



Review

Smart Biomimetic Nanozymes for Precise Molecular Imaging: Application and Challenges

Qiao Luo [†], Ni Shao [†], Ai-Chen Zhang, Chun-Fang Chen, Duo Wang, Liang-Ping Luo ^{*} and Ze-Yu Xiao ^{*}

The Guangzhou Key Laboratory of Molecular and Functional Imaging for Clinical Translation, The First Affiliated Hospital of Jinan University, Guangzhou 510632, China

^{*} Correspondence: tluolp@jnu.edu.cn (L.-P.L.); zeyuxiao@jnu.edu.cn (Z.-Y.X.)

[†] These authors contributed equally to this work.

Abstract: New nanotechnologies for imaging molecules are widely being applied to visualize the expression of specific molecules (e.g., ions, biomarkers) for disease diagnosis. Among various nanoplateforms, nanozymes, which exhibit enzyme-like catalytic activities in vivo, have gained tremendously increasing attention in molecular imaging due to their unique properties such as diverse enzyme-mimicking activities, excellent biocompatibility, ease of surface tenability, and low cost. In addition, by integrating different nanoparticles with superparamagnetic, photoacoustic, fluorescence, and photothermal properties, the nanoenzymes are able to increase the imaging sensitivity and accuracy for better understanding the complexity and the biological process of disease. Moreover, these functions encourage the utilization of nanozymes as therapeutic agents to assist in treatment. In this review, we focus on the applications of nanozymes in molecular imaging and discuss the use of peroxidase (POD), oxidase (OXD), catalase (CAT), and superoxide dismutase (SOD) with different imaging modalities. Further, the applications of nanozymes for cancer treatment, bacterial infection, and inflammation image-guided therapy are discussed. Overall, this review aims to provide a complete reference for research in the interdisciplinary fields of nanotechnology and molecular imaging to promote the advancement and clinical translation of novel biomimetic nanozymes.

Keywords: nanozymes; molecular imaging; magnetic resonance imaging; photoacoustic imaging; positron emission tomography; multimodal imaging



Citation: Luo, Q.; Shao, N.; Zhang, A.-C.; Chen, C.-F.; Wang, D.; Luo, L.-P.; Xiao, Z.-Y. Smart Biomimetic Nanozymes for Precise Molecular Imaging: Application and Challenges. *Pharmaceuticals* **2023**, *16*, 249. <https://doi.org/10.3390/ph16020249>

Academic Editor: Serge Mordon

Received: 21 December 2022

Revised: 1 February 2023

Accepted: 2 February 2023

Published: 7 February 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

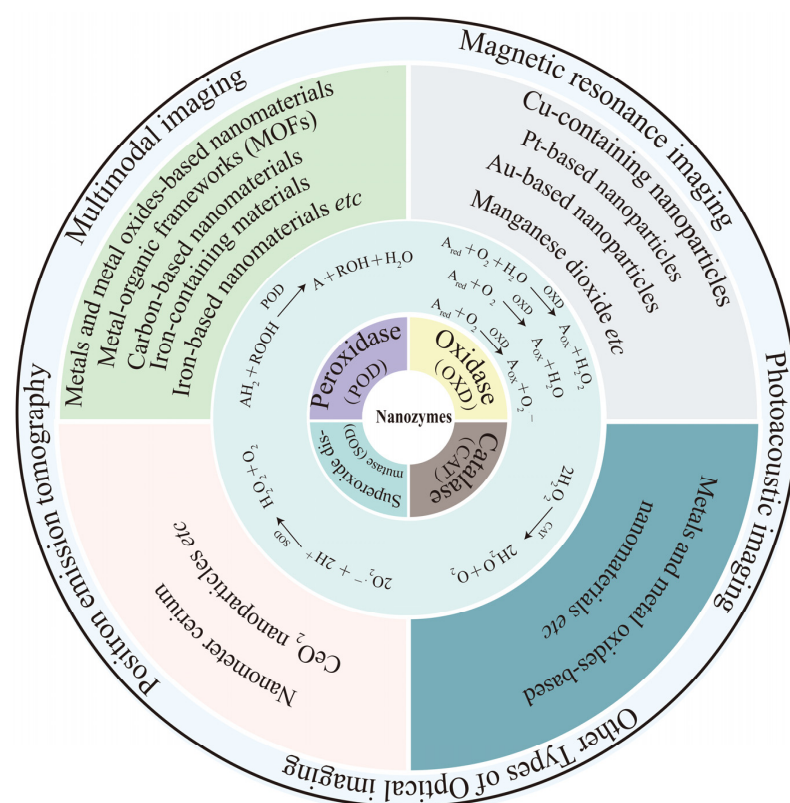
As an emerging discipline at the intersection of molecular biology and traditional medical imaging, molecular imaging (MI) promotes the development of precision medicine by visualizing specific molecular and cellular targets related to disease diagnosis and treatment [1–3]. Various imaging modalities, including positron emission tomography (PET), single-photon emission computed tomography (SPECT), optical imaging, magnetic resonance imaging (MRI), computed tomography (CT), and ultrasound imaging, have been successfully applied in the field of molecular imaging. To obtain precise MI in vivo, highly sensitive and specific molecular targeted imaging probes have aroused researchers' interest [4,5]. The ideal molecular probe can facilitate MI with enhanced metabolic stability, favorable pharmacokinetics, improved binding affinity and selectivity, better imaging ability as well as biosafety [6–8]. The determination and analysis of these parameters are crucial for the discovery, treatment, monitoring, and prognosis of early diseases [9,10]. Therefore, molecular probes are an indispensable part of a precise MI.

To date, connecting diverse imaging components (radioisotopes, fluorophores, nanoparticles) and targeted ligands (small molecules, peptides, proteins, antibodies, cells) has led to the development of imaging probes suitable for various imaging models [9,11–13]. These imaging probes have shown improved imaging performance to a certain extent. However, their development still faces some obstacles, such as the low affinity of molecular imaging

probes, low efficiency of molecular probe transcellular transport, insufficient intracellular delivery, and low concentration of the probes in the targeted tissues, leading to inaccurate molecular imaging [14,15].

The interdisciplinary nature of nanotechnology has improved the above problems [16,17]. Nanoprobes have unprecedented potential in personalized medicine, affording many advantages, including dramatic signal amplification, enhanced affinity and specificity, and bypassing of biological barriers [18,19]. Researchers have developed various nanoprobes with specific enzyme-like activities based on the inherent physiochemical properties of materials such as metal oxides [20,21]. In addition to metal oxides, nanozymes can also be composed of metals, metal-organic frameworks, semiconductors, biomimetic materials, and carbon [22,23]. These artificial enzymes based on nanomaterials (nanozymes) have been widely used in many fields, such as biosensors, immunoassays, neuroprotection, and stem cell growth [24,25]. Notably, some studies have also applied nanozymes in imaging [26]. Benefiting from the inherent characteristics of nanozymes (such as the magnetism of Fe, the X-ray absorptivity of Zr, and the optics of Au), MRI, CT, and optical imaging can be used to track the behavior of nanozymes in vivo [27–29]. Taking advantage of the catalytic properties of nanozymes, scientists can utilize fluorescent products for imaging [30]. Some studies have also suggested that nanozymes can improve imaging sensitivity [31–33].

Despite the significant advancements in nanozymes for MI applications, few studies have systematically reviewed these novel emerging nanomaterials. Hence, in this review, nanozymes, including those with peroxidase (POD)-, oxidase (OXD)-, catalase (CAT)-, and superoxide dismutase (SOD)-like properties, are collectively summarized and their applications in MI, such as with MRI, PET, etc., is comprehensively discussed (Scheme 1). In addition, the current challenges and future directions to exploit and develop nanozymes for disease imaging and diagnosis are included. Overall, this review aims to provide a complete reference for research in the interdisciplinary fields of nanotechnology and molecular imaging to better promote the advancement and clinical translation of novel biomimetic nanozymes.



Scheme 1. Schematic illustration of biomimetic nanozymes for precise molecular imaging.

2. Classification of Nanozymes

In general, nanozymes can be roughly divided into peroxidase (POD), oxidase (OXD), catalase (CAT), and superoxide dismutase (SOD) based on the substrates they catalyze [34–36]. Although various nanomaterials with different properties and physiological functions have been exploited, most of those that are being used for cancer diagnosis and theranostics may be classified as oxidoreductase nanozymes [37,38].

2.1. POD-like Nanozymes

PODs usually catalyze substrate oxidation by consuming H_2O_2 or organic peroxides [39]. Heme iron protein is the main component of most natural PODs [40]. Inspired by this, researchers have designed and developed various iron-based nanomaterials exhibiting POD-like catalytic activity [41]. For instance, the pioneering work by Gao and coworkers in 2007 indicated that Fe_3O_4 nanoparticles possessed intrinsic catalytic activity toward classical POD substrates, including 3,3',5,5'-tetramethylbenzidine (TMB), diazo-aminobenzene (DAB), and o-phenylenediamine (OPD) [42]. Later, more iron-containing materials, such as iron–sulfur compounds [41] and Prussian blue [43,44], were proven to exhibit POD catalytic activity [45–47]. From this discovery, transition metal elements such as Au, Ag, Pt, and Pd together with Fe can thus be exploited as POD-like nanozymes for diagnosis, therapeutics, and biosensing [48–50]. Moreover, metal-organic frameworks (MOFs) consisting of coordinating ions or metal clusters (such as copper and iron) of organic ligands are also emerging as next-generation nanozymes considering that the type and contents of metal components in the structure can be tuned to meet the different demands of various imaging technologies [51]. In addition, carbon-based nanomaterials are another kind of peroxidase-active nanomaterial with great significance [52].

Metals and metal oxides can have CAT-like and POD-like activities. According to research, enzyme activity will be dominated by pH or temperature. Specifically, metal nanomaterials display POD-like activity under acidic conditions. In contrast, CAT-like activity can be observed in metal nanomaterials in an alkaline environment [53]. The tumor microenvironment (TME) provides a favorable environment for tumor progression due to its unique characteristics, such as acidity, hypoxia, inflammation, and excessive hydrogen peroxide generation [54]. Accordingly, the acidic conditions of the TME have become a prominent feature in the activation of nanozyme oxidases [55]. Subsequently, the TME will play a catalytic role in decomposing H_2O_2 into O_2 or reactive oxygen species (ROS) at the tumor site, relieving hypoxia or enhancing the effectiveness of tumor treatment, including photodynamic therapy (PDT), sonodynamic therapy (SDT), and chemodynamic therapy (CDT) [56,57]. For example, the microenvironment can activate the composite nanomaterial $\text{Fe}_3\text{O}_4@\text{Carbon}@\text{Ptchlorin Ce}_6$ (MCPT Ce_6) through a POD-like catalytic process and extend PDT/PTT in acidic and H_2O_2 -rich microenvironments [58]. This nanomaterial is a multifunctional tumor theranostic agent that can be used for MRI, PTT, PDT, and catalytic therapy. MCPT Ce_6 simultaneously displayed improved photothermal performance and ROS-generating capacity due to the synergistic effects of the four components (PtNPs, Fe_3O_4 NPs, Ce_6 , and the carbon shell) of the yolk–shell structure.

2.2. OXD-like Nanozymes

Natural oxidases can catalyze the oxidation of a substrate into oxidized products and $\text{H}_2\text{O}/\text{H}_2\text{O}_2/\text{O}_2$ with the assistance of molecular oxygen (or other oxidizing reagents) [59]. Several nanomaterials have been reported to act as oxidases, such as Cu-containing nanoparticles, Pt-based nanoparticles, and manganese dioxide [60].

In Rossi's study, Au NPs could be used as a GOx (glucose oxidase) mimic to oxidize glucose into H_2O_2 and glucose delta-lactone (GDL) [61]. Since then, researchers have begun to explore the potential of Au NP enzyme-like activity in cancer treatment (such as hunger treatment and chemotherapy) [62]. Nanozymes with OXD properties can consume O_2 to produce ROS, considerably meeting the requirements for tumor treatment. For example, Gao's research has proven that dendritic mesoporous silica NPs loaded with

ultrasmall Au and Fe₃O₄ NPs, inorganic nanozymes with multiple enzyme activities, achieve high efficacy and excellent biosafety [63]. Based on the H₂O₂ concentration in the tumor, AuNPs explicitly oxidize glucose to gluconic acid and H₂O₂. Afterward, POD mimics the Fe₃O₄ NPs to catalyze H₂O₂ to release ROS (•OH), and tumor cells die via typical Fenton-catalyzed reactions.

Recently, Yuan and coworkers constructed a TME-responsive nanozyme, DOX@HMSN/Mn₃O₄(R) [64]. This nanozyme shows activities similar to OXD and CAT, alleviating hypoxic conditions in the TME and reversing the Mn³⁺/Mn²⁺ transformation by TME via in situ triggering, thus disrupting the excessive production of intrinsic redox steady-state catalysis (ROS). Under radiotherapy, high-energy X-rays can stimulate the outer electrons in the nanozymes to form photoelectrons that participate in an OXD-like enzyme reaction, thus enhancing the accumulation of ROS and the effect of radiotherapy/chemotherapy.

2.3. CAT-like Nanozymes

CAT can decompose H₂O₂ into H₂O and O₂ [65–67]. Many nanomaterials (e.g., metals and metal oxides) display CAT-like activity [68–70]. As mentioned earlier, these reported nanomaterials have CAT-like and POD-like activity, and pH or temperature will determine which type of activity dominates. Recently, Pt and Pd nanomaterials have attracted much interest due to their excellent simulation of CAT activity, near-infrared (NIR) fluorescence characteristics, and photothermal conversion ability [71]. Sun et al. took advantage of the catalase activity of Pt and Pd to design the nanoplatfrom Pd@Pt-T790 [72]. This nanoplatfrom was successfully applied to eradicate myositis induced by methicillin-resistant *Staphylococcus aureus* (MRSA). Nanozymes can increase ultrasound-driven dynamic therapy by catalyzing the generation of oxygen. This nanosystem skillfully utilizes the convertible ultrasonic enzyme activity to ensure significant oxygen accumulation at the infected site.

2.4. SOD-like Nanozymes

SOD catalyzes the conversion of superoxide free radicals (O₂•) to O₂ and H₂O₂ [73]. Nanometer cerium is one of the earliest reported nanomaterials with SOD-like activity [74,75]. CeO₂ nanoparticles have been widely recognized as candidates for superoxide dismutase [76]. Owing to the existence of the mixed valence states Ce³⁺ and Ce⁴⁺ as well as the presence of oxygen vacancies, CeO₂ shows a variety of enzyme-like catalytic activities, including those that are POD-like, CAT-like, and SOD-like [75]. For example, a hollow mesoporous Mn/Zr-codoped CeO₂ tandem nanozyme (PHMZCO-AT) regulates various enzyme activities and disrupts H₂O₂ homeostasis, ultimately achieving the therapeutic purpose of CDT [77]. Due to paramagnetic Mn²⁺ and the high atomic number of Zr, T₁-weighted MRI and high contrast X-ray computed tomography (CT) imaging of PHMZCO-AT nanozymes are feasible.

3. Nanozymes for Molecular Imaging

Early personalized imaging and diagnosis are crucial for effective cancer treatment with a better prognosis [78]. However, accurately detecting small lesions early and distinguishing and identifying benign and malignant tissues with high sensitivity and specificity are significant obstacles [79]. Therefore, it is necessary and urgent to develop suitable contrast agents with high sensitivity and affinity for malignant tissues for molecular imaging. Currently, many researchers are devoted to improving the detection sensitivity of nanozymes and optimizing the design of nanozyme-like catalytic activity, including size adjustment, composition, heteroatom doping, and specific surface modifications [80–84]. Nanozymes recognize some specific TME biomarkers to improve the diagnostic efficacy of early cancer, such as abnormal fluctuations and expression of metal ions and miRNA [85,86]. Unlike the former, nanozymes for molecular imaging have therapeutic significance in compliance with pharmacokinetics [87,88]. The distribution and activity of nanozymes are tracked with molecular imaging in vivo. Nanozymes aggregate specifically in the TME and

perform different enzyme activities to participate in catalytic biochemical reactions, which can resolve the complexity and dynamic variability of the TME and cancer treatment [89,90]. In addition, nanozymes integrate diagnosis with imaging (including MRI, CT, near-infrared (NIR) imaging, and PAI) and treatment (including PTT, PDT, and CDT) [91–95] to provide not only a personalized diagnosis but also a constructive direction for integrated diagnosis and treatment [96]. Considering the different basic principles of each imaging mode, we will focus on typical examples and the latest progress in molecular imaging of each mode's nanozymes in this section. Characteristic examples of nanozymes with diverse catalytic activities in different imaging modalities are summarized in Table 1.

Table 1. Summary of typical nanozymes applied in different imaging modalities.

	Enzyme-mimicking Activity	Imaging Modalities	Disease	Refs
Fe ₃ O ₄	POD	MRI	Inflammatory cardiovascular disease	[97]
Fe-CDs	POD	MRI	Hepatocarcinoma, Glioblastoma	[98]
Fe ₅ C ₂ @Fe ₃ O ₄	POD, CAT	MRI	Breast cancer	[99]
APMN NPs	POD, CAT	MRI/PAI	Breast cancer	[100]
PBNZs	POD, CAT, SOD	MRI/PAI	Acute renal injury	[101]
RSPN	SOD	PAI	Acute liver failure	[102]
Au ₂ Pt-PEG-Ce6	POD, CAT	PAI/CT	Cervical cancer	[103]
Pt-CuS Janus	CAT	PAI	Colon cancer	[104]
GQDzyme	POD	PAI	Nasopharyngeal carcinoma	[105]
AMP NRs	POD	PAI	Breast cancer	[106]
HSC-2	POD, CAT	PAI/FLI	Breast cancer	[107]
GNPs	POD, CAT	FLI/PAI	Breast cancer	[108]
OMCAPs@rBSA-FA@IR780	CAT	FLI	Gastric carcinoma	[109]
BSA@Au	CAT	FLI	Breast cancer	[110]
HMON-Au-col@Cu-TA	XOD	PET/FLI	Pancreatic ductal adenocarcinoma	[111]
FHNPs	POD, CAT	PET/SPECT	Inflammation	[112]
AgPd PB	CAT	PAI/FLI/CT	Cervical cancer	[113]
Ang-IR780-MnO ₂ -PLGA	POD, CAT	PAI/FLI/MRI	Glioblastoma	[114]
FeWOX NSs	POD	PAI/MRI/CT	Breast cancer	[115]
Fe ₃ O ₄ /Ag/Bi ₂ MoO ₆ NP	POD, CAT, SOD	PAI/MRI/PTI	Breast cancer	[116]

3.1. Magnetic Resonance Imaging

MRI is one of the most effective imaging tools in modern medical imaging owing to its high spatial resolution, excellent soft tissue contrast, imaging in any direction, and low damage to tissues [117,118]. It is widely used in biomedicine, such as central nervous system imaging or cardiac and kidney function evaluation [119,120]. Although MRI is currently the leading imaging technique to detect soft tissues, it is still difficult to discriminate between benign and malignant tissues due to the negligible differences caused by the long relaxation time of water protons [121,122]. Therefore, researchers introduced contrast agents to accelerate the proton relaxation rate to solve this problem, thereby increasing the contrast [123,124]. Notably, with the help of contrast medium, the sensitivity and specificity of MRI in disease diagnosis have been improved [125]. MRI contrast agents can be divided into T_1 —weighted (positive) and T_2 —weighted (negative) agents according to their effects on longitudinal T_1 or transverse T_2 relaxation [126]. Fast T_1 -weighted imaging shows bright contrast in MRI, while the opposite T_2 -weighted imaging shows dark contrast [127]. The most commonly used MRI contrast agents are paramagnetic gadolinium complexes, such as gadolinium diethylene triaminepenta acetate (Gd-DTPA) [128,129]. However, due to the relatively low relaxation characteristics of gadolinium-based contrast agents, high doses of contrast agents are needed, which may lead to serious side effects, such as nephrogenic systemic fibrosis (NSF) [130,131]. In addition, due to its inherently short cycle time, it is more difficult to obtain high-resolution images [132]. Even though conjugated macromolecules can overcome this problem with

gadolinium chelates, live ions may increase the possibility of side effects [133,134]. Therefore, it is vital to develop new MRI contrast agents that can improve the imaging sensitivity and accuracy of disease diagnosis.

