

Review

# Molecular Targets of Fatty Acid Ethanolamides in Asthma

Oxana Kytikova \*, Tatyana Novgorodtseva, Marina Antonyuk, Yulia Denisenko and Tatyana Gvozdenko

Vladivostok Branch of Federal State Budgetary Science Institution, Far Eastern Scientific Center of Physiology and Pathology of Respiration—Institute of Medical Climatology and Rehabilitative Treatment, Russkaya st. 73g, 690105 Vladivostok, Russia; curdeal@mail.ru (T.N.); antonyukm@mail.ru (M.A.); karaman@inbox.ru (Y.D.); vfdnz@mail.ru (T.G.)

\* Correspondence: kytikova@yandex.ru; Tel./Fax: 42-32-788201

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**Abstract:** Asthma is a common allergic pathology of the respiratory tract that requires the study of mechanisms underlying it, due to severe forms of the disease, which are refractory to therapy. The review is devoted to the search for molecular targets of fatty acid ethanolamides in asthma, in particular palmitoylethanolamide (PEA), which has been successfully used in the treatment of chronic inflammatory and neurodegenerative diseases, in the pathogenesis of which the nervous and immune systems are involved. Recently, the potentially important role of neuro-immune interactions in the development of allergic reactions has been established. Many of the clinical symptoms accompanying allergic airway inflammation are the result of the activation of neurons in the airways, so the attention of researchers is currently focused on neuro-immune interactions, which can play an important role in asthma pathophysiology. A growing number of scientific works confirm that the key molecule in the implementation of these inter-systemic interactions is nerve growth factor (NGF). In addition to its classic role in nervous system physiology, NGF is considered as an important factor associated with the pathogenesis of allergic diseases, particularly asthma, by regulating of mast cell differentiation. In this regard, NGF can be one of the targets of PEA in asthma therapy. PEA has a biological effect on the nervous system, and affects the activation and the degranulation of mast cells.

**Keywords:** fatty acid metabolism; palmitoylethanolamide; immunology; neurons; lung; inflammation; nerve growth factor

## 1. Introduction

Asthma is a widespread chronic disease of the respiratory system, and is a significant medical and social problem throughout the world [1]. Between 1% and 18% of the population of different countries suffer from asthma, and there is a tendency to increase the number of the patients [2].

As a chronic, genetically determined, and heterogeneous inflammatory disease of the respiratory tract, asthma is characterized by reversible obstruction and bronchial hyperreactivity caused by a Th2-mediated immune response to immunological, neurogenic, physical, chemical, and environmental stimuli [3]. The majority of patients have disease symptoms that can be controlled by the combined use of inhaled corticosteroids and bronchodilators [4]. However, between 5% and 10% of patients suffer from refractory asthma [5]. Therefore, further study of the mechanisms leading to chronic inflammation and the development of bronchospasm is required to develop new and effective therapeutic approaches to the treatment of this pathology [6].

In most cases, asthma is an allergic disease, so the main mechanism of the formation of the pathological process is immune. Currently, the important role of the nervous system in the development

of allergic reactions in asthma patients has been shown. Lung nociceptors are involved in the pathogenesis of airway inflammation in asthma through the bronchoconstrictor mechanism and the initiation of local neurogenic inflammation, which is the result of an imbalance of the interactions between nerve and immune cells [7]. As a result, in recent years researchers have focused on neuro-immune interactions, which can play an important role in asthma pathophysiology, due to their contribution to allergic inflammation [8]. Connections between neurons and immune cells have been found [9]. This may be explained by their proximity to the mucous membranes. The immune system activates sensory neurons through inflammatory mediators (cytokines, histamine), initiating the development of bronchospasm in asthma. Conversely, neurons interact with immune cells through neurotransmitters or neuropeptides, modulating the development of Th2-mediated immune response and the development of chronic inflammation.

The study of neuro-immune mechanisms in asthma pathophysiology will allow the finding of new therapeutic targets.

## 2. Material and Methods

Bibliographic searches were carried out in PubMed for reports of randomized controlled trials published between 2008 and 2018. In the search for publications, keyword combinations were used: “fatty acid metabolism” OR “palmitoylethanolamide” OR “immunology” OR “neurons” OR “lung” OR “inflammation” OR “nerve growth factor”.

## 3. The Role of Eosinophils and Mast Cells in the Pathophysiology of Asthma

The pathophysiology of allergic asthma is extremely multifaceted, and is characterized by chronic inflammation, which leads to a change in respiratory tract physiology manifested by the formation of hyperresponsiveness and reversible airflow obstruction [10]. Eosinophils and mast cells are cells of the immune system that are key for allergic diseases.

Eosinophils play an important role in asthma pathophysiology. These cells are the main effector cells in this disease [11]. Clinical evidence suggests that eosinophilia may be associated either with allergic forms of early asthma, which respond to inhaled corticosteroids and bronchodilators, or with severe non-allergic forms of late asthma [12].

Research data indicate the important role of mast cells in asthma pathophysiology and reveal the mechanism of their influence on pulmonary function [13]. Mast cell degranulation is accompanied by the release of allergic mediators (histamine, proteases, chemotactic factors, cytokines and arachidonic acid metabolites), which act on the vascular network, smooth muscle, connective tissue, goblet cells, and inflammatory cells of the respiratory tract, thereby causing bronchospasm [14]. In addition, mast cells are located near the main structures of the bronchial wall, such as the epithelium and smooth muscles, and their interaction is fundamental to asthma [15]. Since mast cells synthesize a wide range of pro-inflammatory mediators [16], they are involved in both the early and late stages of allergic reactions in sensitized people [17]. Constant activation of mast cells leads to chronic inflammation of the respiratory tract in asthma [18], therefore the regulation of mast cell degranulation will allow for inhibiting the release of allergic and pro-inflammatory mediators, and may be one of the important methods for the treatment of the disease.

