

Supporting Information

Antimicrobial activity of a library of thioxanthenes and their potential as efflux pump inhibitors

Fernando Durães ^{1,2}, Andreia Palmeira ^{1,2}, Bárbara Cruz ³, Joana Freitas-Silva ^{2,4}, Nikoletta Szemerédi ⁵, Luís Gales ^{6,7}, Paulo Martins da Costa ^{2,4}, Fernando Remião ³, Renata Silva ³, Madalena Pinto ^{1,2}, Gabriella Spengler ^{5 *}, Emília Sousa ^{1,2 *}

¹ Laboratory of Organic and Pharmaceutical Chemistry, Department of Chemical Sciences, Faculty of Pharmacy, University of Porto, Rua de Jorge Viterbo Ferreira, 228, 4050-313 Porto, Portugal; fduraes5@gmail.com (F. D.); apalmeira@ff.up.pt (A.P.).

² CIIMAR-Interdisciplinary Centre of Marine and Environmental Research, University of Porto, Novo Edifício do Terminal de Cruzeiros do Porto de Leixões, Avenida General Norton de Matos, S/N, 4450-208 Matosinhos, Portugal; madalena@ff.up.pt (M.P.), joanafreitasdasilva@gmail.com (J.F.-S.).

³ UCIBIO-REQUIMTE, Laboratory of Toxicology, Faculty of Pharmacy, University of Porto, Rua de Jorge Viterbo Ferreira 228, 4050-313 Porto, Portugal; up201607633@med.up.pt (B.C.); remiao@ff.up.pt (F.R.), rsilva@ff.up.pt (R.S.)

⁴ ICBAS – Institute of Biomedical Sciences Abel Salazar, Universidade do Porto, Rua de Jorge Viterbo Ferreira 228, 4050-313 Porto, Portugal; pmcosta@icbas.up.pt (P.M.C.).

⁵ Department of Medical Microbiology and Immunobiology, Faculty of Medicine, University of Szeged, Dóm tér 10, 6720 Szeged, Hungary; szemeredi.nikoletta@med.u-szeged.hu (N. S.).

⁶ Department of Molecular Biology, ICBAS - Instituto de Ciências Biomédicas Abel Salazar, University of Porto, Porto, Portugal; lgales@ibmc.up.pt (L.G.).

⁷ Bioengineering & Synthetic Microbiology, I3S – Instituto de Investigação e Inovação em Saúde, University of Porto, Porto, Portugal

* Correspondence: spengler.gabriella@med.u-szeged.hu (G. S.); esousa@ff.up.pt (E. S.).

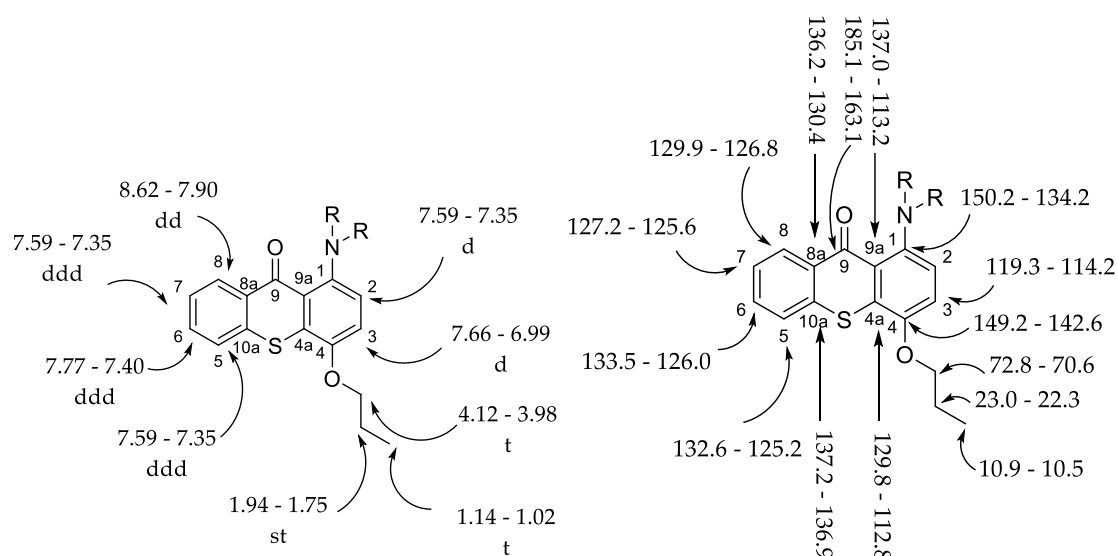


Figure S1. Main ^1H (left) and ^{13}C (right) signals for the 1-nitrogen substituted thioxanthenes **8-14**.

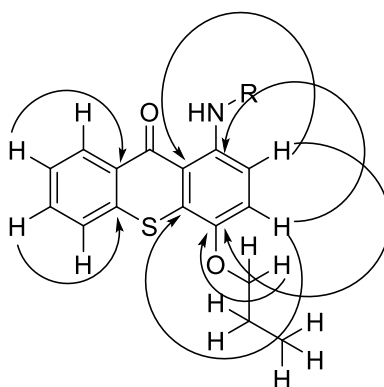


Figure S2. Main connectivities found in the HMBC for the thioxanthone scaffold used in this work.

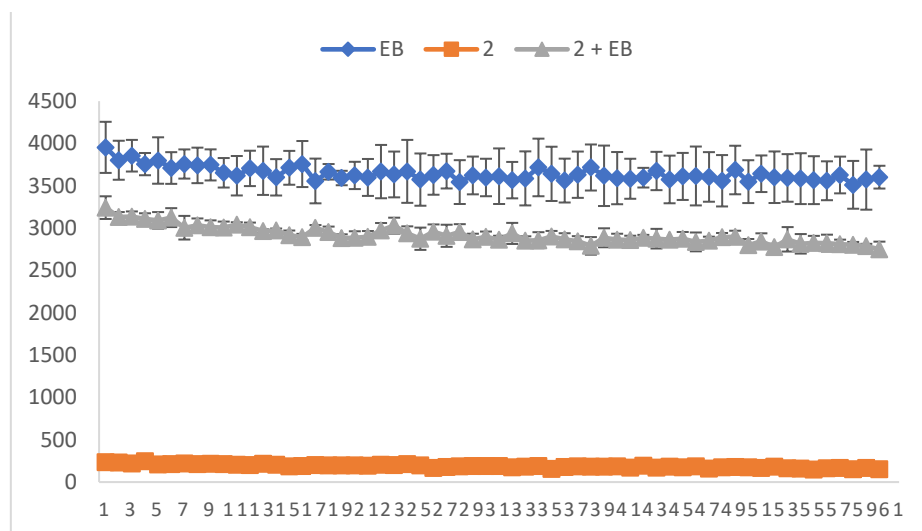


Figure S3. Evaluation of the fluorescence of compound **2** alone and in combination with EB.

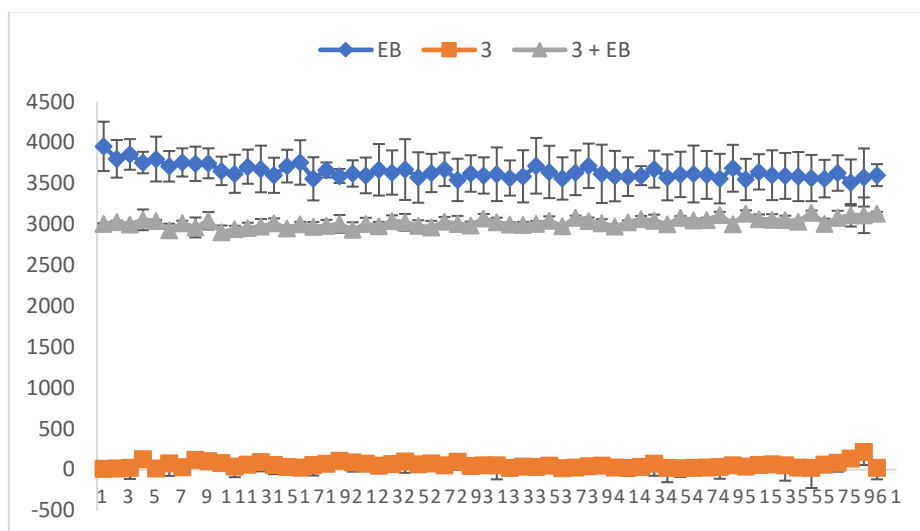


Figure S4. Evaluation of the fluorescence of compound **3** alone and in combination with EB.

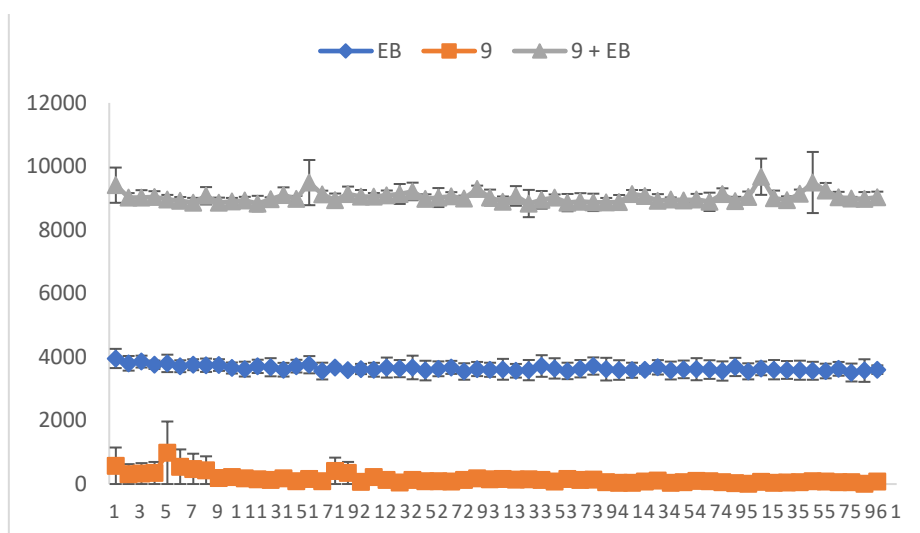


Figure S5. Evaluation of the fluorescence of compound **9** alone and in combination with EB.

