

**Table S1**

Polymerase read statistics

Polymerase Read Bases (G)	Polymerase Reads	Polymerase Read Length(mean)	Polymerase Read N50
28.7	583625	49169	113836

\*Polymerase reads: High-quality sequencing reads produced by a single molecule

**Table S2**

Subreads statistics

Subread base(G)	Subreads number	Average subreads length	N50
27.88	11234648	2483	2747

\*Subreads: Filtered polymerase read

**Table S3**

Classification statistics of CCS, FLNC and Consensus

Read	number	Min length	Max length	Mean length	N50
CCS	466891	57	15000	2847	3079
FLNC	392939	52	14910	2736	2975
Consensus	31073	372	11772	2770	3051

\*CCS: The consensus sequence obtained after self-alignment correction

FLNC: a non-chimeric sequence with 3', 5' primers and a poly A tail

Consensus: Sequence after cluster and remove redundancy FLNC

**Table S4**

Illumina sequence data and mapping efficiency

sample	Raw Base(G)	Clean Base(G)	mapping efficiency
CON_4hS1	7.63	7.33	90.84%
CON_4hS2	6.53	6.23	92.04%
CON_4hS3	6.66	6.41	91.89%
CON_8hS1	6.78	6.47	90.83%
CON_8hS2	6.47	6.24	91.78%
CON_8hS3	6.66	6.39	90.52%
CON_12hS1	6.57	6.33	90.91%
CON_12hS2	6.46	6.19	91.40%
CON_12hS3	6.55	6.22	91.04%
CON_24hS1	6.73	6.44	91.23%
CON_24hS2	6.46	6.08	90.08%
CON_24hS3	6.94	6.64	91.19%
CON_72hS1	7.29	7.02	90.22%
CON_72hS2	6.41	6.18	90.90%
CON_72hS3	6.6	6.31	92.05%
CON_168hS1	7.15	6.89	91.47%

CON_168hS2	7.28	7	91.57%
CON_168hS3	7.28	7.01	91.12%
TTX_4hS1	6.84	6.57	91.56%
TTX_4hS2	7.07	6.67	88.31%
TTX_4hS3	6.83	6.6	91.06%
TTX_8hS1	7.23	6.85	91.88%
TTX_8hS2	6.31	6.03	91.11%
TTX_8hS3	6.49	6.25	91.61%
TTX_12hS1	6.6	6.16	91.50%
TTX_12hS2	8.2	7.99	90.38%
TTX_12hS3	6.28	6.06	91.98%
TTX_24hS1	6.53	6.33	90.42%
TTX_24hS2	7.04	6.8	90.07%
TTX_24hS3	6.27	6.17	90.44%
TTX_72hS1	6.51	6.3	91.38%
TTX_72hS2	6.83	6.52	91.07%
TTX_72hS3	6.67	6.4	91.79%
TTX_168hS1	6.41	6.09	90.84%
TTX_168hS2	6.73	6.49	91.16%
TTX_168hS3	6.62	6.37	91.48%

**Table S5**

Consensus sequence length statistics before and after transcript correction

Type	Total nucleotides	Total number	Mean length	Min length	Max length	N50	N90
Before correct	86046645	31073	2770	372	11772	3051	1820
After correct	86036010	31073	2769	372	11774	3050	1820

**Table S6**

Polished consensus sequence map to the reference genome

Total reads	Total mapped	Unmapped	Multiple mapped	Uniquely mapped	Reads map to “+”	Reads map to “-”
31073	29502 (91.73%)	2671 (8.27%)	67 (0.22%)	28435 (91.51%)	14418 (46.4%)	14017 (45.11%)

**Table S7**

Number of peptides and proteins identified

MS/MS spectrum database search analysis summary				
Total spectra	Matched spectrum	Peptide	Identified proteins	Quantifiable proteins
341245	46001	25204	4368	4358

**Table S8**

Number of differentially expressed protein

Differentially expressed protein summary		
	regulated type	fold change>1.5
TTX/CON	up-regulated	50
	down-regulated	150

