



Process Optimization for the Bioinspired Synthesis of Gold Nanoparticles Using *Cordyceps militaris*, Its Characterization, and Assessment of Enhanced Therapeutic Efficacy

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Abstract: The promising therapeutic implications of nanoparticles have spurred their development for biomedical applications. An eco-friendly methodology synthesizes gold nanoparticles using *Cordyceps militaris*, an edible mushroom (*Cord*-Au-NPs), using a quality-by-design approach (central composite design). UV-visible spectroscopy analysis revealed an absorption peak in the 540–550 nm, thus confirming the synthesis of gold nanoparticles. *Cord*-Au-NPs have a crystalline structure, as evidenced by the diffraction peaks. The zeta potential value of -19.42 mV signifies the stability of *Cord*-Au-NPs. XRD study shows gold facets and EDX analysis revealed a strong peak of spherical nanoparticles in the gold region with a mean particle size of 7.18 nm and a polydispersity index of 0.096. The obtained peaks are closely associated with phenolic groups, lipids, and proteins, as examined by FTIR, suggesting that they function as the reducing agent. *Cord*-Au-NPs exhibited dose-dependent antioxidant, antidiabetic, and antibacterial activity. The method is eco-friendly, non-toxic, less time-consuming, and does not use synthetic materials, leading to higher capabilities in biomedical applications.

Keywords: green synthesis; central composite design; antioxidant activity; antidiabetic activity; process optimization

Supplementary File

2.4 Physicochemical characterization

2.4.1 UV- Visible Spectroscopy

The synthesis of *Cord*-Au-NPs was confirmed by taking an absorption spectrum of the reaction mixture in the scanning wavelength range of 200 to 800nm using a UV-visible spectrophotometer (Shimadzu, UV-2700).

2.4.2 Fourier Transform Infrared (FT-IR) Spectroscopy

Fourier Transform Infrared spectrophotometer (Make: Shimadzu, IR Affinity-1) was used to distinguish the bioreduction compounds involved in synthesizing *Cord*-Au-NPs. The spectrum was obtained using the KBr pellet method in the scanning range of 400 to 4000cm⁻¹.

2.4.3 X-ray Diffraction Study (XRD)

The synthesized *Cord*-Au-NPs were analyzed to examine the structure and composition using an X-ray diffractometer (XRD) (Rigaku, Ultima IV) at an angular range of 10°<2θ>80° at 40kV and 20mA having a 10mm divergence slit in 2θ/θ continuous scanning mode.

2.4.4 Scanning Electron Microscopy (SEM) and Energy Dispersive X-ray (EDX) Analysis

The microstructure and elemental composition of the *Cord*-Au-NPs were analyzed using SEM with EDX analysis (Carl ZEISS EVO 18). Magnification ranges of 1μm, 2μm, and 200nm were used to observe and visualize the *Cor*-ZnONPs morphology during SEM.

2.4.5 High-Resolution Transmission Electron Microscopy (HRTEM) and Selected Area Electron Diffraction (SAED)

HRTEM analysis and SAED studies of the *Cord*-Au-NPs were carried out using Transmission Electron Microscope (Make: Joel, Model: JEM 2100) to determine the morphology of the *Cor*-ZnONPs.

2.4.6 Particle Size Analysis (PSA) and Zeta Potential Analysis

The average particle size, polydispersity index, mobility and stability profile of *Cor*-ZnONPs were ascertained by performing PSA using Nanophox, NX0088 (Sympatec, Germany).



Fig. S1. Green synthesis of gold nanoparticles (a) initial color (b) color change after 1 h of incubation

