

**Figure S1: Principle of the assay**

The first round of amplification used allele specific primers designed to obtain different fragment sizes for each genetic location. The last nucleotide of the 3'flanking forward primers was specific to the genotype (wild-type or a variant allele). The reverse primer was the same for both allele (wild-type or a variant allele). A Pigtail GTTTCTT was added to 5'flanking reverse primers to improve the amplicon migration. A second round of amplification used universal primers combined with fluorescent probes.

A M13 sequence (-20) GTAAAACGACGGCCAGT in the 5' flanking forward wild-type primers allowed hybridization and amplification of the wild type allele and a M13 sequence (-40) GTTTTCCCAGTCACGAC in the 5' flanking forward variant primers allowed hybridization and amplification of the variant allele. After denaturation, PCR products were loaded on a sequencer. The color of the fluorescence of the fragment allowed allelic discrimination and the size of the fragment allowed genetic position discrimination.

**Table S1: Repeatability results.** Eight internal control samples were analyzed in duplicate 4 different days.

QC1

QC2

ABCB1-rs2032582-GT	DM	100%							
ABCB1-rs2229109	WT	100%							
CYP1A2-1F	HM	100%							
CYP2C9-2	WT	100%							
CYP2C9-3	WT	100%							
CYP2C19-2	WT	100%							
CYP2C19-3	WT	100%							
CYP2C19-17	DM	100%							
CYP2D6-3	WT	100%							
CYP2D6-4	DM	100%							
CYP2D6-6	HM	100%							
CYP2D6-9	WT	100%							
CYP3A4-22	WT	100%							
CYP3A5	DM	100%							
VKORC1-rs9923231	HM	100%							

### QC3

position	Day1		Day2		Day3		Day4		repeatability
	status R1	status R2							
ABCB1-rs1045642	DM	100%							
ABCB1-rs1128503	DM	100%							
ABCB1-rs2032582-A	-	-	-	-	-	-	-	-	100%
ABCB1-rs2032582-GT	DM	100%							
ABCB1-rs2229109	WT	100%							
CYP1A2-1F	DM	100%							
CYP2C9-2	DM	100%							
CYP2C9-3	WT	100%							
CYP2C19-2	WT	100%							
CYP2C19-3	WT	100%							
CYP2C19-17	WT	100%							
CYP2D6-3	WT	100%							
CYP2D6-4	WT	100%							
CYP2D6-6	WT	100%							
CYP2D6-9	WT	100%							
CYP3A4-22	HM	100%							
CYP3A5	DM	100%							
VKORC1-rs9923231	HM	100%							

### QC4

QC5

QC6

QC7

CYP3A4-22	WT	100%								
CYP3A5	HM	100%								
VKORC1-rs9923231	HM	100%								

QC8

position	Day1		Day2		Day3		Day4		repeatability
	status R1	status R2							
ABCB1-rs1045642	WT	100%							
ABCB1-rs1128503	WT	100%							
ABCB1-rs2032582-A	-	-	-	-	-	-	-	-	100%
ABCB1-rs2032582-GT	WT	100%							
ABCB1-rs2229109	HM	100%							
CYP1A2-1F	HM	100%							
CYP2C9-2	WT	100%							
CYP2C9-3	WT	100%							
CYP2C19-2	WT	100%							
CYP2C19-3	HM	100%							
CYP2C19-17	HM	100%							
CYP2D6-3	WT	100%							
CYP2D6-4	WT	100%							
CYP2D6-6	WT	100%							
CYP2D6-9	WT	100%							
CYP3A4-22	HM	100%							
CYP3A5	DM	100%							
VKORC1-rs9923231	DM	100%							

WT: wild-type allele; varHz: heterozygous for the variant allele; varHm: homozygous for the variant allele

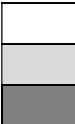
QC: internal quality control DNA

R: replicate

**Table S2: Genotyping results of clinical cases and genes involved in the pharmacokinetics of treatments**

	ABCB 1- rs1045 642	ABCB 1- rs1128 503	ABCB 1- rs2032 582-A	ABCB 1- rs2032 582-GT	ABC B1- rs222 9109	CYP1 A2*1F	CYP2 C9*2	CYP2 C9*3	CYP2 C19*2	CYP2 C19*3	CYP2C 19*17	CYP2 D6*3	CYP2 D6*4	CYP2 D6*6	CYP2 D6*9	CYP3 A4*22	CYP3 A5*3	VKO RC1- rs992 3231
P1	WT	WT	<b>var Hm</b>	WT	WT	<b>var Hm</b>	WT	WT	WT	WT	<b>varH z</b>	<b>var Hz</b>	WT	WT	WT	WT	var Hm	<b>var Hz</b>
Haloperidol																		
Clozapine																		
P2	<b>var Hz</b>	<b>var Hz</b>	-	<b>var Hz</b>	WT	varH m	WT	WT	<b>varH z</b>	WT	WT	WT	WT	WT	WT	<b>var Hz</b>	var Hm	WT
P3	<b>var Hm</b>	<b>var Hm</b>	-	<b>var Hm</b>	WT	WT	var Hz	WT	WT	WT	WT	-	-	-	-	WT	var Hm	var Hz
Vincristine																		