**Table S9**

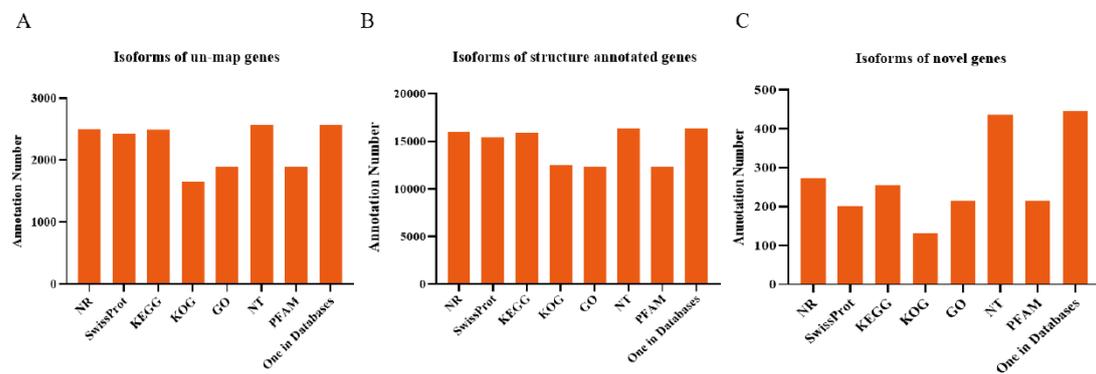
The primer sequence of differential expressed genes

Gene Name	Primer Sequence
EF1 $\alpha$	ACTGAGGTGAAGTCTGTGGAAA TTTGGTGGGTTCGTTCTTGCTGTC
FOS	TGCGACTCTTCTTCCCGTTG CTTGATGCTGTCAGGTCGGT
Satb2	CCAGCGCTACAACGTCAAAC GCAACGCTGCACATGATTCT
IER2	ATCCTGGCAGTGTGATCAG CGGGCACTTCGCATAACCAT
LMAN1	TGATGGCGAAGAACGCAGAG CACAGGCGGGTTTGATCCAT
HOOK3	GGAGGAACTTCGAAAGCCA AGAGTCTCGTTCCTCA
EGR1	AGGCTCACGTCCAAAATCA TGCGGATGTGAGTGGTAAGG
AHNAK	ATCACTCGCCGATCACTAC CTCCAGACTCATGCCGTAGC
MYL4	ATGTGGAGACGTGATGCGAG TGAGCCCCTCCACAAAATCC
CKM	TCCTGACTTGTGCCGGTATG GAATCGCCTGAAGACCTCCC
TNNI2	TGCTGACGCCATGTTGAAAG CCTCCACATTCTTACGCCAGT
TPM3	CGGGGTCCATTCAGCCTTTT TTTGTCCAGCTCGTCCTCAG
MYH7	GGAGGACCGGAAGAATGTGCG GAGATGAGTGTGGCCTGCT
CRYAB	AAGGATGGTCCCGGCTTTAC CAGCATCGGAGCGGATATGA
TCAP	AGAGCTACAACGCCGAATGG AAGTGCCCACTCTGAAGACC
TXLNB	CAGTGACCCAGTTCAGGTCC TGCTTCTTAGCGGAGGTGTC
PYGM	CCCTTGTCAGACCACGACAG TGTGGGCGAGAGCAAAGTAG

PFKM	ATATGCAACAGGGCGGAACA
	GAGTTGGCAAAGATGCGACC
KPNA2	AGAACCCCTCACCCCTTTA
	GCACCGTCCGTCAAGTATGA
ATP2A1	CAACTTCTGGCTGCTTGCTG
	GCGACAACTTCAGCACCTC
Flnc	CCGTAACTTCCTCGTCCCC
	ATGGTGATTGTCCGTCCTT