Since the discovery of the POD-like activity of Fe_3O_4 , an increasing number of iron-based nanozymes (including iron-based nanomaterials and their composites) have attracted extensive attention [135]. Because of the superparamagnetism of Fe_3O_4 nanoparticles (Fe_3O_4 NPs), the relaxation time of the surrounding protons is shortened, thus providing the possibility of employing this material as T_2 -weighted MRI agent media [97,136]. However, owing to the inherent dark signal of T_2 -weighted MRI, it is impossible to accurately distinguish tumors from other low-signal areas, such as calcifications, metal deposits, or hemorrhage [98,137]. Researchers found that when the size of an Fe_3O_4 NP is less than 5 nm, it works with T_1 -weighted MRI because the reduced magnetic moment will strongly inhibit the T_2 effect [138], which provides the possibility of preparing responsive T_2/T_1 switched MRI contrast agents [139–141]. Benefiting from their catalytic activity similar to that of peroxidase, Fe_3O_4 NPs can promote iron-mediated apoptosis of tumor cells through the Fenton reaction for cancer treatment [142–144].

Yu et al. prepared a T_1/T_2 convertible MRI-guided cancer therapeutic agent based on ROS generated by $\text{Fe}_5\text{C}_2@ \text{Fe}_3\text{O}_4$ NPs (Figure 1) [99]. In endogenous tumor environments, $\text{Fe}_5\text{C}_2@ \text{Fe}_3\text{O}_4$ NPs release ferrous ions due to the low pH, which is disproportionate to the excessive H_2O_2 at the tumor site, and generate $\bullet\text{OH}$ radicals through the Fenton reaction, thus effectively acting against tumors. The high magnetic properties of $\text{Fe}_5\text{C}_2@ \text{Fe}_3\text{O}_4$ NPs make them capable of T_2 -weighted MRI. Moreover, the ionization of these NPs in an acidic environment improves the T_1 signal. This switchable T_2/T_1 process visualizes ferrous ion release and ROS generation for cancer therapy. $\text{Fe}_5\text{C}_2@ \text{Fe}_3\text{O}_4$ NPs have high efficiency and tumor specificity, showing great therapeutic potential in the TME under the guidance of MRI. Further modification of Fe_3O_4 NPs with photothermal agents or sonosensitizers may provide multimodal cancer treatment with photothermal therapy (PTT) or photodynamic therapy (PDT). For example, altering magnetic Fe_3O_4 NPs with the classical photothermal agent (AuNPs), in addition to MRI and magnetic targeting functions, can bring additional PTT capabilities and improve the accuracy of Fe_3O_4 NPs to diagnose and treat tumors. As a result, Fe_3O_4 NPs have become a novel nanoplatform for personalized tumor theranostics.

Likewise, as another transition metal element, manganese can also be exploited as an MRI contrast agent [145–147]. Manganese oxide nanoparticles (MONs) are potential T_1 -weighted MR contrast agent candidates. Mechanistically, the five unpaired electrons in the three-dimensional orbital of this ion can produce significant magnetic moments and cause the relaxation of nearby water protons [148,149]. Studies have shown that MONs can mimic the characteristics of POD, CAT, and SOD. Among various MONs, MnO_2 nanostructures have attracted considerable attention as stimuli-responsive and biodegradable materials [150,151]. In the TME, MnO_2 can react with GSH, H^+ ions, and H_2O_2 to generate Mn^{2+} ions, playing multiple roles in tumor diagnosis and treatment. MnO_2 can also be an MR probe that is specifically activated in the TME to accurately target tumors. Taking advantage of their CAT-like activity, MnO_2 nanostructures can decompose excessive H_2O_2 into O_2 in situ, relieving hypoxia and enhancing other treatments. As a drug vehicle, MnO_2 nanoshells can also be decomposed by GSH and H^+ ions in the tumor microenvironment. Thus, the loaded chemotherapeutic drugs can be unleashed in response to achieve precise treatment [152–155]. MnO_2 nanostructures are regarded as CDT agents because they catalyze H_2O_2 in the TME to produce $\bullet\text{OH}$ via Fenton-like reactions.

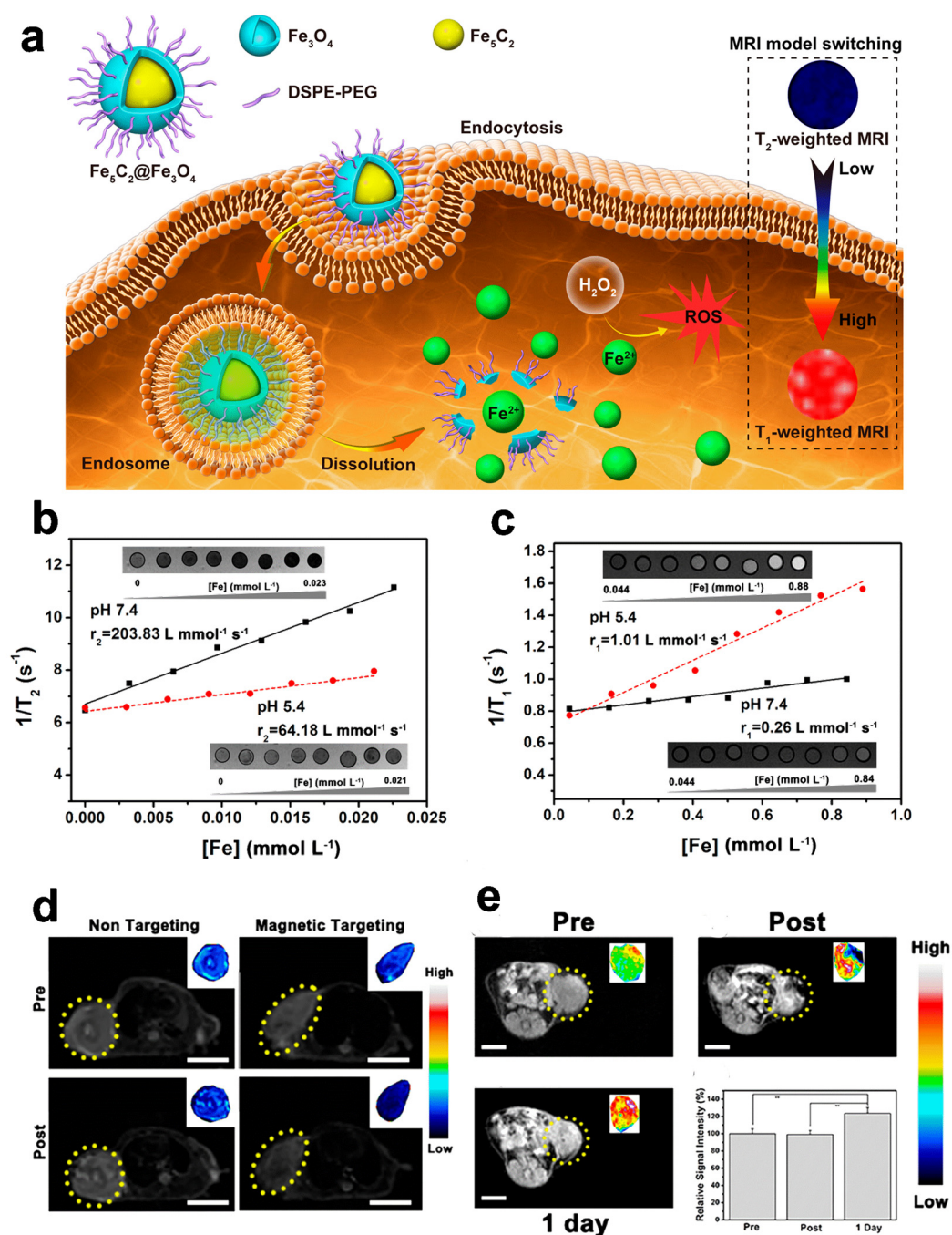


Figure 1. $\text{Fe}_5\text{C}_2@ \text{Fe}_3\text{O}_4$ NPs for MRI. (a) Schematic illustration of $\text{Fe}_5\text{C}_2@ \text{Fe}_3\text{O}_4$ NPs with Ph-responsive Fe^{2+} release, ROS generation, and T_2/T_1 signal-conversion abilities. (b,c) pH-dependent MRI mode switching of PEG/ $\text{Fe}_5\text{C}_2@ \text{Fe}_3\text{O}_4$ nanoparticles. (d) Representative T_2 —weighted MR images. (e) Representative T_1 —weighted MR images. Reproduced with permission from Ref. [99]. Copyright © 2019 American Chemical Society. ** $p < 0.05$.

For example, Huang et al. designed the multifunctional bionic thermal nanoplatform $\text{Au}@ \text{Pd}@ \text{MnO}_2$ (APMN NPs) and further coated it with a natural killer cell membrane to form a core-shell structure (Figure 2) [100]. The $\text{Au}@ \text{Pd}$ nucleus was determined to have a high-temperature effect and nanozyme catalytic activity under near-infrared (NIR) activation. The mesoporous MnO_2 shell provides MR imaging and high drug-loading ability and endows the nanoplatform with various enzyme-like activities, such as those of POD and XOD. In addition, NK-92 cell membrane camouflage endows APMN NPs with enhanced tumor-targeting ability, immune escape functions, and membrane-protein-

mediated tumor uptake characteristics. The TME-responsive MR imaging and drug-release characteristics make APMN NPs an intelligent integrated nanoplatform. The developed NPs showed high therapeutic efficacy in MCF-7 tumor-bearing mice for antitumor therapy and biosafety in histological and blood biochemical experiments.

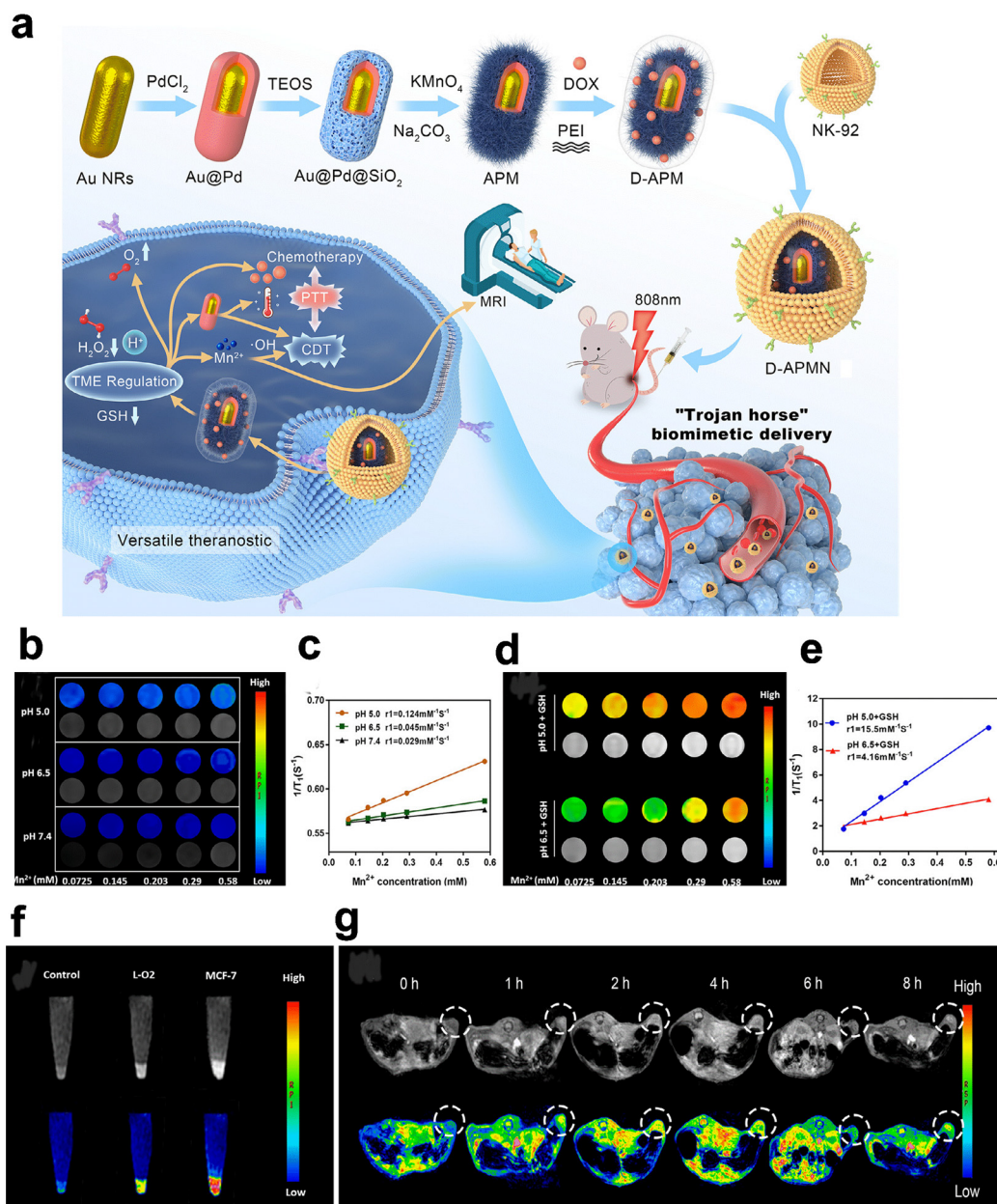


Figure 2. APMN NPs for MRI. (a) Schematic illustration of the main synthetic process of APMN NPs and the mechanism of this biomimetic theranostic nanoplatform for TME-responsive MR imaging and efficient induction of multimodal synergistic treatments. (b–e) MR images of APMN NPs under different conditions. (f) T₁-weighted MR images of APMN NPs in cells. (g) T₁-weighted MR images of MCF-7 tumor-bearing nude mice. Reproduced with permission from Ref. [100]. Copyright © 2022 Acta Materialia Inc. Published by Elsevier Ltd. All rights reserved.

Metal-organic frameworks (MOFs) containing transition elements are also emerging as MRI contrast agents [156,157]. Prussian blue (PB), as a vital MOF, is a promising material with unique physical and chemical properties [158]. The carbon-bonded Fe(II) (S = 0) and nitrogen-bonded Fe(III) (S = 5/2) in PB produce five unpaired electron $\pm C \equiv N - Fe^{3+}$ units for each Fe^{2+} , which can shorten the relaxation time of protons and

enable them to be used as contrast agents for magnetic resonance imaging (MRI) [159]. In recent years, researchers have also found that PBNPs exhibit catalytic properties such as those of POD, CAT, and SOD [160]. PBNPs can potentially treat symptoms caused by ROS due to their multienzyme activities and ROS-clearance abilities [161]. Zhang's team successfully fabricated multifunctional ultrasmall nanozymes based on PB (PB NZs) [101]. The as-prepared PB NZs exhibited efficient cellular protection from H₂O₂—induced damage in vitro because of their excellent multienzyme-mimicking capabilities to remove excessive RONS [e.g., H₂O₂, the hydroxyl radical (\bullet OH), the superoxide anion (O₂ \bullet^-), nitric oxide (NO), and peroxynitrite (ONOO $^-$)]. Based on their MR and PA imaging properties, PB NZs accumulate rapidly and efficiently in the kidneys of AKI mice. The ultrasmall size of PB NZs produced negligible systemic toxicity in mice. In addition, serum and biomarker testing, histological staining, and mouse survival studies have demonstrated that ultrasmall PB NZs have good biocompatibility. Consequently, ultrasmall PB NZs have excellent potential as a nanozyme therapy agent to treat AKI.

3.2. Photoacoustic Imaging

At present, photoacoustic imaging (PAI) has become a promising noninvasive imaging method that combines spectral selectivity from laser excitation with the high resolution of ultrasonic imaging [162,163]. Sensitive light absorption, high spatial resolution, and imaging depth are the advantages of PAI [164,165]. Compared with ultraviolet and visible light, the interaction of near-infrared light with biological tissues is relatively weak, which is more conducive to clinical diagnosis [166,167]. The near-infrared region includes the first (NIR-I, 650–995 nm) and the second (NIR-II, 1000–1700 nm) [168,169] windows. NIR-II window PAI has apparent advantages in terms of penetration depth and signal-to-noise ratio (SNR) [169,170]; thus, it has aroused increasing interest from researchers [171].

In addition to the excitation light source, the photothermal effect based on photothermal transduction agents (PTAs) is also a focus of PAI [164,172]. PTAs absorb light energy and convert it into heat to raise the temperature of the surrounding environment, and then the tissue undergoes temporary thermoelastic expansion to generate ultrasonic waves [173,174]. Eventually, the generated ultrasonic signals are gathered with broadband ultrasonic transducers and transformed into PA images [172]. Generally, PTAs can be used for PTT and PAI [175]. PTT involves increasing the temperature of the surrounding environment to cause cancer cell death through the photothermal effect [176]. Thus, PTAs lay a solid foundation for the integration of diagnosis and treatment [177]. The perfect PTA should have a high photothermal conversion efficiency, an excellent signal-to-noise ratio, and satisfactory tumor accumulation [178]. PTAs can be classified as inorganic materials or organic materials. Inorganic materials include noble metal materials (Au, Ag, Pt, Pd), metal chalcogenide materials, carbon-based nanomaterials (e.g., graphene and carbon nanotubes), and other two-dimensional (2D) materials (such as nanosheets, boron nitride, and graphite carbonitride) [179–181]. Typically, inorganic PTAs have higher photothermal conversion efficiency and better stability than organic agents. Regardless, organic PTAs are superior in terms of biodegradability and biocompatibility.

Specifically, after the laser light is absorbed by noble metal PTAs, the electrons are converted from the ground state to the excited state. Finally, they release energy via nonradiative attenuation as heat [102,182]. Additionally, these PTAs can simulate the catalytic activities of POD, OXD, CAT, etc. In the microenvironment, nanozymes have one or more enzyme activities and can effectively solve cancer treatment problems with PTT, PDT, or CDT. As proof, Wang et al. manufactured a combined treatment nanoplatfrom (Au₂Pt-PEG-Ce6), with the photosensitizer chlorin e6 (Ce6) covalently linked to convert O₂ into singlet oxygen (¹O₂), which significantly enhanced the stability of ¹O₂ and avoided its premature release in complex organisms [103]. This platform has CAT and POD dual catalytic activities, which can generate O₂ and \bullet OH to enhance PDT and CDT. Additionally, Au₂Pt-PEG-Ce6 can effectively achieve PDT and PTT under 650 nm and 808 nm laser irradiation. Furthermore, due to the strong absorption in the NIR region, Au₂Pt-PEG-

Ce6 can serve as a photothermal transition agent for NIR-responsive PTT and provide the possibility of PA and photothermal (PT) imaging. Moreover, the extraordinary X-ray attenuation capabilities of Au and Pt endow Au2Pt-PEG-Ce6 nanomaterials with the ability to be CT imaging contrast agents. Importantly, image-guided tumor therapy has confirmed that Au2PtPEG-Ce6 can significantly inhibit tumor growth without side effects to normal tissues. The imaging results were consistent and the tumor showed an enhanced signal, while only a weak signal was observed in the control group. Notably, the image-guided tumor treatment results demonstrated that Au2PtPEG-Ce6 can effectively inhibit tumor growth with almost no side effects to normal tissues. Additionally, in vivo toxicity experiments showed that Au2Pt-PEG-Ce6 has outstanding biocompatibility, making it promising for biological applications.

