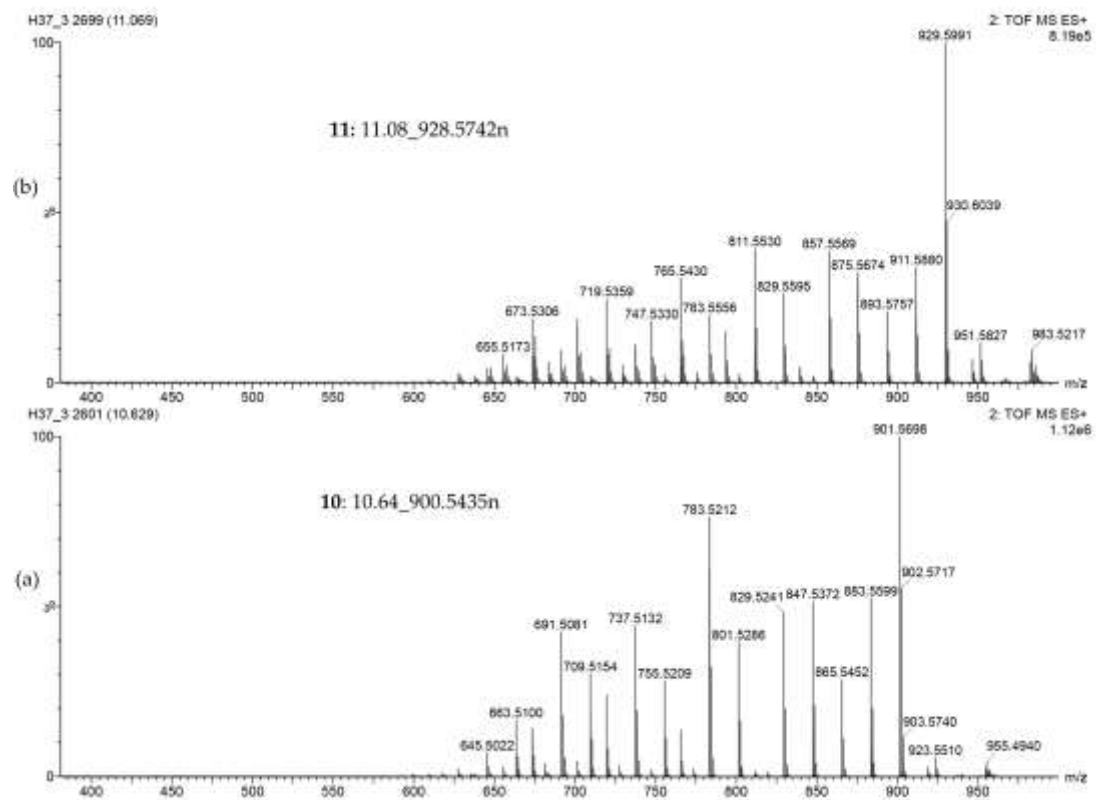


Figure S8. The MS/MS spectra of two compounds in H37 acquired by MSE method (a, 10.64_900.5435n (10); b, 11.08_928.5742n (11)).

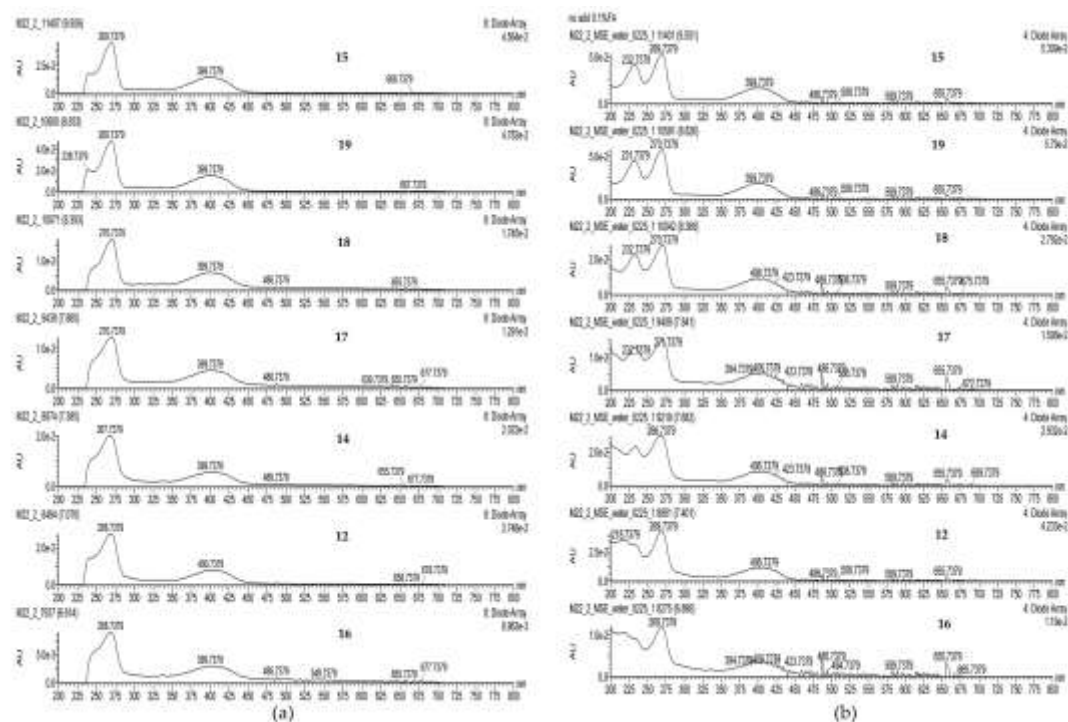


Figure S9. The UV spectra of seven trioxacarcin-type compounds in the UPLC-UV-HRMS chromatograms of M22. (a, LC conditions: ACN and H₂O containing with 0.1% HCOOH; b, LC conditions: ACN and H₂O; **12**, 7.16_1028.3600m/z; **14**, 7.47_1028.3592m/z; **15**, 9.55_876.2958n, trioxacarcin A; **16**, 6.69_1030.3751m/z; **17**, 7.94_1013.3486m/z; **18**, 8.43_894.3132n, trioxacarcin B; **19**, 8.89_878.3168n, trioxacarcin C).

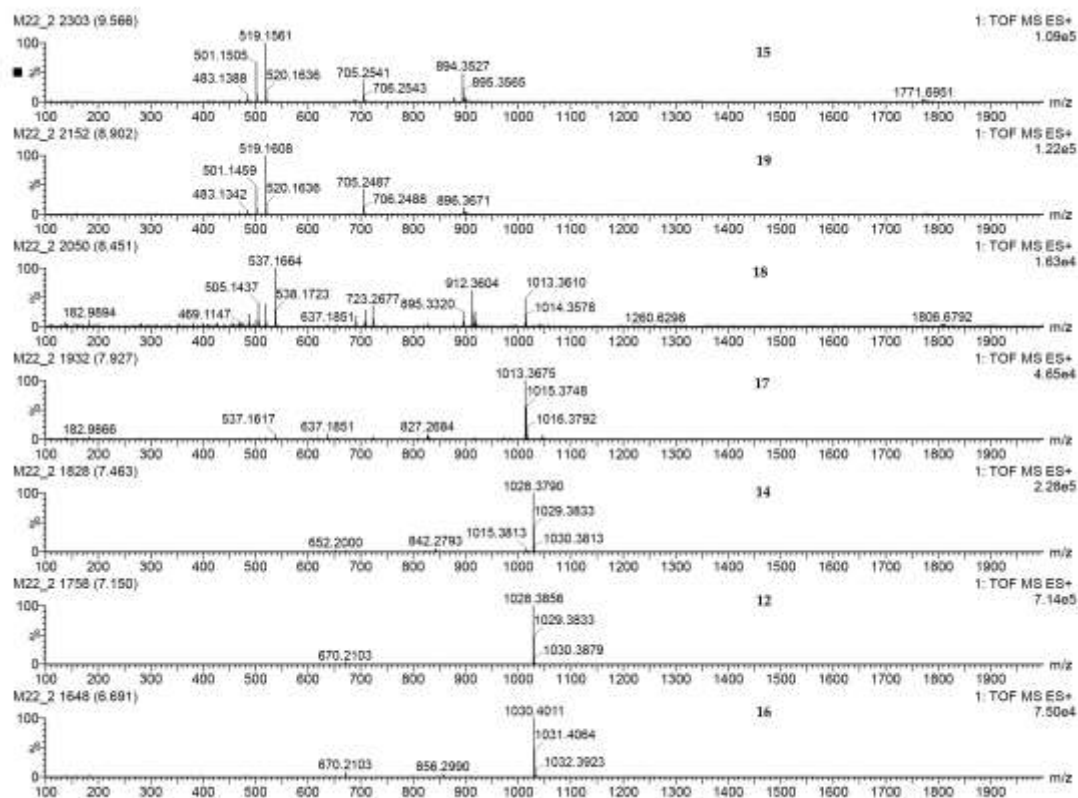


Figure S10. The MS spectra of seven trioxacarcin-type compounds in the UPLC-UV-HRMS chromatograms of M22. (**12**, 7.16_1028.3600 m/z ; **14**, 7.47_1028.3592 m/z ; **15**, 9.55_876.2958n, trioxacarcin A; **16**, 6.69_1030.3751 m/z ; **17**, 7.94_1013.3486 m/z ; **18**, 8.43_894.3132n, trioxacarcin B; **19**, 8.89_878.3168n, trioxacarcin C).