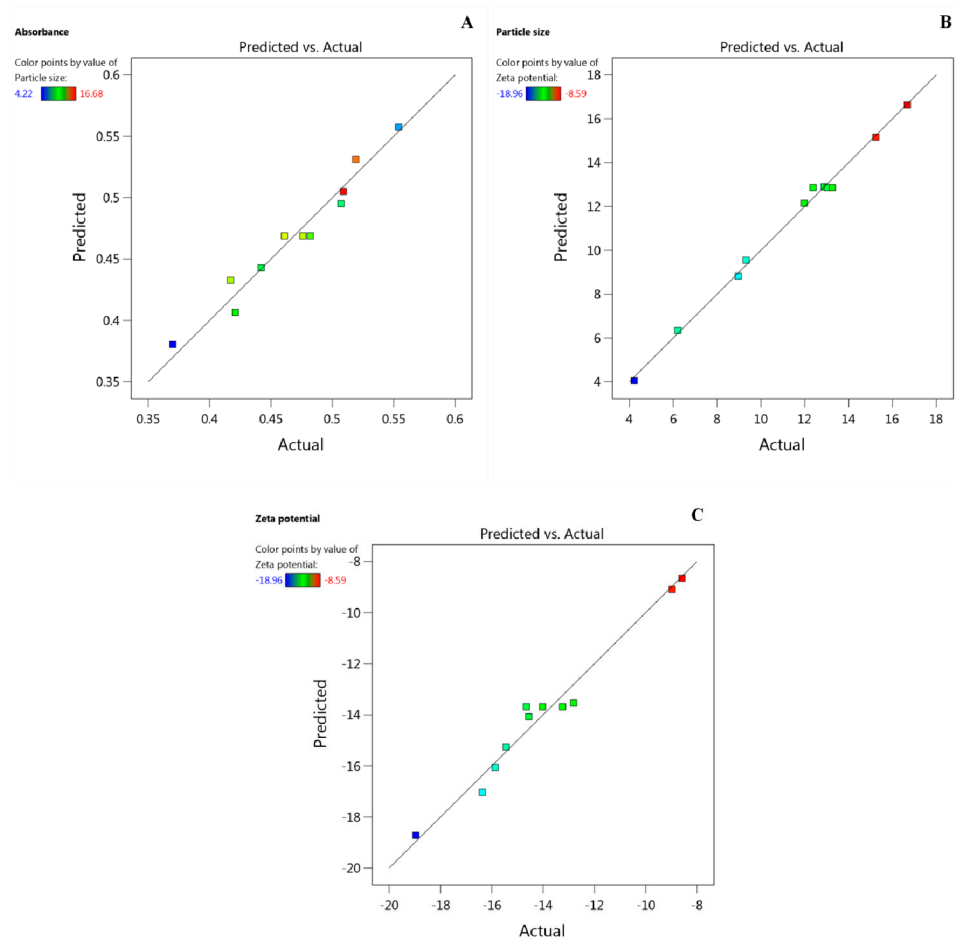
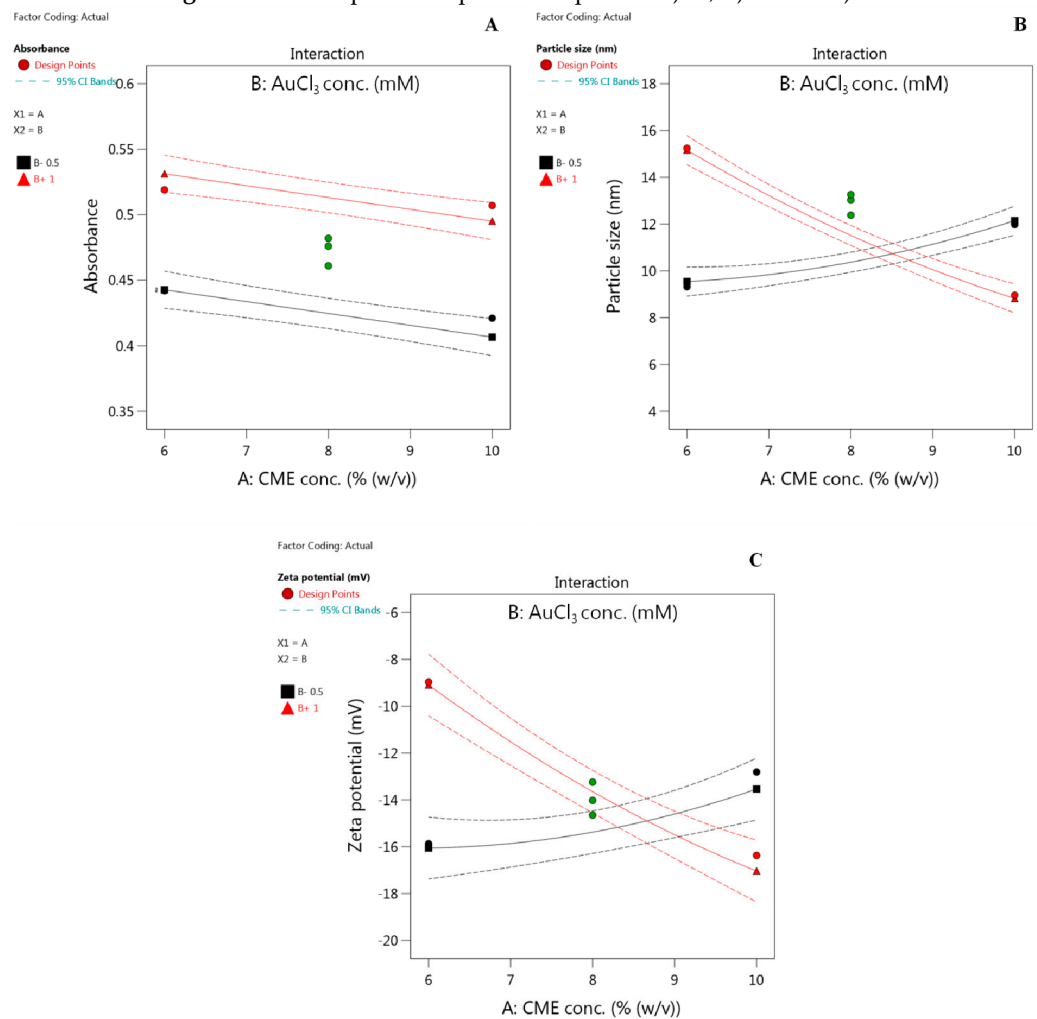