Liang and coworkers designed and synthesized the excellent hollow structure Pt CuS Janus (Figure 3) [104]. The hollow interior of CuS supplies a great space for TAPP molecule packing to enforce sonodynamic therapy. Moreover, Pt endowed with the catalytic property of CAT modulates the engagement of O₂ to alleviate tumor hypoxia and boost sonodynamic-therapy-induced ROS production. Significantly, the CAT-like enzyme activity of Pt can be accelerated by the heat generated under 808 nm laser irradiation to generate more O₂, thereby enhancing the effect of sonodynamic therapy. Finally, after the temperature-sensitive polymer poly(oligo(ethylene oxide) methacrylic acid co-2-(2-methoxy ethoxy)methacrylate) (p-(OEOMA-co-MEMA)) was covered with Pt-CuS NPs (Pt-CuS-P-TAPP, labeled PCPT), the nanozyme activity and drug-release rate were effectively controlled by temperature. Moreover, the nanosystems, acting as attractive PA imaging agents and NIR thermal imaging agents, were further optimized to guide the therapeutic options for cancer PA imaging. The tumor signal from CT26 tumor-bearing mice was evident and increased to a maximum value at 12 h. Under 808 nm laser irradiation, a rapid increase in tumor temperature via the effective accumulation of PCPT in tumors not only destroys tumor tissue but also ensures its potential clinical transformation due to its high biosafety.

However, PTAs based on precious metals face the problems of high cost and non-degradability, which are complex problems for clinical conversion. Thus, scientists looked for other inorganic materials for use as PTAs. Carbon-based materials with broad optical absorption and reasonable photothermal properties, such as graphene, graphene oxide, and carbon nanotubes, have attracted significant attention. The intrinsic POD-like activity of graphene quantum dot nanozymes (GQDzymes) effectively converts 2,2'-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) into its oxidized form in the presence of H₂O₂. Oxidized ABTS exhibits strong near-infrared (NIR) absorbance, rendering it an ideal contrast agent for PAI. For instance, Ding and coworkers designed a novel H₂O₂-responsive exosome-like nanozyme vesicle for PAI detection of nasopharyngeal carcinoma [105]. They developed an approach to construct exosome-like nanozyme vesicles via biomimetic functionalization of GQDzyme/ABTS nanoparticles with natural erythrocyte membranes modified with folate acid. This biomimetic decoration contributes to prolonging circulation time, improving tumor accumulation, and facilitating tumor uptake, which were superior to those exhibited by traditional nonbiological material modification strategies. Notably, due to the POD activity, light absorption, and photothermal properties of GQDzyme, the PA signal is enhanced in the H₂O₂-containing tumor environment. In addition, nanovesicles also have good biocompatibility and long-term blood circulation invisibility.

Based on Ding's research [103], Liu et al. loaded ABTS into the POD-like metal-organic framework (MOF) MIL-100. They coated the frame with polyvinylpyrrolidone (PVP) to design activated ABTS@MIL-100/PVP nanoreactors (AMP NRs) (Figure 4) [106]. The tumor microenvironment can activate these AMP NRs to display their photoacoustic imaging signal and perform photothermal therapy (PTT). In addition, the high levels of H₂O₂ in the tumor microenvironment undergoes a reaction to produce hydroxyl radicals and destroy intracellular glutathione (GSH), giving AMP NRs the ability to carry out enhanced chemodynamic therapy (ECDT). After intravenous injection of AMP NRs, 4T1

tumor-bearing mice at different tumor-growth stages (5 and 12 days after inoculation) were imaged, and the data highlighted the excellent ability of the nanoreactor to detect tumors in the early stage. Additionally, AMP NRs completely inhibited cancer growth in 4T1 tumor-bearing mice by satisfactory ECD and PTT. The low toxicity and high biocompatibility of AMP NRs ensure their clinical transformation.

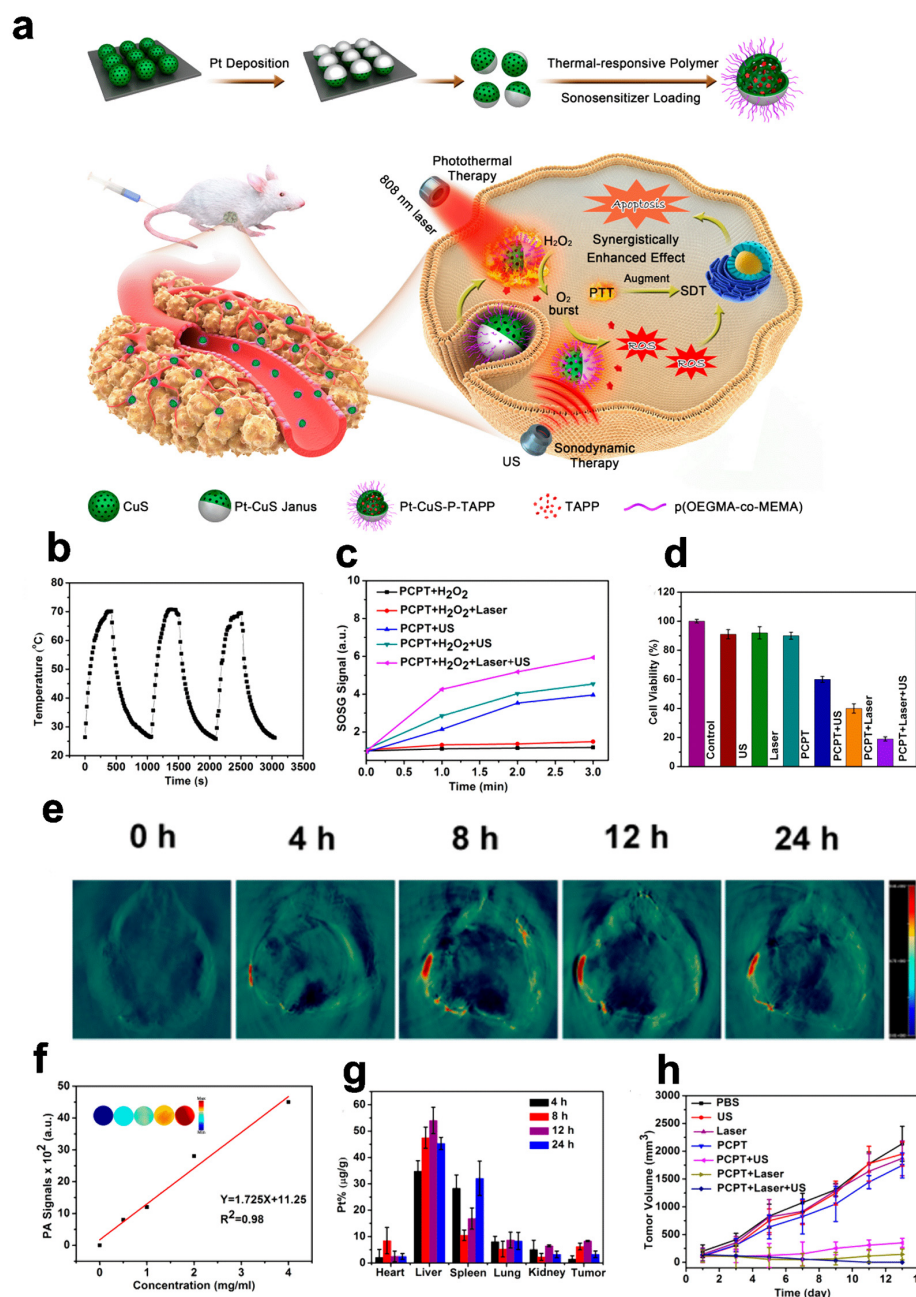


Figure 3. Pt-CuS Janus for PAI. (a) Schematic illustration of the main synthesis procedures and antitumor mechanism of PCPT. (b) Photothermal stability of the Pt-CuS solution. (c) The production of ¹O₂ in the presence of PCPT (50 ppm) under different conditions. (d) The viability of CT26 cells after different treatments. (e) In vivo PA imaging of tumor-bearing mice. (f) In vitro PA imaging of different concentrations of Pt-CuS NPs. (g) Biodistribution of Pt in the tumor and main organs after injection of PCPT. (h) Tumor volume change curves during therapy. Reproduced with permission from Ref. [104]. Copyright © 2019 American Chemical Society.

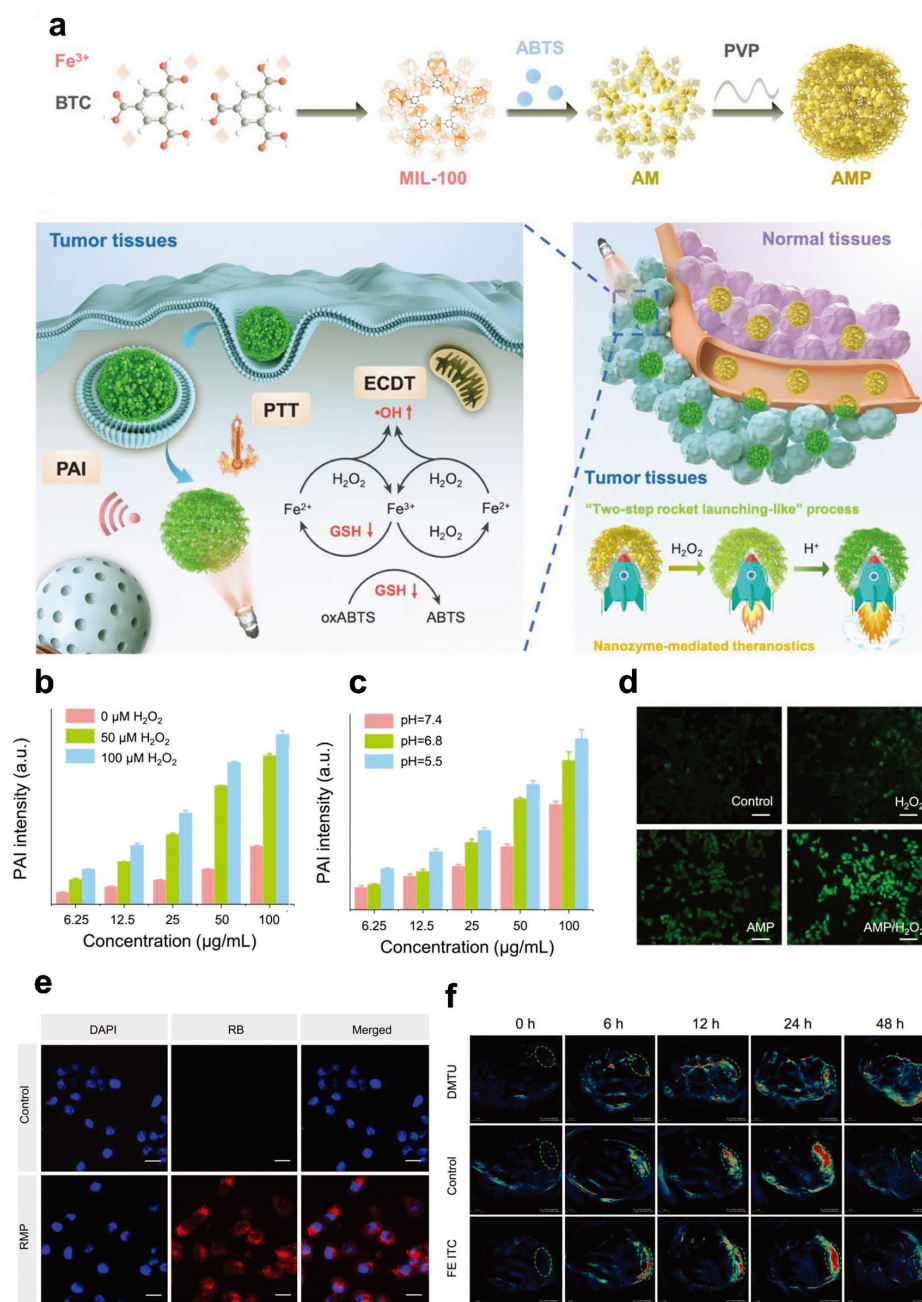


Figure 4. AMP NRs for PAI. (a) Preparation procedure and schematic illustration of AMP NRs for tumor-microenvironment-activated nanozyme-mediated therapeutics. (b) PAI signal intensity of AMP NRs with different concentrations of H_2O_2 . (c) PAI signal intensity of AMP NRs at different pH values in the presence of $100 \times 10^{-6} \text{ M H}_2\text{O}_2$. (d) Detection of ROS levels in 4T1 cells treated with different formulations. (e) CLSM images of 4T1 cells treated with AMP NPs (B@MIL-100/PVP). (f) Representative PAI images of tumors after different treatments. Reproduced with permission from Ref. [106]. Copyright © 2019 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim.

NIR-II PA imaging has been extensively discussed and studied due to its excellent accuracy in diagnosing diseases and the noninvasive antitumor performance of PTT under imaging guidance [183]. Single PAI cannot effectively provide tumor information with distinct spatial resolution and scale. NIR-II fluorescence imaging stands out in optical imaging because of its good spatial resolution, high sensitivity, and negligible background interference. Accordingly, researchers have constructed dual-modal imaging techniques by integrating NIR-II PA and NIR-II FL imaging technology to supply accurate and successful imaging

details and guide precise antitumor treatment. For instance, Zheng and coworkers developed a new nanozyme, HSC-2 (Figure 5) [107]. HSC-2 has an adjustable dual-modal NIR-II PA/NIR-II FL imaging ability to guide precise, synergistic, catalytic photothermal therapy. In this system, by adjusting the band gap and surface N-doped carbon content to change the carbon-silicon ratio in the frame, the light absorption and emission capabilities in the NIR-II window were optimized simultaneously. Due to its optimum silicon-carbon ratio, HSC-2 produced strong NIR-II PA absorption and excellent NIR-II fluorescence emission performance. As a dual-modal imaging probe, HSC-2 not only detects deep tumor tissues but can also display comprehensive information on tumors with high resolution. The POD-like property of HSC-2 can transform endogenous H_2O_2 in tumor areas into highly toxic ROS ($\bullet\text{OH}$ and $\text{O}_2\bullet^-$). Additionally, because of its excellent photothermal conversion efficiency and POD-like catalytic ability, HSC-2 generates stable high temperatures and sufficient ROS in tumor tissues to produce antitumor effects.

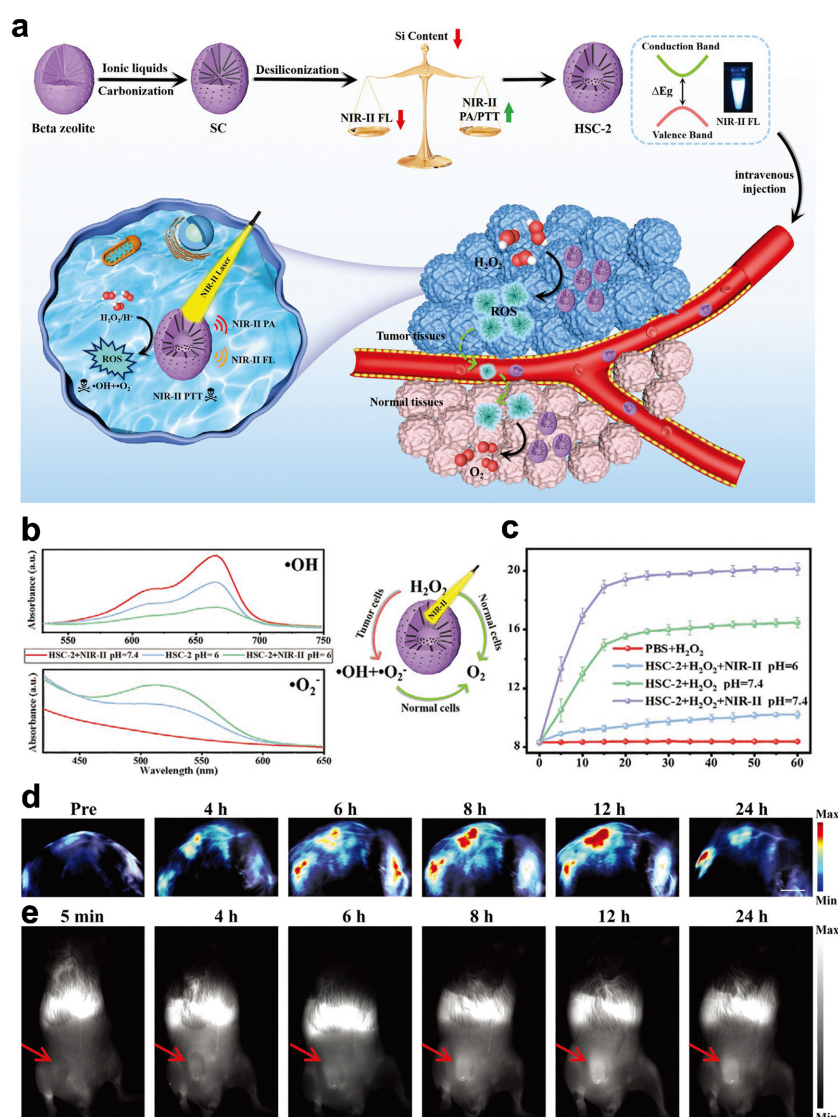


Figure 5. HSC-2 for NIR-II PA/NIR-II FL imaging. (a) Schematic illustration of adjustable photoacoustic/fluorescence image-guided photothermal/catalytic therapy in the NIR-II window. (b) POD-like activity and (c) CAT-like activity of HSC-2 in different systems over time. (d) PA images of tumors in mice at different time points after intratumoral injection of HSC-2 (808 nm laser (0.3 W cm^{-2}) irradiation; 1064 nm). (e) NIR-II FL images of 4T1 tumor-bearing mice after systemic administration of HSC-2 (1000 LP and 100 ms). Reproduced with permission from Ref. [107]. Copyright © 2021 Wiley-VCH GmbH.

3.3. Other Types of Optical Imaging

Optical imaging is an indispensable component of molecular imaging [184]. In recent years, the benefits of optical imaging, such as high sensitivity, great cost-effectiveness, nonionizing effects, and real-time imaging capabilities, have attracted much attention in various studies of *in vivo* and *in vitro* systems [185]. Nevertheless, optical imaging *in vivo* has many limitations caused by light scattering, intrinsic fluorescence, and absorption by adjacent tissues, water, and lipids [186,187]. Hence, some advanced imaging technologies (e.g., fluorescence, bioluminescence, diffusion optical tomography, and optical coherence tomography) have been developed to improve the status quo [188–190]. In particular, near-infrared fluorescence (NIRF) imaging is a current method that is widely used for imaging small animals *in vivo* [191]. This section will focus on NIRF imaging probes.

The primary principle of fluorescence imaging is that external light excites specific fluorophores and a susceptible charge-coupled device camera is used to detect emission [192]. The fluorophores can be endogenous molecules, such as hemoglobin, or exogenous molecules, such as synthetic optical probes (e.g., fluorescein isothiocyanate and rhodamine) [193]. Additionally, because optical probes based on nanozymes were initially designed to be turned off, they can only respond to specific signals when subjected to complex microenvironments [194]. Therefore, this kind of optical probe imaging has the advantages of low background noise and a high signal-to-noise ratio.