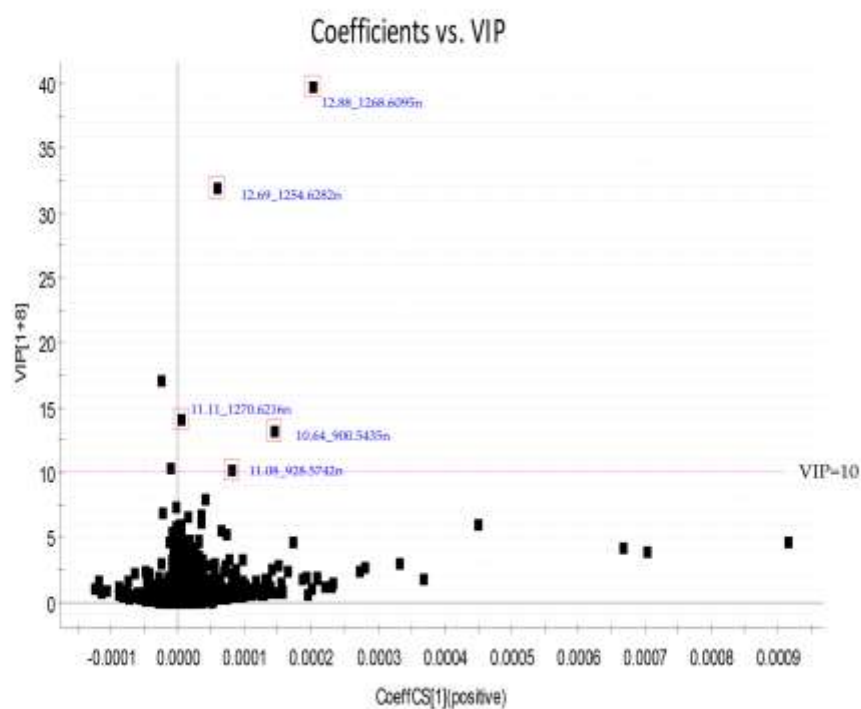


Figure S11. The VIP score of selected markers in the OPLS-DA model.

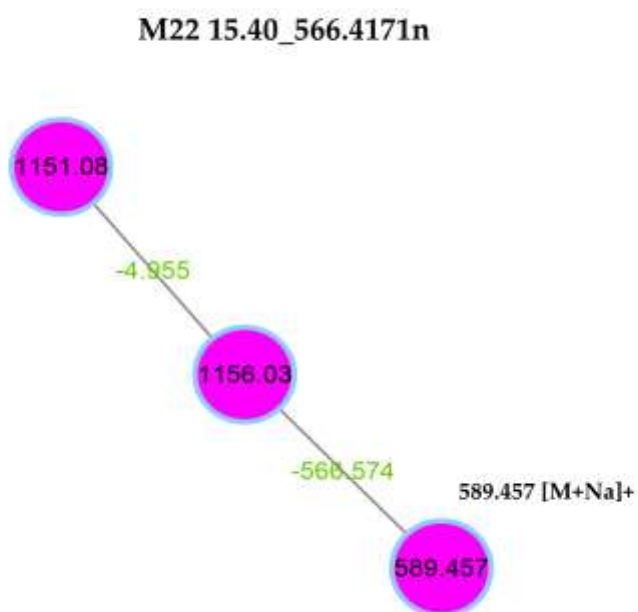


Figure S12. Molecular network of the cluster containing the compound 15.40_566.4171n (**13**) in M22 extract.

H37 10.64_900.5435n and 11.08_928.5742n

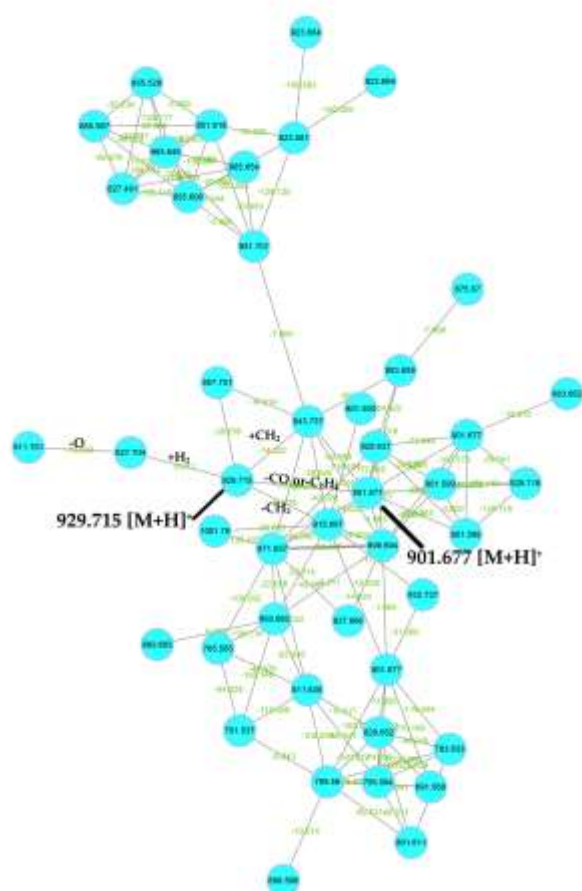


Figure S13. Molecular network of the cluster containing the compounds 10.64_900.5435n (**10**) and 11.08_928.5742n (**11**) in H37 extract.

