

# Supplemental Material

## Population pharmacokinetic method to predict within-subject variability using single-period clinical data

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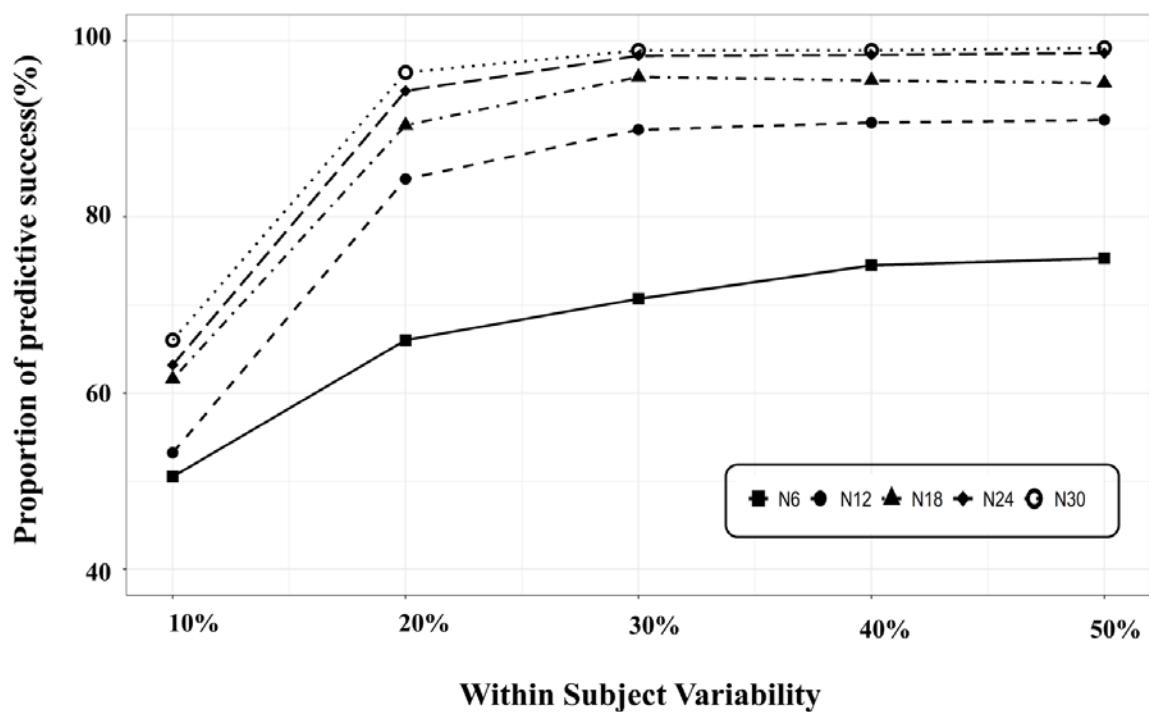
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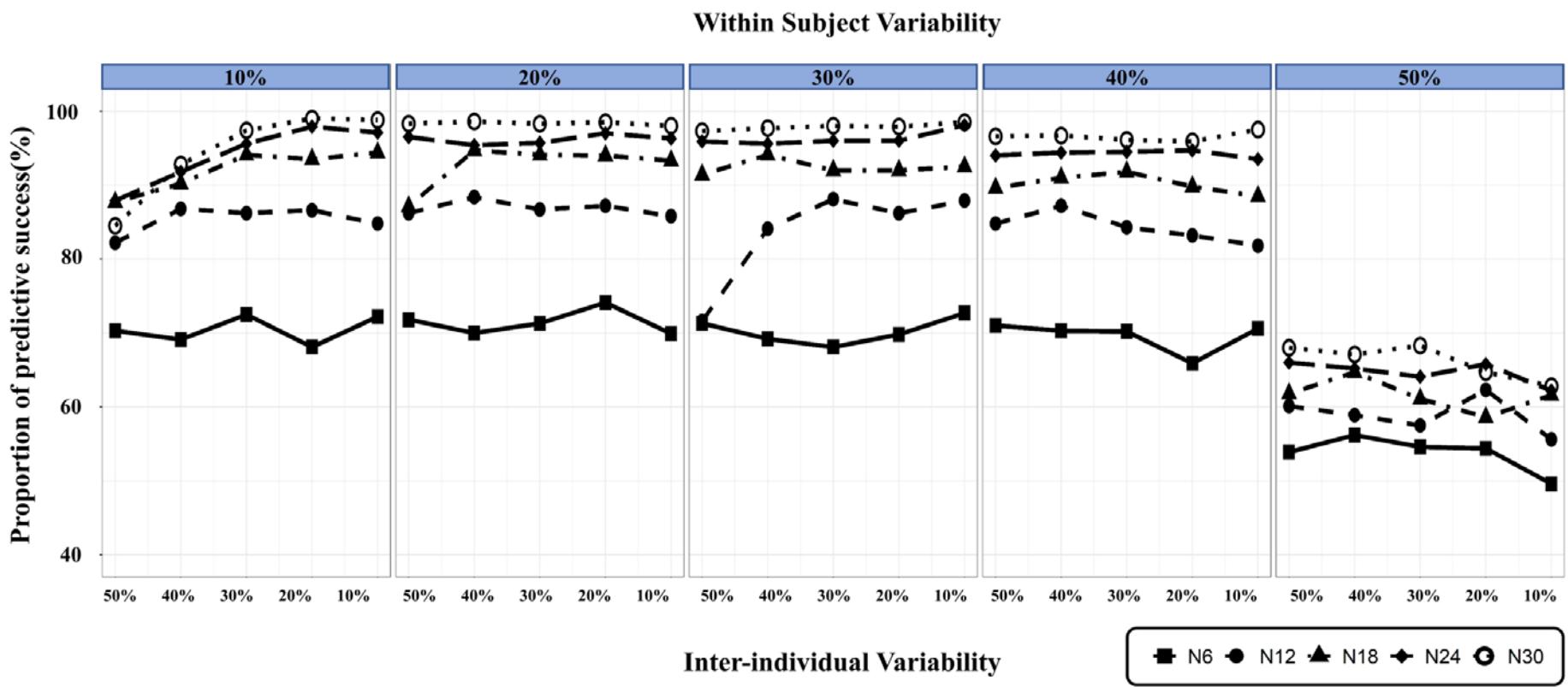
\* Those of authors were contributed equally for this work as corresponding author