Both mast cells and eosinophils are involved in the pathophysiological mechanisms of asthma. Despite the fact that these cells differ in a number of parameters, which determine their connection with various stages of an allergic reaction, they are always detected in chronic inflammation [19]. These cells can form an “allergic effector unit” in asthma [20]. M. Elishmereni et al. first present evidence for the relationship between mast cells and eosinophils [20]. In addition, *in vitro* studies conducted by the same group of authors indicate that the interaction between these cells leads to the release of pro-inflammatory mediators, which may play an important role in chronic allergic inflammation. Both activated mast cells and eosinophils can modulate each other’s functions by producing a number of mediators, such as stem cell factor, osteopontin, interleukin 5, prostaglandin D2, platelet activating

factor, vascular endothelial growth factor (VEGF-A), histamine, and nerve growth factor (NGF) [21]. It can be concluded that therapeutic impact on mast cells will allow for modulating the functions of eosinophils through the interaction of specific receptors and blocking the existing interaction between mast cells and eosinophils, which plays an important role in the process of mediator release. At the same time, little is known about endogenous molecules and mechanisms modulating the functions of these cells.

#### 4. The Role of Nerve Growth Factor in Asthma Pathophysiology

A number of mediators produced by mast cells and eosinophils are involved in the implementation of functions of the immune and nervous system, and also play an important role in asthma pathophysiology.

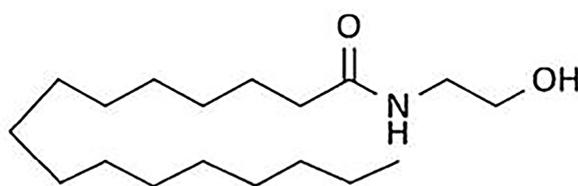
NGF is produced by both mast cells [22] and eosinophils [23]. Kim et al. have shown that NGF level correlates with the number of eosinophils, which are the main effector cells in asthma [24]. The interaction between immune cells and NGF was first found for mast cells [25]. It has been shown that NGF acts as a mast cell differentiation factor, affecting their functioning and survival by protecting against apoptosis [26]. A growing body of research on the studied problem confirms existing views that NGF is a key molecule in the interaction between the immune and nervous systems. According to the scientific literature, changes in NGF synthesis influence on both neuron physiology and immune cell activity [27]. NGF has pronounced biological action on mast and nerve cells, which is mediated by receptors expressed by these cells. Despite the fact that NGF is known primarily for its classical role in the physiology and pathology of the nervous system it level is considered as an important factor associated with the pathogenesis of allergic diseases, in particular asthma [28,29]. Using experimental animal models of asthma, Yang et al. have discovered that NGF may aggravate inflammation and airway remodeling through the Th2 immune response [30]. Besides that, under experimental conditions Qin et al. have shown the role of NGF in the Th2-mediated immune response to asthma [31]. The study of asthma pathogenesis in ovalbumin-sensitized mice provides evidence that NGF blocking decreases inflammation in the airways [32]. Shi et al. have confirmed, in an experimental asthma model, that NGF blocking inhibits airways allergic inflammation by modulating the balance of the Th1 and Th2 responses of T-cells [33]. An elevated NGF level was observed in patients with asthma after allergen bronchial provocation, and was closely related to the severity of the disease [24,34]. A high expression of NGF and tissue inhibitor of metalloproteinase-1 (TIMP-1), as well as the correlation between parameters in asthma patients, indicate a possible relationship between NGF and TIMP-1, which may play an important role in asthma pathogenesis. Renz et al. have demonstrated that NGF contributes to airway remodeling in asthma [34]. However, it is worth noting that the mechanism for the participation of NGF in the pathophysiology of chronic inflammation in asthma remains not fully understood, and is relevant for further research. It is obvious that the modulation of NGF level, which affects the activity of immune and mast cells, will allow for controlling the inter-systemic relationships in asthma. It is one of the current areas of search for approaches to the treatment of this pathology.

There is a large database of fatty acid ethanolamides, which have pronounced anti-inflammatory and analgesic activity for neuropathic pain and neurodegenerative diseases [35,36], as well as the ability to regulate the activation and degranulation of mast cells (this action is known as autacoid local injury antagonism (ALIA)) [37]. These data open prospects for the use of fatty acid ethanolamides for asthma treatment, taking into account the close relationship between the nervous and immune systems.

#### 5. Palmitoylethanolamide

Palmitoylethanolamide (N-palmitoylethanolamine, or PEA) was discovered in the late 1950s. It is an endogenous lipid mediator belonging to the class of fatty acid ethanolamides [38], acts as a signal compound, and is synthesized on demand from membrane phospholipids Figure 1. In addition, PEA is a natural lipid component found in nutritional products and food additives. It has been shown that egg yolk, peanut oil, and soy lecithin exhibit anti-allergic and anti-inflammatory properties, which

are mediated by PEA fraction [39]. After studying the prophylactic and therapeutic efficacy of PEA in viral infections [40], its importance in veterinary medicine was considered (Redonyl) and then it was described as a nutritional supplement for the treatment of chronic pain and chronic inflammation (Normast, Pelvilen, Peapure, Epitech Srl, and Recoclix) [41]. At present, there is overwhelming evidence that PEA metabolism is disturbed in inflammatory, neurodegenerative diseases and pain syndromes [36]. Recent data obtained by Portavella et al. confirm that PEA has a neuroprotective effect [42]. It allows considering PEA as an anti-inflammatory, neuroprotective and analgesic mediator, and opens new prospects for its use [43]. Although the biological effects of PEA are well documented, the molecular mechanisms and targets for its action remain a subject for discussion. The most studied organ-targets are the intestine [44], the bladder [45], the skin [46], kidneys [47], and the central nervous system [48]. At the same time, the possible effect of PEA on the respiratory system has been studied in singular works [49].