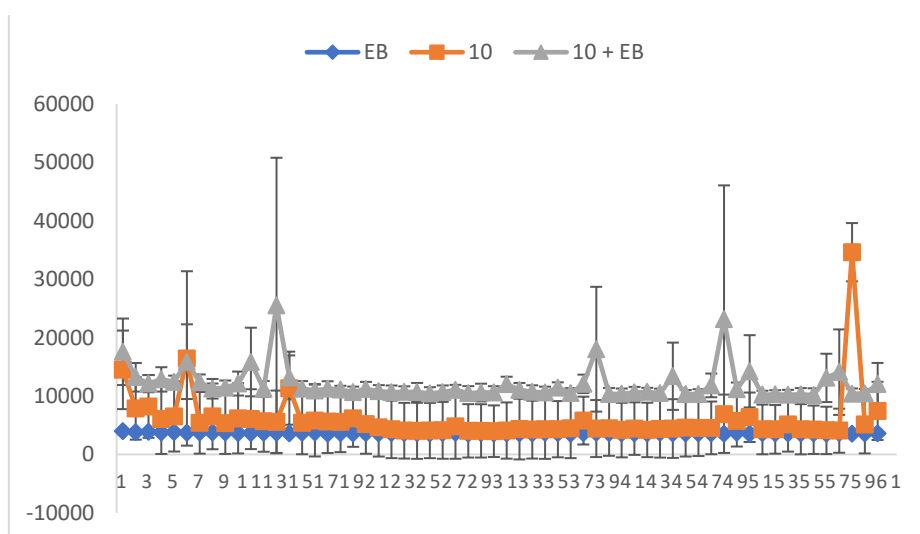


Figure S6. Evaluation of the fluorescence of compound **10** alone and in combination with EB.

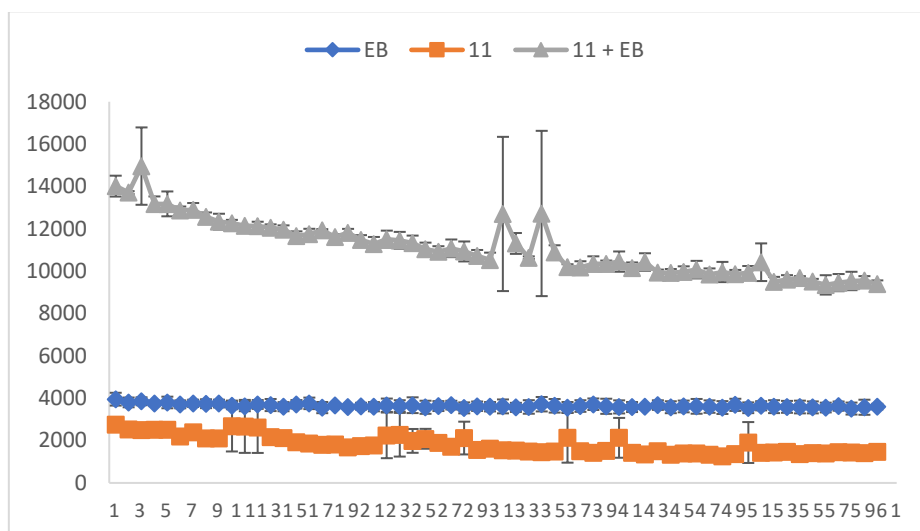


Figure S7. Evaluation of the fluorescence of compound **11** alone and in combination with EB.

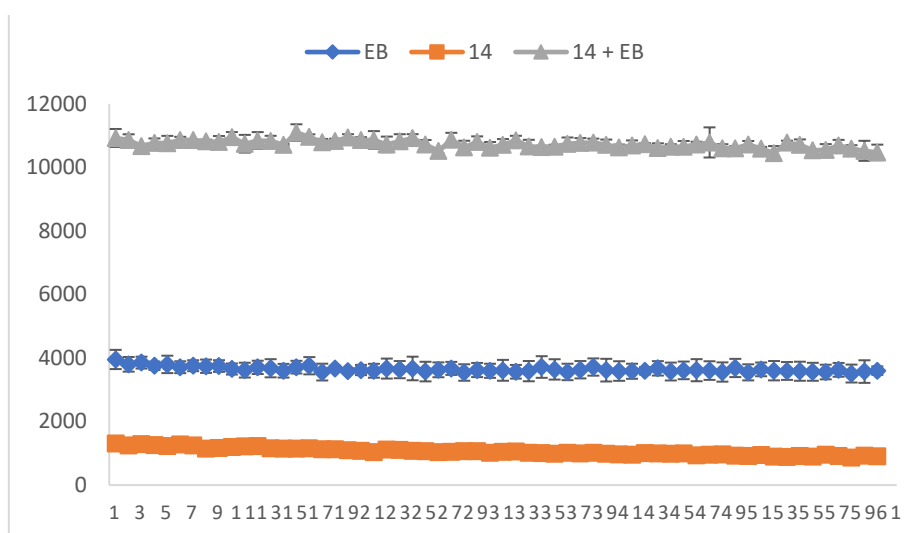


Figure S8. Evaluation of the fluorescence of compound **14** alone and in combination with EB.

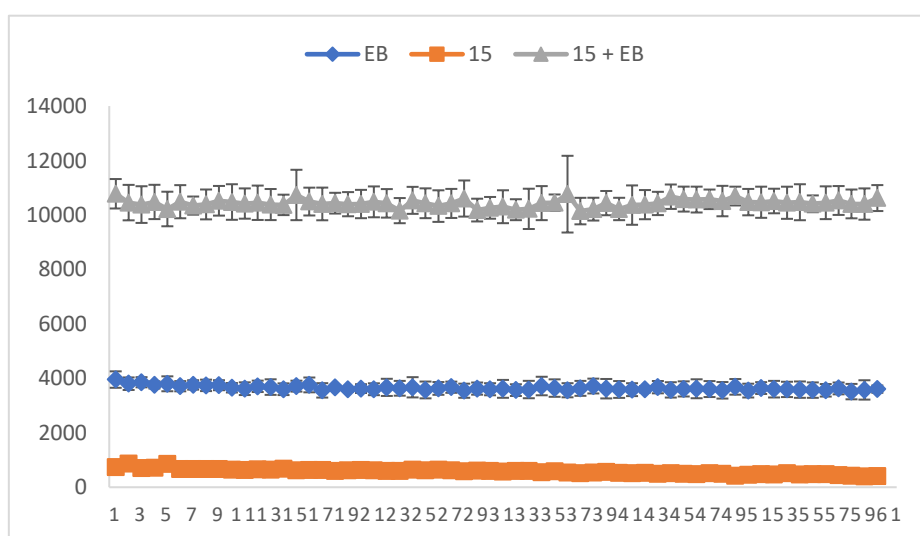


Figure S9. Evaluation of the fluorescence of compound **15** alone and in combination with EB.

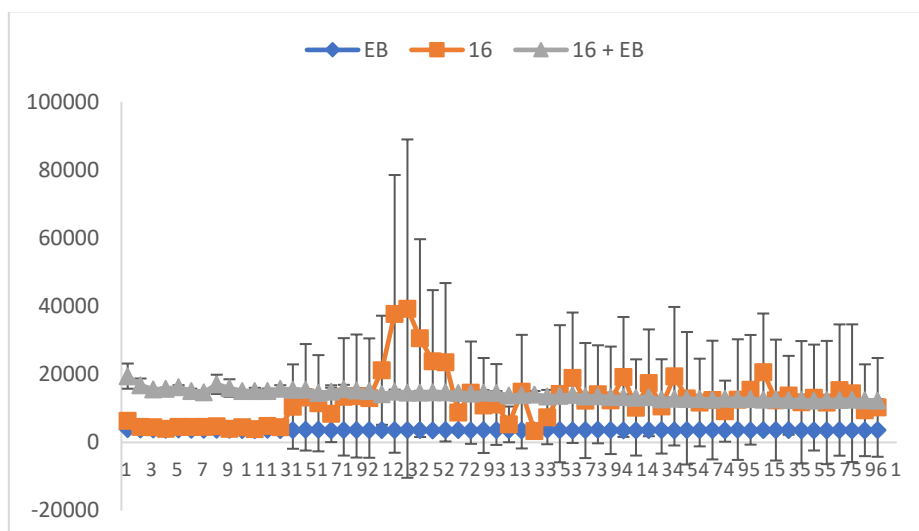


Figure S10. Evaluation of the fluorescence of compound **16** alone and in combination with EB.

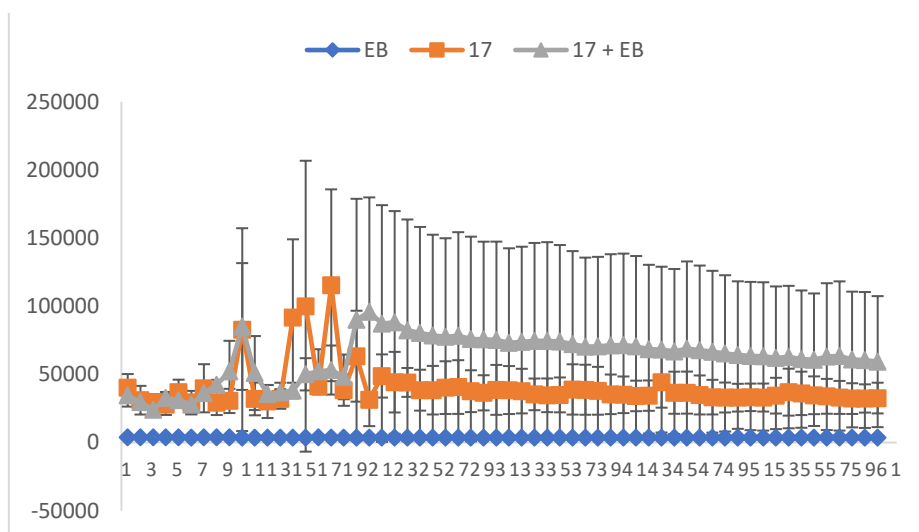


Figure S11. Evaluation of the fluorescence of compound **17** alone and in combination with EB.