## Contents

<b>Figure S1</b> .....	<b>1</b>
<b>Figure S2</b> .....	<b>2</b>
<b>Table S1</b> .....	<b>3</b>
<b>Text S1</b> .....	<b>4</b>
<b>Text S2</b> .....	<b>6</b>
<b>Text S3</b> .....	<b>8</b>



**Figure S1.** The proportion of predictive success for the first experiment.



**Figure S2.** The proportion of predictive success for the second experiment.

**Table S1. Tabulated summary for results of comparison with and without covariance between omegas**

Evaluation scenario			Proportion of predictive success for RV(%)	
WV(%)	IIV(%)	No. of subject	With Covariance between $\omega_1$ and $\omega_2^*$	Without Covariance between $\omega_1$ and $\omega_2^{**}$
50	10	18	52	62
50	50	18	50	62
10	50	18	89	88

\* Preform of NONMEM PK model for 100 simulation dataset generated by mrgsolve R package,

\*\*Perform of NONMEM PK model for 1000 simulation dataset generated by basic R code. The  $\omega_1$  and  $\omega_2$  meant IIV on CL and Vd, respectively.

**Text S1. Example as a R code for generating simulation dataset**

```

## Individual PK parameters(IIV 10%)
data.sample <- 1:1000
start.number <- min(data.sample)
i <- start.number
for (i in data.sample) {
  set.seed(seed[i])
  x <- rlnorm(100000, meanlog = 10, sdlog = 1)
  y <- rlnorm(100000, meanlog = 50, sdlog = 5)
  CL <- sample(log(x)[log(x)>0],12)    # Preventing for generating negative number at CL
  Vd <- sample(log(y)[log(y)>0],12)    # Preventing for generating negative number at Vd
  id <-seq(1:12)
  data <- data.frame(id,CL,Vd)
  write.table(data,paste("data",i,".csv",sep=""),sep=",",row.names = F)
}
## Calculation the concentration using parameters(CL, Vd)
total <- 1:1000
min.total <- min(total)
k <- min.total
for(k in total){
  one <- read.table(paste("data",k,".csv",sep=""),sep = ",",header=TRUE)
  kel <- one$CL/one$Vd
  one <- cbind(one,kel)
  number <- 1:12
  begin <- min(number)
  i <- begin
  for(i in number){
    special.ID <- DO[DO$ID==i,]
    timenumber <- 1:12
    time.start <- min(timenumber)
    j <- time.start
    for(j in timenumber){
      #iv PK equation(WV = 0.3)
      conco <- (100*exp(-one$kel[i]*special.ID$TIME[j])/(one$Vd[i]))
      eps <- rnorm(12, mean = 0, sd = 0.3)
      eps.sample <- sample(eps,1)
      conco2 <- conco*(1+eps.sample)    ##Proportional model
      # Preventing for generating negative number at plasma concentration
      while(conco2<0){
        eps <- rnorm(12, mean = 0, sd = 0.3)
        eps.sample <- sample(eps,1)
        conco2 <- conco*(1+eps.sample)    ##Proportional model
      }
      if(j==time.start)
        DV <- conco2
      else DV <- rbind(DV,conco2)
    }
    real <- cbind(special.ID, DV)
    if(i==begin)
      set <- real
  }
}