In Zhang's research, smart macrophages loaded with GOx nanoparticles expressed by IR820 macrophages were successfully constructed for synergistic photothermal activation hunger therapy in tumors (Figure 6) [108]. Because macrophages have excellent phagocytosis abilities, H₂O₂-sensitive GOx nanozymes were loaded onto the macrophages to load drugs effectively. Second, macrophages carrying therapeutic agents naturally accumulate at the tumor site due to their inherent tumor tropism. This process creates the fluorescence imaging ability obtained by the photosensitizer IR820 to track the transportation process. Finally, the external laser and special TME trigger the spatiotemporal unpacking of the loaded therapeutic agent at the tumor site. Under fluorescence guidance, the GOx enzyme activity is no longer shielded, and starvation treatment begins. At this time, positive feedback is initiated in the tumor microenvironment, and the newly generated H₂O₂ further induces the activation of GNPs. In addition, laser-triggered PTT induces apoptosis of tumor cells for local tumor hyperthermia, accelerates the decomposition of H₂O₂ into O₂, and enhances GOx enzyme activity for hunger therapy. Therefore, IRG@RC plays a variety of roles, such as those that promote laser-induced drug release and activation, the response of the tumor microenvironment, and circular amplification characteristics, realizing synergistic PTT and hunger therapy *in vitro* and *in vivo*.

The intrinsic catalytic properties of noble metal nanomaterials have been combined with different tumor therapies [109]. In addition, noble metals have optical imaging abilities due to their light stability and strong light signal. Researchers developed an image-guided treatment to overcome the dilemma of tumor chemotherapy tolerance. For example, approximately 100 kinds of NIR-II gold nanoclusters smaller than 3 nm have drawn substantial awareness for their intrinsic FL emission, renal clearance, and high biocompatibility. Along with their FL imaging capability, Au-NCs may also produce toxic ¹O₂ under NIR laser irradiation to enhance PDT to obliterate cancer cells and bacteria. Au-NCs exhibit CAT-like qualities to decompose the endogenous hydrogen peroxide (H₂O₂) in tumor tissue into O₂ to relieve tumor hypoxia. Hence, Dan et al. designed a new photosensitizer to improve PDT by utilizing the NIR-II fluorescence property and CAT activity of a gold NC (BSA@Au) [110]. Because of its bright NIR-II fluorescence, BSA@Au can pinpoint the tumor location with a high signal background ratio (SBR = 7.3) in 4T1 tumor-bearing mouse models and conduct efficient PDT. Second, the biological distribution and metabolic pathway of BSA@Au *in vivo* ensure safety for clinical conversion. Compared with the control group, the survival time of tumor-bearing mice treated with the BSA@Au-based PDT strategy was five times longer. Furthermore, BSA@Au-based PDT can protect against bacterial infections.

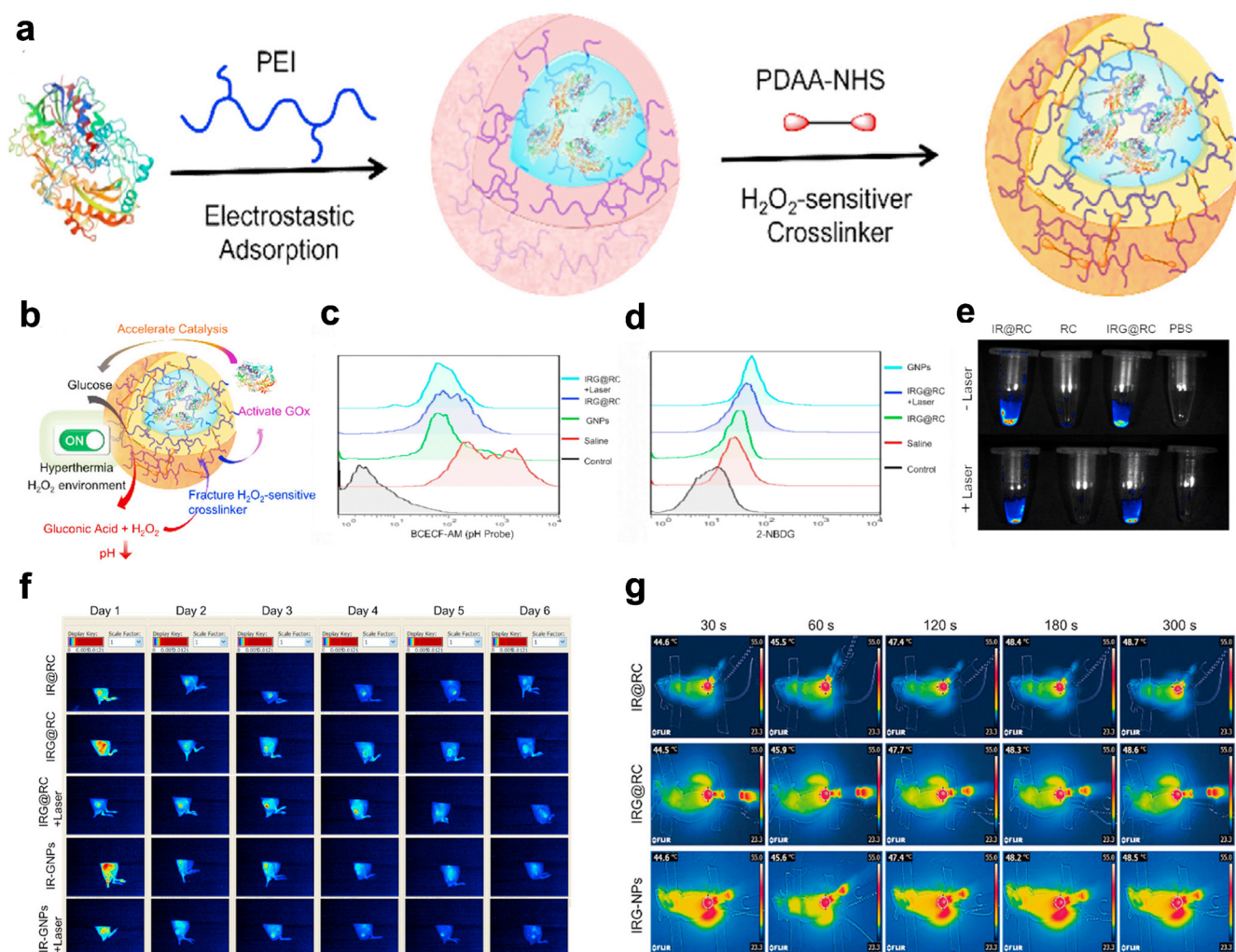


Figure 6. GNPs for FLI and infrared thermography. (a) Fabrication process of GNPs. (b) Schematic illustration of the catalytic activity of GNPs. (c,d) H_2O_2 content in tumor tissue determined with an H_2O_2 detection kit. Flow cytometry images of the pH probes BCECF-AM (c) and 2-NBDG (d) to determine the cellular pH value and glucose consumption, respectively. (e) Ex vivo fluorescence images of RC, IR@RC, and IRG@RC with/without laser irradiation. (f) In vivo fluorescence images of tumor-bearing mice. (g) Thermal imaging at the tumor site after laser irradiation by in vivo infrared thermography. Reproduced with permission from Ref. [108]. Copyright © 2022 The Authors. Published by Elsevier Ltd.

3.4. Positron Emission Tomography

PET has been widely used in clinical practice for nearly half a century due to its extremely high sensitivity, excellent penetration depth, and signal that can be quantitatively analyzed [195]. The positrons emitted by the accumulated radionuclides in living subjects are annihilated and detected [196]. PET scanners detect isotopes in deep tissues at picomolar concentrations [197]. This unique sensitivity can be used to obtain quantitative information from drugs that have accumulated at target sites [198]. In addition, the amount of nanomaterial needed for PET imaging is minimal, and the substance will not interfere with biological systems and is unlikely to induce toxic reactions [199]. Combined imaging modes, including PET/CT and PET/MRI, have been explored to overcome inherent restrictions, such as the lack of anatomical information and low resolution [200]. The joint development of PET and nanotechnology has mutual benefits [201]. The application of nanozymes in PET is expected to become unprecedented thanks to their unique physical and chemical properties [197]. Furthermore, PET is an excellent choice for scientists

to optimize the pharmacokinetics, intratumoral penetration, tumor bioavailability, and cargo-delivery mechanisms of nanoparticulate agents [202,203].

Nanozymes that have become PET probes include iron oxide nanoparticles, manganese oxide nanoparticles, Au nanoparticles, and graphene oxide nanoparticles [204,205]. Gold nanoparticles (Au NPs) are glucose OXD-like nanozymes that can efficiently catalyze glucose into H_2O_2 and gluconic acid. However, suppose that a large amount of surfactant is added during the preparation process to stabilize the ultrasmall AuNPs. In this case, the ultrasmall AuNPs cannot be used directly for biomedical applications. Additionally, these small Au NPs are rapidly cleared from systemic circulation by the kidneys. Therefore, an ideal nanocarrier is needed to immobilize ultrasmall Au NPs to ensure their biological safety and catalytic effect in vivo. Li et al. reported the in situ design of polymeric hollow mesoporous organosilicon nanoparticles (HMONs) to immobilize ultrasmall Au NPs and provide CDT with self-supplied H_2O_2 (Figure 7) [111]. Additionally, a Cu^{2+} tannic acid complex was deposited on the surface of the HMONs to initiate a Fenton-like reaction and convert the self-provided H_2O_2 into $\bullet\text{OH}$, a highly toxic ROS. Last, collagenase (Col), which can degrade collagen I fibers in the extracellular matrix (ECM), was loaded into the biocatalytic nanoreactor to increase the permeability of the HMONs and O_2 to enhance PDT. To determine the targeting effects of the nanomaterial HMON-Au-Col@Cu-TA-PVP in vivo, researchers performed PET imaging and NIR-II imaging. The results showed that the nanoprobe was clustered at the tumor site and reached its peak value in 24 h. The determined biological distribution in vivo lays a foundation for the clinical transformation of this nanomaterial. After CDT, approximately 80% of the pancreatic cancer xenograft tumor model animals experienced rapid tumor regression and long-term tumor-free survival.

Recently, Yuan et al. radiolabeled FHNPs using a thermally induced radiolabeling strategy (PET $^{89}\text{Zr}^{4+}$ or $^{64}\text{Cu}^{2+}$, SPECT $^{111}\text{In}^{3+}$) for PET or single photon emission computed tomography (SPECT) imaging [112]. Feraheme nanoparticles (FHNPs), a very small superparamagnetic iron oxide, have been widely used in the treatment of iron anemia (due to its slow release of ionic iron in acidic environments) and MRI contrast agents. In addition, injected FHNPs are internalized by monocytes and serve as MRI biomarkers for the pathological accumulation of monocytes in diseases. PET/SPECT has essential benefits for FHNPs imaging over MRI due to the low iron content and the increased ability to quantify the NP concentration in tissues. Radioactively labeling FHNPs maintains the physical and biological characteristics of the NPs. ^{89}Zr -FHNPs showed slow accumulation in mouse lymph nodes and inflammatory sites, which were similar to the MRI uptake results. Finally, by comparing the biological distribution of radiolabeled FH with that of $^{89}\text{Zr}^{4+}$ or $^{64}\text{Cu}^{2+}$ cations, the in vivo stability of FH can be determined. Radionuclide-labeled FHNPs are functional radioactive nanomaterials for lymph node mapping and PET imaging of inflammation and related inflammatory diseases.

Single imaging technologies cannot meet the current needs of personalized diagnosis and treatment; however, multimodal imaging can be used to compensate for the deficiencies [206]. Distinct imaging technologies combine multiple imaging modalities to overcome the intrinsic drawbacks of individual imaging technologies and convey more comprehensive and specific imaging information [207,208]. Until now, PET/CT has been the only multimodal molecular imaging strategy demonstrated to be clinically transformable. Other bimodal modalities, such as MRI-optics, MRI-PET/SPECT, MRI-CT, MRI-MPI, MRI-MMUI, and MRI-MPA, are still in the preclinical stage and should be further studied. With the continuous development of multifunctional imaging probes, the robust expansion of multimodal imaging is inevitable [209–212]. Nanozymes with various composite materials have high imaging efficiency under endogenous and exogenous stimuli and can enhance treatment [113,114,213].

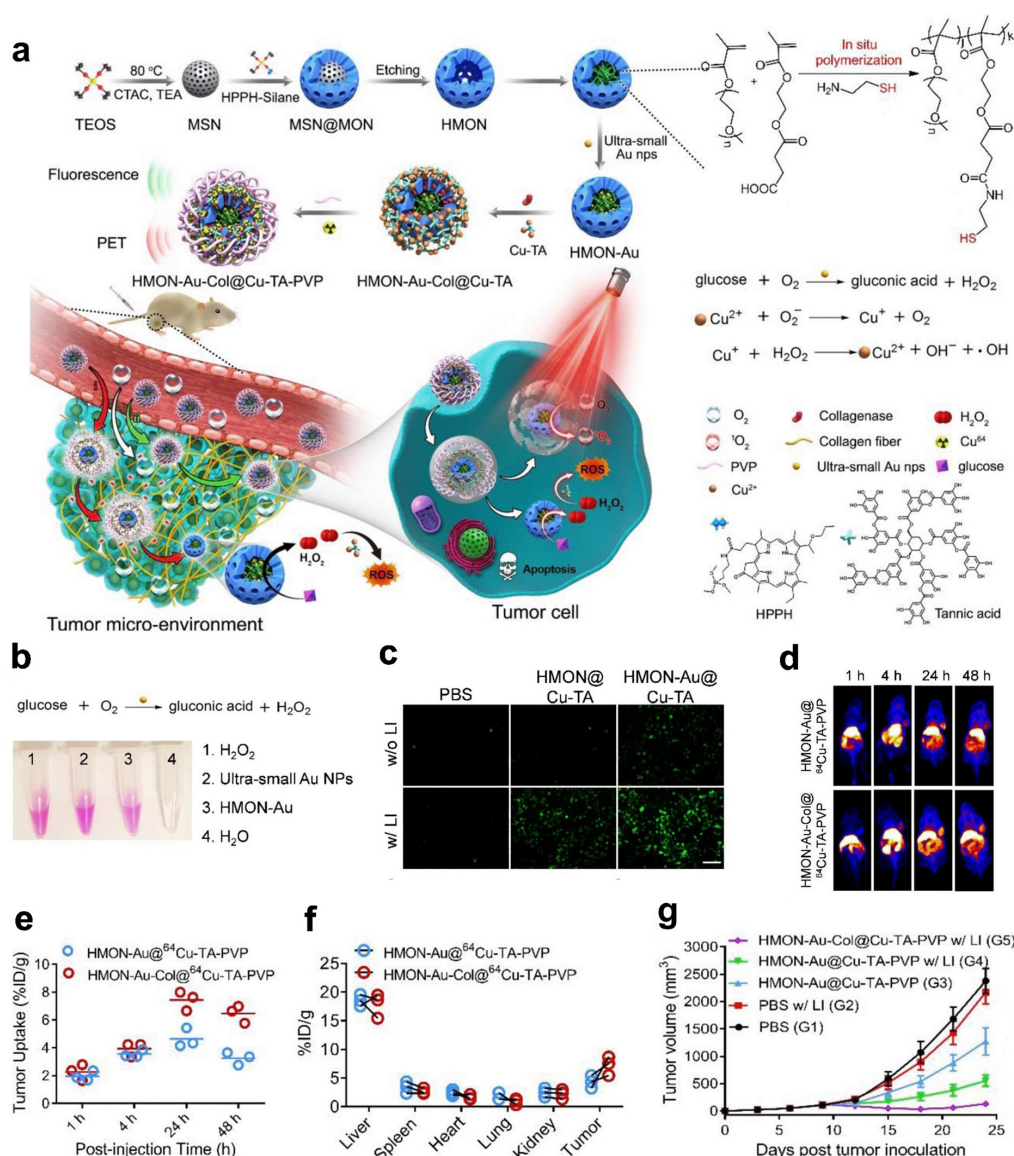


Figure 7. HMONs-Au-Col@⁶⁴Cu-TA-PVP for PET. (a) Schematic showing the process of preparing the in situ polymerized hollow mesoporous organosilica biocatalytic nanoreactor for synergistic PDT/CDT. (b) Detection of H₂O₂ production after the indicated treatment detected with a hydrogen peroxide assay kit. (c) Fluorescence images of ROS generation by BxPC-3 cells after the indicated treatment in the culture medium with glucose. (d) Representative in vivo PET imaging of tumor-bearing mice. (e) Quantification of HMON-Au@⁶⁴Cu-TA-PVP and HMON-Au-Col@⁶⁴Cu-TA-PVP tumor uptake. (f) Quantification of the PET signal intensities in the main organs of tumor-bearing mice. (g) The average tumor growth curves after treatment. Reproduced with permission from Ref. [111]. Copyright © 2019 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim.

3.5. Multimodal Imaging

In Gong's research, a unique, favorably efficient, and stable POD-mimicking nanozyme, FeWOX, was constructed (Figure 8) [115]. The flaky structure with considerable exposure of iron atoms and positively distributed oxygen vacancies (catalytic sites) on its surface endow FeWOX nanozymes with significant activity to decompose H₂O₂ into hydroxyl radicals (•OH). In addition, based on its effective enzyme activity and sheet structure, the FeWOX nanozyme was used to construct a ratiometric nanoprobe (FeTIR) based on H₂O₂-activated nanozymes, which coanchored TMB and IR780 for PA imaging and displayed high sensitivity and stability. The subcutaneous 4T1 xenotransplantation tumor model and LPS-induced inflammation

model proved that the obtained FeTIR nanoprobe had excellent endogenous H_2O_2 ratio measurement PA imaging performance in vivo after local injection. Second, FeTIR mainly works because the IR780 dye becomes a fluorescent probe. Moreover, the CT imaging ability comes from vital X-ray attenuation of tungsten (W). Finally, FeTIR can also be used as an MR T_2 -weighted imaging contrast agent due to the presence of iron. This probe integrates multiple imaging functions into a single nanoplatform. Different imaging modes can verify the specificity of the probe to target tumors and compensate for the inherent defects of the other's imaging methods.