Molecular visualization of compounds **3, **7**, **8**, and **13** into the substrate-binding site of AcrB:**

Compounds **3**, **7**, **8** and **13** were visualized in the SBS of AcrB. As it can be seen in **Figure 4, D**, the compounds are predicted to bind in the approximate site, but it is suggested that they interact with different residues. This is clarified when the compounds are analyzed individually.

Compound **3** (**Figure S11, A**) is predicted to only interact with Arg-620, through a hydrogen bond between the oxygen in the propoxy chain and a nitrogen in this residue. While it is true that all the compounds present the propoxy chain, it is also noteworthy that compound **3** has the simplest and most flexible substituent out of all this group of compounds, which can possibly allow it to get to this site within the SBS.

Compound **7** (**Figure S12, B and C**), hydrogen bonds can be seen between the carbonyl in C-9 and Gln-176, the oxygen in the propoxy chain in C-4 and Asn-274, and between the ketone of the substituent in C-1 and Gln-89.

For compound **8** (**Figure S12, D and E**), it can be noted that the nitrogens within the pyrimidine moiety form dipole interactions with Gln-89 and Gln-176, and the carbonyl in C-9 of the thioxanthone moiety forms a hydrogen bond with the amine of Gln-89. Additionally, there is a T-shaped π - π interaction between the pyrimidine substituent and Phe-615.

Lastly, for compound **13** (**Figure S12, F and G**), many different hydrogen interactions can be seen. The oxygen in the propoxy chain interacts with Gln-89, like **7** and **8**. The oxygens in the sulfamide interact with Thr-87 and Arg-620, and the amine directly bonded to the aromatic ring

in the substituent can also interact with Ser-46. The carbonyl in C-9 can also interact with a water molecule, that can show hydrogen interactions with Gln-176 and Leu-177.

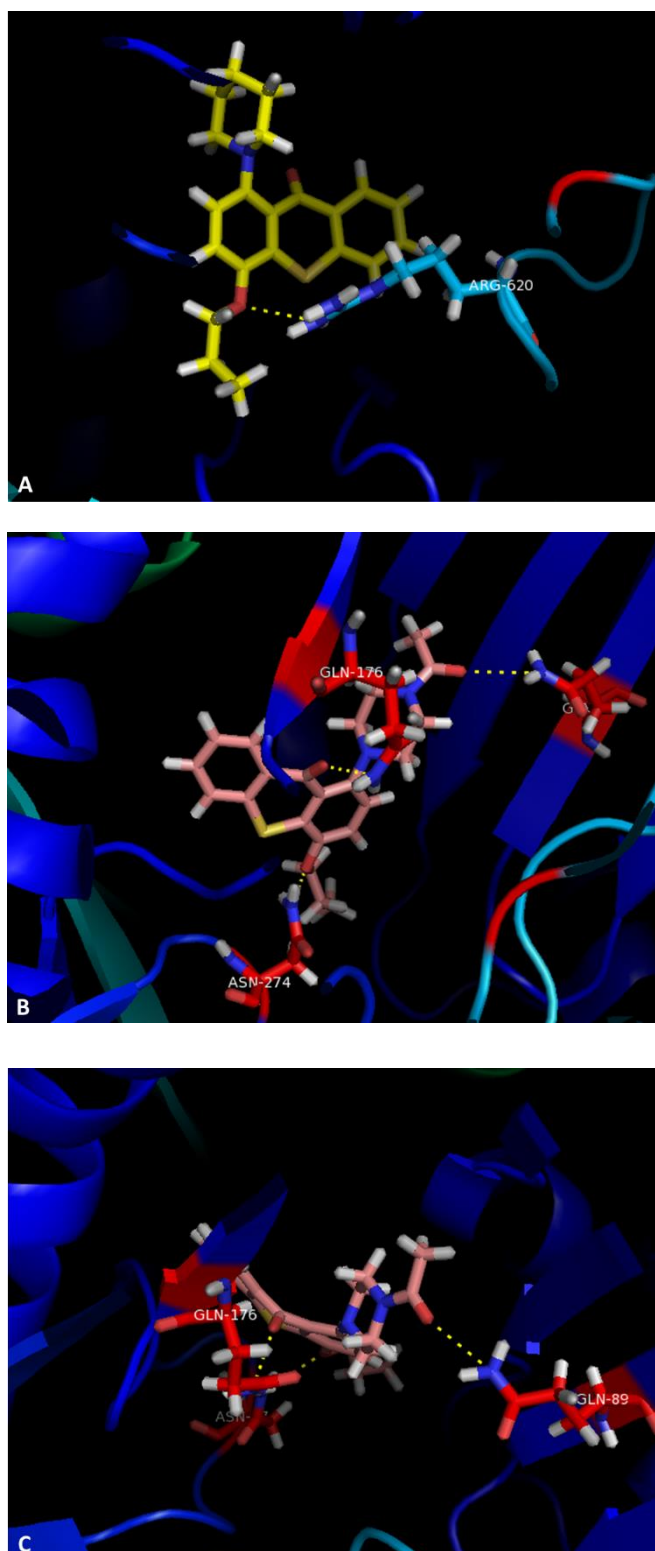


Figure S12. Molecular visualization in the SBS of AcrB. (A) Interaction between compound **3** and the SBS; (B) and (C) Different perspectives of the interactions between compound **7** and the SBS; (D) and (E) Different perspectives of the interactions between compound **8** and the SBS; (G) and (H) Different perspectives of the interactions between **13** and the SBS.

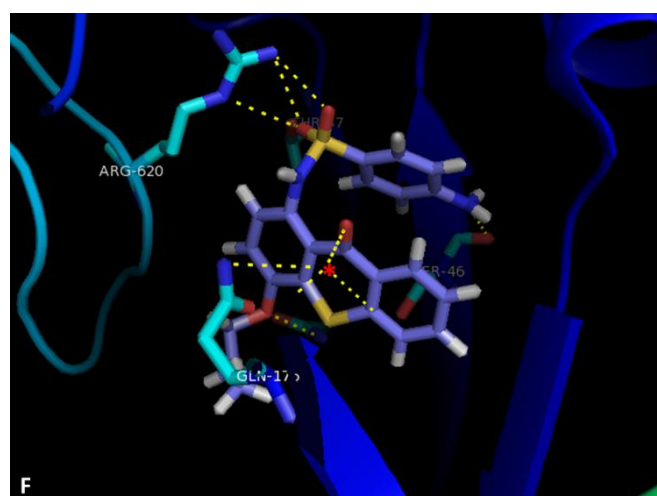
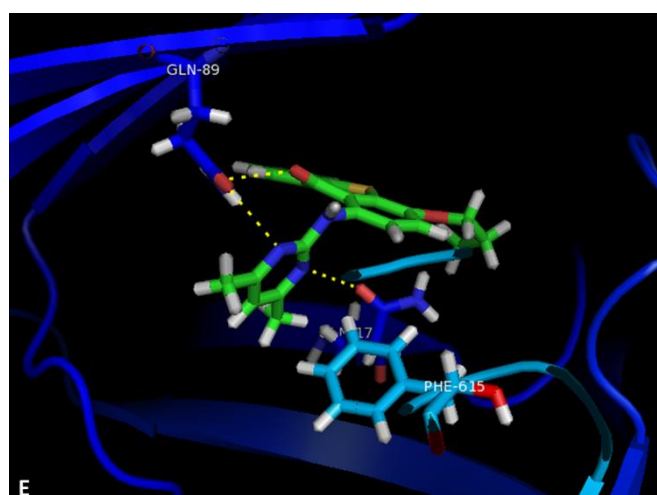
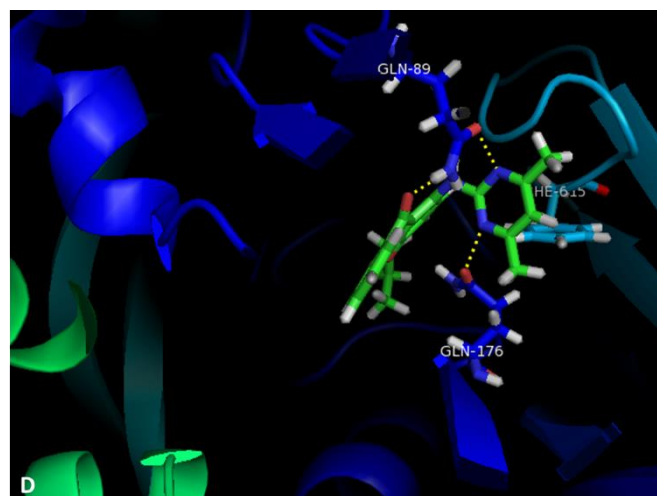


Figure S12. (cont'd)