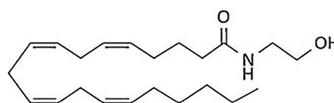


**Palmitoylethanolamide (PEA)**

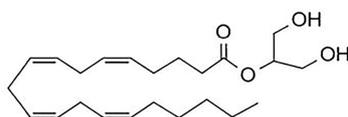
**Figure 1.** Chemical structures of palmitoylethanolamide (PEA).

## 6. Molecular Targets and the Mechanisms of Action of Palmitoylethanolamide

PEA is a structural analogue of anandamide (AEA), and has similarity to the ligands of N-acylethanolamine (NAE) of cannabinoid receptors (Figure 2). The direct or indirect (through action on endogenous ligands or through receptor expression) molecular targets of PEA are G protein-coupled receptors (GPCR), as well as cannabinoid receptors 1 (CB1) and 2 (CB2) belonging to their family; a new cannabinoid receptor orphan G protein-coupled receptor (GPR 55); transient receptor potential (TRP) ion channels, in particular TRP vanilloid receptor 1 (TRPV1); nuclear peroxisome proliferator-activated receptors (PPAR $\alpha/\gamma/\delta$ ); and ATP-sensitive potassium channels (K<sub>ATP</sub> channel) [50] Figure 3.

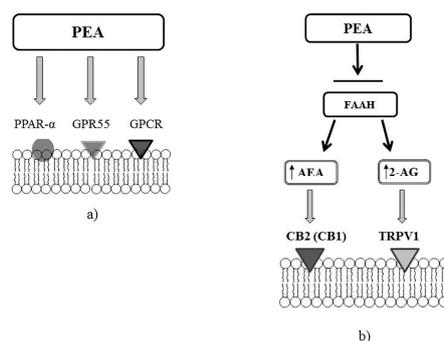


Anandamide (*N*-arachidonylethanolamine, AEA)



2-Arachidonoyl-glycerol (2-AG)

**Figure 2.** Chemical structures of *N*-arachidonylethanolamine or anandamide (AEA), and 2-arachidonylethanolamine (2-AG).



**Figure 3.** Molecular targets of PEA. (a) PEA can directly activate G protein-coupled receptor  $\alpha$  (PPAR $\alpha$ ) or a cannabinoid receptor G protein-coupled receptor (GPR55). (b) PEA through the inhibition of the expression of fatty acid amide hydrolase (FAAH), may increase the endogenous levels of AEA and 2-AG, which directly activate cannabinoid receptor 2 (CB2) (or cannabinoid receptor 1, CB1) and transient receptor potential vanilloid receptor 1 (TRPV1) channels.

PEA has low affinity for CB1 and CB2 receptors; as a result, this lipid mediator realizes its biological effect through the endocannabinoid 2-arachidonoylglycerol (2-AG), which is the endogenous ligand of CB2 receptors [36] (see Figure 2). PEA acts as an endogenous CB2 cannabinoid receptor agonist under in vitro conditions. The analgesic effect of PEA is inhibited by CB2 antagonist -SR144528 under in vivo conditions [51]. It has been shown that PEA increases 2-AG level two times [46], under both in vitro and in vivo conditions [52]. There are findings that indicate that 2-AG is an agonist for TRPV1 channel [36]. TRPV1 desensitization is the basis of the analgesic and anti-inflammatory effects of TRPV1 agonists, which currently are used to treat neuropathic pain [53]. In addition, there are indications that PEA can significantly enhance TRPV1 activation under exposure to 2-AG in vitro [52]. These results have allowed the uncovering of another mechanism of action for this lipid mediator, opening new prospects for the realization of powerful, TRPV1-mediated anesthetic and anti-inflammatory action. Thus, the sensitivity of nociceptors, the number of which significantly increases in the lungs of asthma patients, is associated with the activity of sensory neural TRP ion channels.

NGF is a stimulus for the development of a prolonged pain syndrome. It is known that its level rises in chronic pain conditions. Recent studies have shown that NGF can modulate TRPV1 activity in rats [54]. Eskander et al., have established that NGF causes a stable, nociceptive state mediated by TRPV1 activity and oxidative mechanisms [55]. Consequently, PEA can have an analgesic effect by influencing TRPV1 activity and the NGF level, which mediate the development of a prolonged pain syndrome.

The multifaceted biological effects of PEA are also mediated by its interaction with receptors GPR55 [56], GPR119 [57], and PPAR $\alpha$  [51].

There are three isoforms of PPAR: PPAR $\alpha$  (NR1C1), PPAR $\beta/\delta$  (NR1C2), and PPAR $\gamma$  (NR1C3). Peroxisome proliferator-activated receptors can be activated by different types of fatty acids (FA) that are part of the PAE molecule. Linolenic acid (C18:2 *n*-6; LA) and alpha-linolenic acid (C18:3 *n*-3, ALA), which are long-chain polyunsaturated FAs (LCPUFAs) and play a key role in bronchopulmonary diseases, have the highest affinity for PPAR.