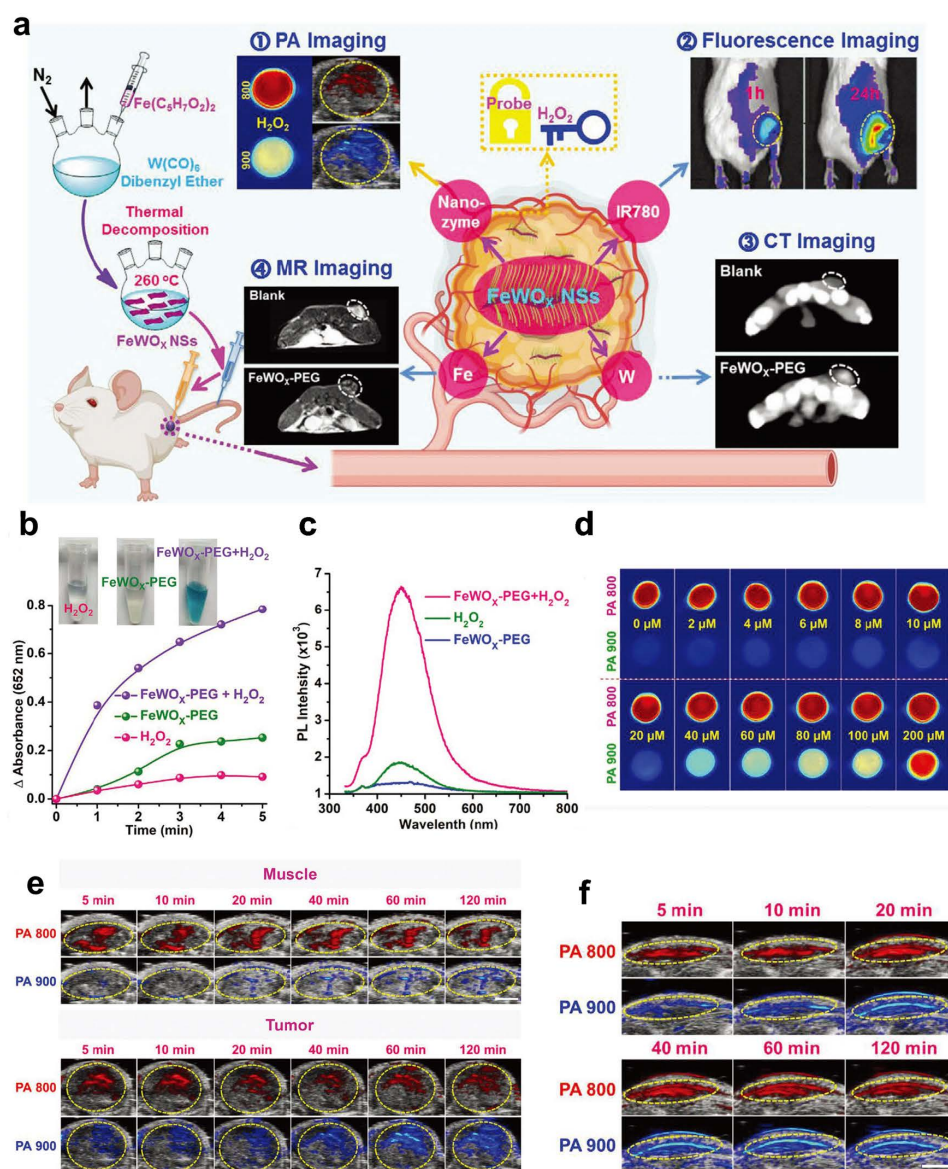


Figure 8. FeTIR for multimodal imaging. (a) Schematic illustration showing the preparation of oxygen-deficient bimetallic oxide FeWO_x nanosheets as peroxidase-like nanozymes for sensing tumors via multimodal imaging. (b) Comparison of TMB oxidation by FeWO_x-PEG, H₂O₂, and FeWO_x-PEG/H₂O₂. (c) Fluorescence spectra obtained from different treatment groups containing H₂O₂, FeWO_x-PEG, and FeWO_x-PEG/H₂O₂. (d) In vitro ratiometric PA imaging at different concentrations of H₂O₂ (0–200 $\times 10^{-6}$ M) with the FeTIR nanoprobe. (e) In vivo ratiometric PA imaging of muscle and 4T1 tumor tissues of mice after injection of the FeTIR nanoprobe. (f) In vivo ratiometric PA imaging of inflammation sites in mice after FeTIR nanoprobe injection. Reproduced with permission from Ref. [115]. Copyright © 2020 Wiley-VCH GmbH.

Currently, the types of nanozymes are limited, their efficiency is usually impaired, and their presence is not sustainable in the tumor microenvironment (TME) [214,215]. For example, some nanozymes mimic POD and CAT to assist with CDT or PDT, but the supply of H_2O_2 in the TME is limited and not maintainable and their efficiency is usually compromised [216]. Even if combined treatment with other therapies is carried out, the use of nanozymes is not practical because of the strong side effects, high cost, and low efficiency [217]. For instance, Cao et al. designed unprecedented “integrated” $Fe_3O_4/Ag/Bi_2MoO_6$ nanoparticles (FAB-NPs), which are highly efficient and specific and have few side effects [116]. FAB-NPs have high POD, CAT, SOD, glutathione oxidase (GSHOD), and photodynamic activity. The doping of Fe_3O_4 and AgNPs gives Bi_2MoO_6 strong NIR-II absorption, enhanced photocatalytic activity, and ferromagnetic and photothermal effects. These properties enable MR, PA, and PT imaging to guide nanocatalytic therapy. The multimodal imaging results showed that FAB-NPs accumulated specifically at the 4T1 breast cancer tumor site and increased with time. After laser irradiation twice a day, the tumor disappeared entirely within 16 days. The blood biochemical and complete blood group analyses after treatment showed no abnormalities. In vitro and in vivo experiments proved that this “integrated” nanoparticle system can achieve synergistic CDT, PDT, and PTT. Therefore, the self-enhancement from cascade nanocatalytic reactions and various nanozyme activities can become a new direction of nanozyme research.

4. Conclusions and Outlook

As a unique functional material, nanozymes have been extensively explored and have shown remarkable benefits and potential applications in the biomedical field, mainly diagnosis and treatment. Nanozyme-integrated diagnosis and therapy have unprecedented potential to achieve precise treatment of glioblastoma, breast cancer, hepatocarcinoma, colon cancer, pancreatic ductal adenocarcinoma, cervical cancer, and nasopharyngeal carcinoma. Due to their inherent physical properties, nanozymes can react to external stimuli, which is conducive to regulating their biological behaviors and processes in vivo (e.g., targeted imaging, treatment, and drug release). Adjustment of the metal ion proportion, changes in enzyme catalytic activity, and other types of structural adjustments can specifically enhance imaging sensitivity. As rising stars, nanozymes have made precision medicine a reality.

Nevertheless, flourishing nanozymes still experience many challenges.

1. Poor dispersion, uncontrollable precipitation after surface modification, limited types of catalytic activity, poor substrate selectivity, and potential nanotoxicity due to the drawbacks of current preparation technologies and methods.
2. Systemic toxicity derived from the overaccumulation of metal elements; for instance, the direct toxic reaction caused by gold metal overaccumulation after long-term application and the indirect toxic response, such as ion-induced oxidative stress, DNA damage, and normal tissue apoptosis. Reducing potential adverse side effects and improving the accuracy and efficacy of diagnosis and treatment are significant problems that must be solved for the viable and sustainable development of metal nanozymes.
3. There is a lack of a comprehensive monitoring and evaluation system for nanozymes after administration in living systems, such as histocompatibility, blood compatibility, immunogenicity, biological distribution, cytotoxicity, in vivo uptake, and metabolism. These factors affect the effective clinical conversion of nanozymes.

Author Contributions: Z.-Y.X. and L.-P.L. designed and supervised the work; Q.L., N.S. and Z.-Y.X. wrote the original draft; A.-C.Z., C.-F.C. and D.W. edited the manuscript. All authors discussed and reviewed the final manuscript; All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the National Natural Science Foundation of China (82102005, 82271943 and 81971672), the Fundamental Research Funds for the Central Universities (21622102), and the Medical Joint Fund of Jinan University (YXJC2022008).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Massoud, T.F.; Gambhir, S.S. Molecular Imaging in Living Subjects: Seeing Fundamental Biological Processes in a New Light. *Genes Dev.* **2003**, *17*, 545–580. [\[CrossRef\]](#)
- James, M.L.; Gambhir, S.S. A Molecular Imaging Primer: Modalities, Imaging Agents, and Applications. *Physiol. Rev.* **2012**, *92*, 897–965. [\[CrossRef\]](#) [\[PubMed\]](#)
- Böhmer, V.I.; Szymanski, W.; Feringa, B.L.; Elsinga, P.H. Multivalent Probes in Molecular Imaging: Reality or Future? *Trends Mol. Med.* **2021**, *27*, 379–393. [\[CrossRef\]](#) [\[PubMed\]](#)
- van Duijnhooven, S.M.J.; Robillard, M.S.; Langereis, S.; Grüll, H. Bioresponsive Probes for Molecular Imaging: Concepts and in Vivo Applications: Bioresponsive Molecular Imaging Probes. *Contrast Media Mol. Imaging* **2015**, *10*, 282–308. [\[CrossRef\]](#) [\[PubMed\]](#)
- Reynolds, F.; Kelly, K.A. Techniques for Molecular Imaging Probe Design. *Mol. Imaging* **2011**, *10*, 407–419. [\[CrossRef\]](#) [\[PubMed\]](#)
- Chen, K.; Chen, X. Design and Development of Molecular Imaging Probes. *CTMC* **2010**, *10*, 1227–1236. [\[CrossRef\]](#)
- Nerella, S.G.; Singh, P.; Sanam, T.; Digwal, C.S. PET Molecular Imaging in Drug Development: The Imaging and Chemistry. *Front. Med.* **2022**, *9*, 812270. [\[CrossRef\]](#)
- Bouleau, A.; Lebon, V.; Truillet, C. PET Imaging of Immune Checkpoint Proteins in Oncology. *Pharmacol. Ther.* **2021**, *222*, 107786. [\[CrossRef\]](#)
- Massoud, T.F.; Gambhir, S.S. Integrating Noninvasive Molecular Imaging into Molecular Medicine: An Evolving Paradigm. *Trends Mol. Med.* **2007**, *13*, 183–191. [\[CrossRef\]](#)
- Hussain, T.; Nguyen, Q.T. Molecular Imaging for Cancer Diagnosis and Surgery. *Adv. Drug Deliv. Rev.* **2014**, *66*, 90–100. [\[CrossRef\]](#)
- Lee, S.; Xie, J.; Chen, X. Peptides and Peptide Hormones for Molecular Imaging and Disease Diagnosis. *Chem. Rev.* **2010**, *110*, 3087–3111. [\[CrossRef\]](#) [\[PubMed\]](#)
- Wang, W.; Hu, Z. Targeting Peptide-Based Probes for Molecular Imaging and Diagnosis. *Adv. Mater.* **2019**, *31*, 1804827. [\[CrossRef\]](#)
- He, H.; Zhang, X.; Du, L.; Ye, M.; Lu, Y.; Xue, J.; Wu, J.; Shuai, X. Molecular Imaging Nanoprobes for Theranostic Applications. *Adv. Drug Deliv. Rev.* **2022**, *186*, 114320. [\[CrossRef\]](#)
- Zhang, Y.; Zhang, G.; Zeng, Z.; Pu, K. Activatable Molecular Probes for Fluorescence-Guided Surgery, Endoscopy and Tissue Biopsy. *Chem. Soc. Rev.* **2022**, *51*, 566–593. [\[CrossRef\]](#)
- Li, K.C.P.; Pandit, S.D.; Guccione, S.; Bednarski, M.D. Molecular Imaging Applications in Nanomedicine. *Biomed. Microdevices* **2004**, *6*, 113–116. [\[CrossRef\]](#) [\[PubMed\]](#)
- Liu, G.; Swierczewska, M.; Lee, S.; Chen, X. Functional Nanoparticles for Molecular Imaging Guided Gene Delivery. *Nano Today* **2010**, *5*, 524–539. [\[CrossRef\]](#) [\[PubMed\]](#)
- Toy, R.; Bauer, L.; Hoimes, C.; Ghaghada, K.B.; Karathanasis, E. Targeted Nanotechnology for Cancer Imaging. *Adv. Drug Deliv. Rev.* **2014**, *76*, 79–97. [\[CrossRef\]](#) [\[PubMed\]](#)
- Thakor, A.S.; Gambhir, S.S. Nanooncology: The Future of Cancer Diagnosis and Therapy. *CA A Cancer J. Clin.* **2013**, *63*, 395–418. [\[CrossRef\]](#)
- Cai, W.; Chen, X. Nanoplatforams for Targeted Molecular Imaging in Living Subjects. *Small* **2007**, *3*, 1840–1854. [\[CrossRef\]](#)
- Golchin, J.; Golchin, K.; Alidadian, N.; Ghaderi, S.; Eslamkhah, S.; Eslamkhah, M.; Akbarzadeh, A. Nanozyme Applications in Biology and Medicine: An Overview. *Artif. Cells Nanomed. Biotechnol.* **2017**, *45*, 1069–1076. [\[CrossRef\]](#)
- Estelrich, J.; Busquets, M.A. Prussian Blue: A Nanozyme with Versatile Catalytic Properties. *IJMS* **2021**, *22*, 5993. [\[CrossRef\]](#) [\[PubMed\]](#)
- Wei, H.; Wang, E. Nanomaterials with Enzyme-like Characteristics (Nanozymes): Next-Generation Artificial Enzymes. *Chem. Soc. Rev.* **2013**, *42*, 6060. [\[CrossRef\]](#)
- Wu, J.; Wang, X.; Wang, Q.; Lou, Z.; Li, S.; Zhu, Y.; Qin, L.; Wei, H. Nanomaterials with Enzyme-like Characteristics (Nanozymes): Next-Generation Artificial Enzymes (II). *Chem. Soc. Rev.* **2019**, *48*, 1004–1076. [\[CrossRef\]](#)
- Meng, X.; Fan, K.; Yan, X. Nanozymes: An Emerging Field Bridging Nanotechnology and Enzymology. *Sci. China Life Sci.* **2019**, *62*, 1543–1546. [\[CrossRef\]](#) [\[PubMed\]](#)
- Sindhu, R.K.; Najda, A.; Kaur, P.; Shah, M.; Singh, H.; Kaur, P.; Cavalu, S.; Jaroszuk-Sierocińska, M.; Rahman, M.H. Potentiality of Nanoenzymes for Cancer Treatment and Other Diseases: Current Status and Future Challenges. *Materials* **2021**, *14*, 5965. [\[CrossRef\]](#) [\[PubMed\]](#)
- Wang, P.; Wang, T.; Hong, J.; Yan, X.; Liang, M. Nanozymes: A New Disease Imaging Strategy. *Front. Bioeng. Biotechnol.* **2020**, *8*, 15. [\[CrossRef\]](#)
- Vallabani, N.V.S.; Singh, S.; Karakoti, A.S. Magnetic Nanoparticles: Current Trends and Future Aspects in Diagnostics and Nanomedicine. *CDM* **2019**, *20*, 457–472. [\[CrossRef\]](#)

28. Fan, W.; Tang, W.; Lau, J.; Shen, Z.; Xie, J.; Shi, J.; Chen, X. Breaking the Depth Dependence by Nanotechnology-Enhanced X-Ray-Excited Deep Cancer Theranostics. *Adv. Mater.* **2019**, *31*, 1806381. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Darabdhara, G.; Das, M.R.; Singh, S.P.; Rengan, A.K.; Szunerits, S.; Boukherroub, R. Ag and Au Nanoparticles/Reduced Graphene Oxide Composite Materials: Synthesis and Application in Diagnostics and Therapeutics. *Adv. Colloid Interface Sci.* **2019**, *271*, 101991. [\[CrossRef\]](#)
30. Sarbadhikary, P.; George, B.P.; Abrahamse, H. Recent Advances in Photosensitizers as Multifunctional Theranostic Agents for Imaging-Guided Photodynamic Therapy of Cancer. *Theranostics* **2021**, *11*, 9054–9088. [\[CrossRef\]](#)
31. Yun, S.-W.; Kang, N.-Y.; Park, S.-J.; Ha, H.-H.; Kim, Y.K.; Lee, J.-S.; Chang, Y.-T. Diversity Oriented Fluorescence Library Approach (DOFLA) for Live Cell Imaging Probe Development. *Accounts Chem. Res.* **2014**, *47*, 1277–1286. [\[CrossRef\]](#) [\[PubMed\]](#)
32. Sharkey, R.M.; Goldenberg, D.M. Targeted Therapy of Cancer: New Prospects for Antibodies and Immunoconjugates. *CA A Cancer J. Clin.* **2006**, *56*, 226–243. [\[CrossRef\]](#)
33. Jiang, D.; Ni, D.; Rosenkrans, Z.T.; Huang, P.; Yan, X.; Cai, W. Nanozyme: New Horizons for Responsive Biomedical Applications. *Chem. Soc. Rev.* **2019**, *48*, 3683–3704. [\[CrossRef\]](#) [\[PubMed\]](#)
34. Huang, Y.; Ren, J.; Qu, X. Nanozymes: Classification, Catalytic Mechanisms, Activity Regulation, and Applications. *Chem. Rev.* **2019**, *119*, 4357–4412. [\[CrossRef\]](#) [\[PubMed\]](#)
35. Wang, H.; Wan, K.; Shi, X. Recent Advances in Nanozyme Research. *Adv. Mater.* **2019**, *31*, 1805368. [\[CrossRef\]](#)
36. Robert, A.; Meunier, B. How to Define a Nanozyme. *ACS Nano* **2022**, *16*, 6956–6959. [\[CrossRef\]](#)
37. Zhang, X.; Chen, X.; Zhao, Y. Nanozymes: Versatile Platforms for Cancer Diagnosis and Therapy. *Nano-Micro Lett.* **2022**, *14*, 95. [\[CrossRef\]](#)
38. Liang, M.; Yan, X. Nanozymes: From New Concepts, Mechanisms, and Standards to Applications. *Accounts Chem. Res.* **2019**, *52*, 2190–2200. [\[CrossRef\]](#)
39. Cao, C.; Zhang, T.; Yang, N.; Niu, X.; Zhou, Z.; Wang, J.; Yang, D.; Chen, P.; Zhong, L.; Dong, X.; et al. POD Nanozyme Optimized by Charge Separation Engineering for Light/PH Activated Bacteria Catalytic/Photodynamic Therapy. *Signal Transduct. Target Ther.* **2022**, *7*, 86. [\[CrossRef\]](#)
40. Chen, G.-Q.; Benthani, F.A.; Wu, J.; Liang, D.; Bian, Z.-X.; Jiang, X. Artemisinin Compounds Sensitize Cancer Cells to Ferroptosis by Regulating Iron Homeostasis. *Cell Death Differ.* **2020**, *27*, 242–254. [\[CrossRef\]](#)
41. Shi, C.; Li, Y.; Gu, N. Iron-Based Nanozymes in Disease Diagnosis and Treatment. *ChemBioChem* **2020**, *21*, 2722–2732. [\[CrossRef\]](#)
42. Gao, L.; Zhuang, J.; Nie, L.; Zhang, J.; Zhang, Y.; Gu, N.; Wang, T.; Feng, J.; Yang, D.; Perrett, S.; et al. Intrinsic Peroxidase-like Activity of Ferromagnetic Nanoparticles. *Nat. Nanotechnol.* **2007**, *2*, 577–583. [\[CrossRef\]](#)
43. Chen, J.; Wang, Q.; Huang, L.; Zhang, H.; Rong, K.; Zhang, H.; Dong, S. Prussian Blue with Intrinsic Heme-like Structure as Peroxidase Mimic. *Nano Res.* **2018**, *11*, 4905–4913. [\[CrossRef\]](#)
44. Gautam, M.; Poudel, K.; Yong, C.S.; Kim, J.O. Prussian Blue Nanoparticles: Synthesis, Surface Modification, and Application in Cancer Treatment. *Int. J. Pharm.* **2018**, *549*, 31–49. [\[CrossRef\]](#)
45. Cheng, N.; Li, J.; Liu, D.; Lin, Y.; Du, D. Single-Atom Nanozyme Based on Nanoengineered Fe–N–C Catalyst with Superior Peroxidase-Like Activity for Ultrasensitive Bioassays. *Small* **2019**, *15*, 1901485. [\[CrossRef\]](#)
46. Wang, X.; Wang, H.; Zhou, S. Progress and Perspective on Carbon-Based Nanozymes for Peroxidase-like Applications. *J. Phys. Chem. Lett.* **2021**, *12*, 11751–11760. [\[CrossRef\]](#)
47. Pan, T.; Chen, H.; Gao, X.; Wu, Z.; Ye, Y.; Shen, Y. Engineering Efficient Artificial Nanozyme Based on Chitosan Grafted Fe-Doped-Carbon Dots for Bacteria Biofilm Eradication. *J. Hazard. Mater.* **2022**, *435*, 128996. [\[CrossRef\]](#)
48. Lu, C.; Tang, L.; Gao, F.; Li, Y.; Liu, J.; Zheng, J. DNA-Encoded Bimetallic Au–Pt Dumbbell Nanozyme for High-Performance Detection and Eradication of Escherichia Coli O157:H7. *Biosens. Bioelectron.* **2021**, *187*, 113327. [\[CrossRef\]](#)
49. Wu, S.-C.; Tsai, T.-T.; Li, T.-H.; Tung, C.-Y.; Chiu, P.-Y.; Lin, J.-H.; Chen, C.-F. Palladium-Platinum Bimetallic Nanomaterials and Their Application in Staphylococcus Aureus Detection on Paper-Based Devices. *Biosens. Bioelectron.* **2022**, *216*, 114669. [\[CrossRef\]](#) [\[PubMed\]](#)
50. Dadigala, R.; Bandi, R.; Alle, M.; Park, C.-W.; Han, S.-Y.; Kwon, G.-J.; Lee, S.-H. Effective Fabrication of Cellulose Nanofibrils Supported Pd Nanoparticles as a Novel Nanozyme with Peroxidase and Oxidase-like Activities for Efficient Dye Degradation. *J. Hazard. Mater.* **2022**, *436*, 129165. [\[CrossRef\]](#) [\[PubMed\]](#)
51. Cao, Z.; Wang, H.; Chen, J.; Zhang, Y.; Mo, Q.; Zhang, P.; Wang, M.; Liu, H.; Bao, X.; Sun, Y.; et al. Silk-Based Hydrogel Incorporated with Metal-Organic Framework Nanozymes for Enhanced Osteochondral Regeneration. *Bioact. Mater.* **2023**, *20*, 221–242. [\[CrossRef\]](#) [\[PubMed\]](#)
52. Weng, Q.; Hu, X.; Zheng, J.; Xia, F.; Wang, N.; Liao, H.; Liu, Y.; Kim, D.; Liu, J.; Li, F.; et al. Toxicological Risk Assessments of Iron Oxide Nanocluster- and Gadolinium-Based T1MRI Contrast Agents in Renal Failure Rats. *ACS Nano* **2019**, *13*, 6801–6812. [\[CrossRef\]](#)
53. Li, J.; Liu, W.; Wu, X.; Gao, X. Mechanism of PH-Switchable Peroxidase and Catalase-like Activities of Gold, Silver, Platinum and Palladium. *Biomaterials* **2015**, *48*, 37–44. [\[CrossRef\]](#) [\[PubMed\]](#)
54. Sahu, A.; Kwon, I.; Tae, G. Improving Cancer Therapy through the Nanomaterials-Assisted Alleviation of Hypoxia. *Biomaterials* **2020**, *228*, 119578. [\[CrossRef\]](#) [\[PubMed\]](#)
55. Zhang, Y.; Jin, Y.; Cui, H.; Yan, X.; Fan, K. Nanozyme-Based Catalytic Theranostics. *RSC Adv.* **2019**, *10*, 10–20. [\[CrossRef\]](#) [\[PubMed\]](#)