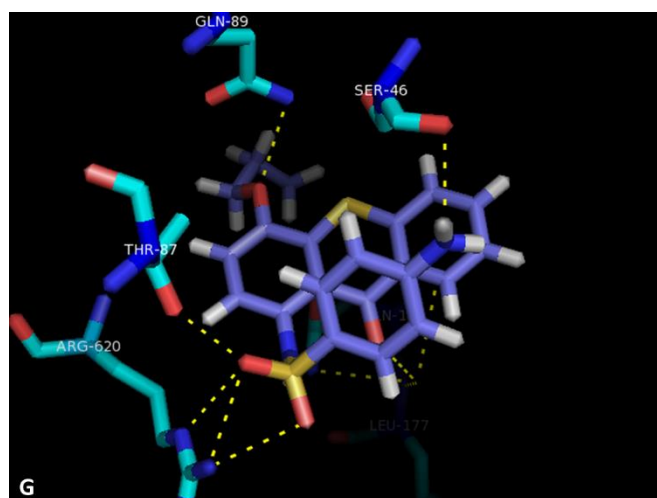


Figure S12. (cont'd)

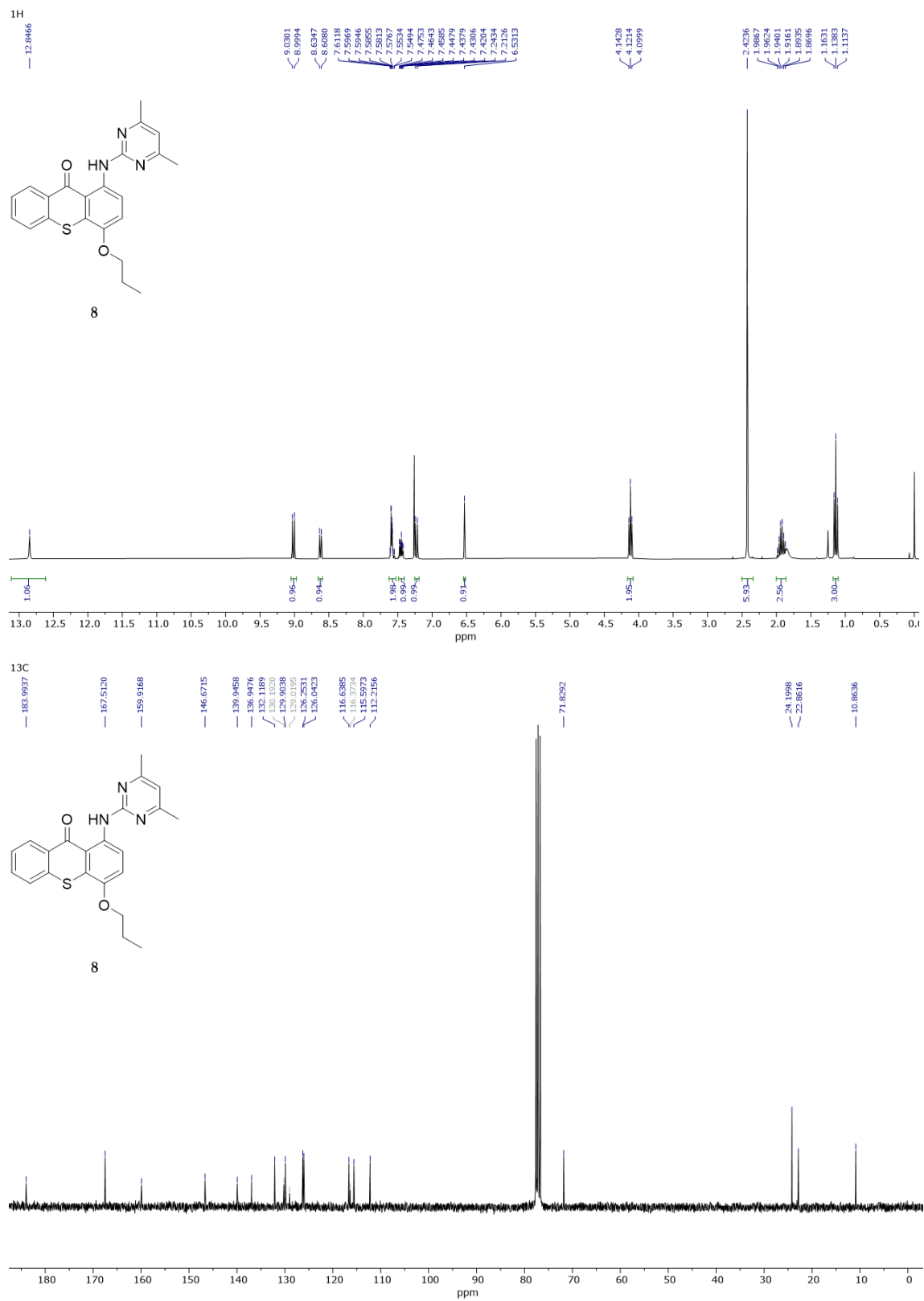


Figure S13. ¹H NMR (top, 300.13 MHz, CDCl₃) and ¹³C NMR (bottom, 75.48 MHz, CDCl₃) for compound **8**.

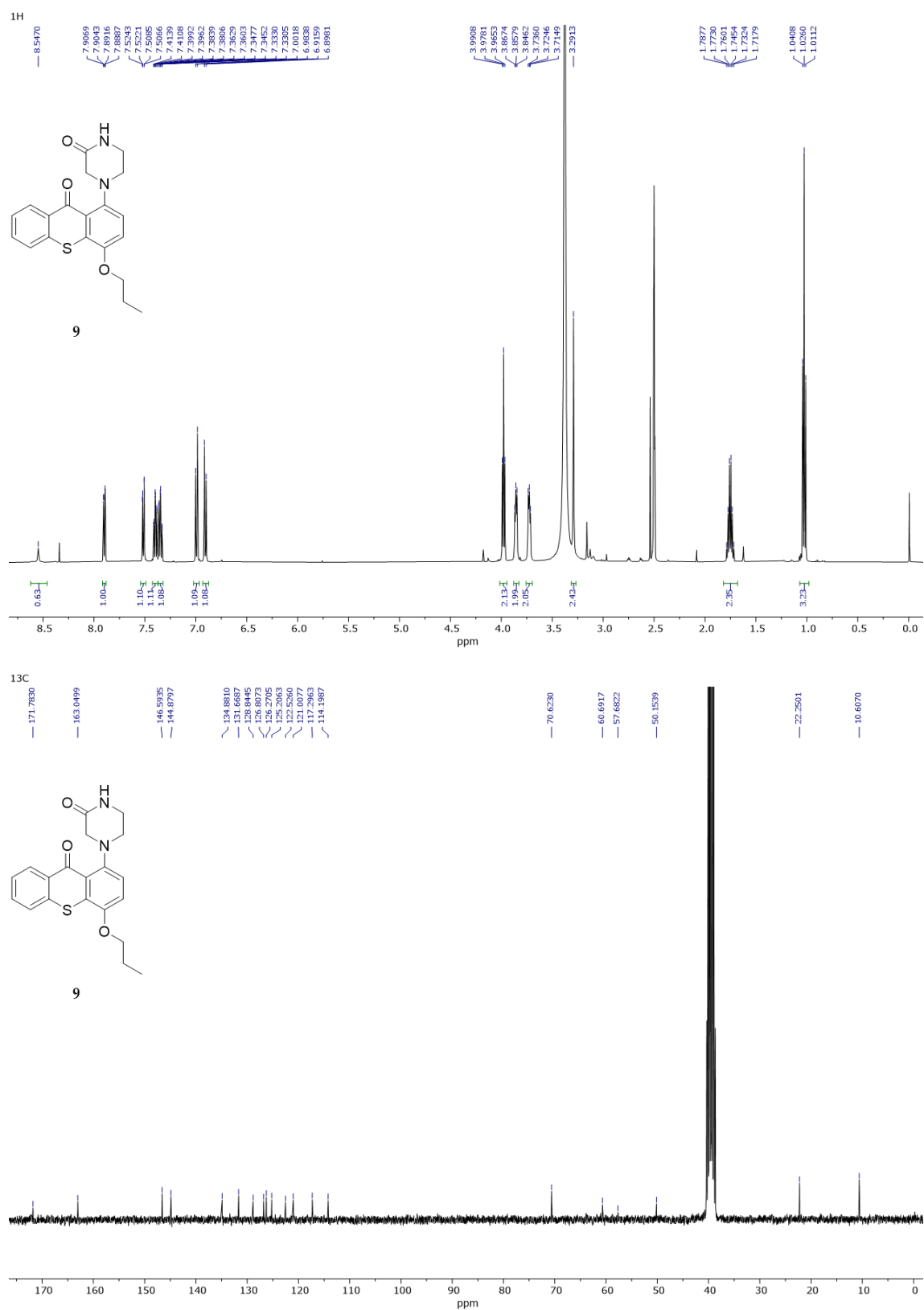
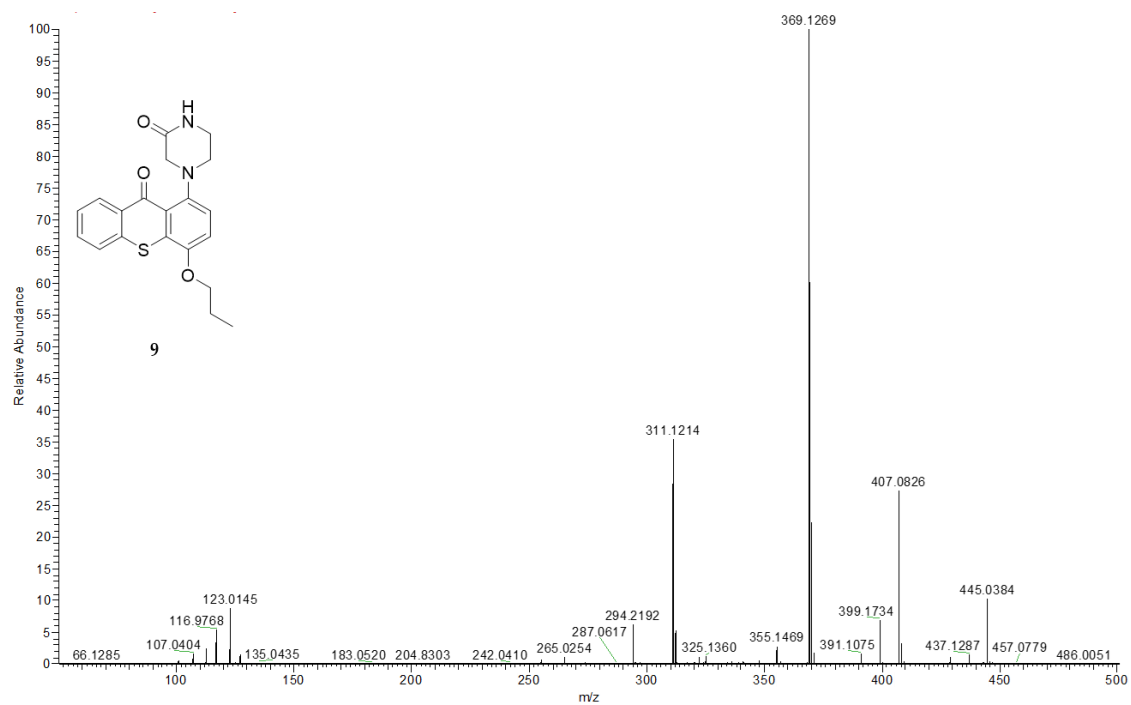


Figure S14. ¹H NMR (top, 500.13 MHz, DMSO) and ¹³C NMR (bottom, 75.48 MHz, DMSO) for compound 9.