PPAR $\alpha$  is expressed in tissues with a high FA oxidation and is involved in cell proliferation, apoptosis, and differentiation, as well as the normalization of the balance between lipid peroxidation and antioxidant protection by increasing the activity of antioxidant enzymes. PPAR $\alpha$  activation by LCPUFA (mainly C18:3 *n*-3, or ALA) appears to play a key role in the prevention and treatment of asthma. The action of PEA via PPAR $\alpha$  leads to the activation of the TRPV1 channel [58]. Thus, the activation of the PPAR $\alpha$  receptor also mediates the anti-inflammatory and analgesic effects of PEA [47]. In addition, the activation of two other isoforms of the PPAR receptor family, including PPAR $\beta/\delta$  and PPAR $\gamma$ , which initiates peroxisome proliferation [59], is not excluded. PPAR $\alpha$  impacts fatty acid metabolism, and its activation reduces lipid levels, whereas PPAR $\gamma$  is involved in the regulation of adipogenesis,

energy balance, and lipid biosynthesis. PPAR $\beta$  and PPAR $\delta$  are involved in the oxidation of fatty acids and the regulation of glucose and cholesterol levels in the blood.

As noted above, NGF level mediated by TRPV1 activity increases in chronic pain conditions [55]. In turn, PEA action is mediated through 2-AG, which acts as an agonist of the TRPV1 channel [36]. These facts indicate the possibility of the realization of analgesic and anti-inflammatory effects of PEA-mediated TRPV1.

The analgesic effect of PEA has been discussed by Hesselink [60].

## 7. Palmitoylethanolamide and Asthma

NGF is considered in close connection with asthma pathogenesis [29]. At the same time, there is evidence of the simultaneous increase in the number of mast cells and the NGF level in chronic inflammation [61]. These indicators also play an important role in asthma pathophysiology [13,14]. Taking into account the targets for biological action of PEA, we can assume that PEA can contribute to the regulation of the NGF level, which is elevated in chronic bronchopulmonary diseases. In addition, a decrease in the activity of mast cells by regulating the NGF level will affect the interaction between mast cells and eosinophils, which results in the release of soluble mediators in chronic allergic inflammation [20].

The existing view of the potentially significant role of PEA in the airways is confirmed by the high expression of this lipid mediator in the bronchi (6 nmol/g), which is about ten times higher than its level in the brain of mouse (100–550 pmol/g) [47]. Besides that, all molecular targets of PEA (PPAR $\alpha$ , GRP55, TRPV1, CB1, and CB2) are localized in the bronchi of mice, which suggests an extremely important role of PEA in the physiology and the pathology of the lungs [36]. Zhou et al. have detected a change in CB2 and GPR55 levels in sensitized mice [62]. Perhaps the activation of these receptors is associated with increased mast cell infiltration. The research of the Nobel Laureate R. Levi-Montalcini have demonstrated the importance of inactivating the inflammatory processes through the influence of PEA on mast cells [63]. According to Calignano et al., PEA reduces the migration and the degranulation of mast cells [64]. Current scientific literature suggests that the endogenous lipid mediator PEA, which is chemically bound to the endogenous cannabinoid anandamide and acts as the local inhibitor of mast cell inflammation [65]. Facci et al. have shown that PEA suppresses cell degranulation mediated by the CB2 cannabinoid receptor [66]. Therefore, CB2 receptors may be involved in the mechanism of the action of PEA on mast cells. The importance of GPR55 for the most of the inhibitory effects of PEA on activated mast cells has been shown [67]. Additionally, Rinne et al., have revealed that PEA increases the expression of proto-oncogene tyrosine protein kinase (Mer tyrosine kinase (Mer)) by activating GPR55, and therefore affects the function of macrophages [68]. The possible contribution of PEA to allergic sensitization has been investigated in the study of Roviezzo et al. [69]. This research group was the first to describe a decrease in bronchial level of PEA in mice with allergic reactions (after the sensitization of allergens). A reduction in the regulation of PEA expression is specific for the bronchi, since no differences in plasma level of PEA between control and sensitized mice were detected. A decrease in the PEA level in sensitized mice is a loss of the endogenous anti-inflammatory mechanism, in light of the known PEA-mediated mechanism (ALIA). This hypothesis is supported by other studies demonstrating the therapeutic effect of some anti-inflammatory drugs associated with the restoration of the endogenous level of PEA [70]. In particular, Di Paola et al. has found that the use of ultra-micronized palmitoylethanolamide (PEA-um) reduces the severity of inflammation in the lungs in a mouse model of idiopathic pulmonary fibrosis [49].

Thus, based on the relationship between the nervous and immune systems, and existing data on the mechanisms of the biological action of PEA and its role in asthma, we can talk about the prospects for the use of fatty acid ethanolamides, in particular PEA, in asthma therapy.

## 8. Conclusions

Allergic airway inflammation is associated with complex neuro-immune interactions that can play a crucial role in chronic inflammation and asthma progression. Although immune-mediated therapy for asthma has shown some success, patients with severe forms of the disease are often resistant to this treatment. Thus, the nervous system and neuro-immune interrelations may become a new and promising therapeutic target in asthma, in particular its allergic form.

A key molecule mediating neuro-immune interactions is NGF. In addition to its classic role in the pathophysiology of the nervous system, NGF is considered to be an important factor associated with the pathogenesis of allergic diseases, particularly asthma, due to its influence on the differentiation of mast cells. In this regard, NGF can be one of the targets for the action of PEA, which has a proven biological effect on the nervous system, as well as on the activation and the degranulation of mast cells. These data open prospects for the use of fatty acid ethanolamides in asthma treatment, taking into account the close relationship between the nervous and immune systems.