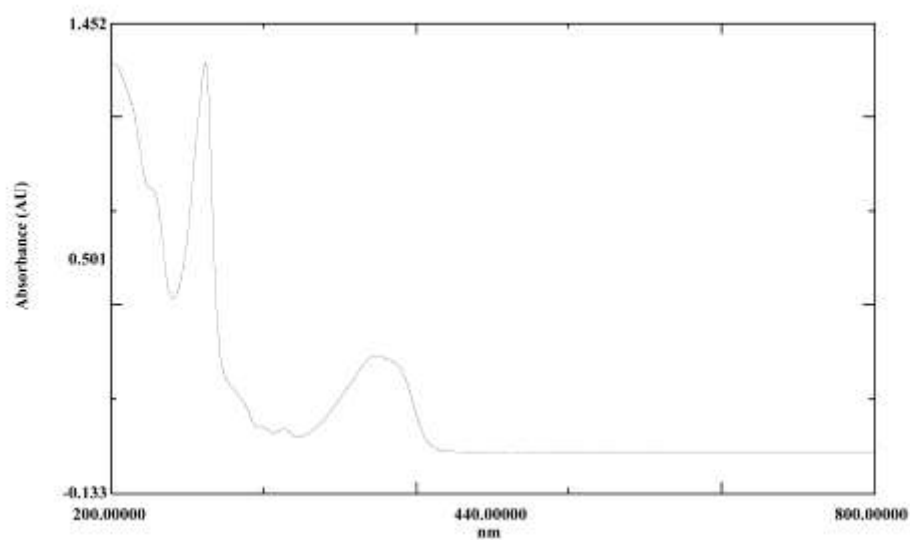


Figure S14. The UV spectrum of compound **16**

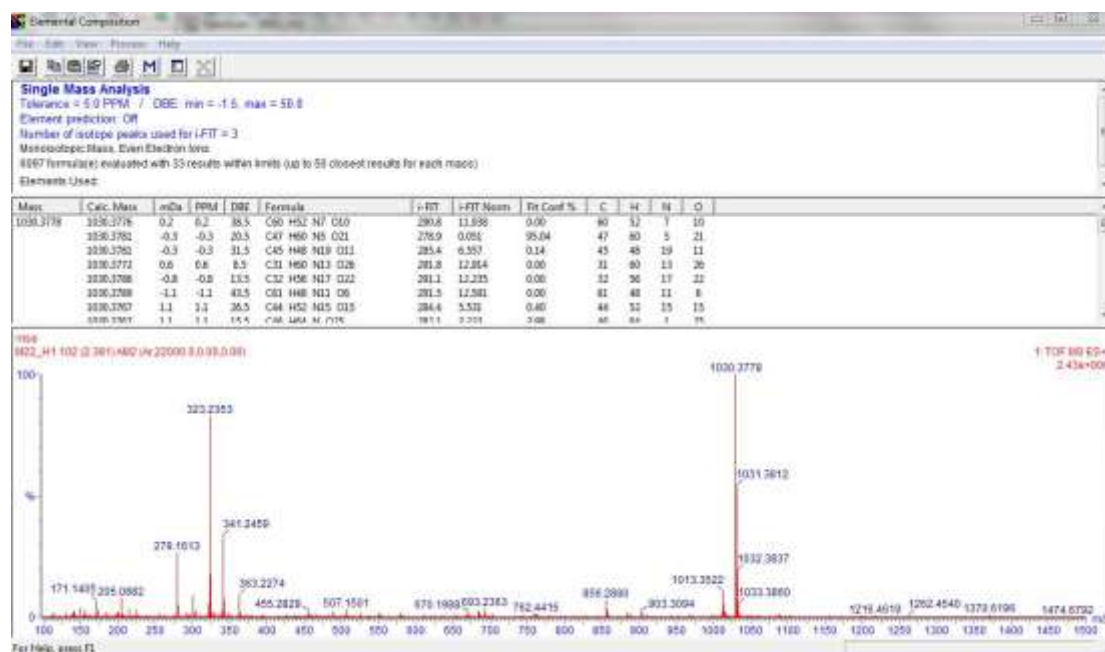


Figure S15. The HRESIMS of compound 16

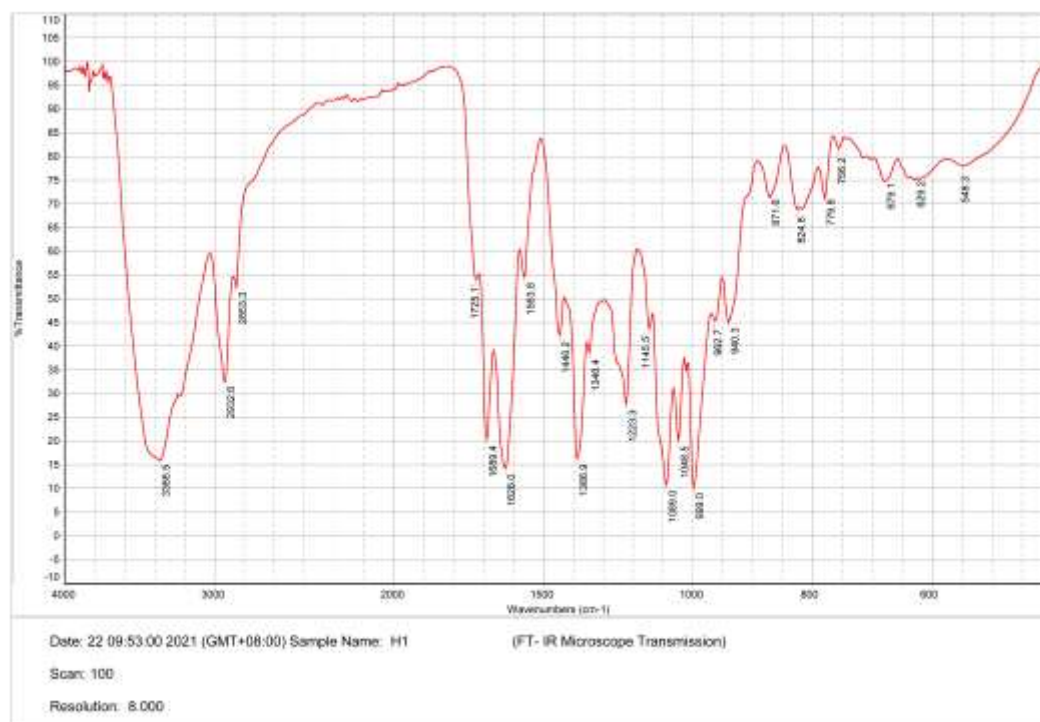


Figure S16. The IR spectrum of compound 16

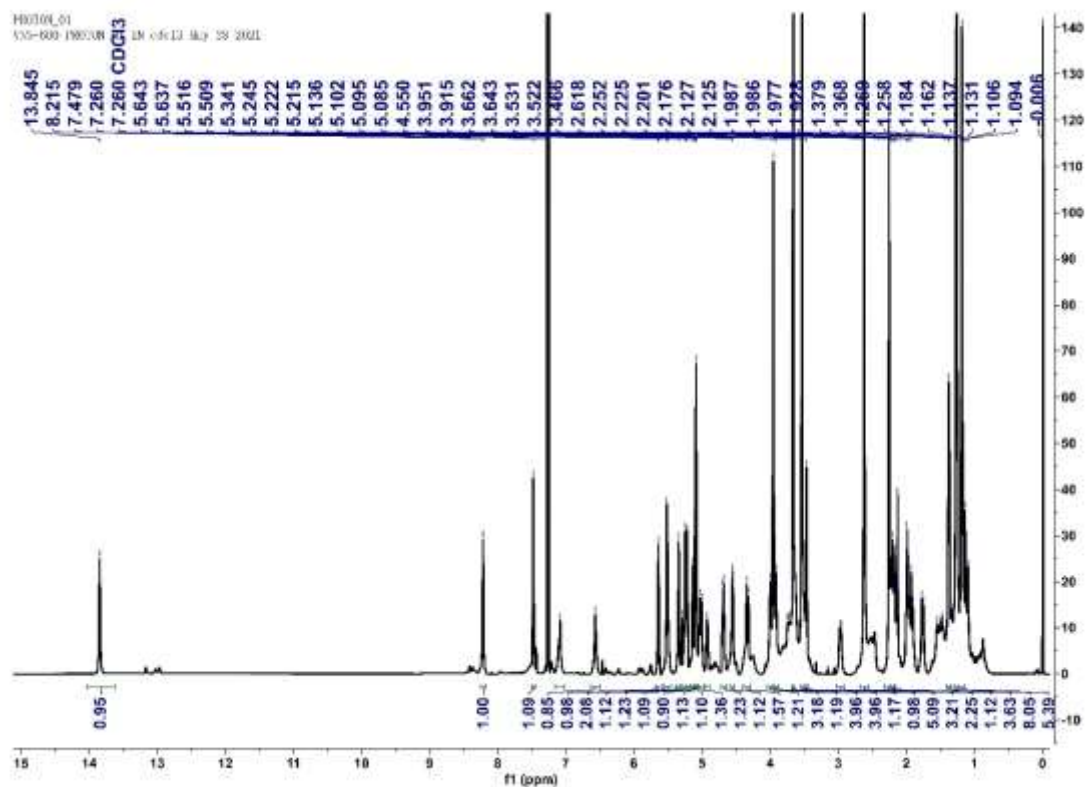


Figure S17. The ¹H NMR (600 MHz) spectrum of compound **16** in CDCl₃

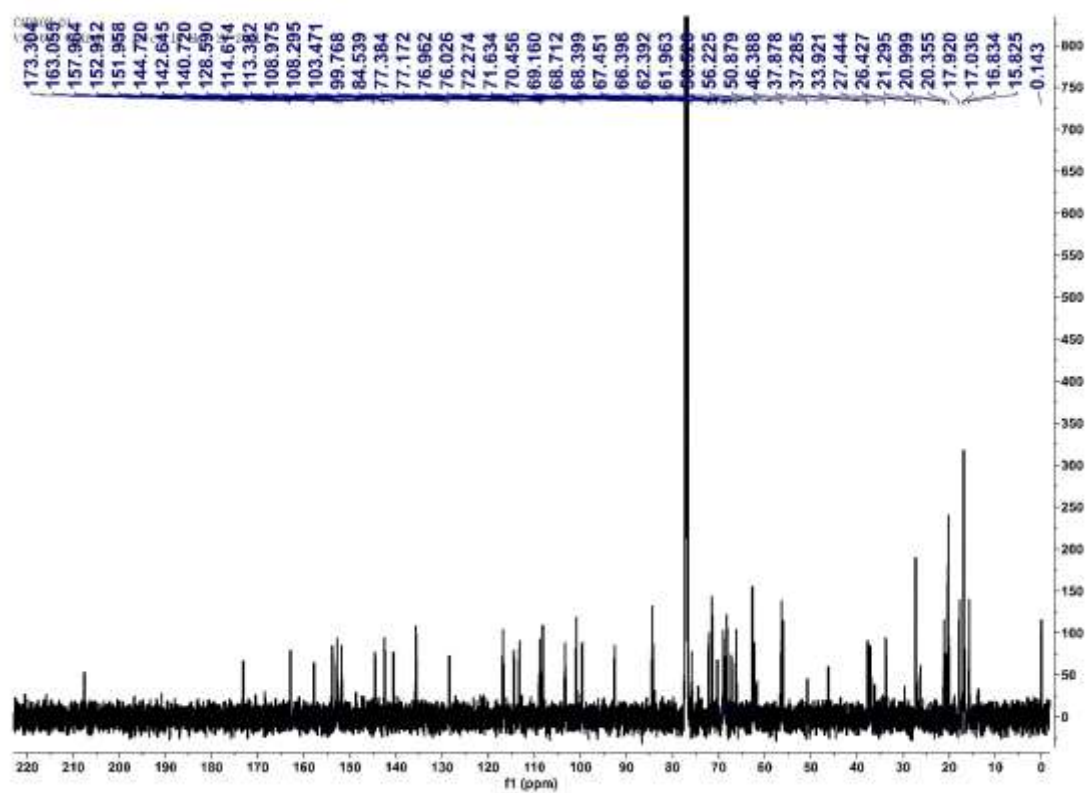


Figure S18. The ¹³C NMR (150 MHz) spectrum of compound **16** in CDCl₃

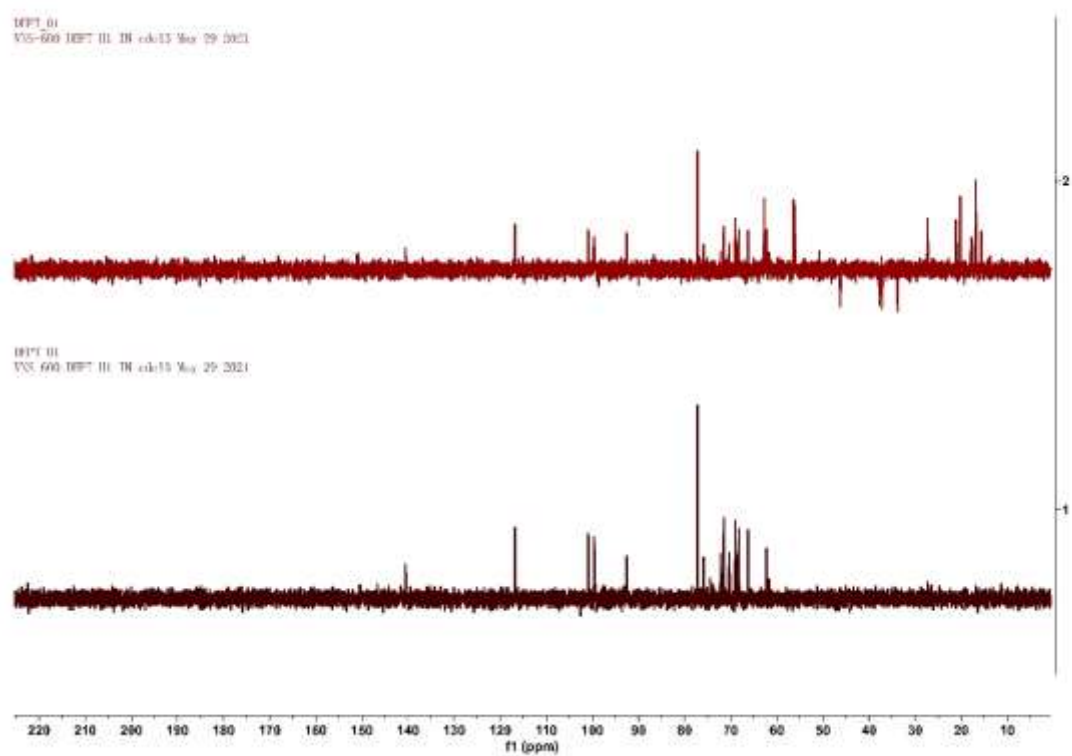


Figure S19. The DEPT spectrum of compound 16

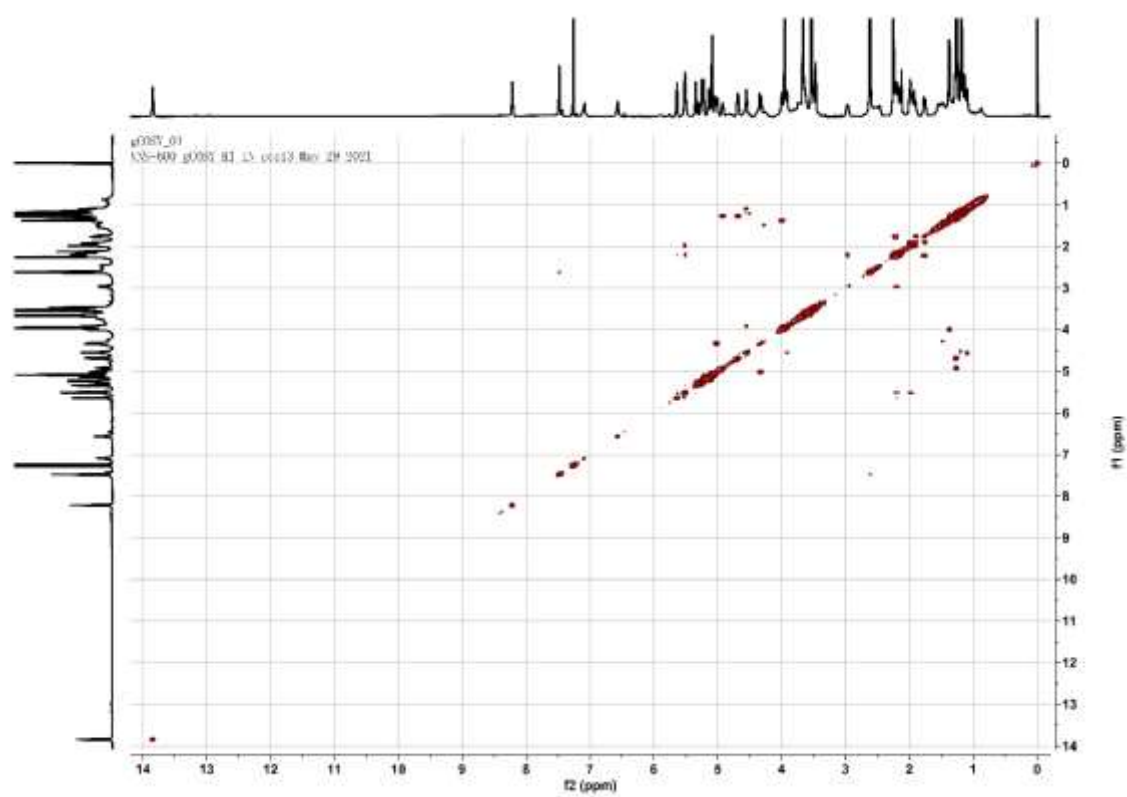


Figure S20. The ^1H - ^1H COSY spectrum of compound 16