56. Wang, Z.; Liu, B.; Sun, Q.; Feng, L.; He, F.; Yang, P.; Gai, S.; Quan, Z.; Lin, J. Upconverted Metal–Organic Framework Janus Architecture for Near-Infrared and Ultrasound Co-Enhanced High Performance Tumor Therapy. *ACS Nano* **2021**, *15*, 12342–12357. [\[CrossRef\]](#)
57. Sang, Y.; Cao, F.; Li, W.; Zhang, L.; You, Y.; Deng, Q.; Dong, K.; Ren, J.; Qu, X. Bioinspired Construction of a Nanozyme-Based H₂O₂ Homeostasis Disruptor for Intensive Chemodynamic Therapy. *J. Am. Chem. Soc.* **2020**, *142*, 5177–5183. [\[CrossRef\]](#)
58. Xu, Z.; Chen, J.; Li, Y.; Hu, T.; Fan, L.; Xi, J.; Han, J.; Guo, R. Yolk-Shell Fe₃O₄@Carbon@Platinum-Chlorin E6 Nanozyme for MRI-Assisted Synergistic Catalytic-Photodynamic-Photothermal Tumor Therapy. *J. Colloid Interface Sci.* **2022**, *628*, 1033–1043. [\[CrossRef\]](#)
59. Wang, M.; Wang, D.; Chen, Q.; Li, C.; Li, Z.; Lin, J. Recent Advances in Glucose-Oxidase-Based Nanocomposites for Tumor Therapy. *Small* **2019**, *15*, 1903895. [\[CrossRef\]](#)
60. Nagvenkar, A.P.; Gedanken, A. Cu 0.89 Zn 0.11 O, A New Peroxidase-Mimicking Nanozyme with High Sensitivity for Glucose and Antioxidant Detection. *ACS Appl. Mater. Interfaces* **2016**, *8*, 22301–22308. [\[CrossRef\]](#)
61. Comotti, M.; Della Pina, C.; Matarrese, R.; Rossi, M. The Catalytic Activity of “Naked” Gold Particles. *Angew. Chem. Int. Ed.* **2004**, *43*, 5812–5815. [\[CrossRef\]](#) [\[PubMed\]](#)
62. Wu, C.-Y.; Hsu, Y.-H.; Chen, Y.; Yang, L.-C.; Tseng, S.-C.; Chen, W.-R.; Huang, C.-C.; Wan, D. Robust O₂ Supplementation from a Trimetallic Nanozyme-Based Self-Sufficient Complementary System Synergistically Enhances the Starvation/Photothermal Therapy against Hypoxic Tumors. *ACS Appl. Mater. Interfaces* **2021**, *13*, 38090–38104. [\[CrossRef\]](#) [\[PubMed\]](#)
63. Gao, S.; Lin, H.; Zhang, H.; Yao, H.; Chen, Y.; Shi, J. Nanocatalytic Tumor Therapy by Biomimetic Dual Inorganic Nanozyme-Catalyzed Cascade Reaction. *Adv. Sci.* **2019**, *6*, 1801733. [\[CrossRef\]](#)
64. Yuan, Z.; Liu, X.; Ling, J.; Huang, G.; Huang, J.; Zhu, X.; He, L.; Chen, T. In Situ-Transition Nanozyme Triggered by Tumor Microenvironment Boosts Synergistic Cancer Radio-/Chemotherapy through Disrupting Redox Homeostasis. *Biomaterials* **2022**, *287*, 121620. [\[CrossRef\]](#)
65. Xu, D.; Wu, L.; Yao, H.; Zhao, L. Catalase-Like Nanozymes: Classification, Catalytic Mechanisms, and Their Applications. *Small* **2022**, *18*, 2203400. [\[CrossRef\]](#)
66. Yuan, B.; Chou, H.-L.; Peng, Y.-K. Disclosing the Origin of Transition Metal Oxides as Peroxidase (and Catalase) Mimetics. *ACS Appl. Mater. Interfaces* **2022**, *14*, 22728–22736. [\[CrossRef\]](#) [\[PubMed\]](#)
67. Meng, X.; Zare, I.; Yan, X.; Fan, K. Protein-protected Metal Nanoclusters: An Emerging Ultra-small Nanozyme. *WIREs Nanomed. Nanobiotechnol.* **2020**, *12*, e1602. [\[CrossRef\]](#)
68. Zhang, R.; Xue, B.; Tao, Y.; Zhao, H.; Zhang, Z.; Wang, X.; Zhou, X.; Jiang, B.; Yang, Z.; Yan, X.; et al. Edge-Site Engineering of Defective Fe–N 4 Nanozymes with Boosted Catalase-Like Performance for Retinal Vasculopathies. *Adv. Mater.* **2022**, *34*, 2205324. [\[CrossRef\]](#)
69. Li, Y.; Yang, L.; Xu, J.; Zhou, H.; Gao, Z.; Song, Y. Pt Nanoparticle-Coupled WO₂.72 Nanoplates as Multi-Enzyme Mimetics for Colorimetric Detection and Radical Elimination. *Anal. Bioanal. Chem.* **2020**, *412*, 521–530. [\[CrossRef\]](#)
70. Chen, F.; Song, T. AuPt Bimetallic Nanozymes for Enhanced Glucose Catalytic Oxidase. *Front. Chem.* **2022**, *10*, 854516. [\[CrossRef\]](#)
71. Dong, P.; Wang, W.; Pan, M.; Yu, W.; Liu, Y.; Shi, T.; Hu, J.; Zhou, Y.; Yu, S.; Wang, F.; et al. Cascaded Amplifier Nanoreactor for Efficient Photodynamic Therapy. *ACS Appl. Mater. Interfaces* **2021**, *13*, 16075–16083. [\[CrossRef\]](#) [\[PubMed\]](#)
72. Sun, D.; Pang, X.; Cheng, Y.; Ming, J.; Xiang, S.; Zhang, C.; Lv, P.; Chu, C.; Chen, X.; Liu, G.; et al. Ultrasound-Switchable Nanozyme Augments Sonodynamic Therapy against Multidrug-Resistant Bacterial Infection. *ACS Nano* **2020**, *14*, 2063–2076. [\[CrossRef\]](#)
73. Singh, N.; NaveenKumar, S.K.; Geethika, M.; Mughesh, G. A Cerium Vanadate Nanozyme with Specific Superoxide Dismutase Activity Regulates Mitochondrial Function and ATP Synthesis in Neuronal Cells. *Angew. Chem. Int. Ed.* **2021**, *60*, 3121–3130. [\[CrossRef\]](#)
74. Korschelt, K.; Ragg, R.; Metzger, C.S.; Kluecker, M.; Oster, M.; Barton, B.; Panthöfer, M.; Strand, D.; Kolb, U.; Mondeshki, M.; et al. Glycine-Functionalized Copper(ii) Hydroxide Nanoparticles with High Intrinsic Superoxide Dismutase Activity. *Nanoscale* **2017**, *9*, 3952–3960. [\[CrossRef\]](#)
75. Korsvik, C.; Patil, S.; Seal, S.; Self, W.T. Superoxide Dismutase Mimetic Properties Exhibited by Vacancy Engineered Ceria Nanoparticles. *Chem. Commun.* **2007**, 1056–1058. [\[CrossRef\]](#)
76. Landa, P. Positive Effects of Metallic Nanoparticles on Plants: Overview of Involved Mechanisms. *Plant Physiol. Biochem.* **2021**, *161*, 12–24. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Dong, S.; Dong, Y.; Liu, B.; Liu, J.; Liu, S.; Zhao, Z.; Li, W.; Tian, B.; Zhao, R.; He, F.; et al. Guiding Transition Metal-Doped Hollow Cerium Tandem Nanozymes with Elaborately Regulated Multi-Enzymatic Activities for Intensive Chemodynamic Therapy. *Adv. Mater.* **2022**, *34*, 2107054. [\[CrossRef\]](#) [\[PubMed\]](#)
78. Rowe, S.P.; Pomper, M.G. Molecular Imaging in Oncology: Current Impact and Future Directions. *CA A Cancer J. Clin.* **2022**, *72*, 333–352. [\[CrossRef\]](#) [\[PubMed\]](#)
79. Zhang, L.; Bhatnagar, S.; Deschenes, E.; Thurber, G.M. Mechanistic and Quantitative Insight into Cell Surface Targeted Molecular Imaging Agent Design. *Sci. Rep.* **2016**, *6*, 25424. [\[CrossRef\]](#)
80. Zhu, X.; Xu, N.; Zhang, L.; Wang, D.; Zhang, P. Novel Design of Multifunctional Nanozymes Based on Tumor Microenvironment for Diagnosis and Therapy. *Eur. J. Med. Chem.* **2022**, *238*, 114456. [\[CrossRef\]](#)

81. Khramtsov, P.; Kropaneva, M.; Minin, A.; Bochkova, M.; Timganova, V.; Maximov, A.; Puzik, A.; Zamorina, S.; Rayev, M. Prussian Blue Nanozymes with Enhanced Catalytic Activity: Size Tuning and Application in ELISA-like Immunoassay. *Nanomaterials* **2022**, *12*, 1630. [\[CrossRef\]](#)
82. Chang, Y.; Gao, S.; Liu, M.; Liu, J. Designing Signal-on Sensors by Regulating Nanozyme Activity. *Anal. Methods* **2020**, *12*, 4708–4723. [\[CrossRef\]](#)
83. Li, S.; Zhao, X.; Gang, R.; Cao, B.; Wang, H. Doping Nitrogen into Q-Graphene by Plasma Treatment toward Peroxidase Mimics with Enhanced Catalysis. *Anal. Chem.* **2020**, *92*, 5152–5157. [\[CrossRef\]](#) [\[PubMed\]](#)
84. Meng, Y.; Chen, Y.; Zhu, J.; Qi, Y.; Ding, J.; Zhou, W. Polarity Control of DNA Adsorption Enabling the Surface Functionalization of CuO Nanozymes for Targeted Tumor Therapy. *Mater. Horiz.* **2021**, *8*, 972–986. [\[CrossRef\]](#) [\[PubMed\]](#)
85. Iqbal, M.A.; Arora, S.; Prakasam, G.; Calin, G.A.; Syed, M.A. MicroRNA in Lung Cancer: Role, Mechanisms, Pathways and Therapeutic Relevance. *Mol. Asp. Med.* **2019**, *70*, 3–20. [\[CrossRef\]](#) [\[PubMed\]](#)
86. Hu, P.; Wang, R.; Zhou, L.; Chen, L.; Wu, Q.; Han, Y.; El-Toni, A.M.; Zhao, D.; Zhang, F. Near Infrared-Activated Upconversion Nanoprobes for Sensitive Endogenous Zn²⁺ Detection and Selective On-Demand Photodynamic Therapy. *Anal. Chem.* **2017**, *89*, 34923500. [\[CrossRef\]](#)
87. Hori, S.S.; Tummers, W.S.; Gambhir, S.S. On-Target Probes for Early Detection. *Nat. Biomed. Eng.* **2017**, *1*, 0062. [\[CrossRef\]](#)
88. Ren, X.; Chen, D.; Wang, Y.; Li, H.; Zhang, Y.; Chen, H.; Li, X.; Huo, M. Nanozymes-Recent Development and Biomedical Applications. *J. Nanobiotechnol.* **2022**, *20*, 92. [\[CrossRef\]](#)
89. Ma, Q.; Liu, Y.; Zhu, H.; Zhang, L.; Liao, X. Nanozymes in Tumor Theranostics. *Front. Oncol.* **2021**, *11*, 666017. [\[CrossRef\]](#)
90. Wang, P.; Min, D.; Chen, G.; Li, M.; Tong, L.; Cao, Y. Inorganic Nanozymes: Prospects for Disease Treatments and Detection Applications. *Front. Chem.* **2021**, *9*, 773285. [\[CrossRef\]](#)
91. Wang, Z.; Li, Z.; Sun, Z.; Wang, S.; Ali, Z.; Zhu, S.; Liu, S.; Ren, Q.; Sheng, F.; Wang, B.; et al. Visualization Nanozyme Based on Tum-or Microenvironment “Unlocking” for Intensive Combination Therapy of Breast Cancer. *Sci. Adv.* **2020**, *6*, eabc8733. [\[CrossRef\]](#)
92. Yang, F.; Hu, S.; Zhang, Y.; Cai, X.; Huang, Y.; Wang, F.; Wen, S.; Teng, G.; Gu, N. A Hydrogen Peroxide-Responsive O₂ Nanogenerator for Ultrasound and Magnetic-Resonance Dual Modality Imaging. *Adv. Mater.* **2012**, *24*, 5205–5211. [\[CrossRef\]](#)
93. Huang, Y.; Hsu, J.C.; Koo, H.; Cormode, D.P. Repurposing Ferumoxytol: Diagnostic and Therapeutic Applications of an FDA-Approved Nanoparticle. *Theranostics* **2022**, *12*, 796–816. [\[CrossRef\]](#) [\[PubMed\]](#)
94. Huang, X.; Lan, N.; Zhang, Y.; Zeng, W.; He, H.; Liu, X. Benzobisthiadiazole and Its Derivative-Based Semiconducting Polymer Nanoparticles for Second Near-Infrared Photoacoustic Imaging. *Front. Chem.* **2022**, *10*, 842712. [\[CrossRef\]](#) [\[PubMed\]](#)
95. Jia, X.; Fan, K.; Zhang, R.; Zhang, D.; Zhang, J.; Gao, Y.; Zhang, T.; Li, W.; Li, J.; Yan, X.; et al. Precise Visual Distinction of Brain Glioma from Normal Tissues via Targeted Photoacoustic and Fluorescence Navigation. *Nanomed. Nanotechnol. Biol. Med.* **2020**, *27*, 102204. [\[CrossRef\]](#)
96. Khan, S.; Sharifi, M.; Bloukh, S.H.; Edis, Z.; Siddique, R.; Falahati, M. In Vivo Guiding Inorganic Nanozymes for Biosensing and Therapeutic Potential in Cancer, Inflammation and Microbial Infections. *Talanta* **2021**, *224*, 121805. [\[CrossRef\]](#)
97. Que, E.L.; Chang, C.J. Responsive Magnetic Resonance Imaging Contrast Agents as Chemical Sensors for Metals in Biology and Medicine. *Chem. Soc. Rev.* **2010**, *39*, 51–60. [\[CrossRef\]](#) [\[PubMed\]](#)
98. Toth, G.B.; Varallyay, C.G.; Horvath, A.; Bashir, M.R.; Choyke, P.L.; Daldrup-Link, H.E.; Dosa, E.; Finn, J.P.; Gahramanov, S.; Harisinghani, M.; et al. Current and Potential Imaging Applications of Ferumoxytol for Magnetic Resonance Imaging. *Kidney Int.* **2017**, *92*, 47–66. [\[CrossRef\]](#) [\[PubMed\]](#)
99. Angelovski, G.; Tóth, É. Strategies for Sensing Neurotransmitters with Responsive MRI Contrast Agents. *Chem. Soc. Rev.* **2017**, *46*, 324–336. [\[CrossRef\]](#) [\[PubMed\]](#)
100. Dadfar, S.M.; Roemhild, K.; Drude, N.I.; von Stillfried, S.; Knüchel, R.; Kiessling, F.; Lammers, T. Iron Oxide Nanoparticles: Diagnostic, Therapeutic and Theranostic Applications. *Adv. Drug Deliv. Rev.* **2019**, *138*, 302–325. [\[CrossRef\]](#) [\[PubMed\]](#)
101. Li, Y.; Zhao, X.; Liu, X.; Cheng, K.; Han, X.; Zhang, Y.; Min, H.; Liu, G.; Xu, J.; Shi, J.; et al. A Bioinspired Nanoprobe with Multilevel Responsive T₁—Weighted MR Signal-Amplification Illuminates Ultrasmall Metastases. *Adv. Mater.* **2020**, *32*, 1906799. [\[CrossRef\]](#)
102. Yue, H.; Yuan, L.; Zhang, W.; Zhang, S.; Wei, W.; Ma, G. Macrophage Responses to the Physical Burden of Cell-Sized Particles. *J. Mater. Chem. B* **2018**, *6*, 393–400. [\[CrossRef\]](#)
103. Zhou, C.; Zhang, L.; Sun, T.; Zhang, Y.; Liu, Y.; Gong, M.; Xu, Z.; Du, M.; Liu, Y.; Liu, G.; et al. Activatable NIR-II Plasmonic Nanotheranostics for Efficient Photoacoustic Imaging and Photothermal Cancer Therapy. *Adv. Mater.* **2021**, *33*, 2006532. [\[CrossRef\]](#)
104. Chen, Y.-S.; Zhao, Y.; Yoon, S.J.; Gambhir, S.S.; Emelianov, S. Miniature Gold Nanorods for Photoacoustic Molecular Imaging in the Second Near-Infrared Optical Window. *Nat. Nanotechnol.* **2019**, *14*, 465–472. [\[CrossRef\]](#) [\[PubMed\]](#)
105. Knox, H.J.; Chan, J. Acoustogenic Probes: A New Frontier in Photoacoustic Imaging. *Accounts Chem. Res.* **2018**, *51*, 2897–2905. [\[CrossRef\]](#) [\[PubMed\]](#)
106. Liu, Y.; Bhattarai, P.; Dai, Z.; Chen, X. Photothermal Therapy and Photoacoustic Imaging via Nanotheranostics in Fighting Cancer. *Chem. Soc. Rev.* **2019**, *48*, 2053–2108. [\[CrossRef\]](#) [\[PubMed\]](#)
107. Yang, X.; Stein, E.W.; Ashkenazi, S.; Wang, L.V. Nanoparticles for Photoacoustic Imaging. *WIREs Nanomed. Nanobiotechnol.* **2009**, *1*, 360–368. [\[CrossRef\]](#)