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## References

1. Kytikova, O.; Gvozdenko, T.; Antonyuk, M. Modern aspects of prevalence of chronic bronchopulmonary diseases. *Bull. Physiol. Pathol. Respir.* **2017**, *64*, 94–100. (In Russian) [[CrossRef](#)]
2. Global Strategy for Asthma Management and Prevention, Global Initiative for Asthma (GINA). 2012. Available online: [http://www.ginasthma.org/documents/5/documents\\_variants/37](http://www.ginasthma.org/documents/5/documents_variants/37) (accessed on 30 December 2012).
3. Bosnjak, B.; Stelzmueller, B.; Erb, K.J.; Epstein, M.M. Treatment of allergic asthma: modulation of Th2 cells and their responses. *Respir. Res.* **2011**, *12*, 114. [[CrossRef](#)]
4. Lang, D.M. Severe asthma: Epidemiology, burden of illness, and heterogeneity. *Allergy Asthma Proc.* **2015**, *36*, 418–424. [[CrossRef](#)]
5. Kudo, M.; Ishigatsubo, Y.; Aoki, I. Pathology of asthma. *Front. Microbiol.* **2013**, *4*. [[CrossRef](#)]
6. Casciano, J.; Krishnan, J.; Dotiwala, Z.; Li, C.; Sun, S.X. Clinical and economic burden of elevated blood eosinophils in patients with and without uncontrolled asthma. *J. Manag. Care Spec. Pharm.* **2017**, *23*, 85–91. [[CrossRef](#)] [[PubMed](#)]
7. Barnes, P.J. Neurogenic inflammation in the airways. *Respir. Physiol.* **2001**, *125*, 145–154. [[CrossRef](#)]
8. Voisin, T.; Bouvier, A.; Chiu, I.M. Neuro-immune interactions in allergic diseases: Novel targets for therapeutics. *Int. Immunol.* **2017**, *29*, 247–261. [[CrossRef](#)]
9. Thornton, M.A.; Akasheh, N.; Walsh, M.T.; Moloney, M.; Sheahan, P.O.; Smyth, C.M.; Walsh, R.M.; Morgan, R.M.; Curran, D.R.; Walsh, M.T.; et al. Eosinophil recruitment to nasal nerves after allergen challenge in allergic rhinitis. *Clin. Immunol.* **2013**, *147*, 50. [[CrossRef](#)] [[PubMed](#)]
10. Chung, K.F. Precision medicine in asthma. *Curr. Opin. Pulm. Med.* **2018**, *24*, 4–10. [[CrossRef](#)] [[PubMed](#)]
11. Yancey, S.W.; Keene, O.N.; Albers, F.C.; Ortega, H.G.; Bates, S.A.; Bleecker, E.; Pavord, I.D. Biomarkers for severe eosinophilic asthma. *J. Allergy Clin. Immun.* **2017**, *140*, 1509–1518. [[CrossRef](#)]
12. Moore, W.C.; Meyers, D.A.; Wenzel, S.E. Identification of asthma phenotypes using cluster analysis in the severe asthma research program. *Am. J. Respir. Crit. Care Med.* **2010**, *181*, 315–323. [[CrossRef](#)]
13. Galli, S.J.; Tsai, M. IgE and mast cells in allergic disease. *Nat. Med.* **2012**, *18*, 693–704. [[CrossRef](#)]
14. Bradding, P.; Arthur, G. Mast cells in asthma—state of the art. *Clin. Exp. Allergy* **2016**, *46*, 194–263. [[CrossRef](#)]
15. Carter, R.J.; Bradding, P. The role of mast cells in the structural alterations of the airways as a potential mechanism in the pathogenesis of severe asthma. *Curr. Pharm. Des.* **2011**, *17*, 685–698. [[CrossRef](#)]