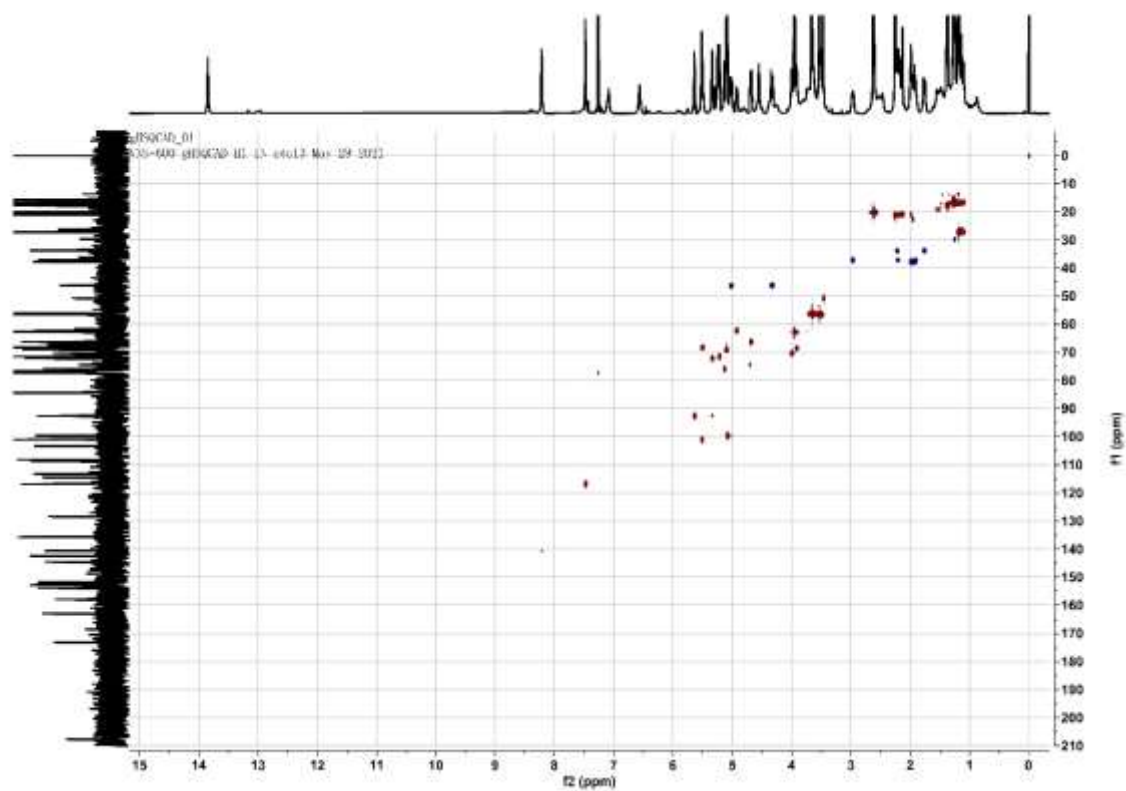


Figure S21. The HSQC spectrum of compound 16

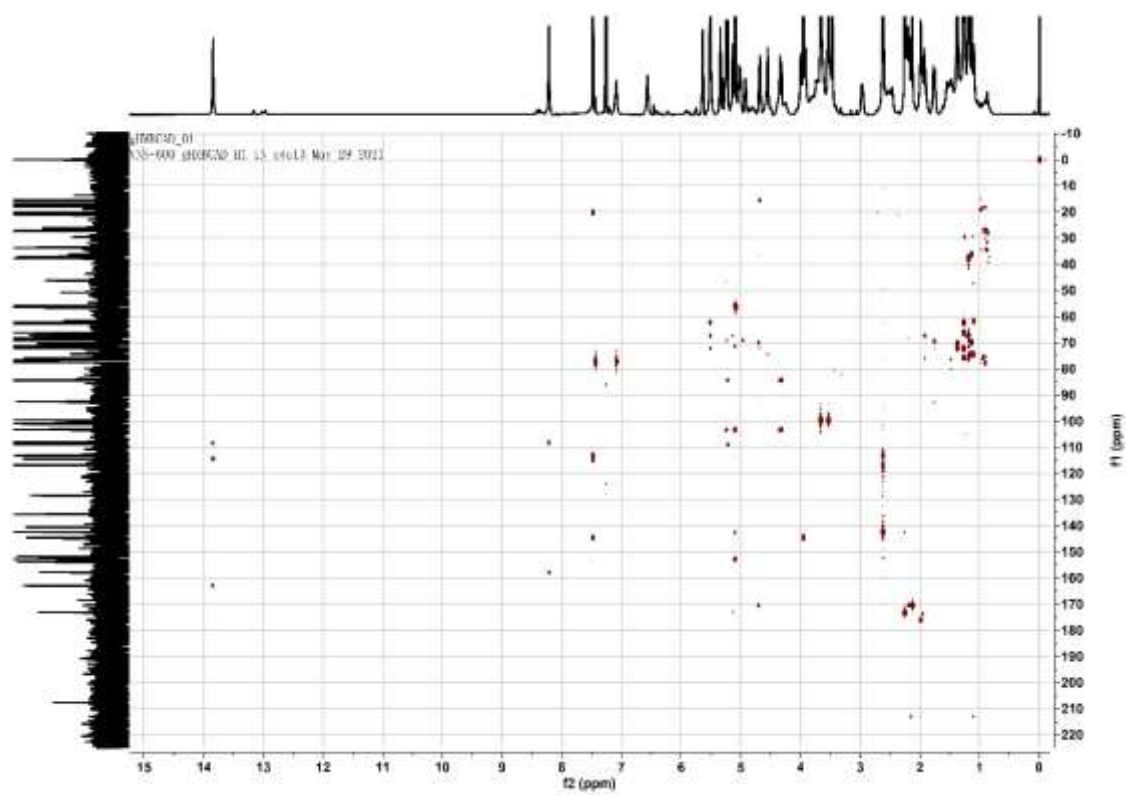


Figure S22. The HMBC spectrum of compound 16

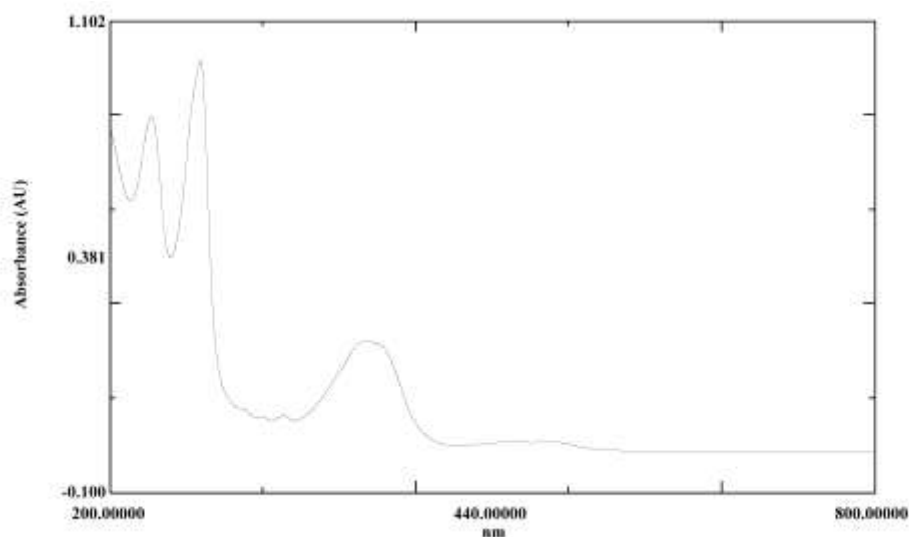


Figure S23. The UV spectrum of compound 20

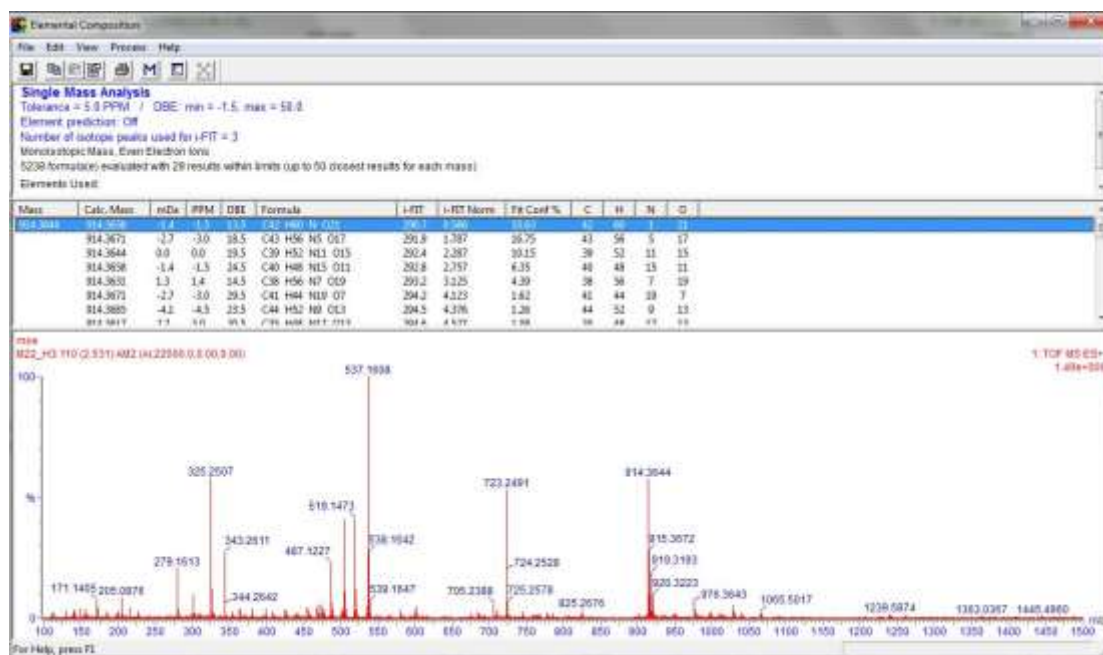


Figure S24. The HRESIMS of compound 20

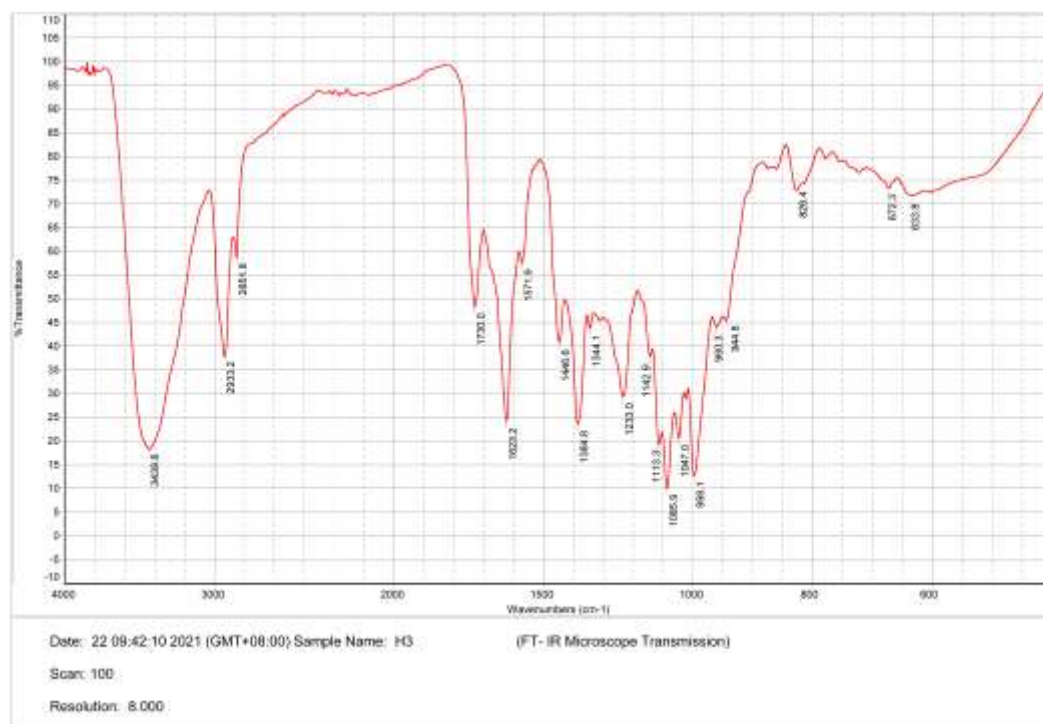


Figure S25. The IR spectrum of compound 20

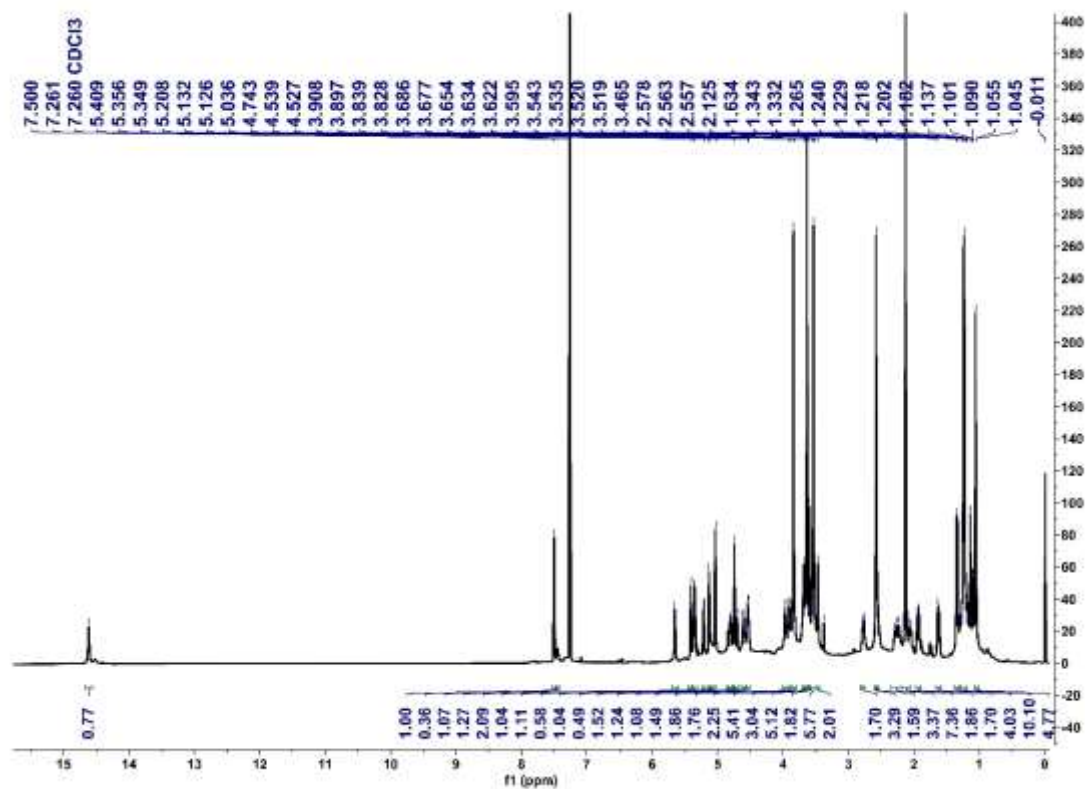


Figure S26. The ^1H NMR (600 MHz) spectrum of compound 20 in CDCl_3

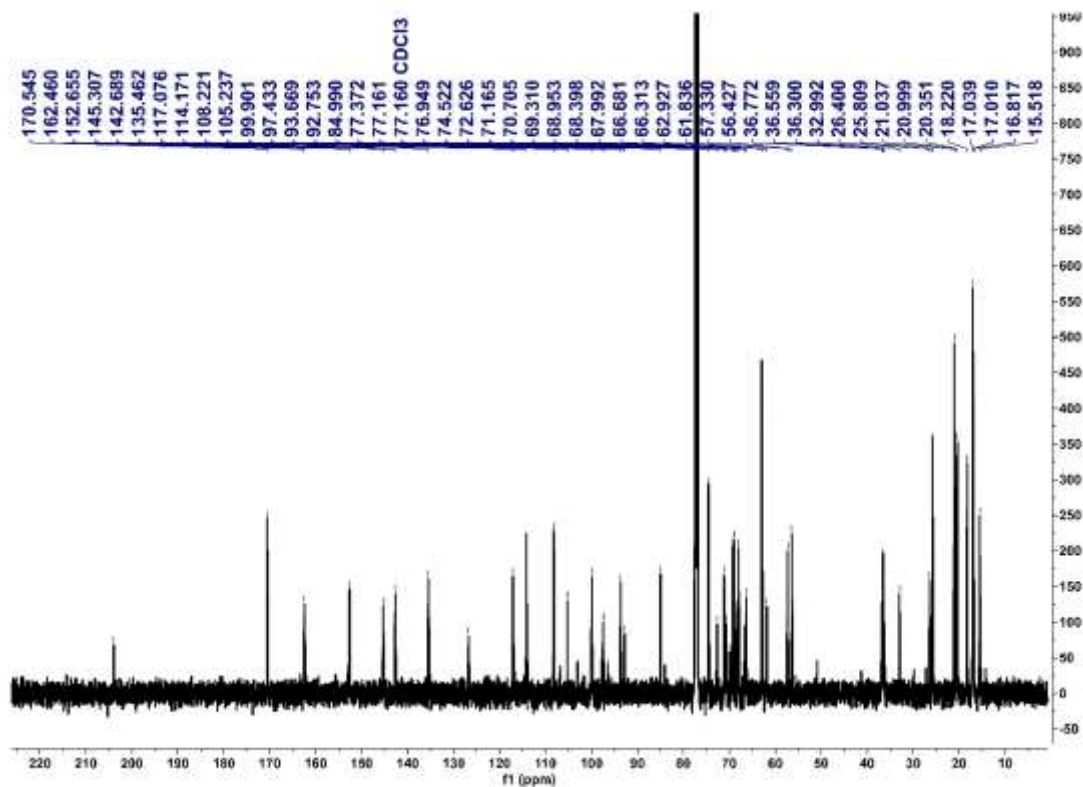


Figure S27. The ^{13}C NMR (150 MHz) spectrum of compound **20** in CDCl_3

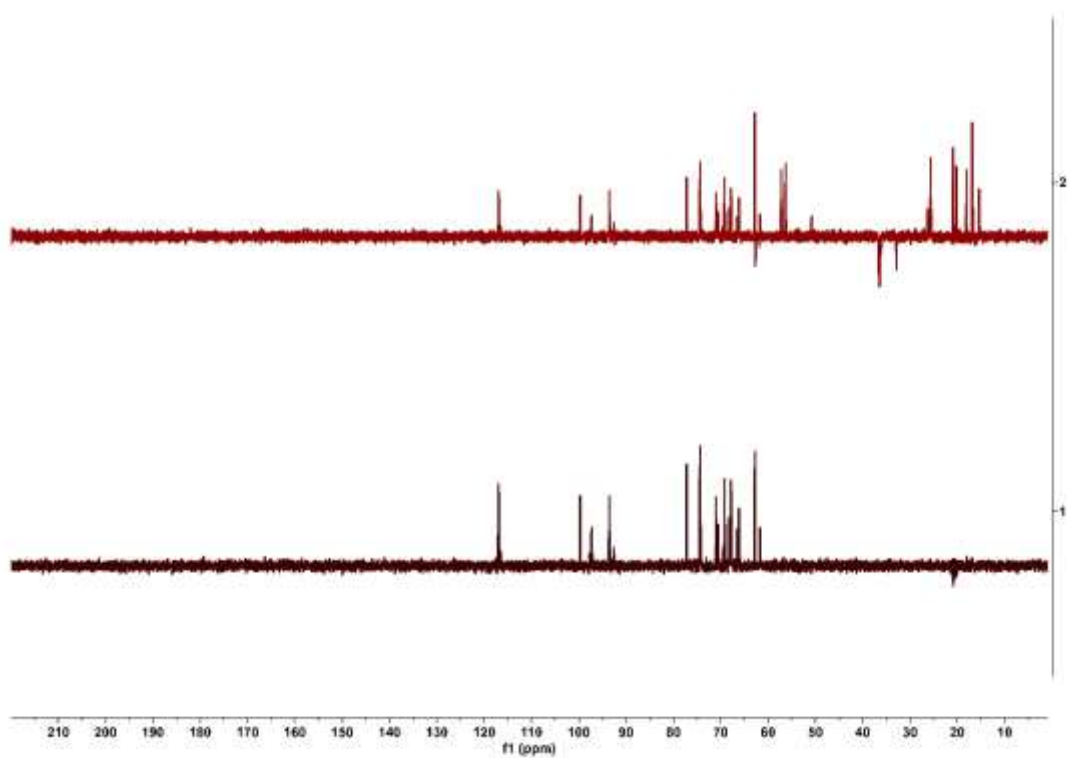