Fig. S2: Actual vs predicted plot for responses A) Y_1 , B) Y_2 and C) Y_3 Fig. S3: Interaction plot for responses A) Y_1 , B) Y_2 and C) Y_3

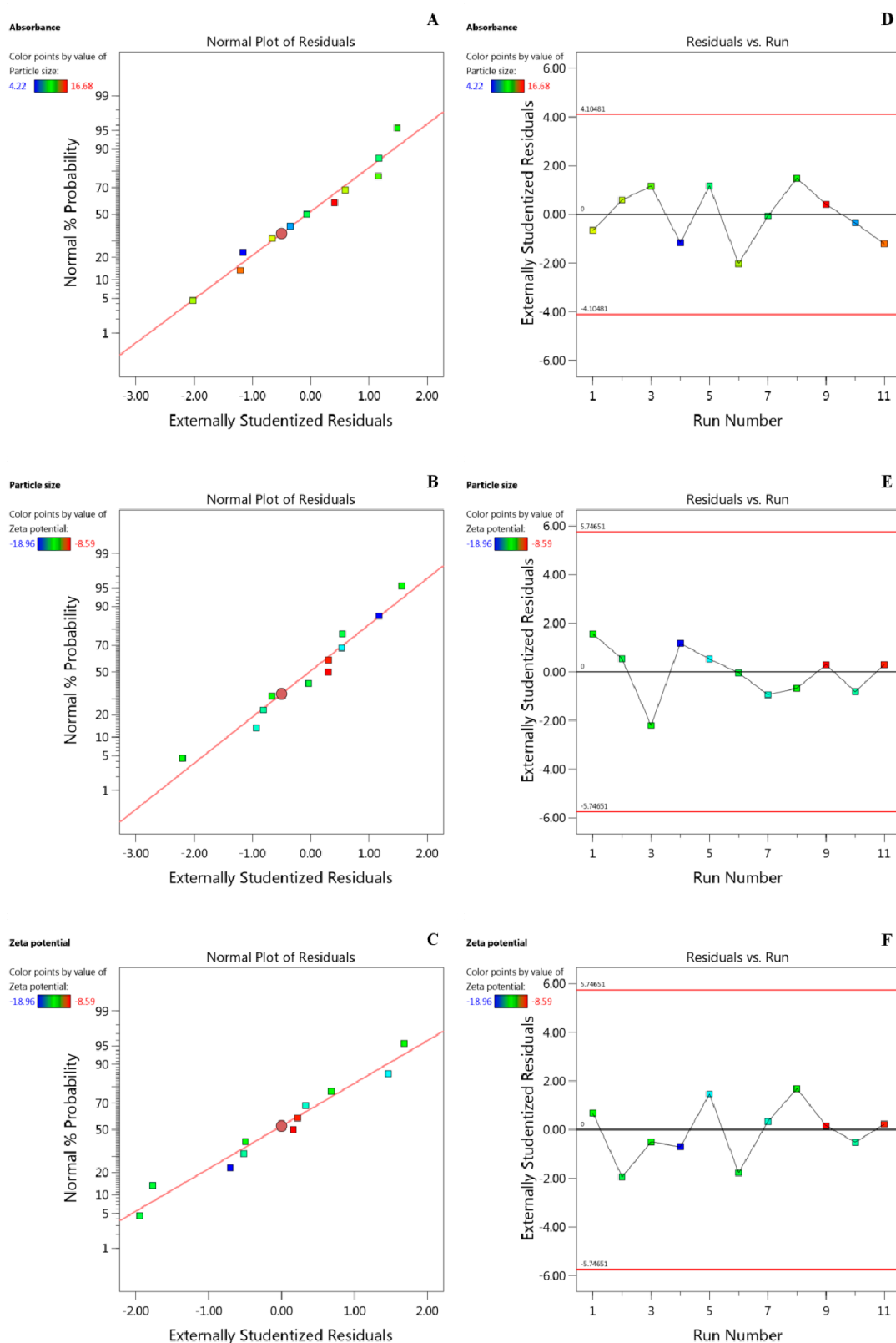


Fig. S4: Normal plots of residuals (A–C) and residual *vs* run plots (D–F) for responses Y₁ (A and D), Y₂ (B and E) and Y₃ (C and F)