16. Oskeritzian, C.A. Mast cell plasticity and sphingosine-1-phosphate in immunity, inflammation and cancer. *Mol. Immunol.* **2015**, *63*, 104–112. [[CrossRef](#)]
17. Thiriou, D.; Morianos, I.; Xanthou, G.; Samitas, K. Innate immunity as the orchestrator of allergic airway inflammation and resolution in asthma. *Int. Immunopharmacol.* **2017**, *48*, 43–54. [[CrossRef](#)]
18. Bulfone-Paus, S.; Bahri, R. Mast cells as regulators of T cell responses. *Front. Immunol.* **2015**, *6*, 394. [[CrossRef](#)]
19. Gangwar, R.S.; Landolina, N.; Arpinati, L.; Levi-Schaffer, F. Mast cell and eosinophil surface receptors as targets for anti-allergic therapy. *Pharmacol. Ther.* **2017**, *170*, 37–63. [[CrossRef](#)]
20. Elishmereni, M.; Bachelet, I.; Nissim Ben-Efraim, A.H.; Mankuta, D.; Levi-Schaffer, F. Interacting mast cells and eosinophils acquire an enhanced activation state in vitro. *Allergy* **2013**, *68*, 171–179. [[CrossRef](#)]
21. Galdiero, M.R.; Varricchi, G.; Seaf, M.; Marone, G.; Levi-Schaffer, F.; Marone, G. Bidirectional mast cell-eosinophil interactions in inflammatory disorders and cancer. *Front. Med.* **2017**, *4*, 103. [[CrossRef](#)]
22. Nilsson, G.; Forsberg-Nilsson, K.; Xiang, Z.; Hallbook, F.; Nilsson, K.; Metcalfe, D.D. Human mast cells express functional TrkA and are a source of nerve growth factor. *Eur. J. Immunol.* **1997**, *27*, 2295–2301. [[CrossRef](#)]
23. Solomon, A.; Aloe, L.; Pe'er, J.; Frucht-Pery, J.; Bonini, S.; Levi-Schaffer, F. Nerve growth factor is preformed in and activates human peripheral blood eosinophils. *J. Allergy Clin. Immunol.* **1998**, *102*, 454–460. [[CrossRef](#)]
24. Kim, J.S.; Kang, J.Y.; Ha, J.H.; Lee, H.Y.; Kim, S.J.; Kim, S.C.; Ahn, J.H.; Kwon, S.S.; Kim, Y.K.; Lee, S.Y. Expression of nerve growth factor and matrix metalloproteinase-9/tissue inhibitor of metalloproteinase-1 in asthmatic patients. *J. Asthma* **2013**, *50*, 712–717. [[CrossRef](#)]
25. Böhm, A.; Aloe, L.; Levi-Montalcini, R. Nerve growth factor enhances precocious differentiation and numerical increase in mast cells in cultures of rat splenocytes. *Accad. Naz. Lincei.* **1986**, *80*, 1–6.
26. Matsuda, H.; Kannan, Y.; Ushio, H.; Kiso, Y.; Kanemoto, T.; Suzuki, H. Nerve growth factor induces development of connective tissue-type mast cells in vitro from murine bone marrow cells. *J. Exp. Med.* **1991**, *174*, 7–14. [[CrossRef](#)]
27. Sofroniew, M.V.; Howe, C.L.; Mobley, W.C. Nerve growth factor signaling, neuroprotection, and neural repair. *Annu. Rev. Neurosci.* **2001**, *24*, 1217–1281. [[CrossRef](#)]
28. Bonini, S.; Lambiase, A.; Angelucci, F.; Magrini, L.; Manni, L.; Aloe, L. Circulating nerve growth factor levels are increased in humans with allergic diseases and asthma. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 10955–10960. [[CrossRef](#)]
29. Nassenstein, C.; Schulte-Herbruggen, O.; Renz, H.; Braun, A. Nerve growth factor: the central hub in the development of allergic asthma? *Eur. J. Pharmacol.* **2006**, *533*, 195–206. [[CrossRef](#)]
30. Yang, Y.G.; Tian, W.M.; Zhang, H.; Li, M.; Shang, Y.X. Nerve growth factor exacerbates allergic lung inflammation and airway remodeling in a rat model of chronic asthma. *Exp. Ther. Med.* **2013**, *6*, 1251–1258. [[CrossRef](#)]
31. Qin, Q.; Wang, Z.; Pan, P.; Cao, Z.; Xia, Q.; Tan, H.; Hu, C. Lung dendritic cells undergo maturation and polarization towards a T helper type 2-stimulating phenotype in a mouse model of asthma: Role of nerve growth factor. *Exp. Ther. Med.* **2014**, *8*, 1402–1408. [[CrossRef](#)]
32. Chen, Y.L.; Huang, H.Y.; Lee, C.C.; Chiang, B.L. Small interfering RNA targeting nerve growth factor alleviates allergic airway hyperresponsiveness. *Mol. Ther. Nucl. Acids* **2014**, *3*, 158. [[CrossRef](#)] [[PubMed](#)]
33. Shi, Y.; Jin, Y.; Guo, W.; Chen, L.; Liu, C.; Lv, X. Blockage of nerve growth factor modulates T cell responses and inhibits allergic inflammation in a mouse model of asthma. *Inflamm. Res.* **2012**, *61*, 1369–1378. [[CrossRef](#)] [[PubMed](#)]
34. Renz, H.; Kiliç, A. Neurotrophins in chronic allergic airway inflammation and remodeling. *Chem. Immunol. Allergy* **2012**, *98*, 100–117.
35. Iannotti, F.A.; Di Marzo, V.; Petrosino, S. Endocannabinoids and endocannabinoid-related mediators: targets, metabolism and role in neurological disorders. *Prog. Lipid Res.* **2016**, *62*, 107–128. [[CrossRef](#)] [[PubMed](#)]
36. Petrosino, S.; Di Marzo, V. The pharmacology of palmitoylethanolamide and first data on the therapeutic efficacy of some of its new formulations. *Br. J. Pharmacol.* **2017**, *174*, 1349–1365. [[CrossRef](#)]
37. Aloe, L.; Leon, A.; Levi-Montalcini, R. A proposed autacoid mechanism controlling mastocyte behaviour. *Agents Actions* **1993**, *39*, 145–147. [[CrossRef](#)]
38. Ueda, N.; Tsuboi, K.; Uyama, T. Metabolism of endocannabinoids and related N-acyl ethanolamines: Canonical and alternative pathways. *FEBS J.* **2013**, *280*, 1874–1894. [[CrossRef](#)]