Figure S28. The DEPT spectrum of compound **20**

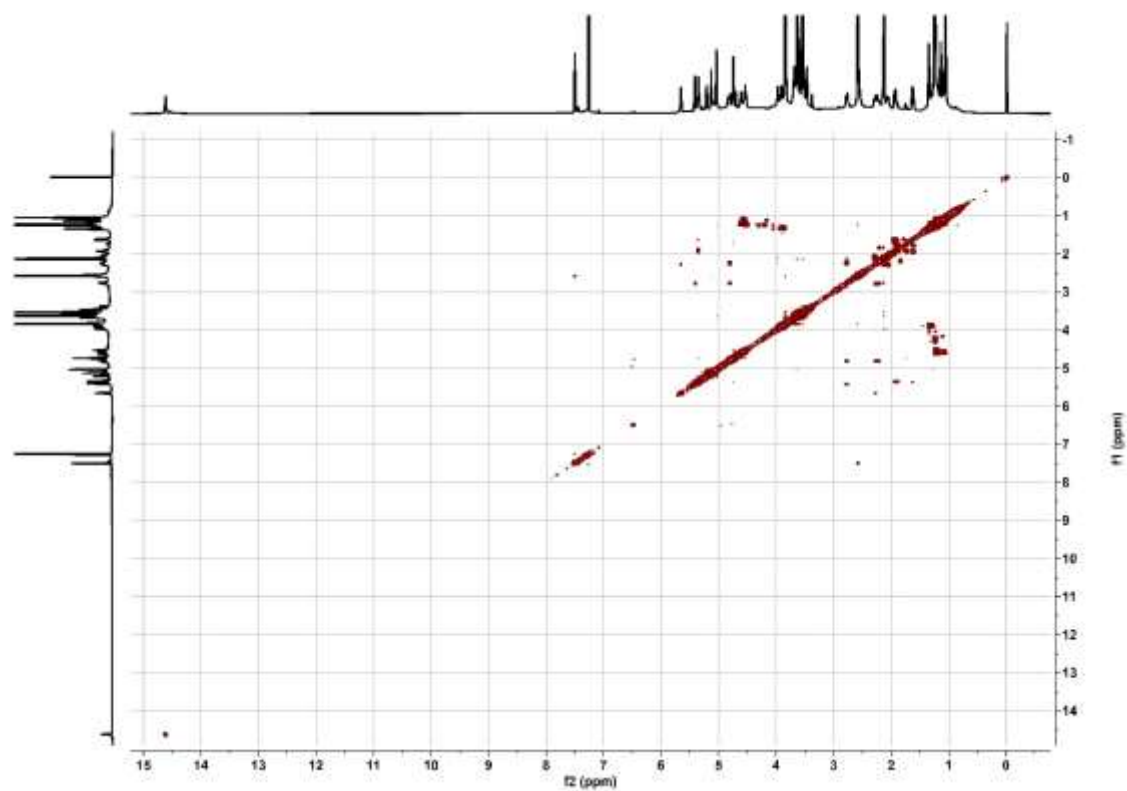


Figure S29. The ^1H - ^1H COSY spectrum of compound 20

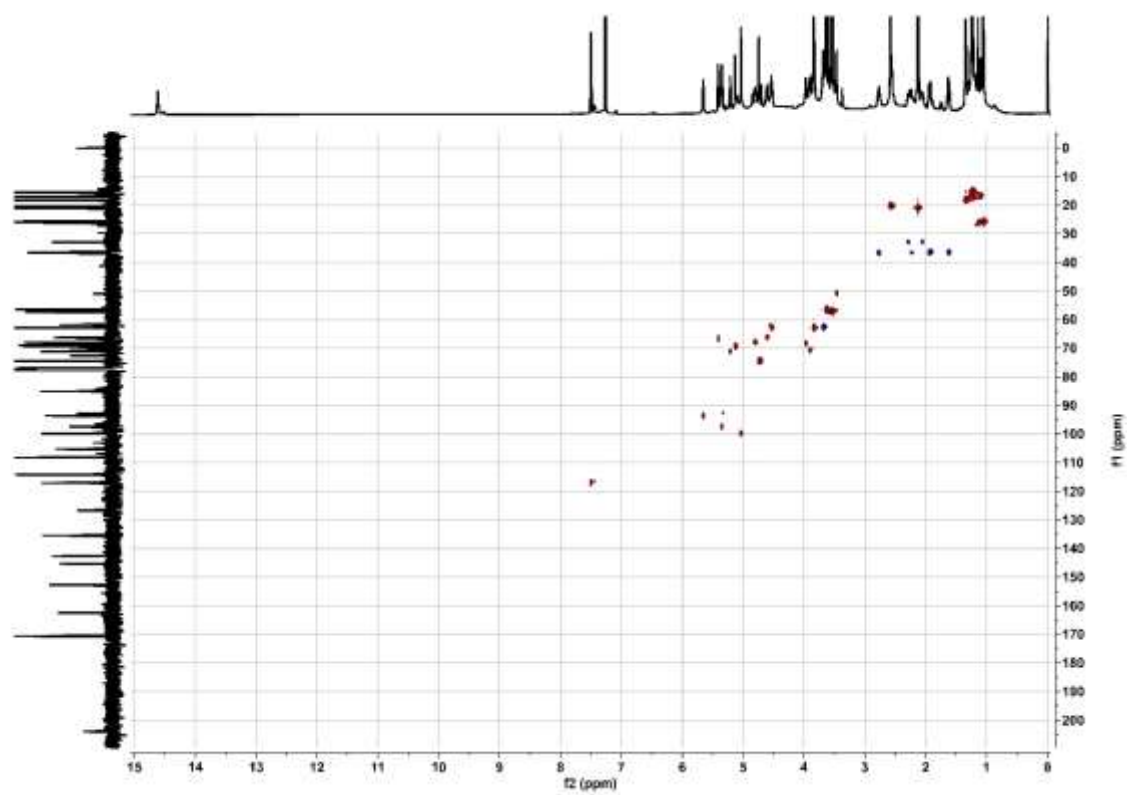


Figure S30. The HSQC spectrum of compound 20

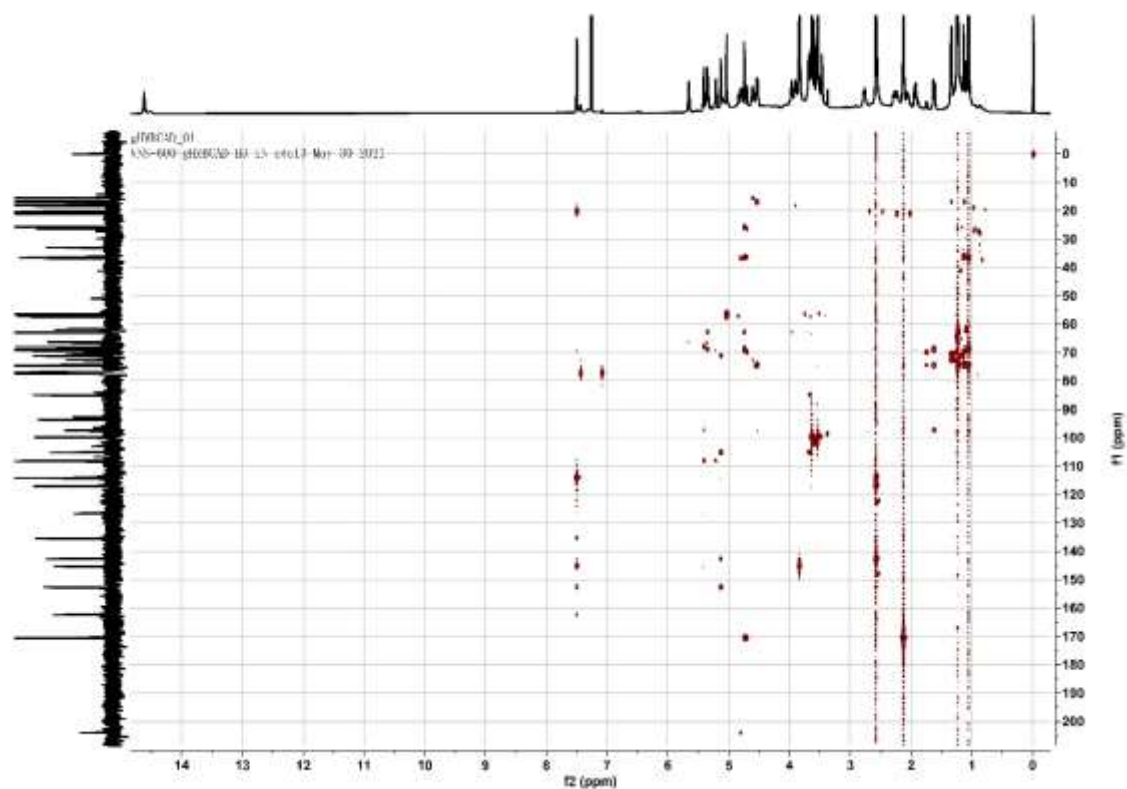


Figure S31. The HMBC spectrum of compound 20

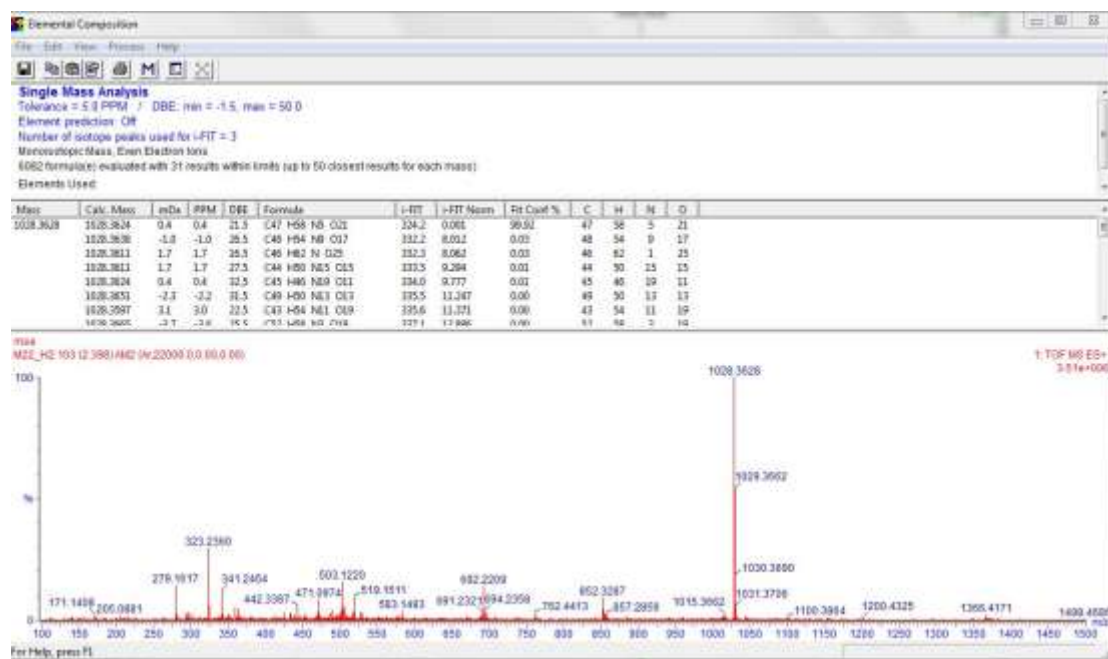


Figure S32. The HRESIMS of Compound 12

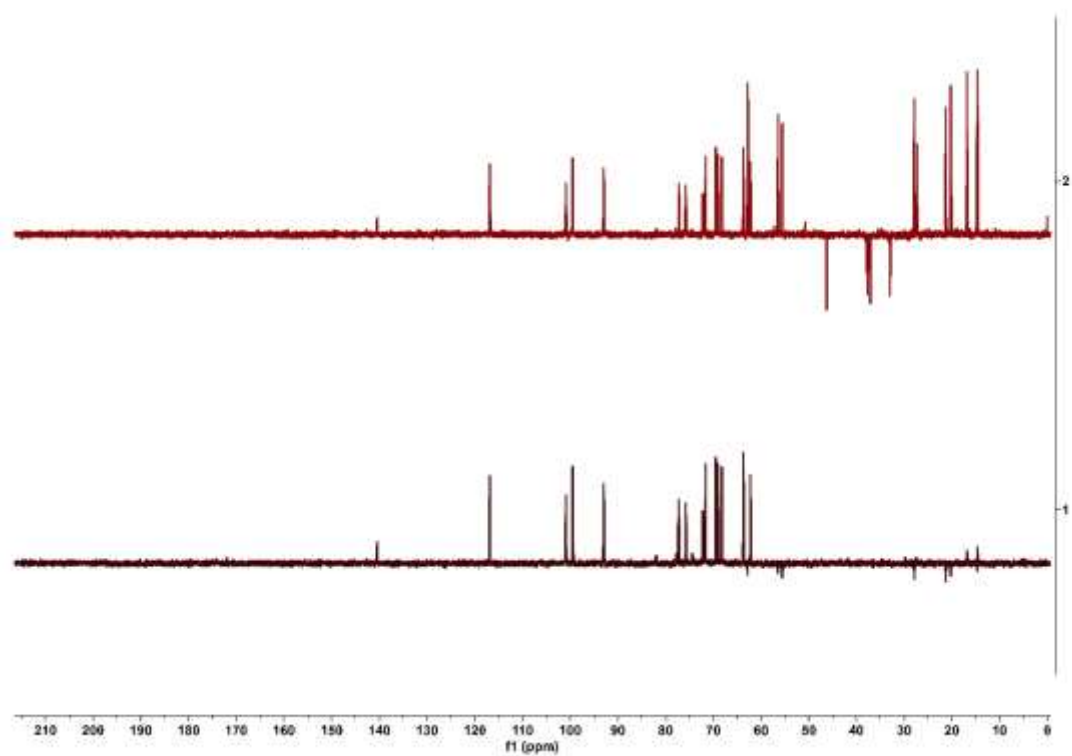


Figure S35. The DEPT spectrum of compound 12

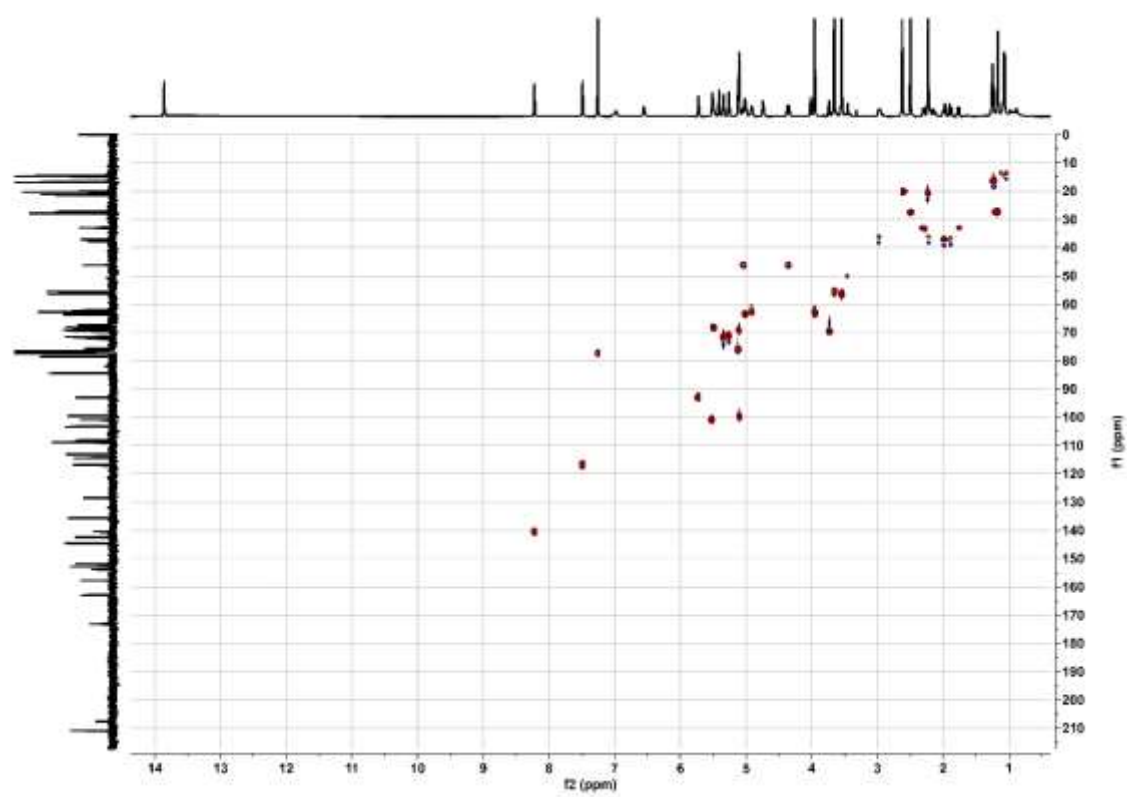


Figure S36. The HSQC spectrum of compound 12

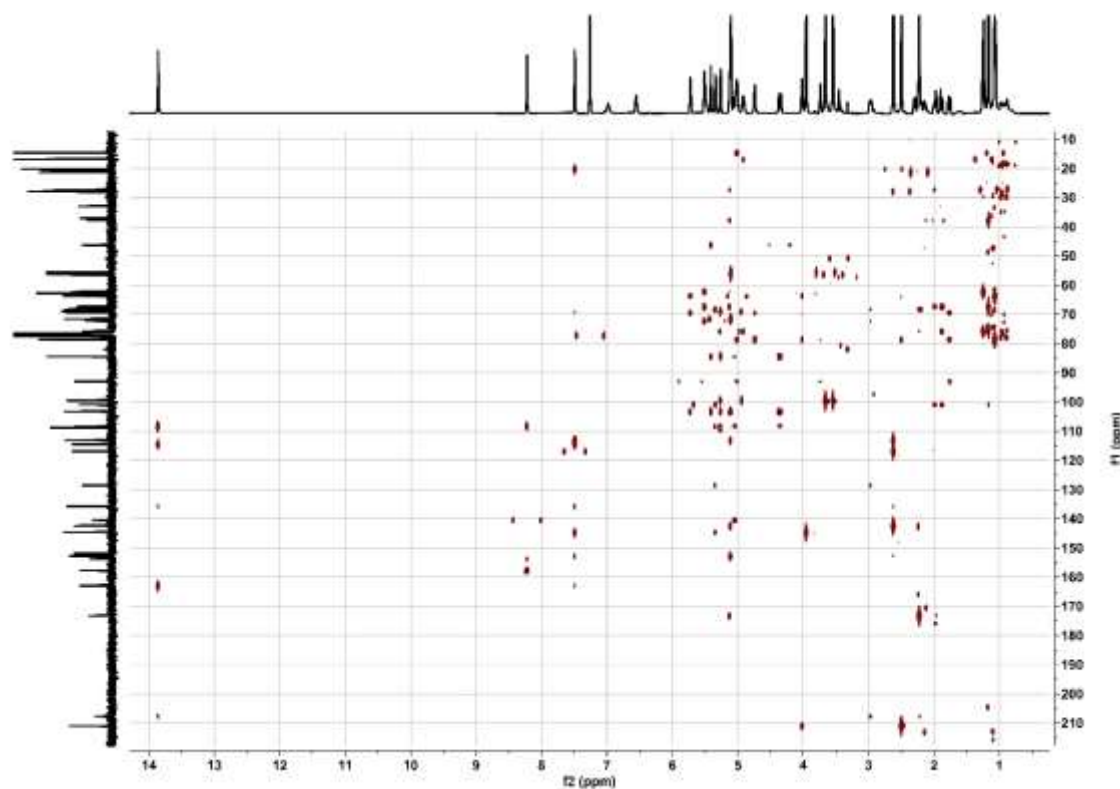


Figure S37. The HMBC spectrum of compound **12**

References