Results of the PCR-Multiplex analysis



Polymorphism on a gene not significantly involved in the drug pharmacokinetics

Polymorphism on a gene moderately involved in the drug pharmacokinetics

Polymorphism on a gene strongly involved in the drug pharmacokinetics

P1, P2, P3: patient ID

WT: wild-type allele; varHz: heterozygous for the variant allele; varHm: homozygous for the variant allele  
(-): no amplification because of full gene deletion

**Table S3. Design of primers used in the assay**

Gene and variant	Primer ID	Primer sequences (5'-3')	amplicon size (pb)	Primer final concentration in the mix (pmol/μL)
CYP1A2*1F rs762551	F-CYP1A2*1F - WT	<u>GTAAAACGACGGCCAGTCAAAGGG</u> TGAGCTCTGTGGTCC	160	0.02
	F-CYP1A2*1F - var	<u>GTTTCCCAGTCACGACCAAAGGG</u> TGAGCTCTGTGGACA		0.02
	R-CYP1A2*1F	<u>GTTTCTTGGAGACATTCAATTCAATT</u> ATTGCC		0.3
CYP2C9*2 rs1799853	F-CYP2C9*2-WT	<u>GTAAAACGACGGCCAGTGGGAAGA</u> GGAGCATTGAGCACC	127	0.02
	F-CYP2C9*2-var	<u>GTTTCCCAGTCACGACGGGAAGA</u> GGAGCATTGAGCACT		0.01
	R-CYP2C9*2	<u>GTTTCTTAGGTCACTGATATGGAGT</u> AGGGT		0.2
CYP2C9*3 rs1057910	F-CYP2C9*3-WT	<u>GTAAAACGACGGCCAGTGTGCACG</u> AGGTCCAGAGAAACA	328	0.03
	F-CYP2C9*3-var	<u>GTTTCCCAGTCACGACGTGCACGA</u> GGTCCAGAGAAACC		0.01
	R-CYP2C9*3	<u>GTTTCTTAAATCTGGAGAACACAC</u> ACTGCC		0.4
CYP2C19*2 rs4244285	F-CYP2C19*2-WT	<u>GTAAAACGACGGCCAGTTCCC</u> ACTATTGATTATTGCCG	164	0.01
	F-CYP2C19*2-var	<u>GTTTCCCAGTCACGACTTCCC</u> ACTATTGATTATTGCCA		0.01
	R-CYP2C19*2	<u>GTTTCTTATCAATAAAAGTCCCGAGG</u> GTTGT		0.4
CYP2C19*3 rs4986893	F-CYP2C19*3-WT	<u>GTAAAACGACGGCCAGTGGATTGT</u> AAGCACCCCGTGG	183	0.01
	F-CYP2C19*3-var	<u>GTTTCCCAGTCACGACGGATTGT</u> AGCACCCCGTGA		0.005
	R-CYP2C19*3	<u>GTTTCTTCACCCCCATGGCTGTCTAG</u> G		0.2
CYP2C19*17 rs12248560	F-CYP2C19*17-WT	<u>GTAAAACGACGGCCAGTGGCGCAT</u> TATCTTACATCAGACATG	202	0.04
	F-CYP2C19*17-var	<u>GTTTCCCAGTCACGACGGCGCATT</u> ATCTTACATCAGACATA		0.01
	R-CYP2C19*17	<u>GTTTCTTGTTCTATTAAATGTGA</u> AGCCTGTTTATG		0.8
CYP2D6*3 rs35742686	F-CYP2D6*3-WT	<u>GTAAAACGACGGCCAGTGTGAGC</u> TGCTAACTGAGGACA	478	0.02
	F-CYP2D6*3-var	<u>GTTTCCCAGTCACGACGTGAGCT</u> GCTAACTGAGGACG		0.005
	R-CYP2D6*3	<u>GTTTCTCGGCCCCCTGCACTGTTTC</u>		0.2
CYP2D6*4 rs3892097	F-CYP2D6*4-WT	<u>GTAAAACGACGGCCAGTCCGCATC</u> TCCCACCCCTCAG	450	0.02
	F-CYP2D6*4-var	<u>GTTTCCCAGTCACGACCCGCATCT</u> CCCACCCCTCA		0.01
	R-CYP2D6*4	<u>GTTTCTTCTGACGTGGATAGGA</u> GGTACA		1.6
CYP2D6*6 rs5030655	F-CYP2D6*6-WT	<u>GTAAAACGACGGCCAGTCCTCCTCG</u> GTCAGCCA	227	0.04
	F-CYP2D6*6-var	<u>GTTTCCCAGTCACGACCCCTCCTCG</u> GTCAGCCC		0.01
	R-CYP2D6*6	<u>GTTTCTTCAGGGGGAGCATAGGGT</u> T		2
CYP2D6*9 rs5030656	F-CYP2D6*9-WT	<u>GTAAAACGACGGCCAGTCTCCTGG</u> CAGAGATGGACAAAG	410	0.02
	F-CYP2D6*9-var	<u>GTTTCCCAGTCACGACCCCTCCTG</u> GCAGAGATCGAG		0.01
	R-CYP2D6*9	<u>GTTTCTCGGCCCCCTGCACTGTTTC</u>		1.6