Meas. m/z	Formula	m/z	err [ppm]
369.1269	C ₂₀ H ₂₁ N ₂ O ₃ S	369.1273	-1.08

Figure S15. Electrospray ESI data for compound 9.

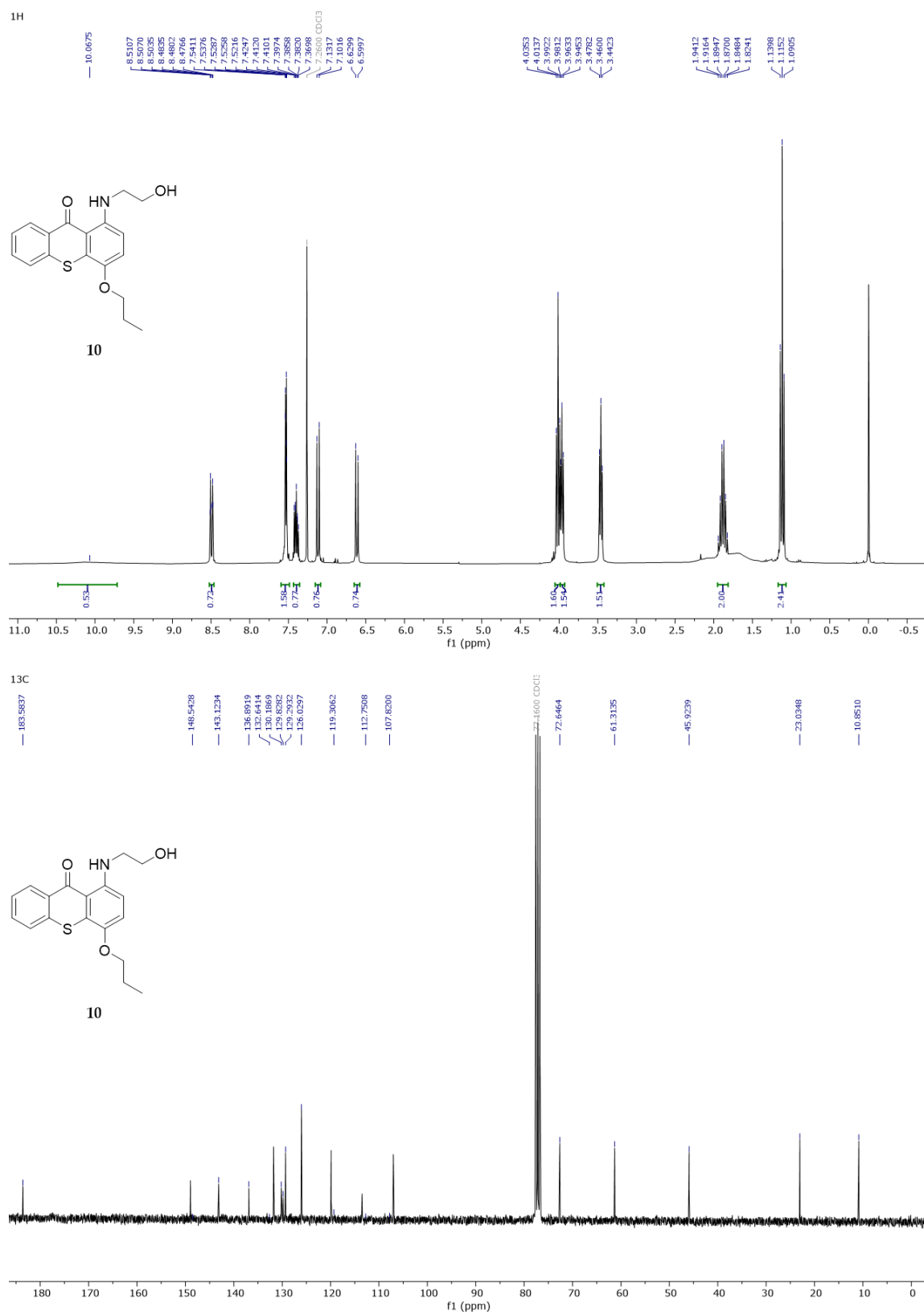
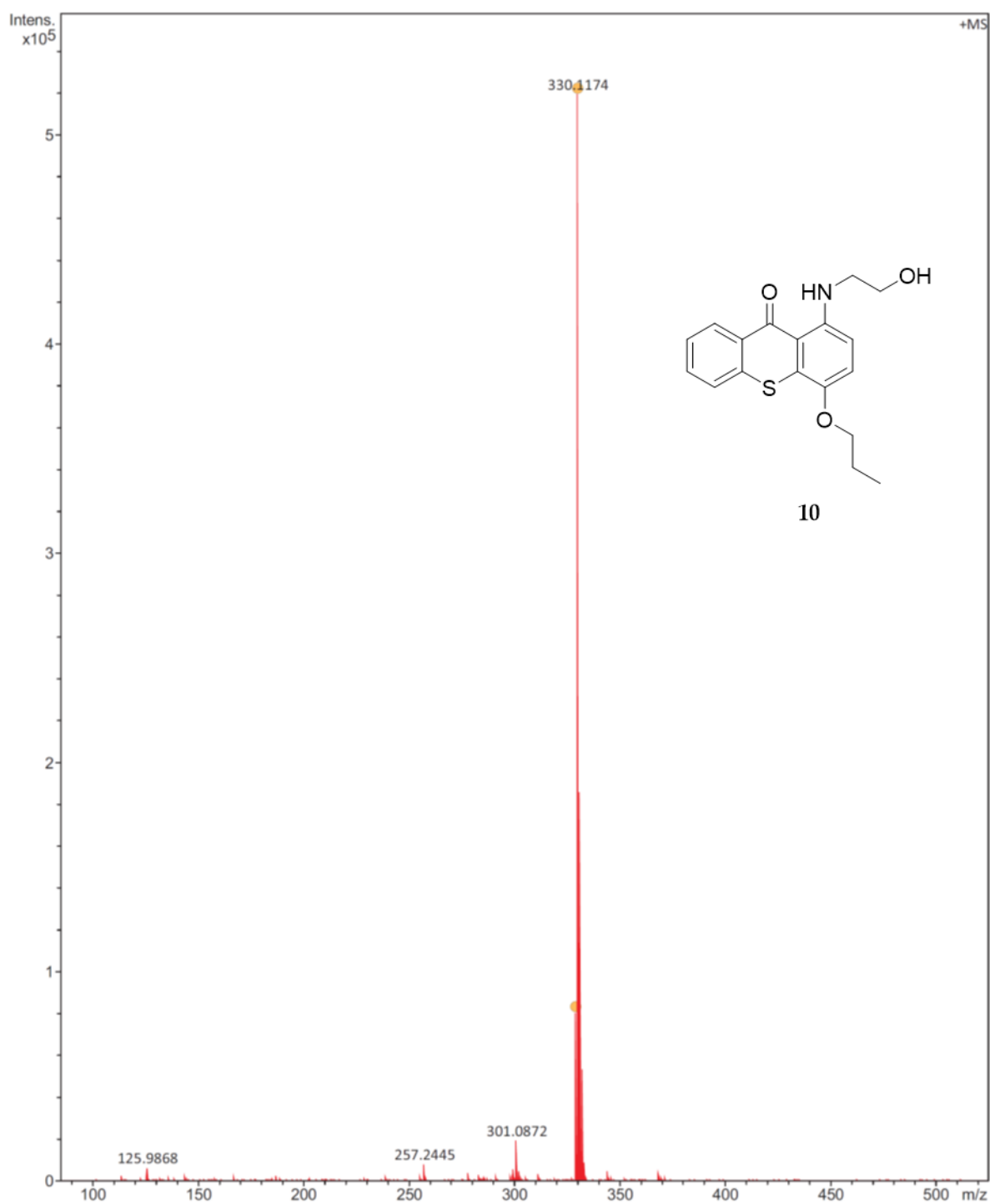


Figure S16. ¹H NMR (top, 300.13 MHz, CDCl₃) and ¹³C NMR (bottom, 75.48 MHz, CDCl₃) for compound 10.



Meas. m/z	Formula	m/z	err [ppm]
329.1066	C ₁₈ H ₁₉ NO ₃ S	329.10866	4.40

Figure S17. Electrospray ESI data for compound 10.