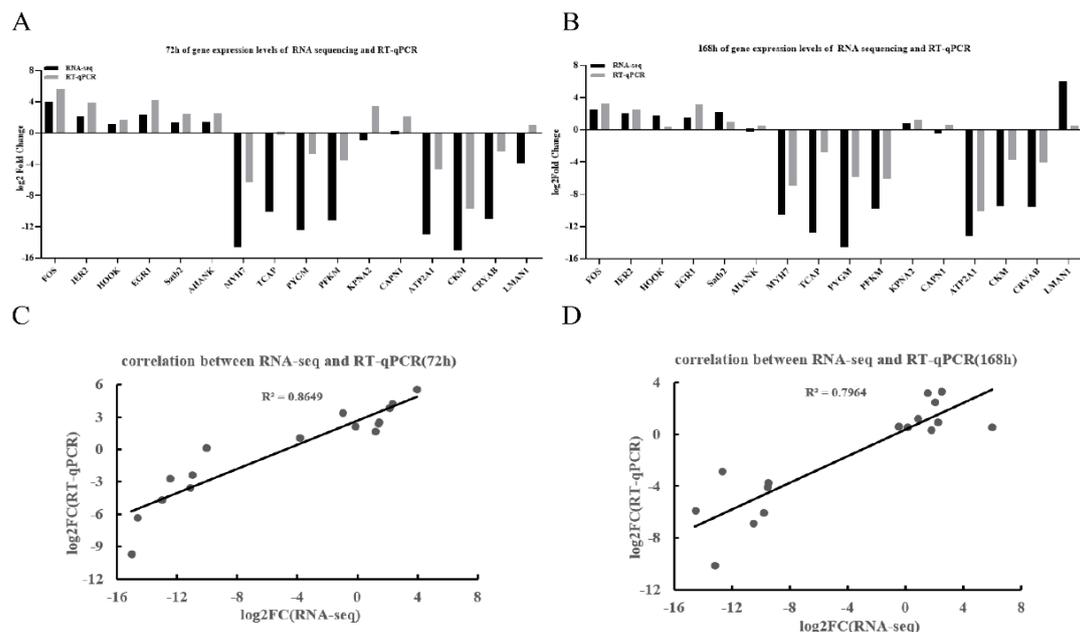
## Supplementary Figures

**Figure S1**



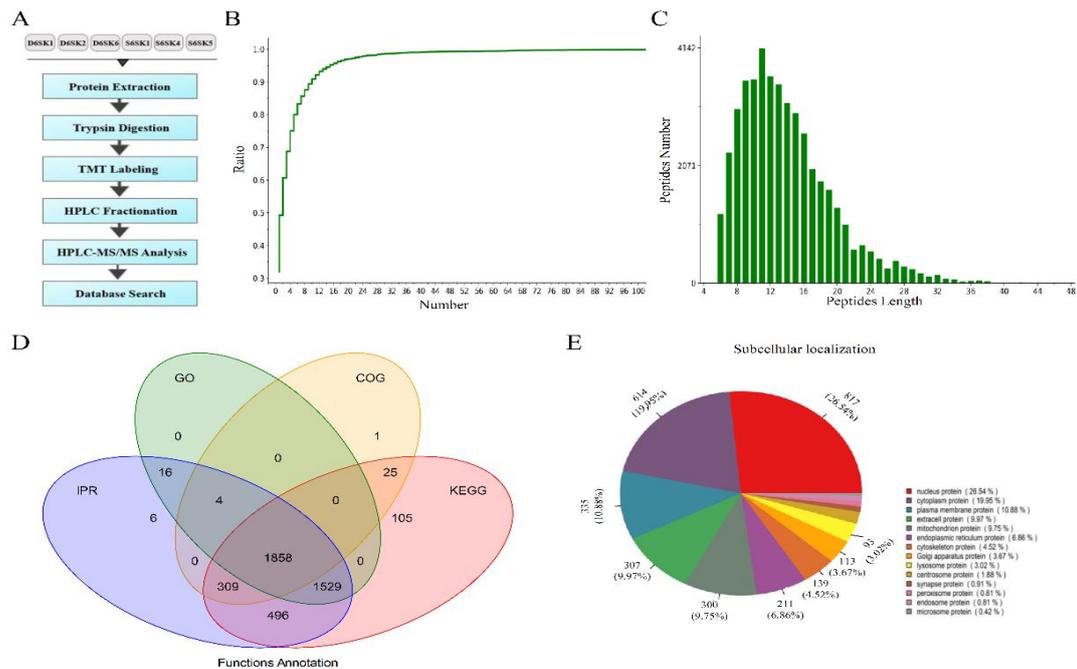
**Fig.S1** Functional annotation in seven databases (NR, NT, PFAM, KOG/COG, SwissProt, KEGG, GO). A. unmapped isoforms. B. the structurally annotated isoforms. C. the novel genes.

**Figure S2**



**Fig.S2** qPCR verify the RNA-seq result of DEGs. A(B). Comparison of gene expression levels of 16 DEGs of 72h (168h) between RNA sequencing and RT-qPCR analyses. C(D). Correlation analysis between RT-qPCR and RNA-Seq results of 16 DEGs of 72h (168h) indicating a positive correlation (linear-fitting method (R<sup>2</sup>)).

**Figure S3**



**Fig.S3** Quality control and functional annotation of identified proteins. A The procedure of the proteomic study. B. The distribution of Unique peptide number. C. Peptide length distribution identified by quantitative proteomic analysis. D. Functional annotation of the identified proteins. E. Subcellular localization of DEPs.