108. Biffi, S.; Voltan, R.; Rampazzo, E.; Prodi, L.; Zauli, G.; Secchiero, P. Applications of Nanoparticles in Cancer Medicine and beyond: Optical and Multimodal in Vivo Imaging, Tissue Targeting and Drug Delivery. *Expert Opin. Drug Deliv.* **2015**, *12*, 1837–1849. [\[CrossRef\]](#)
109. Serkova, N.J.; Glunde, K.; Haney, C.R.; Farhoud, M.; De Lille, A.; Redente, E.F.; Simberg, D.; Westerly, D.C.; Griffin, L.; Mason, R.P. Preclinical Applications of Multi-Platform Imaging in Animal Models of Cancer. *Cancer Res.* **2021**, *81*, 1189–1200. [\[CrossRef\]](#)
110. Leary, J.; Key, J. Nanoparticles for Multimodal in Vivo Imaging in Nanomedicine. *IJN* **2014**, 711. [\[CrossRef\]](#)
111. Ametamey, S.M.; Honer, M.; Schubiger, P.A. Molecular Imaging with PET. *Chem. Rev.* **2008**, *108*, 1501–1516. [\[CrossRef\]](#)
112. Ge, J.; Zhang, Q.; Zeng, J.; Gu, Z.; Gao, M. Radiolabeling Nanomaterials for Multimodality Imaging: New Insights into Nuclear Medicine and Cancer Diagnosis. *Biomaterials* **2020**, *228*, 119553. [\[CrossRef\]](#)
113. You, Q.; Zhang, K.; Liu, J.; Liu, C.; Wang, H.; Wang, M.; Ye, S.; Gao, H.; Lv, L.; Wang, C.; et al. Persistent Regulation of Tumor Hypoxia Microenvironment via a Bioinspired Pt-Based Oxygen Nanogenerator for Multimodal Imaging-Guided Synergistic Phototherapy. *Adv. Sci.* **2020**, *7*, 1903341. [\[CrossRef\]](#)
114. Wu, M.; Shu, J. Multimodal Molecular Imaging: Current Status and Future Directions. *Contrast Media Mol. Imaging* **2018**, *2018*, 1382183. [\[CrossRef\]](#) [\[PubMed\]](#)
115. Vermeulen, I.; Isin, E.M.; Barton, P.; Cillero-Pastor, B.; Heeren, R.M.A. Multimodal Molecular Imaging in Drug Discovery and Development. *Drug Discov. Today* **2022**, *27*, 2086–2099. [\[CrossRef\]](#)
116. Li, X.; Lovell, J.F.; Yoon, J.; Chen, X. Clinical Development and Potential of Photothermal and Photodynamic Therapies for Cancer. *Nat. Rev. Clin. Oncol.* **2020**, *17*, 657–674. [\[CrossRef\]](#) [\[PubMed\]](#)
117. Shin, T.-H.; Choi, Y.; Kim, S.; Cheon, J. Recent Advances in Magnetic Nanoparticle-Based Multi-Modal Imaging. *Chem. Soc. Rev.* **2015**, *44*, 4501–4516. [\[CrossRef\]](#)
118. Qin, R.; Feng, Y.; Ding, D.; Chen, L.; Li, S.; Deng, H.; Chen, S.; Han, Z.; Sun, W.; Chen, H. Fe-Coordinated Carbon Nanozyme Dots as Peroxidase-Like Nanozymes and Magnetic Resonance Imaging Contrast Agents. *ACS Appl. Bio Mater.* **2021**, *4*, 5520–5528. [\[CrossRef\]](#) [\[PubMed\]](#)
119. Yu, J.; Zhao, F.; Gao, W.; Yang, X.; Ju, Y.; Zhao, L.; Guo, W.; Xie, J.; Liang, X.; Tao, X.; et al. Magnetic Reactive Oxygen Species Nanoreactor for Switchable Magnetic Resonance Imaging Guided Cancer Therapy Based on PH-Sensitive Fe₅C₂@Fe₃O₄ Nanoparticles. *ACS Nano* **2019**, *13*, 10002–10014. [\[CrossRef\]](#)
120. Huang, Q.; Pan, Y.; Wang, M.; Liu, Z.; Chen, H.; Wang, J.; Zhao, Z.; Zhang, Y. Tumor Microenvironment-Responsive Versatile “Trojan Horse” Theranostic Nanoplatform for Magnetic Resonance Imaging-Guided Multimodal Synergistic Antitumor Treatment. *Acta Biomater.* **2022**, *147*, 270–286. [\[CrossRef\]](#)
121. Zhang, D.-Y.; Liu, H.; Zhu, K.S.; He, T.; Younis, M.R.; Yang, C.; Lei, S.; Wu, J.; Lin, J.; Qu, J.; et al. Prussian Blue-Based Theranostics for Ameliorating Acute Kidney Injury. *J. Nanobiotechnol.* **2021**, *19*, 266. [\[CrossRef\]](#) [\[PubMed\]](#)
122. Wu, H.; Xia, F.; Zhang, L.; Fang, C.; Lee, J.; Gong, L.; Gao, J.; Ling, D.; Li, F. A ROS-Sensitive Nanozyme-Augmented Photoacoustic Nanoprobe for Early Diagnosis and Therapy of Acute Liver Failure. *Adv. Mater.* **2022**, *34*, 2108348. [\[CrossRef\]](#) [\[PubMed\]](#)
123. Wang, M.; Chang, M.; Chen, Q.; Wang, D.; Li, C.; Hou, Z.; Lin, J.; Jin, D.; Xing, B. Au₂Pt-PEG-Ce6 Nanoformulation with Dual Nanozyme Activities for Synergistic Chemodynamic Therapy/Phototherapy. *Biomaterials* **2020**, *252*, 120093. [\[CrossRef\]](#)
124. Liang, S.; Deng, X.; Chang, Y.; Sun, C.; Shao, S.; Xie, Z.; Xiao, X.; Ma, P.; Zhang, H.; Cheng, Z.; et al. Intelligent Hollow Pt-CuS Janus Architecture for Synergistic Catalysis-Enhanced Sonodynamic and Photothermal Cancer Therapy. *Nano Lett.* **2019**, *19*, 4134–4145. [\[CrossRef\]](#)
125. Ding, H.; Cai, Y.; Gao, L.; Liang, M.; Miao, B.; Wu, H.; Liu, Y.; Xie, N.; Tang, A.; Fan, K.; et al. Exosome-like Nanozyme Vesicles for H₂O₂-Responsive Catalytic Photoacoustic Imaging of Xenograft Nasopharyngeal Carcinoma. *Nano Lett.* **2019**, *19*, 203–209. [\[CrossRef\]](#) [\[PubMed\]](#)
126. Liu, F.; Lin, L.; Zhang, Y.; Wang, Y.; Sheng, S.; Xu, C.; Tian, H.; Chen, X. A Tumor-Microenvironment-Activated Nanozyme-Mediated Theranostic Nanoreactor for Imaging-Guided Combined Tumor Therapy. *Adv. Mater.* **2019**, *31*, 1902885. [\[CrossRef\]](#)
127. Zheng, Z.; Jia, Z.; Qin, Y.; Dai, R.; Chen, X.; Ma, Y.; Xie, X.; Zhang, R. All-in-One Zeolite–Carbon-Based Nanotheranostics with Adjustable NIR-II Window Photoacoustic/Fluorescence Imaging Performance for Precise NIR-II Photothermal-Synergized Catalytic Antitumor Therapy. *Small* **2021**, *17*, 2103252. [\[CrossRef\]](#)
128. Zhang, Y.; Wang, K.; Xing, G.; Dong, X.; Zhu, D.; Yang, W.; Mei, L.; Lv, F. Nanozyme-Laden Intelligent Macrophage EXPRESS Amplifying Cancer Photothermal-Starvation Therapy by Responsive Stimulation. *Mater. Today Bio* **2022**, *16*, 100421. [\[CrossRef\]](#)
129. Zhang, A.; Pan, S.; Zhang, Y.; Chang, J.; Cheng, J.; Huang, Z.; Li, T.; Zhang, C.; de la Fuentea, J.M.; Zhang, Q.; et al. Carbon-Gold Hybrid Nanoprobes for Real-Time Imaging, Photothermal/Photodynamic and Nanozyme Oxidative Therapy. *Theranostics* **2019**, *9*, 3443–3458. [\[CrossRef\]](#)
130. Dan, Q.; Yuan, Z.; Zheng, S.; Ma, H.; Luo, W.; Zhang, L.; Su, N.; Hu, D.; Sheng, Z.; Li, Y. Gold Nanoclusters-Based NIR-II Photosensitizers with Catalase-like Activity for Boosted Photodynamic Therapy. *Pharmaceutics* **2022**, *14*, 1645. [\[CrossRef\]](#)
131. Li, L.; Yang, Z.; Fan, W.; He, L.; Cui, C.; Zou, J.; Tang, W.; Jacobson, O.; Wang, Z.; Niu, G.; et al. In Situ Polymerized Hollow Mesoporous Organosilica Biocatalysis Nanoreactor for Enhancing ROS-Mediated Anticancer Therapy. *Adv. Funct. Mater.* **2020**, *30*, 1907716. [\[CrossRef\]](#)
132. Yuan, H.; Wilks, M.Q.; Normandin, M.D.; El Fakhri, G.; Kaittanis, C.; Josephson, L. Heat-Induced Radiolabeling and Fluorescence Labeling of Feraheme Nanoparticles for PET/SPECT Imaging and Flow Cytometry. *Nat. Protoc.* **2018**, *13*, 392–412. [\[CrossRef\]](#)

133. Jia, T.; Li, D.; Du, J.; Fang, X.; Gerasimov, V.; Ågren, H.; Chen, G. A Bimodal Type of AgPd Plasmonic Blackbody Nanozyme with Boosted Catalytic Efficacy and Synergized Photothermal Therapy for Efficacious Tumor Treatment in the Second Biological Window. *J. Nanobiotechnol.* **2022**, *20*, 424. [[CrossRef](#)] [[PubMed](#)]
134. Liu, S.; Zhang, W.; Chen, Q.; Hou, J.; Wang, J.; Zhong, Y.; Wang, X.; Jiang, W.; Ran, H.; Guo, D. Multifunctional Nanozyme for Multimodal Imaging-Guided Enhanced Sonodynamic Therapy by Regulating the Tumor Microenvironment. *Nanoscale* **2021**, *13*, 14049–14066. [[CrossRef](#)] [[PubMed](#)]
135. Gong, F.; Yang, N.; Wang, Y.; Zhuo, M.; Zhao, Q.; Wang, S.; Li, Y.; Liu, Z.; Chen, Q.; Cheng, L. Oxygen-Deficient Bimetallic Oxide FeWO₃ Nanosheets as Peroxidase-Like Nanozyme for Sensing Cancer via Photoacoustic Imaging. *Small* **2020**, *16*, 2003496. [[CrossRef](#)] [[PubMed](#)]
136. Cao, C.; Zou, H.; Yang, N.; Li, H.; Cai, Y.; Song, X.; Shao, J.; Chen, P.; Mou, X.; Wang, W.; et al. Fe₃O₄/Ag/Bi₂MoO₆ Photoactivatable Nanozyme for Self-Replenishing and Sustainable Cascaded Nanocatalytic Cancer Therapy. *Adv. Mater.* **2021**, *33*, 2106996. [[CrossRef](#)] [[PubMed](#)]
137. Zhou, I.Y.; Catalano, O.A.; Caravan, P. Advances in Functional and Molecular MRI Technologies in Chronic Liver Diseases. *J. Hepatol.* **2020**, *73*, 1241–1254. [[CrossRef](#)]
138. Zijlstra, L.E.; Trompet, S.; Jukema, J.W.; Kroft, L.J.M.; de Bresser, J.; van Osch, M.J.P.; Hammer, S.; Witjes, M.-N.; van Buren, M.; Mooijart, S.P. Association of Cardiovascular Structure and Function with Cerebrovascular Changes and Cognitive Function in Older Patients with End-Stage Renal Disease. *Aging* **2020**, *12*, 1496–1511. [[CrossRef](#)]
139. Datta, A.; Raymond, K.N. Gd-Hydroxypyridinone (HOPO)-Based High-Relaxivity Magnetic Resonance Imaging (MRI) Contrast Agents. *Accounts Chem. Res.* **2009**, *42*, 938–947. [[CrossRef](#)]
140. Sukstanskii, A.L.; Yablonskiy, D.A. Microscopic Theory of Spin–Spin and Spin–Lattice Relaxation of Bound Protons in Cellular and Myelin Membranes—A Lateral Diffusion Model (LDM). *Magn. Reson. Med.* **2023**, *89*, 370–383. [[CrossRef](#)]
141. Aime, S.; Castelli, D.D.; Crich, S.G.; Gianolio, E.; Terreno, E. Pushing the Sensitivity Envelope of Lanthanide-Based Magnetic Resonance Imaging (MRI) Contrast Agents for Molecular Imaging Applications. *Accounts Chem. Res.* **2009**, *42*, 822–831. [[CrossRef](#)] [[PubMed](#)]
142. Li, Y.; Dong, H.; Tao, Q.; Ye, C.; Yu, M.; Li, J.; Zhou, H.; Yang, S.; Ding, G.; Xie, X. Enhancing the Magnetic Relaxivity of MRI Contrast Agents via the Localized Supercacid Microenvironment of Graphene Quantum Dots. *Biomaterials* **2020**, *250*, 120056. [[CrossRef](#)] [[PubMed](#)]
143. Yoo, D.; Lee, J.-H.; Shin, T.-H.; Cheon, J. Theranostic Magnetic Nanoparticles. *Accounts Chem. Res.* **2011**, *44*, 863–874. [[CrossRef](#)]
144. Wahsner, J.; Gale, E.M.; Rodríguez-Rodríguez, A.; Caravan, P. Chemistry of MRI Contrast Agents: Current Challenges and New Frontiers. *Chem. Rev.* **2019**, *119*, 957–1057. [[CrossRef](#)] [[PubMed](#)]
145. Levine, D.; McDonald, R.J.; Kressel, H.Y. Gadolinium Retention After Contrast-Enhanced MRI. *JAMA* **2018**, *320*, 1853. [[CrossRef](#)]
146. Verwilt, P.; Park, S.; Yoon, B.; Kim, J.S. Recent Advances in Gd-Chelate Based Bimodal Optical/MRI Contrast Agents. *Chem. Soc. Rev.* **2015**, *44*, 1791–1806. [[CrossRef](#)]
147. Sieber, M.A.; Pietsch, H.; Walter, J.; Haider, W.; Frenzel, T.; Weinmann, H.-J. A Preclinical Study to Investigate the Development of Nephrogenic Systemic Fibrosis: A Possible Role for Gadolinium-Based Contrast Media. *Investig. Radiol.* **2008**, *43*, 65–75. [[CrossRef](#)]
148. Mitka, M. MRI Contrast Agents May Pose Risk for Patients With Kidney Disease. *JAMA* **2007**, *297*, 249. [[CrossRef](#)]
149. Choi, S.H.; Kim, S.Y.; Lim, Y.-S. Selection of MRI Contrast Agent and Diagnostic Criteria for HCC to Maximize the Advantages of Contrast Agents. *J. Hepatol.* **2020**, *73*, 714–715. [[CrossRef](#)]
150. Gale, E.M.; Atanasova, I.P.; Blasi, F.; Ay, I.; Caravan, P. A Manganese Alternative to Gadolinium for MRI Contrast. *J. Am. Chem. Soc.* **2015**, *137*, 15548–15557. [[CrossRef](#)]
151. Lu, Z.-R.; Laney, V.; Li, Y. Targeted Contrast Agents for Magnetic Resonance Molecular Imaging of Cancer. *Accounts Chem. Res.* **2022**, *55*, 2833–2847. [[CrossRef](#)]
152. Shen, Z.; Chen, T.; Ma, X.; Ren, W.; Zhou, Z.; Zhu, G.; Zhang, A.; Liu, Y.; Song, J.; Li, Z.; et al. Multifunctional Theranostic Nanoparticles Based on Exceedingly Small Magnetic Iron Oxide Nanoparticles for T₁-Weighted Magnetic Resonance Imaging and Chemotherapy. *ACS Nano* **2017**, *11*, 10992–11004. [[CrossRef](#)] [[PubMed](#)]
153. Li, Y.; Zhang, H. Fe₃O₄-Based Nanotheranostics for Magnetic Resonance Imaging-Synergized Multifunctional Cancer Management. *Nanomedicine* **2019**, *14*, 1493–1512. [[CrossRef](#)] [[PubMed](#)]
154. Peters, J.A. Relaxivity of Manganese Ferrite Nanoparticles. *Prog. Nucl. Magn. Reson. Spectrosc.* **2020**, *120–121*, 72–94. [[CrossRef](#)] [[PubMed](#)]
155. Petralia, G.; Summers, P.E.; Agostini, A.; Ambrosini, R.; Cianci, R.; Cristel, G.; Calistri, L.; Colagrande, S. Dynamic Contrast-Enhanced MRI in Oncology: How We Do It. *Radiol. Med.* **2020**, *125*, 1288–1300. [[CrossRef](#)]
156. Lee, N.; Yoo, D.; Ling, D.; Cho, M.H.; Hyeon, T.; Cheon, J. Iron Oxide Based Nanoparticles for Multimodal Imaging and Magnetoresponsive Therapy. *Chem. Rev.* **2015**, *115*, 10637–10689. [[CrossRef](#)] [[PubMed](#)]
157. Zhao, S.; Yu, X.; Qian, Y.; Chen, W.; Shen, J. Multifunctional Magnetic Iron Oxide Nanoparticles: An Advanced Platform for Cancer Theranostics. *Theranostics* **2020**, *10*, 6278–6309. [[CrossRef](#)] [[PubMed](#)]
158. Liu, Y.; Wang, X.; Li, X.; Qiao, S.; Huang, G.; Hermann, D.M.; Doeppner, T.R.; Zeng, M.; Liu, W.; Xu, G.; et al. A Co-Doped Fe₃O₄ Nanozyme Shows Enhanced Reactive Oxygen and Nitrogen Species Scavenging Activity and Ameliorates the Deleterious Effects of Ischemic Stroke. *ACS Appl. Mater. Interfaces* **2021**, *13*, 46213–46224. [[CrossRef](#)]