39. Esposito, E.; Cuzzocrea, S. Palmitoylethanolamide in homeostatic and traumatic central nervous system injuries. *CNS Neurol. Disord. Drug Targets* **2013**, *12*, 55–61. [[CrossRef](#)]
40. Kahlich, R.; Klima, J.; Cihla, F.; Frankov'a, V.; Masek, K.; Rosick'y, M. Studies on prophylactic efficacy of N-2-hydroxyethyl palmitamide [Impulsin] in acute respiratory infections, serologically controlled field trials. *J. Hyg. Epidemiol. Microbiol. Immunol.* **1979**, *23*, 11–24.
41. Keppel Hesselink, J.M.; Kopsky, D.J.; Witkamp, R.F. Palmitoylethanolamide (PEA)—'Promiscuous' anti-inflammatory and analgetic molecule at the interface between nutrition and pharma. *PharmaNutrition* **2014**, *2*, 19–25. [[CrossRef](#)]
42. Portavella, M.; Rodriguez-Espinosa, N.; Galeano, P.; Blanco, E.; Romero, J.I.; Holubiec, M.I.; Rodriguez de Fonseca, F.; Fernández-Espejo, E. Oleoylethanolamide and palmitoylethanolamide protect cultured cortical neurons against hypoxia. *Cannabis Cannabinoid Res.* **2018**, *3*, 171–178. [[CrossRef](#)]
43. Boccella, S.; Cristiano, C.; Romano, R.; Iannotta, M.; Belardo, C.; Farina, A.; Guida, F.; Piscitelli, F.; Palazzo, E.; Mazzitelli, M.; et al. Ultra-micronized palmitoylethanolamide rescues the cognitive decline-associated loss of neural plasticity in the neuropathic mouse entorhinal cortex-dentate gyrus pathway. *Neurobiol. Dis.* **2018**, *121*, 106–119. [[CrossRef](#)]
44. Borrelli, F.; Romano, B.; Petrosino, S.; Pagano, E.; Capasso, R.; Coppola, D.; Battista, G.; Orlando, P.; Di Marzo, V.; Izzo, A. Palmitoylethanolamide, a naturally occurring lipid, is an orally effective intestinal anti-inflammatory agent. *Br. J. Pharmacol.* **2015**, *172*, 142–158. [[CrossRef](#)]
45. Pessina, F.; Capasso, R.; Borrelli, F.; Aveta, T.; Buono, L.; Valacchi, G.; Fiorenzani, P.; Di Marzo, V.; Orlando, P.; Izzo, A.A. Protective effect of palmitoylethanolamide in a rat model of cystitis. *J. Urol.* **2015**, *193*, 1401–1408. [[CrossRef](#)]
46. Vaia, M.; Petrosino, S.; De Filippis, D.; Negro, L.; Guarino, A.; Carnuccio, R.; Di Marzo, V.; Iuvone, T. Palmitoylethanolamide reduces inflammation and itch in a mouse model of contact allergic dermatitis. *Eur. J. Pharmacol.* **2016**, *791*, 669–674. [[CrossRef](#)]
47. Mattace Raso, G.; Russo, R.; Calignano, A.; Meli, R. Palmitoylethanolamide in CNS health and disease. *Pharmacol. Res.* **2014**, *86*, 32–41. [[CrossRef](#)]
48. Citraro, R.; Russo, E.; Scicchitano, F.; van Rijn, C.M.; Cosco, D.; Avagliano, C.; Russo, R.; D'Agostino, G.; Petrosino, S.; Guida, F.; et al. Antiepileptic action of N-palmitoylethanolamine through CB1 and PPAR- $\alpha$  receptor activation in a genetic model of absence epilepsy. *Neuropharmacology* **2012**, *69*, 115–126. [[CrossRef](#)]
49. Di Paola, R.; Impellizzeri, D.; Fusco, R.; Cordaro, M.; Siracusa, R.; Crupi, R.; Esposito, R.; Salvatore, E. Ultramicronized palmitoylethanolamide (PEA-um<sup>(®)</sup>) in the treatment of idiopathic pulmonary fibrosis. *Pharmacol. Res.* **2016**, *111*, 405–412. [[CrossRef](#)]
50. Pertwee, R.G.; Howlett, A.C.; Abood, M.E.; Alexander, S.P.; Di Marzo, V.; Elphick, M.R.; Greasley, P.J.; Hansen, H.S.; Kunos, G.; Mackie, K.; et al. International union of basic and clinical pharmacology. LXXIX. Cannabinoid receptors and their ligands: beyond CB<sub>1</sub> and CB<sub>2</sub>. *Pharmacol. Rev.* **2010**, *62*, 588–631. [[CrossRef](#)]
51. Lo Verme, J.; Fu, J.; Astarita, G.; La Rana, G.; Russo, R.; Calignano, A.; Piomelli, D. The nuclear receptor peroxisome proliferator-activated receptor- $\alpha$  mediates the anti-inflammatory actions of palmitoylethanolamide. *Mol. Pharmacol.* **2005**, *67*, 15–19. [[CrossRef](#)]
52. Zygmunt, P.M.; Ermund, C.; Movahed, P.; Andersson, D.A.; Simonsen, C.; Jönsson, B.A. Monoacylglycerols activate TRPV1—A link between phospholipase C and TRPV1. *PLoS ONE* **2013**, *8*, e81618. [[CrossRef](#)] [[PubMed](#)]
53. Brederson, J.D.; Kym, P.R.; Szallasi, A. Targeting TRP channels for pain relief. *Eur. J. Pharmacol.* **2013**, *716*, 61–76. [[CrossRef](#)]
54. Mills, C.D.; Nguyen, T.; Tanga, F.Y.; Zhong, C.; Gauvin, D.M.; Mikusa, J.; Gomez, E.J.; Salyers, A.K.; Bannon, A.W. Characterization of nerve growth factor-induced mechanical and thermal hypersensitivity in rats. *Eur. J. Pain* **2013**, *17*, 469–479. [[CrossRef](#)]
55. Eskander, M.A.; Ruparel, S.; Green, D.P.; Chen, P.B.; Por, E.D.; Jeske, N.A.; Gao, X.; Flores, E.R.; Hargreaves, K.M. Persistent nociception triggered by nerve growth factor (NGF) is mediated by TRPV1 and oxidative mechanisms. *J. Neurosci.* **2015**, *35*, 8593–8603. [[CrossRef](#)]
56. Ryberg, E.; Larsson, N.; Sjögren, S.; Hjorth, S.; Hermansson, N.O.; Leonova, J.; Elebring, T.; Nilsson, K.; Drmota, T.; Greasley, P.J. The orphan receptor GPR55 is a novel cannabinoid receptor. *Br. J. Pharmacol.* **2007**, *152*, 1092–1101. [[CrossRef](#)]