CYP3A4*22 rs35599367	F-CYP3A4*22-WT	<b>GTAAAACGACGGCCAGT</b> AGTGTCT CCATCACACCGAG <b>C</b>	118	0.02
	F-CYP3A4*22-var	<b>GTTTCCCCAGTCACGAC</b> AGTGTCTC CATCACACCGAG <b>T</b>		0.01
	R-CYP3A4*22	<b>GTTTCTT</b> GATCTACTAGATCACCTTC TATCACACTCCA		0.4
CYP3A5*3 rs776746	F-CYP3A5-WT	<b>GTAAAACGACGGCCAGT</b> TGTGGTC CAAACAGGGAAAGAGTT <b>A</b>	305	0.02
	F-CYP3A5-var	<b>GTTTCCCCAGTCACGAC</b> ACTGTGGTCC AAACAGGGAAAGAGTT <b>A</b>		0.01
	R-CYP3A5	<b>GTTTCTT</b> AGATGACACAGCTCTAGA TGTC		0.8
VKORC1- rs9923231	F-VKORC1-rs9923231-WT	<b>GTAAAACGACGGCCAGT</b> GACCTGA AAAACAACCATTGG <b>ACG</b>	397	0.01
	F-VKORC1-rs9923231-var	<b>GTTTCCCCAGTCACGAC</b> GACCTGAA AAACAACCATTGG <b>AC</b>		0.04
	R-VKORC1-rs9923231	<b>GTTTCTT</b> CCTGACACCTAGTGGCTG GT		1.6
ABCB1- rs1045642	F-ABCB1-rs1045642-var	<b>GTAAAACGACGGCCAGT</b> CTCCTTTG CTGCCCT <b>GAC</b>	319	0.02
	F-ABCB1-rs1045642-WT	<b>GTTTCCCCAGTCACGAC</b> CTCCTTTG CTGCCCT <b>GAC</b>		0.005
	R-ABCB1-rs1045642	<b>GTTTCTT</b> CACACAAACTTTCTTA ATCTCA		0.8
ABCB1- rs1128503	F-ABCB1-rs1128503-var	<b>GTAAAACGACGGCCAGT</b> ACTCTGC ACCTCAGGTT <b>GAGA</b>	338	0.02
	F-ABCB1-rs1128503-WT	<b>GTTTCCCCAGTCACGAC</b> ACTCTGCA CCTCAGGTT <b>GAGG</b>		0.005
	R-ABCB1-rs1128503	<b>GTTTCTT</b> AGCCAAGTATTGACAGCT ATTCG		0.6
ABCB1- rs2032582	F-ABCB1-rs2032582-var-T	<b>GTAAAACGACGGCCAGT</b> ATTAGTT TGACTCACCTCCGAG <b>A</b>	424	0.02
	F-ABCB1-rs2032582-WT-G	<b>GTTTCCCCAGTCACGAC</b> ATTAGTT GACTCACCTCCGAG <b>C</b>		0.005
	F-ABCB1-rs2032582-var-A	<b>GTTTCCCCAGTCACGAC</b> CATATTAA GTTTGACTCACCTCCCT <b>G</b>	427	0.005
	R-ABCB1-rs2032582	<b>GTTTCTT</b> CTGAAGTCATGGAAATTCT TACTGT	424/427	0.8
ABCB1- rs2229109	F-ABCB1-rs2229109-WT	<b>GTAAAACGACGGCCAGT</b> CCTTAACT TCTTTCGAGATGGAA <b>A</b>	360	0.02
	F-ABCB1-rs2229109-var	<b>GTTTCCCCAGTCACGAC</b> CCTTAACT TCTTTCGAGATGGAA <b>T</b>		0.01
	R-ABCB1-rs2229109	<b>GTTTCTT</b> GGACAGGCATCTCCAAGC AT		0.4

F: forward; R: reverse; WT: wild type; var: variant  
**GTAAAACGACGGCCAGT**: M13 sequence (-20) added to 5' flanking forward primers  
**GTTTCCCCAGTCACGAC**: M13 sequence (-40) added to the 5' flanking forward primers  
**GTTTCTT**: "Pigtail" added to 5' flanking reverse primers  
**X**: mismatch nucleotide in bold  
**X**: position of the polymorphism  
For the design of primers, the term "variant" was assigned to the lowest frequent allele compared to wild type in the total human population and according to the database dbSNP  
<https://www.ncbi.nlm.nih.gov/snp/>