159. Jin, R.; Lin, B.; Li, D.; Ai, H. Superparamagnetic Iron Oxide Nanoparticles for MR Imaging and Therapy: Design Considerations and Clinical Applications. *Curr. Opin. Pharmacol.* **2014**, *18*, 18–27. [\[CrossRef\]](#)
160. Russell, E.; Dunne, V.; Russell, B.; Mohamud, H.; Ghita, M.; McMahon, S.J.; Butterworth, K.T.; Schettino, G.; McGarry, C.K.; Prise, K.M. Impact of Superparamagnetic Iron Oxide Nanoparticles on in Vitro and in Vivo Radiosensitisation of Cancer Cells. *Radiat. Oncol.* **2021**, *16*, 104. [\[CrossRef\]](#)
161. Gale, E.M.; Wey, H.-Y.; Ramsay, I.; Yen, Y.-F.; Sosnovik, D.E.; Caravan, P. A Manganese-Based Alternative to Gadolinium: Contrast-Enhanced MR Angiography, Excretion, Pharmacokinetics, and Metabolism. *Radiology* **2018**, *286*, 865–872. [\[CrossRef\]](#) [\[PubMed\]](#)
162. Cai, X.; Zhu, Q.; Zeng, Y.; Zeng, Q.; Chen, X.; Zhan, Y. Manganese Oxide Nanoparticles As MRI Contrast Agents In Tumor Multimodal Imaging And Therapy. *IJN* **2019**, *14*, 8321–8344. [\[CrossRef\]](#)
163. Ragg, R.; Schilman, A.M.; Korschelt, K.; Wieseotte, C.; Klueker, M.; Viel, M.; Völker, L.; Preiß, S.; Herzberger, J.; Frey, H.; et al. Intrinsic Superoxide Dismutase Activity of MnO Nanoparticles Enhances the Magnetic Resonance Imaging Contrast. *J. Mater. Chem. B* **2016**, *4*, 7423–7428. [\[CrossRef\]](#) [\[PubMed\]](#)
164. Hsu, B.Y.W.; Kirby, G.; Tan, A.; Seifalian, A.M.; Li, X.; Wang, J. Relaxivity and Toxicological Properties of Manganese Oxide Nanoparticles for MRI Applications. *RSC Adv.* **2016**, *6*, 45462–45474. [\[CrossRef\]](#)
165. Sobańska, Z.; Roszak, J.; Kowalczyk, K.; Stepnik, M. Applications and Biological Activity of Nanoparticles of Manganese and Manganese Oxides in In Vitro and In Vivo Models. *Nanomaterials* **2021**, *11*, 1084. [\[CrossRef\]](#)
166. Zhu, D.; Zhu, X.-H.; Ren, S.-Z.; Lu, Y.-D.; Zhu, H.-L. Manganese Dioxide (MnO₂) Based Nanomaterials for Cancer Therapies and Theranostics. *J. Drug Target.* **2021**, *29*, 911–924. [\[CrossRef\]](#)
167. Yang, G.; Ji, J.; Liu, Z. Multifunctional MnO₂ Nanoparticles for Tumor Microenvironment Modulation and Cancer Therapy. *WIREs Nanomed. Nanobiotechnol.* **2021**, *13*, e1720. [\[CrossRef\]](#) [\[PubMed\]](#)
168. He, T.; Jiang, C.; He, J.; Zhang, Y.; He, G.; Wu, J.; Lin, J.; Zhou, X.; Huang, P. Manganese-Dioxide-Coating-Instructed Plasmonic Modulation of Gold Nanorods for Activatable Duplex-Imaging-Guided NIR-II Photothermal-Chemodynamic Therapy. *Adv. Mater.* **2021**, *33*, 2008540. [\[CrossRef\]](#)
169. Fan, W.; Bu, W.; Shen, B.; He, Q.; Cui, Z.; Liu, Y.; Zheng, X.; Zhao, K.; Shi, J. Intelligent MnO₂ Nanosheets Anchored with Upconversion Nanoprobes for Concurrent PH-/H₂O₂ -Responsive UCL Imaging and Oxygen-Elevated Synergetic Therapy. *Adv. Mater.* **2015**, *27*, 4155–4161. [\[CrossRef\]](#)
170. Zhao, M.; Xie, M.; Guo, J.; Feng, W.; Xu, Y.; Liu, X.; Liu, S.; Zhao, Q. Facile Phototherapeutic Nanoplatfrom by Integrating a Multifunctional Polymer and MnO₂ for Enhancing Tumor Synergistic Therapy. *Adv. Healthc. Mater.* **2019**, *8*, 1900414. [\[CrossRef\]](#)
171. Lan, G.; Ni, K.; Lin, W. Nanoscale Metal–Organic Frameworks for Phototherapy of Cancer. *Coord. Chem. Rev.* **2019**, *379*, 65–81. [\[CrossRef\]](#)
172. Chen, Y.; Li, Z.; Pan, P.; Hu, J.; Cheng, S.; Zhang, X. Tumor-Microenvironment-Triggered Ion Exchange of a Metal–Organic Framework Hybrid for Multimodal Imaging and Synergistic Therapy of Tumors. *Adv. Mater.* **2020**, *32*, 2001452. [\[CrossRef\]](#) [\[PubMed\]](#)
173. Kale, S.; Burga, R.; Sweeney, E.; Zun, Z.; Sze, R.; Tuesca, A.; Subramony, A.; Fernandes, R. Composite Iron Oxide—Prussian Blue Nanoparticles for Magnetically Guided T1-Weighted Magnetic Resonance Imaging and Photothermal Therapy of Tumors. *IJN* **2017**, *12*, 6413–6424. [\[CrossRef\]](#)
174. Qin, Z.; Chen, B.; Huang, X.; Mao, Y.; Li, Y.; Yang, F.; Gu, N. Magnetic Internal Heating-Induced High Performance Prussian Blue Nanoparticle Preparation and Excellent Catalytic Activity. *Dalton Trans.* **2019**, *48*, 17169–17173. [\[CrossRef\]](#)
175. Ma, X.; Hao, J.; Wu, J.; Li, Y.; Cai, X.; Zheng, Y. Prussian Blue Nanozyme as a Pyroptosis Inhibitor Alleviates Neurodegeneration. *Adv. Mater.* **2022**, *34*, 2106723. [\[CrossRef\]](#)
176. Fan, S.; Jiang, X.; Yang, M.; Wang, X. Sensitive Colorimetric Assay for the Determination of Alkaline Phosphatase Activity Utilizing Nanozyme Based on Copper Nanoparticle-Modified Prussian Blue. *Anal. Bioanal. Chem.* **2021**, *413*, 3955–3963. [\[CrossRef\]](#)
177. Upputuri, P.K.; Pramanik, M. Recent Advances in Photoacoustic Contrast Agents for in Vivo Imaging. *WIREs Nanomed. Nanobiotechnol.* **2020**, *12*, e1618. [\[CrossRef\]](#) [\[PubMed\]](#)
178. Liu, Y.; Teng, L.; Yin, B.; Meng, H.; Yin, X.; Huan, S.; Song, G.; Zhang, X.-B. Chemical Design of Activatable Photoacoustic Probes for Precise Biomedical Applications. *Chem. Rev.* **2022**, *122*, 6850–6918. [\[CrossRef\]](#)
179. Fu, Q.; Zhu, R.; Song, J.; Yang, H.; Chen, X. Photoacoustic Imaging: Contrast Agents and Their Biomedical Applications. *Adv. Mater.* **2018**, *31*, 1805875. [\[CrossRef\]](#) [\[PubMed\]](#)
180. Li, J.; Pu, K. Development of Organic Semiconducting Materials for Deep-Tissue Optical Imaging, Phototherapy and Photoactivation. *Chem. Soc. Rev.* **2019**, *48*, 38–71. [\[CrossRef\]](#)
181. Jiang, Y.; Pu, K. Advanced Photoacoustic Imaging Applications of Near-Infrared Absorbing Organic Nanoparticles. *Small* **2017**, *13*, 1700710. [\[CrossRef\]](#) [\[PubMed\]](#)
182. Zhao, Z.; Swartzchick, C.B.; Chan, J. Targeted Contrast Agents and Activatable Probes for Photoacoustic Imaging of Cancer. *Chem. Soc. Rev.* **2022**, *51*, 829–868. [\[CrossRef\]](#) [\[PubMed\]](#)
183. Ju, Y.; Zhang, H.; Yu, J.; Tong, S.; Tian, N.; Wang, Z.; Wang, X.; Su, X.; Chu, X.; Lin, J.; et al. Monodisperse Au–Fe₂C Janus Nanoparticles: An Attractive Multifunctional Material for Triple-Modal Imaging-Guided Tumor Photothermal Therapy. *ACS Nano* **2017**, *11*, 9239–9248. [\[CrossRef\]](#) [\[PubMed\]](#)

184. Mu, X.; Tang, Y.; Wu, F.; Ma, H.; Huang, S.; Liang, M.; Yang, J.; Lu, Y.; Zhou, X.; Li, Z. A Simple Small Molecule with Synergistic Passive and Active Dual-Targeting Effects for Imaging-Guided Photothermal Cancer Therapy. *ACS Appl. Mater. Interfaces* **2021**, *13*, 36958–36966. [\[CrossRef\]](#)
185. Wu, Z.; Zhou, J.; Nkanga, C.I.; Jin, Z.; He, T.; Borum, R.M.; Yim, W.; Zhou, J.; Cheng, Y.; Xu, M.; et al. One-Step Supramolecular Multifunctional Coating on Plant Virus Nanoparticles for Bioimaging and Therapeutic Applications. *ACS Appl. Mater. Interfaces* **2022**, *14*, 13692–13702. [\[CrossRef\]](#)
186. Li, C.; Cheng, Y.; Li, D.; An, Q.; Zhang, W.; Zhang, Y.; Fu, Y. Antitumor Applications of Photothermal Agents and Photothermal Synergistic Therapies. *IJMS* **2022**, *23*, 7909. [\[CrossRef\]](#) [\[PubMed\]](#)
187. Duan, Q.; Wang, J.; Zhang, B.; Wang, X.; Xue, J.; Zhang, W.; Sang, S. Polydopamine Coated Au-Pt Nanorods: Enhanced Photothermal Properties and Efficient Reactive Oxygen Scavengers. *Colloids Surf. B Biointerfaces* **2022**, *210*, 112247. [\[CrossRef\]](#)
188. Xia, Y.; Li, W.; Cobley, C.M.; Chen, J.; Xia, X.; Zhang, Q.; Yang, M.; Cho, E.C.; Brown, P.K. Gold Nanocages: From Synthesis to Theranostic Applications. *Accounts Chem. Res.* **2011**, *44*, 914–924. [\[CrossRef\]](#)
189. Jiang, T.; Song, J.; Zhang, W.; Wang, H.; Li, X.; Xia, R.; Zhu, L.; Xu, X. Au–Ag@Au Hollow Nanostructure with Enhanced Chemical Stability and Improved Photothermal Transduction Efficiency for Cancer Treatment. *ACS Appl. Mater. Interfaces* **2015**, *7*, 21985–21994. [\[CrossRef\]](#)
190. Li, J.; Zhang, C.; Gong, S.; Li, X.; Yu, M.; Qian, C.; Qiao, H.; Sun, M. A Nanoscale Photothermal Agent Based on a Metal–Organic Coordination Polymer as a Drug-Loading Framework for Effective Combination Therapy. *Acta Biomater.* **2019**, *94*, 435–446. [\[CrossRef\]](#)
191. Luo, Y.; Qiao, B.; Zhang, P.; Yang, C.; Cao, J.; Yuan, X.; Ran, H.; Wang, Z.; Hao, L.; Cao, Y.; et al. TME-Activatable Theranostic Nanoplatfrom with ATP Burning Capability for Tumor Sensitization and Synergistic Therapy. *Theranostics* **2020**, *10*, 6987–7001. [\[CrossRef\]](#) [\[PubMed\]](#)
192. Huang, K.; Zhang, Y.; Lin, J.; Huang, P. Nanomaterials for Photoacoustic Imaging in the Second Near-Infrared Window. *Biomater. Sci.* **2019**, *7*, 472–479. [\[CrossRef\]](#) [\[PubMed\]](#)
193. Pirovano, G.; Roberts, S.; Kossatz, S.; Reiner, T. Optical Imaging Modalities: Principles and Applications in Preclinical Research and Clinical Settings. *J. Nucl. Med.* **2020**, *61*, 1419–1427. [\[CrossRef\]](#)
194. Qi, M.; Pan, H.; Shen, H.; Xia, X.; Wu, C.; Han, X.; He, X.; Tong, W.; Wang, X.; Wang, Q. Nanogel Multienzyme Mimics Synthesized by Biocatalytic ATRP and Metal Coordination for Bioresponsive Fluorescence Imaging. *Angew. Chem. Int. Ed.* **2020**, *59*, 11748–11753. [\[CrossRef\]](#) [\[PubMed\]](#)
195. Zhen, X.; Jiang, X. Polymer-based Activatable Optical Probes for Tumor Fluorescence and Photoacoustic Imaging. *WIREs Nanomed. Nanobiotechnol.* **2020**, *12*, e1593. [\[CrossRef\]](#)
196. Douma, K.; Megens, R.T.A.; van Zandvoort, M.A.M.J. Optical Molecular Imaging of Atherosclerosis Using Nanoparticles: Shedding New Light on the Darkness. *WIREs Nanomed. Nanobiotechnol.* **2011**, *3*, 376–388. [\[CrossRef\]](#)
197. Li, B.; Zhao, M.; Lin, J.; Huang, P.; Chen, X. Management of Fluorescent Organic/Inorganic Nanohybrids for Biomedical Applications in the NIR-II Region. *Chem. Soc. Rev.* **2022**, *51*, 7692–7714. [\[CrossRef\]](#)
198. Wu, L.; Zou, H.; Wang, H.; Zhang, S.; Liu, S.; Jiang, Y.; Chen, J.; Li, Y.; Shao, M.; Zhang, R.; et al. Update on the Development of Molecular Imaging and Nanomedicine in China: Optical Imaging. *WIREs Nanomed. Nanobiotechnol.* **2021**, *13*, e1660. [\[CrossRef\]](#) [\[PubMed\]](#)
199. Son, J.; Yi, G.; Yoo, J.; Park, C.; Koo, H.; Choi, H.S. Light-Responsive Nanomedicine for Biophotonic Imaging and Targeted Therapy. *Adv. Drug Deliv. Rev.* **2019**, *138*, 133–147. [\[CrossRef\]](#)
200. Yuan, H.; Jiang, A.; Fang, H.; Chen, Y.; Guo, Z. Optical Properties of Natural Small Molecules and Their Applications in Imaging and Nanomedicine. *Adv. Drug Deliv. Rev.* **2021**, *179*, 113917. [\[CrossRef\]](#)
201. Ni, D.; Ehlerding, E.B.; Cai, W. Multimodality Imaging Agents with PET as the Fundamental Pillar. *Angew. Chem. Int. Ed.* **2019**, *58*, 2570–2579. [\[CrossRef\]](#) [\[PubMed\]](#)
202. Pellico, J.; Gawne, P.J.; de Rosales, R.T.M. Radiolabelling of Nanomaterials for Medical Imaging and Therapy. *Chem. Soc. Rev.* **2021**, *50*, 3355–3423. [\[CrossRef\]](#)
203. Goel, S.; England, C.G.; Chen, F.; Cai, W. Positron Emission Tomography and Nanotechnology: A Dynamic Duo for Cancer Theranostics. *Adv. Drug Deliv. Rev.* **2017**, *113*, 157–176. [\[CrossRef\]](#)
204. Gawne, P.J.; Man, F.; Blower, P.J.; de Rosales, R.T.M. Direct Cell Radiolabeling for in Vivo Cell Tracking with PET and SPECT Imaging. *Chem. Rev.* **2022**, *122*, 10266–10318. [\[CrossRef\]](#)
205. Chen, D.; Yang, D.; Dougherty, C.A.; Lu, W.; Wu, H.; He, X.; Cai, T.; Van Dort, M.E.; Ross, B.D.; Hong, H. In Vivo Targeting and Positron Emission Tomography Imaging of Tumor with Intrinsically Radioactive Metal–Organic Frameworks Nanomaterials. *ACS Nano* **2017**, *11*, 4315–4327. [\[CrossRef\]](#) [\[PubMed\]](#)
206. Sun, X.; Cai, W.; Chen, X. Positron Emission Tomography Imaging Using Radiolabeled Inorganic Nanomaterials. *Accounts Chem. Res.* **2015**, *48*, 286–294. [\[CrossRef\]](#)
207. Carregal-Romero, S.; Miguel-Coello, A.B.; Martínez-Parra, L.; Martí-Mateo, Y.; Hernansanz-Agustín, P.; Fernández-Afonso, Y.; Plaza-García, S.; Gutiérrez, L.; del Mar Muñoz-Hernández, M.; Carrillo-Romero, J.; et al. Ultrasmall Manganese Ferrites for In Vivo Catalase Mimicking Activity and Multimodal Bioimaging. *Small* **2022**, *18*, 2106570. [\[CrossRef\]](#)

208. Tang, W.; Fan, W.; Zhang, W.; Yang, Z.; Li, L.; Wang, Z.; Chiang, Y.; Liu, Y.; Deng, L.; He, L.; et al. Wet/Sono-Chemical Synthesis of Enzymatic Two-Dimensional MnO₂ Nanosheets for Synergistic Catalysis-Enhanced Phototheranostics. *Adv. Mater.* **2019**, *31*, 1900401. [[CrossRef](#)] [[PubMed](#)]
209. Lee, J.; Kim, J.; Heo, I.; Kim, S.J.; Jang, S.; Lee, H.-Y.; Jang, K.-S.; Yang, C.-S.; Lee, Y.; Yoo, W.C.; et al. One-Pot Bifunctionalization of Silica Nanoparticles Conjugated with Bioorthogonal Linkers: Application in Dual-Modal Imaging. *Biomater. Sci.* **2022**, *10*, 3540–3546. [[CrossRef](#)]
210. Boehm, K.M.; Khosravi, P.; Vanguri, R.; Gao, J.; Shah, S.P. Harnessing Multimodal Data Integration to Advance Precision Oncology. *Nat. Rev. Cancer* **2022**, *22*, 114–126. [[CrossRef](#)]
211. Wu, Y.; Zeng, F.; Zhao, Y.; Wu, S. Emerging Contrast Agents for Multispectral Optoacoustic Imaging and Their Biomedical Applications. *Chem. Soc. Rev.* **2021**, *50*, 7924–7940. [[CrossRef](#)] [[PubMed](#)]
212. Bax, J.J.; Di Carli, M.; Narula, J.; Delgado, V. Multimodality Imaging in Ischaemic Heart Failure. *Lancet* **2019**, *393*, 1056–1070. [[CrossRef](#)] [[PubMed](#)]
213. Smith, B.R.; Gambhir, S.S. Nanomaterials for In Vivo Imaging. *Chem. Rev.* **2017**, *117*, 901–986. [[CrossRef](#)] [[PubMed](#)]
214. Fan, W.; Yung, B.; Huang, P.; Chen, X. Nanotechnology for Multimodal Synergistic Cancer Therapy. *Chem. Rev.* **2017**, *117*, 13566–13638. [[CrossRef](#)] [[PubMed](#)]
215. Lo, P.-C.; Rodríguez-Morgade, M.S.; Pandey, R.K.; Ng, D.K.P.; Torres, T.; Dumoulin, F. The Unique Features and Promises of Phthalocyanines as Advanced Photosensitisers for Photodynamic Therapy of Cancer. *Chem. Soc. Rev.* **2020**, *49*, 1041–1056. [[CrossRef](#)]
216. Cheng, Y.; Kong, X.; Chang, Y.; Feng, Y.; Zheng, R.; Wu, X.; Xu, K.; Gao, X.; Zhang, H. Spatiotemporally Synchronous Oxygen Self-Supply and Reactive Oxygen Species Production on Z-Scheme Heterostructures for Hypoxic Tumor Therapy. *Adv. Mater.* **2020**, *32*, 1908109. [[CrossRef](#)] [[PubMed](#)]
217. Yang, H.; Xu, B.; Li, S.; Wu, Q.; Lu, M.; Han, A.; Liu, H. A Photoresponsive Nanozyme for Synergistic Catalytic Therapy and Dual Phototherapy. *Small* **2021**, *17*, 2007090. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.