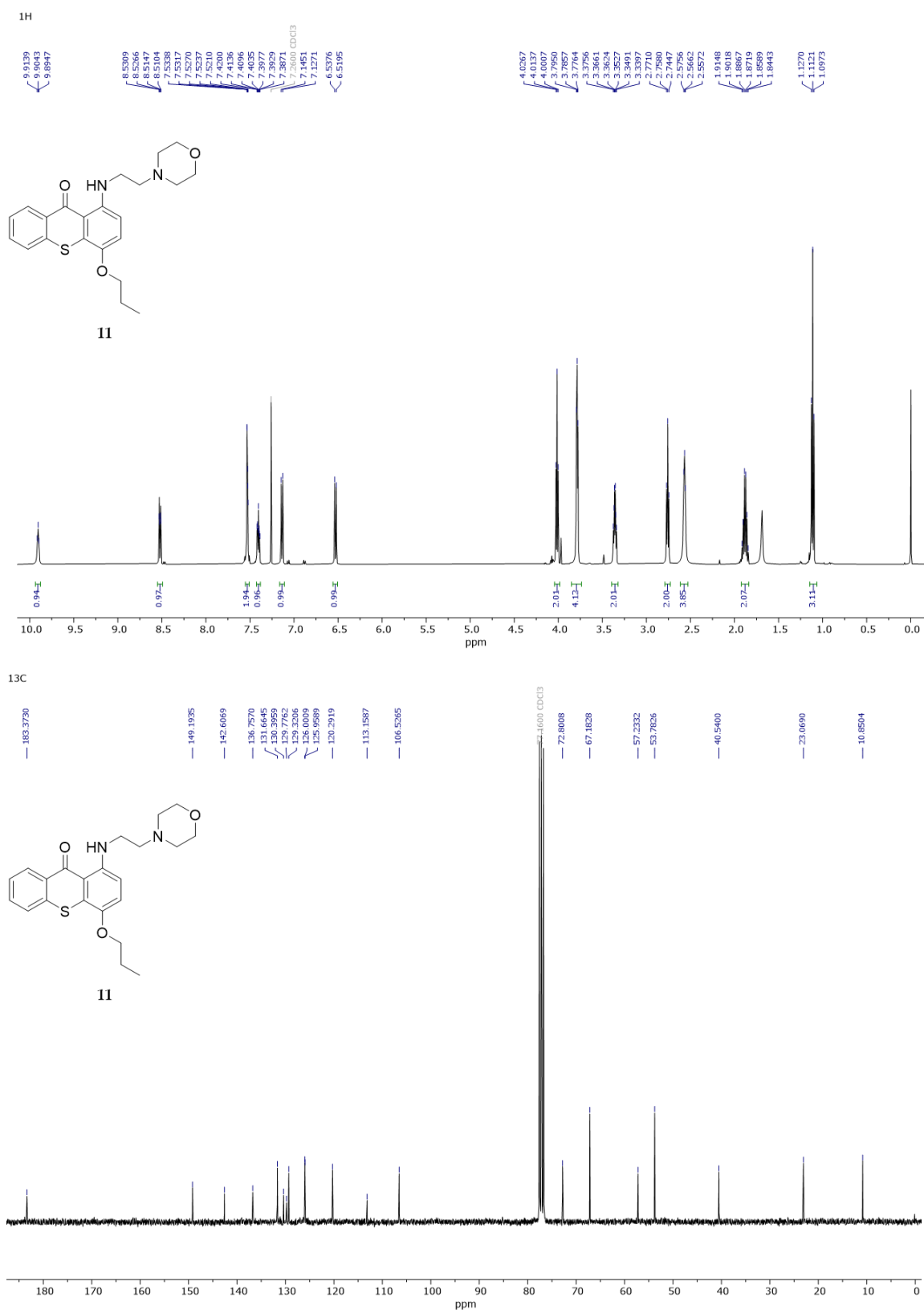
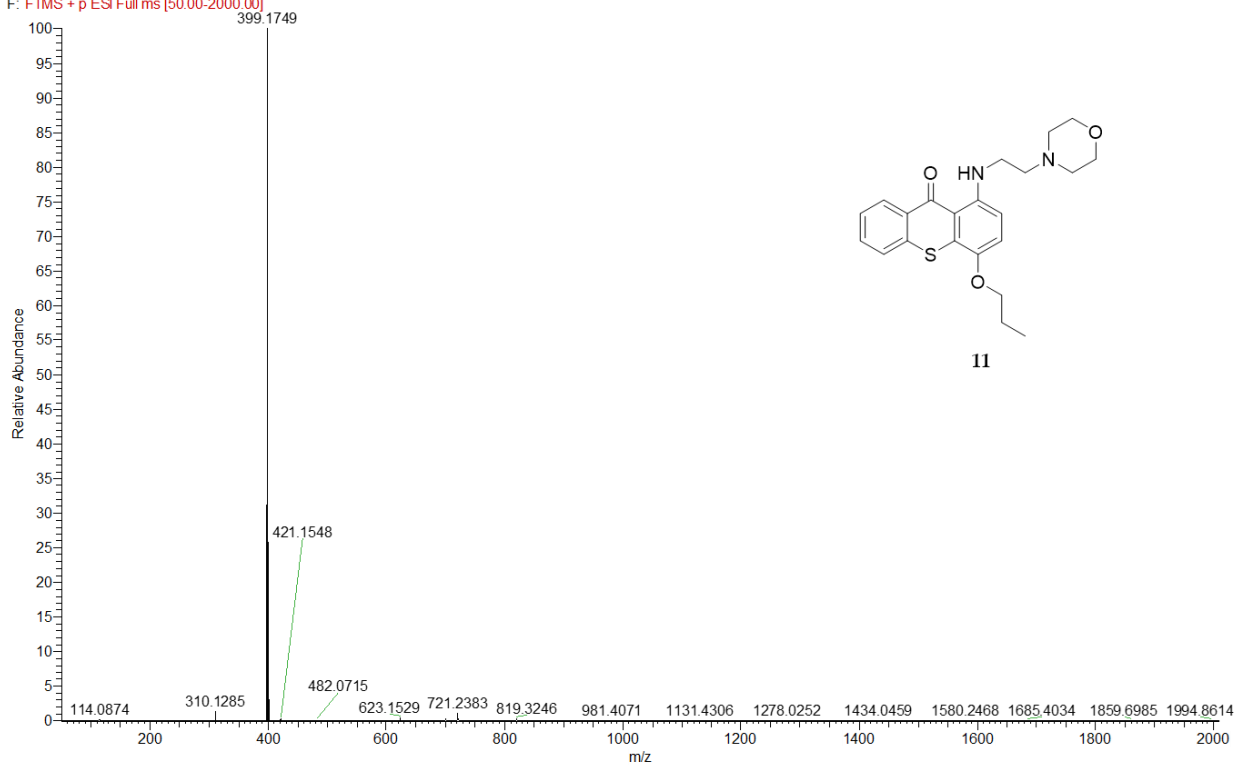


Figure S18. ¹H NMR (top, 500.13 MHz, CDCl₃) and ¹³C NMR (bottom, 75.48 MHz, CDCl₃) for compound **11**.

TX2-4A #3-5 RT: 0.07-0.12 AV: 3 NL: 2.61E8
F: FTMS + p ESI Full ms [50.00-2000.00]



Meas. m/z	Formula	m/z	err [ppm]
399.1749	C ₂₂ H ₂₇ N ₂ O ₃ S	399.1742	1.75

Figure S19. Electrospray ESI data for compound 11.