```

```
    else set <- rbind(set, real)
  }
write.table(set, paste("CONC",k,".csv",sep = ""),sep = ",", row.names = FALSE)}
```

**Text S2. C++ and R script code when used mrgsolve R package**

```
## Example of CPP file
$PROB
- 1 COMP iv PK model
- Random effect : yes
$PARAM
TVCL = 10
TVV = 50
$CMT
CENT
$MAIN
double CL=TVCL*exp(ETA(1));
double V=TVV*exp(ETA(2));
$ODE
dxdt_CENT = -(CL/V)*CENT;
$OMEGA >> annotated=TRUE, block = TRUE
    ECL: ETA on Clearance
    EV: ETA on Volume
0.25
0.0625 0.25
$SIGMA
0.25
$TABLE
double IPRED = CENT/V;
double DV = IPRED*(1+EPS(1));
$CAPTURE
DV IPRED

## Example of R script file
# problem : WV0.5_IIV0.5_omegacovariance0.5
# subject NO. = 18
# load packages
library(mrgsolve)
library(tidyverse)
library(dplyr)
# giving seed number
set.seed(20191203)
# loading cpp file
mod<-mread("WV0.5_IIV0.5_COV0.5","C:/Users/Wonho
Kang/Desktop/TEST_OMBLOCK/WV0.5_IIV0.5_COV0.5/N18")
total <- 1:100
min.total <- min(total)
i <- min.total
for(i in total){
    nn=18
    idata<-tibble(ID=seq(nn))
    tmptolerance = -1
    tmpcount = 0
    while (tmptolerance < 0){
        tmpcount = tmpcount +1
```

```

df<-mod %>%
ev(amt=100) %>%
idata_set(idata) %>%
mrgsim(end=24, delta=0.1) %>%
filter(time==0 | time==0.083 | time==0.167 | time==0.333 | time==0.5 | time==1 | time==2 | time==4 | time==6 | ti
me==8 | time==12 | time==24 )
tmptolerance = min(df$DV)
print(tmpcount)
}
#add MDV
df<-df %>% mutate(MDV=0)
df$MDV[which(df$DV==0)]<-1
#add AMT
df<-df %>% mutate(AMT=0)
df$AMT[which(df$MDV==1)]<-100
#rename TIME
df<-df %>% rename(TIME=time)
df<-df %>% arrange(ID,TIME)
df<-df[c("ID","TIME","DV","MDV","AMT")]
names(df) <- c("#ID", "TIME", "DV", "MDV", "AMT")
write.table(df,paste("datafile",i,".csv",sep = ""),sep = ",",quote=FALSE,row.names=FALSE)
}

```

**Text S3. NONMEM PK Model code**

```
$PROBLEM Simulation dataset2_IIV50%_WV30%_N18
$INPUT ID TIME DV MDV AMT
$DATA datafile1.csv IGNORE=@
$SUBROUTINES ADVAN6 TOL=8
$MODEL
  COMP (CENT, DEFDOSE, DEFOBS)
$PK
  CL =  THETA(1) * EXP(ETA(1))
  V   =  THETA(2) * EXP(ETA(2))
  KEL = CL/V
  S1=V
$DES
  DADT(1) = -KEL*A(1)
$ERROR
  IPRED = F
  DEL=0
  IF(IPRED.EQ.0) DEL=1
  W=IPRED+DEL
  IRES = DV-IPRED
  IWRES = IRES/W
  Y = F+W*EPS(1)
$THETA
(0, 10) ; CL
(0, 50) ; V
$OMEGA
  0.25
  0.25
$SIGMA
  0.09
$EST METHOD=1 MAXEVAL=9999 NOABORT INTER PRINT=5 NSIG=2 SIGL=8
$TABLE ID TIME DV MDV AMT IPRED IWRES CWRES ONEHEADER NOPRINT FILE = sdtab1
$TABLE ID ETA1 ETA2 ONEHEADER NOPRINT FILE = patab1
```