Factor Coding: Actual

Overlay PlotAbsorbance
Particle size
Zeta potential

● Design Points

X1 = A

X2 = B

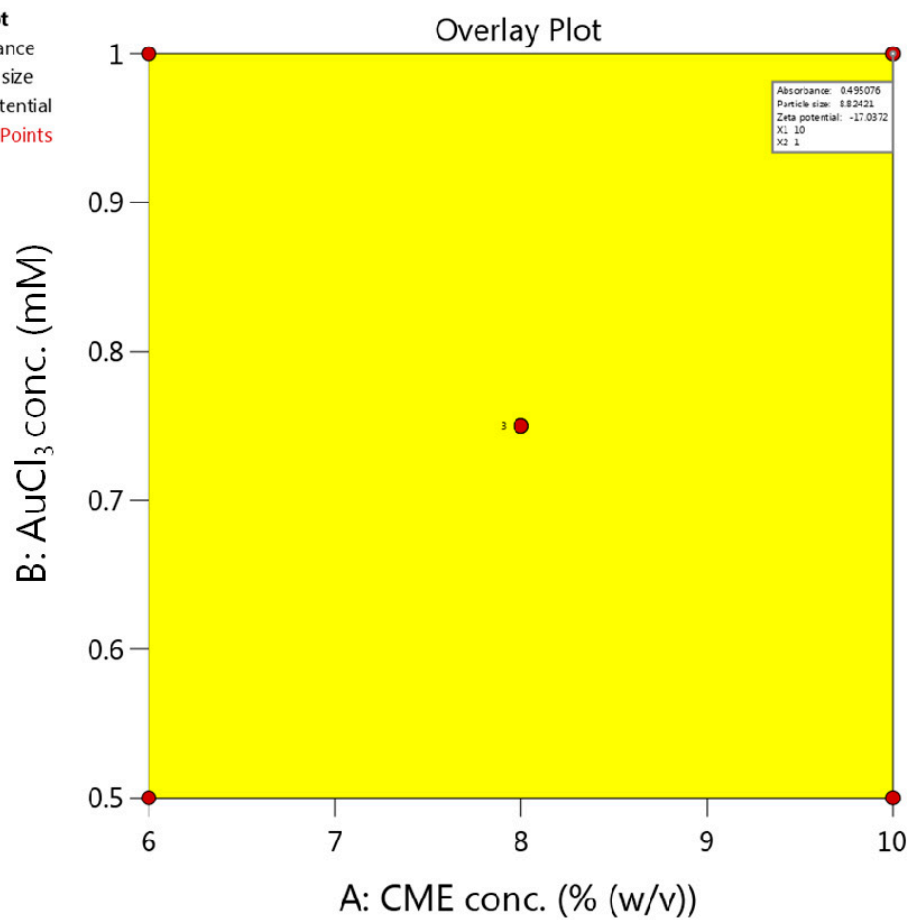


Fig. S5: Overlay contour plot

Table S1. MICs of CME and *Cord*-Au-NPs against the studied bacterial strains

Name of the organisms	MIC (µg/mL)		
	Ampicillin	CME	<i>Cord</i> -Au-NPs
<i>Proteus vulgaris</i> MTCC 426	15.00	46.88	23.44
<i>Staphylococcus epidermis</i> MTCC 435	7.50	187.50	93.75
<i>Bacillus subtilis</i> MTCC 441	15.00	187.50	23.44
<i>Rhodococcus equi</i> MTCC 2558	15.00	93.75	46.88
<i>Shigella flexneri</i> MTCC 1457	30.00	187.50	93.75
<i>Pseudomonas aeruginosa</i> MTCC 1748	7.50	93.75	46.88

Table S2: Brine shrimp lethality results of *Cor*-Au-NPs

Test drug	LC ₅₀ (µg/mL)
<i>Cor</i> -Au-NPs	192.23± 57.34
KCrO ₇ solution (positive control)	<10 µg/mL
Seawater (negative control)	No death was observed