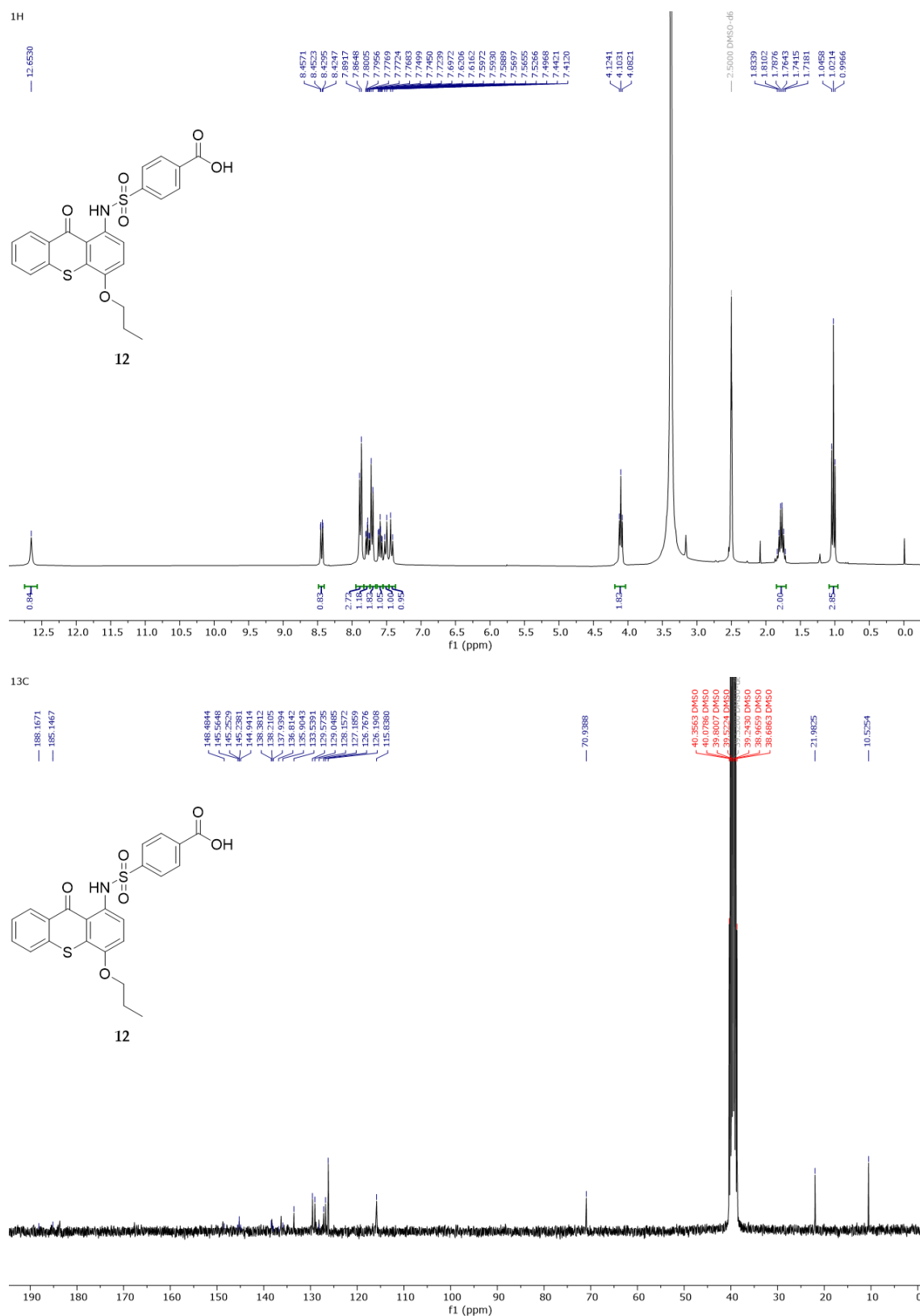
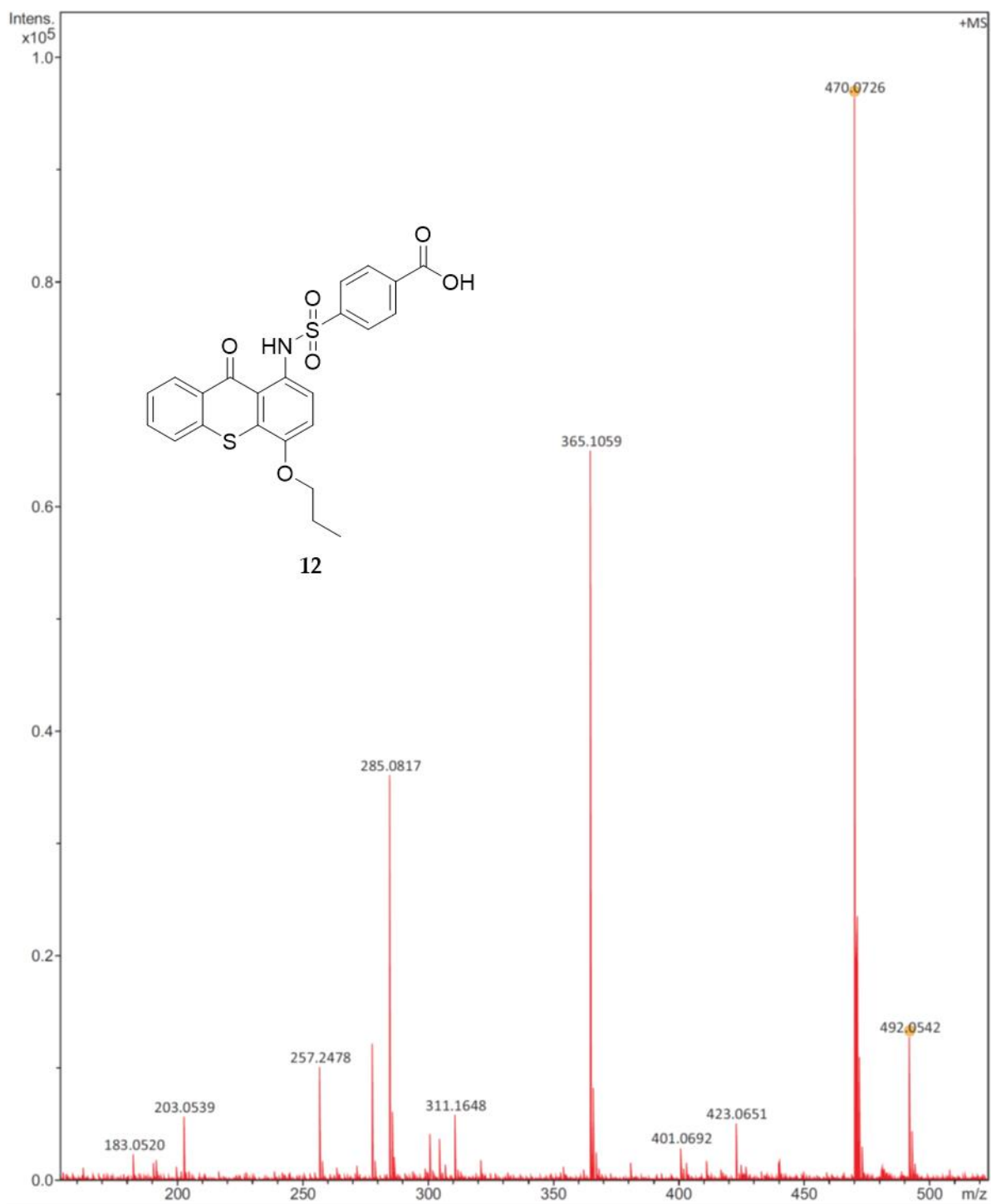


Figure S20. ¹H NMR (top, 300.13 MHz, DMSO) and ¹³C NMR (bottom, 75.48 MHz, DMSO) for compound **12**.



Meas. m/z	Formula	m/z	err [ppm]
470.0726	C ₂₃ H ₂₀ NO ₆ S ₂	470.0732	-1.06

Figure S21. Electrospray ESI data for compound 12.

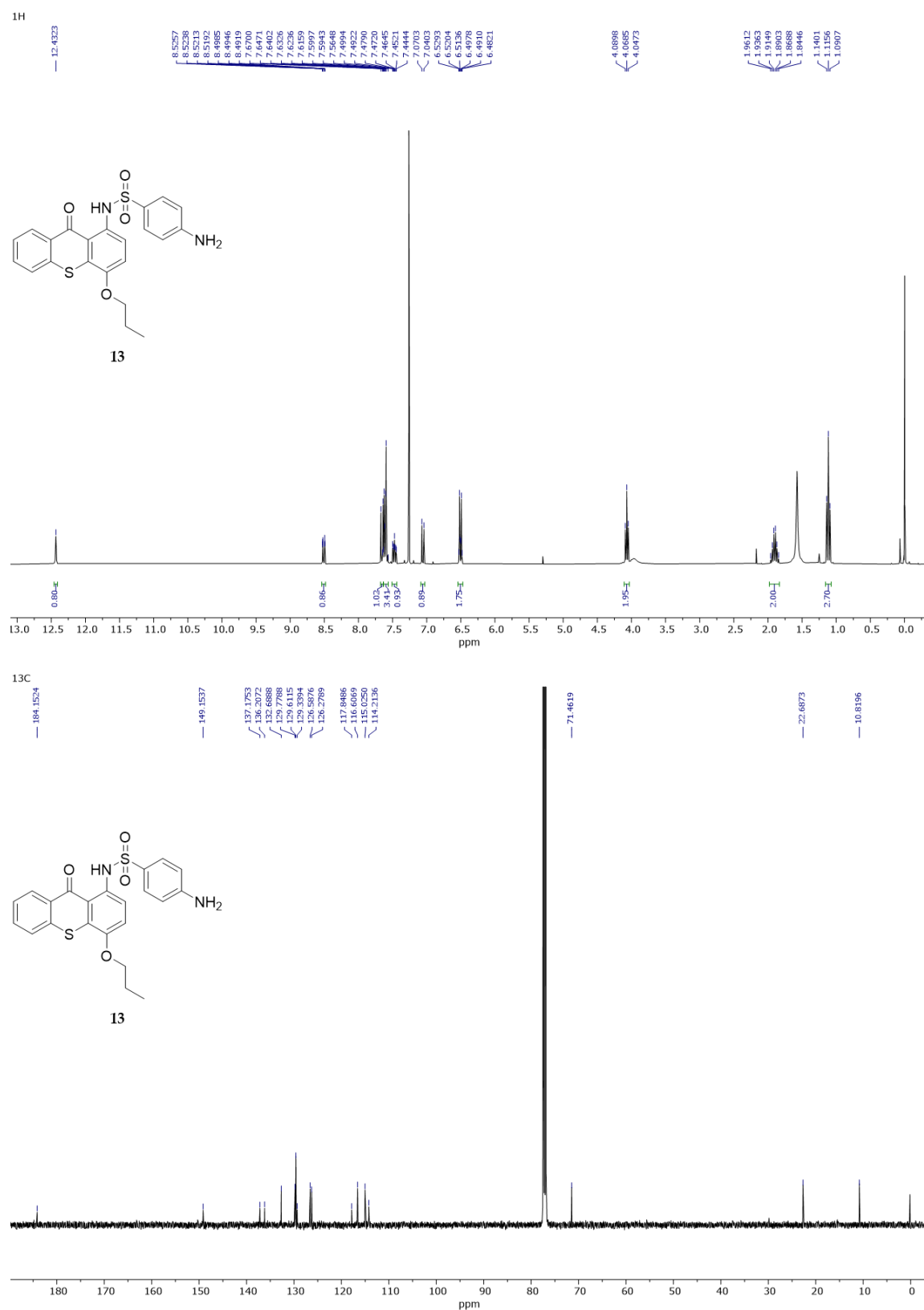
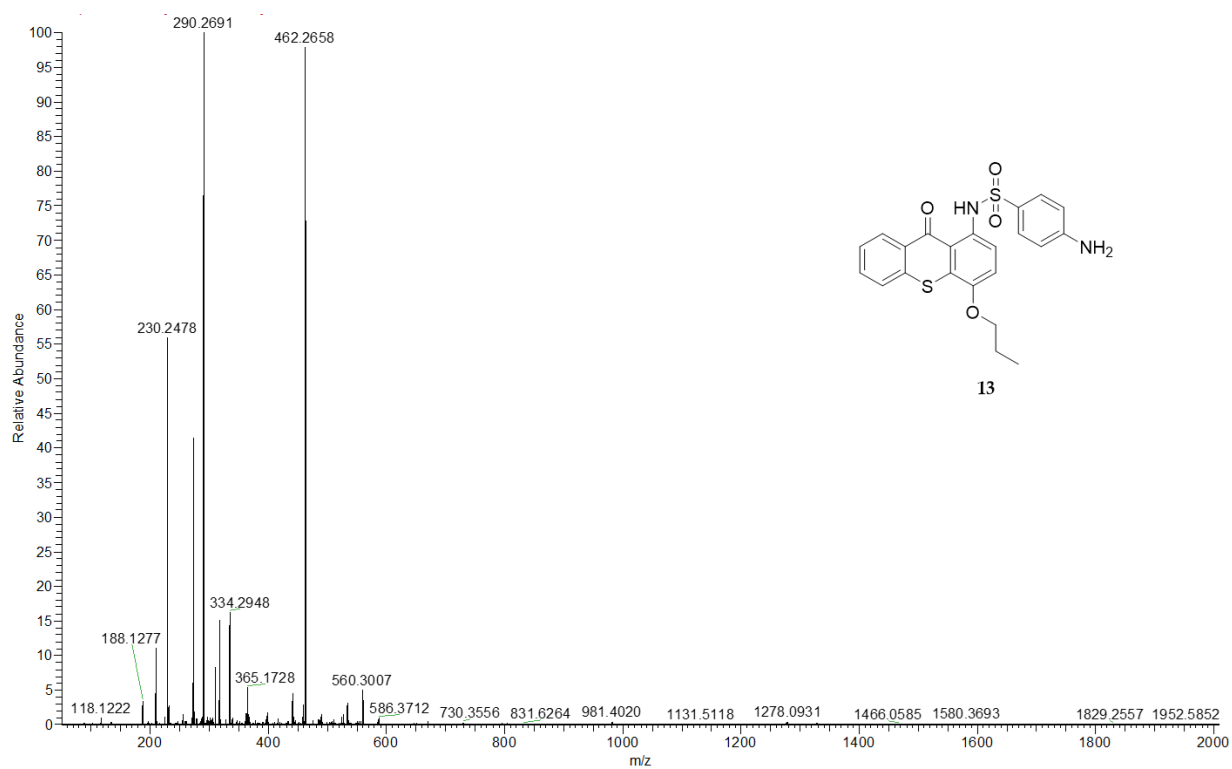