57. Khasabova, I.A.; Xiong, Y.; Coicou, L.; Piomelli, D.; Seybold, V. Peroxisome proliferator-activated receptor alpha mediates acute effects of palmitoylethanolamide on sensory neurons. *J. Neurosci.* **2012**, *32*, 12735–12743. [[CrossRef](#)]
58. Ambrosino, P.; Soldovieri, M.V.; Russo, C.; Tagliatela, M. Activation and desensitization of TRPV1 channels in sensory neurons by the PPAR $\alpha$  agonist palmitoylethanolamide. *Br. J. Pharmacol.* **2013**, *168*, 1430–1444. [[CrossRef](#)]
59. O’Sullivan, S.E.; Kendall, D.A. Cannabinoid activation of peroxisome proliferator-activated receptors: potential for modulation of inflammatory disease. *Immunobiology* **2010**, *215*, 611–616. [[CrossRef](#)] [[PubMed](#)]
60. Keppel Hesselink, J.M. New targets in pain, non-neuronal cells, and the role of palmitoylethanolamide. *Open Pain J.* **2012**, *5*, 12–23. [[CrossRef](#)]
61. Xanthos, D.N.; Gaderer, S.; Drdla, R.; Nuro, E.; Abramova, A.; Ellmeier, W.; Sandkühler, J. Central nervous system mast cells in peripheral inflammatory nociception. *Mol. Pain* **2011**, *7*, 42. [[CrossRef](#)]
62. Zhou, J.; Burkovskiy, I.; Yang, H.; Sardinha, J.; Lehmann, C. CB<sub>2</sub> and GPR55 receptors as therapeutic targets for systemic immune dysregulation. *Front. Pharmacol.* **2016**, *7*, 264. [[CrossRef](#)] [[PubMed](#)]
63. Levi-Montalcini, R.; Skaper, S.D.; dal Toso, R.; Petrelli, L.; Leon, A. Nerve growth factor: From neurotrophin to neurokin. *Trends Neurosci.* **1996**, *19*, 514–520. [[CrossRef](#)]
64. Calignano, A.; La Rana, G.; Piomelli, D. Antinociceptive activity of the endogenous fatty acid amide, palmitoylethanolamide. *Eur. J. Pharmacol.* **2001**, *419*, 191–198. [[CrossRef](#)]
65. Skaper, S.; Facci, L.; Giusti, P. Glia, mast cells as targets for palmitoylethanolamide, an anti-inflammatory and neuroprotective lipid mediator. *Mol. Neurobiol.* **2013**, *48*, 340–352. [[CrossRef](#)] [[PubMed](#)]
66. Facci, L.; Dal Toso, R.; Romanello, S.; Buriani, A.; Skaper, S.D.; Leon, A. Mast cells express a peripheral cannabinoid receptor with differential sensitivity to anandamide and palmitoylethanolamide. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 3376–3380. [[CrossRef](#)]
67. Cantarella, G.; Scollo, M.; Lempereur, L.; Saccani-Jotti, G.; Basile, F.; Bernardini, R. Endocannabinoids inhibit release of nerve growth factor by inflammation-activated mast cells. *Biochem. Pharmacol.* **2011**, *82*, 380–388. [[CrossRef](#)] [[PubMed](#)]
68. Rinne, P.; Guillaumat-Prats, R.; Rami, M.; Bindila, L.; Ring, L.; Lyytikäinen, L.P.; Raitoharju, E.; Oksala, N.; Lehtimäki, T.; Weber, C.; et al. Palmitoylethanolamide promotes a proresolving macrophage phenotype and attenuates atherosclerotic plaque formation. *Arterioscler. Thromb. Vasc. Biol.* **2018**, *38*, 2562–2575. [[CrossRef](#)]
69. Roviezzo, F.; Rossi, A.; Caiazzo, E.; Orlando, P.; Riemma, M.A.; Iacono, V.M.; Guarino, A.; Ialenti, A.; Cicala, C.; Peritore, A.; et al. Palmitoylethanolamide supplementation during sensitization prevents airway allergic symptoms in the mouse. *Front. Pharmacol.* **2017**, *8*, 857. [[CrossRef](#)]
70. Jhaveri, M.D.; Richardson, D.; Robinson, I.; Garle, M.J.; Patel, A.; Sun, Y.; Sagar, D.R.; Bennett, A.J.; Alexander, S.P.; Kendall, D.A.; et al. Inhibition of fatty acid amide hydrolase and cyclooxygenase-2 increases levels of endocannabinoid related molecules and produces analgesia via peroxisome proliferator-activated receptor-alpha in a model of inflammatory pain. *Neuropharmacology* **2008**, *55*, 85–93. [[CrossRef](#)]