1. Lee, M.D.; Fantini, A.A.; Labeda, D.P.; Maiese, W.M.; Testa, R.T.; Borders, D.B. Antitumor agents LL-D49194 α 1, LL-D49194 β 1, LL-D49194 β 2, LL-D49194 β 3, LL-D49194 γ , LL-D49194 δ , LL-D49194 ϵ , LL-D49194 ξ , LL-D49194 η , LL-D49194 ω 1, LL-D49194 ω 2, and LL-D49194 ω 3. Google Patents: 1986.
2. Maiese, W.M.; Labeda, D.P.; Korshalla, J.; Kuck, N.; Fantini, A.A.; Wildey, M.J.; Thomas, J.; Greenstein, M. LL-D49194 antibiotics, a novel family of antitumor agents: taxonomy, fermentation and biological properties. *J Antibiot (Tokyo)* **1990**, *43*, 253-258.
3. Maskey, R.P.; Helmke, E.; Fiebig, H.H.; Laatsch, H. Parimycin: isolation and structure elucidation of a novel cytotoxic 2,3-dihydroquinizarin analogue of gamma-indomycinone from a Marine *Streptomyces* isolate. *J Antibiot (Tokyo)* **2002**, *55*, 1031-1035.
4. Tamaoki, T.; Shirahata, K.; Iida, T.; Tomita, F. Trioxacarcins, novel antitumor antibiotics. II. Isolation, physico-chemical properties and mode of action. *J Antibiot (Tokyo)* **1981**, *34*, 1525-1530.
5. Maskey, R.P.; Helmke, E.; Kayser, O.; Fiebig, H.H.; Maier, A.; Busche, A.; Laatsch, H. Anti-cancer and antibacterial trioxacarcins with high anti-malaria activity from a marine *Streptomyces* and their absolute stereochemistry. *J Antibiot (Tokyo)* **2004**, *57*, 771-779.
6. Maskey, R.P.; Sevvana, M.; Usón, I.; Helmke, E.; Laatsch, H. Gutingimycin: a highly complex metabolite from a marine *Streptomyces*. *Angew. Chem. Int. Ed. Engl.* **2004**, *43*, 1281-1283.

7. Shirahata, K.; Iida, T. Compounds having antibiotic activity, processes for their preparation, pharmaceutical compositions containing them and their use as medicaments. Google Patents: 1984.
8. Nicolaou, K.C.; Cai, Q.; Sun, H.; Qin, B.; Zhu, S. Total Synthesis of Trioxacarcins DC-45-A1, A, D, C, and C7"-epi-C and Full Structural Assignment of Trioxacarcin C. *J. Am. Chem. Soc.* **2016**, *138*, 3118-3124.
9. Nicolaou, K.C.; Cai, Q.; Qin, B.; Petersen, M.T.; Mikkelsen, R.J.; Heretsch, P. Total synthesis of trioxacarcin DC-45-A2. *Angew. Chem. Int. Ed. Engl.* **2015**, *54*, 3074-3078.