Figure S22. ¹H NMR (top, 300.13 MHz, CDCl₃) and ¹³C NMR (bottom, 75.48 MHz, CDCl₃) for compound **13**.



Meas. m/z	Formula	m/z	err [ppm]
462.2658	C ₂₃ H ₁₉ NO ₆ S ₂ Na	462.0683	427.43

Figure S23. Electrospray ESI data for compound 13.

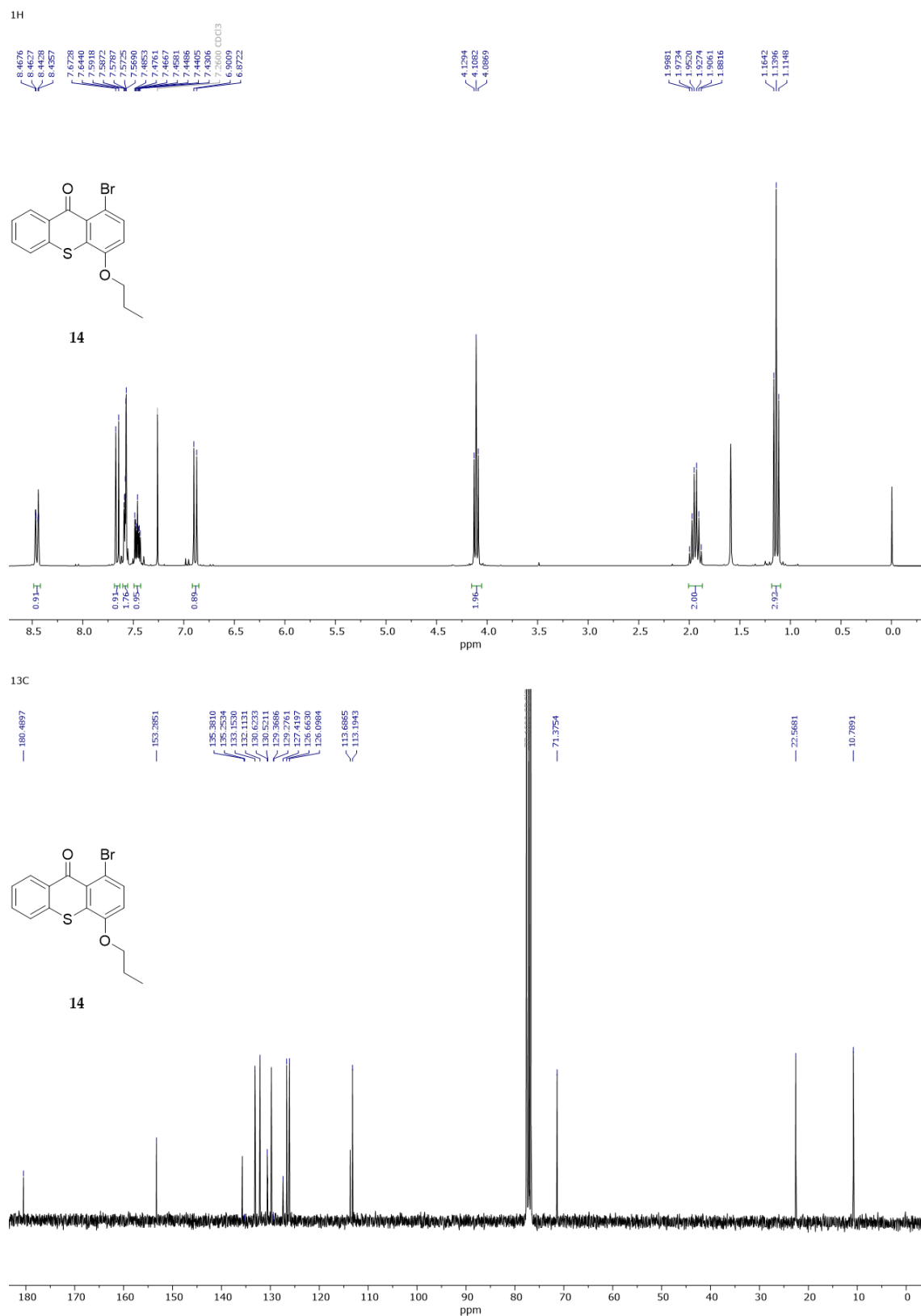


Figure S24. ¹H NMR (top, 300.13 MHz, CDCl₃) and ¹³C NMR (bottom, 75.48 MHz, CDCl₃) for compound **14**.

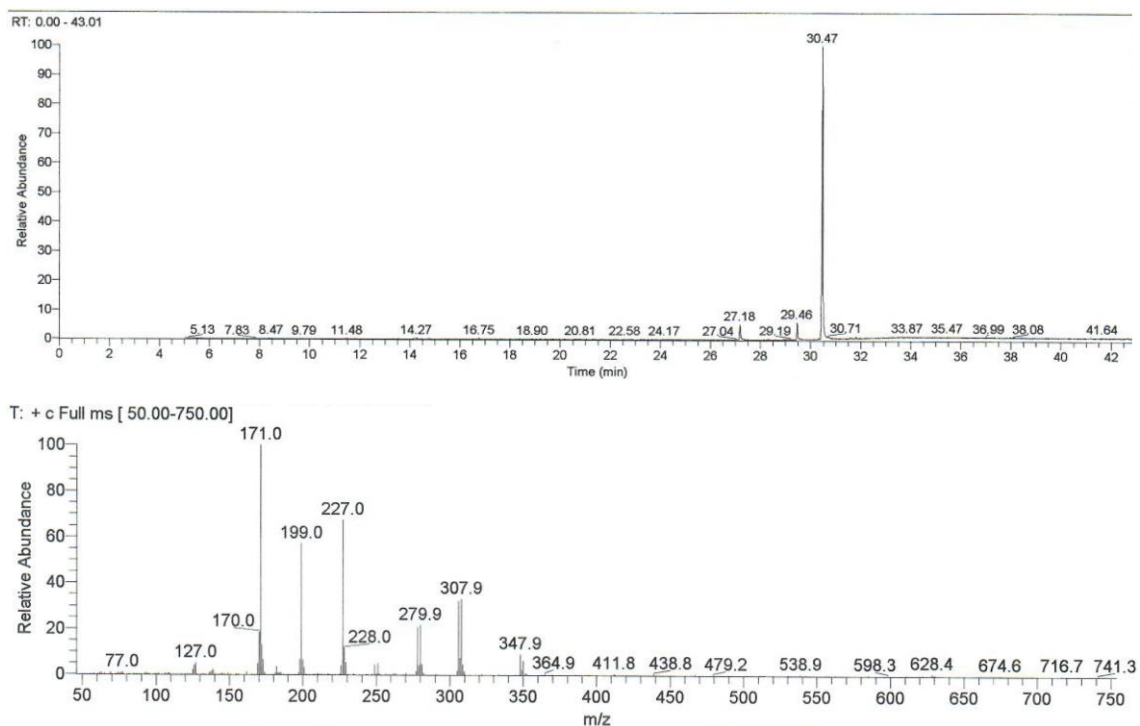


Figure S25. GC-MS spectra of compound 14.

Table S1. Cytotoxicity of the selected compounds in NIH/3T3 cell line.

Compound	IC ₅₀ (μM) ± SD
3	>100
8	>100
12	64.47 ± 4.54
13	>100
17	>100
Doxorubicin	12.05 ± 0.81